R Python, and Ruby clients for GBIF species occurrence data

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- 5 Abstract
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- Background. The number of individuals of each species in a given location forms the basis for many
- 11 sub-fields of ecology and evolution. Data on individuals, including which species, and where they're
- 12 found, can be used for a large number of research questions. Global Biodiversity Information Facility
- 13 (hereafter, GBIF) is the largest of these. Programmatic clients for GBIF would make research dealing
- with GBIF data much easier.
- 15 Methods. We have developed clients to access GBIF data for each of the R, Python, and Ruby
- programming languages: rgbif, pygbif, gbifrb.
- 17 Results. For all clients we describe their design and utility, and demonstrate many use cases.

Discussion. Programmatic access to GBIF will facilitate more open and reproducible science. We hope that all scientists using GBIF data will do so with one of these two clients.

18 Introduction

- Perhaps the most fundamental element in many fields of ecology is the individual organism. The number of individuals of each species in a given location forms the basis for many sub-fields of ecology and evolution. Some research questions necessitate collecting new data, while others can easily take advantage of existing data. In fact, some ecology fields are built largely on existing data, e.g., macro-ecology (Brown, 1995; Beck et al., 2012).
- Data on individuals, including which species, and where they're found, can be used for a large number of research questions. Biodiversity records have been used for a suite of other use cases: validating habitat suitability models with real occurrence data (Ficetola et al., 2014); ancestral range reconstruction (Ferretti et al., 2015; María Mendoza et al., 2015); development of invasive species watch lists (Faulkner et al., 2014); evaluating risk of invasive species spread (Febbraro et al., 2013); and effects of climate change on future biodiversity (Brown et al., 2015).
- In addition to wide utility, this data is important for conservation. Biodiversity loss is one of the greatest challenges of our time (Pimm et al., 2014), and some have called this the sixth great mass extinction (Ceballos et al., 2015). Given this challenge there is a great need for data on specimen records, whether collected from live sightings in the field or specimens in museums.

34 Global Biodiversity Information Facility

- There are many online services that collect and maintain specimen records. However, Global Biodiversity Information Facility (hereafter, GBIF, http://www.gbif.org) is the largest collection of biodiversity records globally, currently with 820 million records, roughly 5.9 million taxa, 36,000 datasets from 1,300 publishers (as of 2016-02-09). Many large biodiversity warehouses such as iNaturalist (http://www.inaturalist.org), VertNet (http://vertnet.org), and USGS's Biodiversity Information Serving Our Nation (BISON; http://bison.usgs.ornl.gov) all feed into GBIF.
- The most important organizational level in GBIF occurrence data is the occurrence record. The fields in a record vary, but include information about taxonomy (kingdom, phylum, genus, species names) and their identifiers, dataset metadata, and locality information including geospatial position. Going upstream, each record is part of a dataset, where each dataset is submitted by an organization, organizations are organized into nodes, datasets are published through institutions (which may be hosted at another organization), and a network is a group of datasets (managed by GBIF).

- Each occurrence record has some taxonomic name associated with it, which itself is linked to a lot of
- 48 other taxonomic data including a master taxonomic backbone that integrates taxonomies across many
- 49 taxonomic authorities.
- 50 The organization of GBIF matters because you can navigate GBIF data through these hierarchical
- organizational levels it helps to be familiar with the terminology and how each group relates to another.

52 The clients

- Although we discuss libraries for R, Python, and Ruby here, we focus mostly on the R library rgbif as
- 54 it has seen the most developer and user attention, and is the most mature.
- 55 rgbif
- Herein, we describe the rgbif software package (Chamberlain et al.) for working with GBIF data in the
- 57 R programming environment (R Core Team, 2014). R is a widely used language in academia, as well as
- non-profit and private sectors. Importantly, R makes it easy to execute all steps of the research process,
- 59 including data management, data manipulation and cleaning, statistics, and visualization. Thus, an R
- 60 client for getting GBIF data is a powerful tool to facilitate reproducible research.
- 61 The rgbif package is nearly completely written in R (a small Javascript library is included for
- reading well known text (Herring, 2011)), uses an MIT license to maximize use everywhere. rgbif is
- developed publicly on GitHub at https://github.com/ropensci/rgbif, where development versions of
- the package can be installed, and bugs and feature requests reported. Stable versions of rgbif can be
- 65 installed from CRAN, the distribution network for R packages. rgbif is part of the rOpenSci project
- 66 (http://ropensci.org), a developer network making R software to facilitate reproducible research.
- 67 pygbif
- ₆₈ pygbif (Chamberlain) is a Python library for working with GBIF data in the Python programming
- 69 environment. Python is a general purpose programming language used widely in all sectors, and for all
- 70 parts of software development including server and client side use cases. Python is used exclusively
- in some scientific disciplines (e.g., astronomy), and has partial usage in other disciplines. A Python
- client for GBIF data is an important tool given the even wider usage of Python than R, though maybe
- ⁷³ slightly less than R for ecology/biology disciplines.

pip install pygbif

import pygbif

- 74 The pygbif library is less mature and complete than the R package. It also uses an MIT license to
- maximize use everywhere. pygbif is developed publicly on GitHub at https://github.com/sckott/pygbif,
- ⁷⁶ where development versions of the package can be installed, and bugs and feature requests reported.
- 577 Stable versions of pygbif can be installed from pypi, the distribution network for Python libraries.
- 78 gbifrb
- 79 gbifrb (Chamberlain) is a library for working with GBIF data in the Ruby programming environment.
- 80 Like Python, Ruby is a general purpose programming language used widely in all sectors. Unlike
- Python, Ruby is not used extensively in scientific disciplines. However, a Ruby client for GBIF data
- s2 can be an important tool given how widely Ruby is used for web and web service development.

gem install gbifrb

require 'gbifrb'

- The gbifrb library is less mature and complete than the R and Python libraries. It also uses
- an MIT license to maximize use everywhere. gbifrb is developed publicly on GitHub at https:
- 25 //github.com/sckott/gbifrb, where development versions of the package can be installed, and bugs and
- feature requests reported. Stable versions of gbifrb can be installed from [Rubygems][gemgbif], the
- 87 distribution network for Ruby libraries.
- 88 Library interfaces
- 89 rgbif, pygbif, and gbifrb are designed following the GBIF Application Programming Interface, or
- 90 API. The GBIF API has four major components: registry, taxonomic names, occurrences, and maps. We
- 91 also include functions to interface with the OAI-PMH GBIF service; only dataset (registry) information
- 92 is available via this service, however. An interface to the GBIF maps API is in development for rgbif,
- but is non-existent for both pygbif and gbifrb. All three libraries have a suite of functions dealing
- 94 with each of registry, taxonomic, names, and occurrences we'll go through each in turn describing
- 95 design of the user interface and example usage.

96 GBIF headers

- 97 With each request rgbif, pygbif, gbifrb make to GBIF's API, we send request headers that tell GBIF
- 98 what library the request is coming from, including what version of the library. This helps GBIF know
- what proportion of requests are coming from which library, and therefore from R vs. Python vs. Ruby;
- this information is helpful for GBIF in thinking about how people are using GBIF data.

101 Registry

102 The GBIF registry API services are spread across five sets of functions via the main GBIF API:

- Datasets
- Installations
- Networks
- Nodes Nodes
- Organizations
- Dataset information in general is available via the OAI-PMH service, functions in rgbif prefixed with gbif_oai_, but not available in pygbif or gbifrb yet.
- Datasets are owned by organizations. Organizations are endorsed by nodes to share datasets with GBIF.
- Datasets are published through institutions, which may be hosted at another organization. A network
- is a group of datasets (managed by GBIF). Datasets are the units that matter the most with respect
- to registry information, while installations, networks, nodes, and organizations are simply higher level
- organizational structure.

115 Datasets

- Dataset functions include search, dataset metadata retrieval, and dataset metrics. Searching for datasets
- is an important part of the discovery process. One can search for datasets on the GBIF web portal.
- However, programmatic searching using any of these libraries is more powerful. Identifying datasets
- appropriate for a research question is helpful as you can get metadata for each dataset, and track down
- dataset specific problems, if any.

The dataset_search() function in rgbif is one way to search for datasets. Here, we search for the term "oregon", which finds any datasets that have words matching that term.

```
res <- dataset_search(query = "oregon")

res$data$datasetTitle[1:10]

#> [1] "Oregon State Ichthyology Collection"

#> [2] "Oregon State University Herpetological Collection"

#> [3] "Mygalomorph spiders from southwestern Oregon, USA, with descriptions of four new specie

#> [4] "A new species of Helobdella (Hirudinida: Glossiphoniidae) from Oregon, USA"

#> [5] "Annotated Checklist of the large branchiopod crustaceans of Idaho, Oregon and Washingto

#> [6] "A new species of Chrysobothris Eschscholtz from Oregon and Washington, with notes on ot

#> [7] "Three new species of Grylloblatta Walker (Insecta: Grylloblattodea: Grylloblattidae), f

#> [8] "A new species of Cladotanytarsus (Lenziella) from Oregon supports the systematic concep

#> [9] "A new monster from southwest Oregon forests: Cryptomasterbehemoth sp. n. (Opiliones, La

#> [10] "Two new species of Fluminicola (Caenogastropoda, Lithoglyphidae) from southwest Oregon,
```

- See also datasets() and dataset_suggest() in rgbif for searching for datasets.
- In Python, we can similarly search for datasets. Here, search for datasets of type OCCURRENCE:

```
from pygbif import registry
registry.datasets(type="OCCURRENCE")
```

125 In Ruby, we can do the same. Here, search for datasets of type OCCURRENCE:

```
require 'gbifrb'
registry = Gbif::Registry
registry.datasets(type: "OCCURRENCE")
```

- Dataset metrics. Dataset metrics are another useful way of figuring out what datasets you may want to use. One drawback is that these metrics data are only available for datasets of type *checklist*, but there
- are quite a lot of them (21522).
- Here, in R we search for dataset metrics for a single dataset, with uuid ec93a739-1681-4b04-b62f-3a687127a17f,
- a checklist of the ants (Hymenoptera: Formicidae) of the World.

rank	count
SPECIES	13710
SUBSPECIES	3234
GENUS	726
TRIBE	53
SUBFAMILY	20
FAMILY	2
KINGDOM	1
PHYLUM	1
CLASS	1
ORDER	1

And in Python, get metrics for the same dataset as above:

```
from pygbif import registry
registry.dataset_metrics(uuid='ec93a739-1681-4b04-b62f-3a687127a17f')
```

132 The same in Ruby:

```
require 'gbifrb'
registry = Gbif::Registry
registry.dataset_metrics(uuid: 'ec93a739-1681-4b04-b62f-3a687127a17f')
```

- Networks, nodes, and installations
- Networks, nodes and installations are at a higher level of organization above datasets, but can be
- useful if you want to explore data from given organizations. Here, in R we search for the first 10 GBIF
- 136 networks, returning just the title field.

```
networks(limit = 10)$data$title

#> [1] "GBIF Backbone Sources"

#> [2] "Canadensys"

#> [3] "Southwest Collections of Arthropods Network (SCAN)"

#> [4] "VertNet"

#> [5] "Dryad"

#> [6] "GBIF Network"

#> [7] "The Knowledge Network for Biocomplexity (KNB) "

#> [8] "Online Zoological Collections of Australian Museums (OZCAM)"

#> [9] "Catalogue of Life"

#> [10] "Ocean Biogeographic Information System (OBIS)"
```

137 And in Python:

```
from pygbif import registry
registry.networks(limit = 10)
```

And in Ruby:

```
require 'gbifrb'
registry = Gbif::Registry
registry.networks(limit: 10)
```

- 139 Taxonomic names
- 140 The GBIF taxonomic names API services are spread across five functions in rgbif:
- Search GBIF name backbone name_backbone()
- Search across all checklists name_lookup()
- Quick name lookup name_suggest()
- Name usage of a name according to a checklist name_usage()
- GBIF name parser parsenames()

- pygbif and gbifrb have all the same functions, except the name parser goes by name_parser() in pygbif and gbifrb.
- The goal of these name functions is often to settle on a taxonomic name known to GBIF's database.
- This serves two purposes: 1) when referring to a taxonomic name, you can point to a URI on the
- 150 Internet, and 2) you can search for metadata on a taxon, and occurrences of that taxon in GBIF.
- 151 Taxonomic names are particularly tricky. Many different organizations have their own unique codes for
- the same taxonomic names, and some taxonomic groups have preferred sources for the definitive names
- for that group. That's why it's best to determine what name GBIF uses, and its associated identifier,
- 154 for the taxon of interest instead of simply searching for occurrences with a taxonomic name.
- When searching for occurrences (see below) you can search by taxonomic name (and other filters, e.g.,
- taxonomic rank), but you're probably better off figuring out the taxonomic key in the GBIF backbone
- taxonomy, and using that to search for occurrences. The taxonkey parameter in the GBIF occurrences
- 158 API expects a GBIF backbone taxon key.
- 159 GBIF Backbone
- The GBIF backbone taxonomy is used in GBIF to have a consistent way to refer to taxonomic names throughout their services. The backbone has 5869207 unique names and 2818534 species names. The backbone taxonomy is also a dataset with key d7dddbf4-2cf0-4f39-9b2a-bb099caae36c
- 163 (https://www.gbif.org/dataset/d7dddbf4-2cf0-4f39-9b2a-bb099caae36c).
- We can search the backbone taxonomy with the function name_backbone() in all thee clients. Here,
- we're searching for the name *Poa*, restricting to genera, and the family *Poaceae*, in R

```
res <- name_backbone(name='Poa', rank='genus', family='Poaceae')
res[c('usageKey', 'kingdom')]
#> $usageKey
#> [1] 2704173
#>
#> $kingdom
#> [1] "Plantae"
```

6 And in Python

```
from pygbif import species

res = species.name_backbone(name='Poa', rank='genus', family='Poaceae')
[ res[x] for x in ['usageKey', 'kingdom'] ]
```

167 And in Ruby

```
require 'gbifrb'
species = Gbif::Species
res = species.name_backbone(name: 'Poa', rank: 'genus', family: 'Poaceae')
res.select { |k,v| k.match(/usageKey|kingdom/) }
```

- 168 Name searching
- One of the quickest ways to search for names is using name_suggest(), which does a very quick search and returns minimal data. Here, we're searching for the query term *Pum*, and we get back many names:

name_suggest(q='Pum', limit = 6)

key	canonicalName	rank
2142856	Althepus pum	SPECIES
8589398	Pumiliopimoidae	FAMILY
8783253	Pumililema	GENUS
4823360	Pumiliopareia	GENUS
4635949	Pumilina	GENUS
4648228	Pumilopaurus	GENUS

171 The same in Python

```
from pygbif import species
species.name_suggest(q='Pum', limit = 6)
```

172 And in Ruby

```
require 'gbifrb'
species = Gbif::Species
species.name_suggest(q: 'Pum', limit: 6)
```

With these results, you can then proceed to search for occurrences with the taxon key(s), or drill down further with other name searching functions to get the exact taxon of interest.

175 Occurrences

GBIF provides two ways to get occurrence data: through the /occurrence/search route (see occ_search in rgbif, occurrences.search in pygbif, Occurrences.search in gbifrb), or via the /occurrence/download route (many functions, see below).

occ_search()/occurrences.search/Occurrences.search are the main functions for the search route, and are more appropriate when you want less data, while the download functions are more appropriate for larger data requests.

Small vs. large amounts of data of course is all relative. GBIF imposes for any given search a limit of 200,000 records in the search service, after which point you can't download any more records for that search. However, you can download more records for different searches.

We think the search service is still quite useful for many people even given the 200,000 limit. For those that need more data, we have created a similar interface in the download functions that should be easy to use with minimal work. Users should take note that using the download service has a few extra steps to get data into R, but is straight-forward.

The download service, like the occurrence search service, is rate-limited. That is, you can only have one to three downloads running simultaneously for your user credentials. However, simply check when a download job is complete, then you can start a new download request. See "Queueing Download Requests" below for help automating many download requests in R.

3 Download API

The download API syntax is similar to the occurrence search API in that the same parameters are used, but the way in which the query is defined is different. For example, in the download API you can do greater than searches (i.e., latitude > 50), whereas you cannot do that in the occurrence search

- API. Thus, unfortunately, we couldn't make the query interface exactly the same for both search and download functions.
- Using the download service can consist of as few as three steps: 1) Request data via a search; 2)
- 200 Download data; 3) Import data into R.
- 201 Request data download given a query. Here, we search for the taxon key 3119195, which is the key for
- 202 Helianthus annuus (http://www.gbif.org/species/3119195).

```
occ_download('taxonKey = 3119195')
#> <<gbif download>>
#> Username: xxxx
#> E-mail: xxxx
#> Download key: 0000840-150615163101818
```

You can check on when the download is ready using the functions occ_download_list() and occ_download_meta(). When it's ready use occ_download_get() to download the dataset to your computer.

```
(res <- occ_download_get("0000840-150615163101818", overwrite = TRUE))
#> <<gbif downloaded get>>
#> Path: ./0000840-150615163101818.zip
#> File size: 3.19 MB
```

- What's printed out above is a very brief summary of what was downloaded, the path to the file, and its size (in human readable form).
- Next, read the data in to R using the function occ_download_import().

#>	2	725767447	NA		NA	NA	NA	NA
#>	3	725767450	NA		NA	NA	NA	NA
#>	4	725767513	NA		NA	NA	NA	NA
#>	5	725767546	NA		NA	NA	NA	NA
#>	6	725767579	NA		NA	NA	NA	NA
#>	7	725767609	NA		NA	NA	NA	NA
#>	8	725767645	NA		NA	NA	NA	NA
#>	9	725767678	NA		NA	NA	NA	NA
#>	10	725767681	NA		NA	NA	NA	NA
#>			• • •					
#>	Var	riables not	shown: availa	ible (lgl),	$bibliographic {\it Cita}$	tion (chr), conform	msTo (lgl)	, contribu
#>		coverage	(lgl), create	ed (chr), c	reator (lgl), date	(lgl), dateAccepte	ed (lgl),	dateCopyri

(lgl), dateSubmitted (lgl), description (lgl), educationLevel (lgl), extent (lgl), forma

hasFormat (lgl), hasPart (lgl), hasVersion (lgl), identifier (chr), instructionalMethod

209 In Python

#>

```
from pygbif import occurrences as occ
occ.download('taxonKey = 3119195')
(res = occ.download_get("0000840-150615163101818", overwrite = True))
```

- We don't have pygbif functionality at the moment for importing data, but it's coming soon.
- The Ruby library gbifrb does not yet have occurrene download functionality.
- Downloaded data format. The downloaded dataset from GBIF is a Darwin Core Archive (DwC-A), an internationally recognized biodiversity informatics standard (http://rs.tdwg.org/dwc/). The DwC-A downloaded is a compressed folder with a number of files, including metadata, citations for each of the datasets included in the download, and the data itself, in separate files for each dataset as well as one single .txt file. In rgbif::occ_download_import(), we simply fetch data from the .txt file. If you want to dig into the metadata, citations, etc., it is easily accessible from the folder on your computer.
- Queueing Download Requests. FILL IN TEXT HERE . . .

```
219 Search API
```

The search API follows the GBIF API and is broken down into the following functions:

- Get a single numeric count of occurrences rgbif: occ_count() / pygbif: occurrences.count

 / gbifrb: Occurrences.count
- Search for occurrences rgbif: occ_search() / pygbif: occurrences.search / gbifrb:

 Occurrences.search
- A simplified and optimized version of rgbif: occ_search() or occ_data() / none / none
- Get occurrences by occurrence identifier rgbif: occ_get() / pygbif: occurrences.get /
 gbifrb: Occurrences.get
- Get occurrence metadata rgbif: occ_metadata() / pygbif: various / gbifrb: various
- Search for occurrences. The main search work-horse is occ_search(). This function allows very flexible search definitions. In addition, this function does paging internally, making it such that the user does not have worry about the 300 records per request limit but of course we can't go over the 200,000 maximum limit.
- The output of occ_search() presents a compact data.frame so that no matter how large the
 data.frame, the output is easily assessed because only a few of the records (rows) are shown, only a few
 columns are shown (with others shown in name only), and metadata is shown on top of the data.frame
 to indicate data found and returned, media records found, unique taxonomic hierarchies returned, and
 the query executed.
- The output of these examples, except one, aren't shown.
- 239 Search by species name, using name_backbone() first to get key

240 \mathbf{R}

```
library(rgbif)
(key <- name_suggest(q = 'Helianthus annuus', rank = 'species')$key[1])
#> [1] 9206251
occ_search(taxonKey = key, limit = 2)
```

```
#> Records found [17858]
#> Records returned [2]
#> No. unique hierarchies [1]
#> No. media records [1]
#> No. facets [0]
#> Args [limit=2, offset=0, taxonKey=9206251, fields=all]
#> # A tibble: 2 x 75
                              key decimalLatitude decimalLongitude
#>
                 <chr>
                            \langle int \rangle
                                            <dbl>
                                                              <dbl>
                                                           16.54257
#> 1 Helianthus annuus 1433793045
                                         59.66859
#> 2 Helianthus annuus 1434024463
                                          63.71622
                                                           20.31247
#> # ... with 71 more variables: issues <chr>, datasetKey <chr>,
      publishingOrgKey <chr>, publishingCountry <chr>, protocol <chr>,
       lastCrawled <chr>, lastParsed <chr>, crawlId <int>, extensions <chr>,
#> #
#> #
     basisOfRecord <chr>, ...
```

241 Python

```
from pygbif import species
from pygbif import occurrences as occ
key = species.name_suggest(q = 'Helianthus annuus', rank = 'species')['data'][0]['key']
occ.search(taxonKey = key, limit = 2)
```

Ruby

```
require 'gbifrb'
species = Gbif::Species
occ = Gbif::Occurrences
key = species.name_suggest(q: 'Helianthus annuus', rank: 'species')['data'][0]['key']
occ.search(taxonKey: key, limit: 2)
```

Instead of getting a taxon key first, you can search for a name directly

 244 ${f R}$

```
occ_search(scientificName = 'Ursus americanus')
245 Python
```

246 Ruby

```
occ.search(scientificName: 'Ursus americanus')
```

Search for many species

occ.search(scientificName = 'Ursus americanus')

248 **R**.

```
splist <- c('Cyanocitta stelleri', 'Junco hyemalis', 'Aix sponsa')
keys <- sapply(splist, function(x) name_suggest(x)$key[1], USE.NAMES = FALSE)
occ_search(taxonKey = keys, limit = 5, return = 'data')</pre>
```

249 Python

```
from pygbif import species
from pygbif import occurrences as occ
splist = ['Cyanocitta stelleri', 'Junco hyemalis', 'Aix sponsa']
keys = [ species.name_suggest(x)['data'][0]['key'] for x in splist ]
occ.search(taxonKey = keys, limit = 5)
```

250 Ruby

```
species = Gbif::Species

occ = Gbif::Occurrences

splist = ['Cyanocitta stelleri', 'Junco hyemalis', 'Aix sponsa']

keys = [ species.name_suggest(x)['data'][0]['key'] for x in splist ]

occ.search(taxonKey: keys, limit: 5)
```

Spatial search, based on well known text format (Herring, 2011), or a bounding box set of four coordinates. The well known text string and the bounding box in the below example specify the same rectangular area in California, centering approximately on Sacramento. Whereas the bounding box format requires longitude SW corner, latitude SW corner, longitude NE corner, latitude NE corner, the well known text string requires an extra long/lat pair to close the polygon.

 \mathbf{R}

```
# well known text
wkt <- 'POLYGON((-122.6 39.9,-120.0 39.9,-120.0 37.9,-122.6 37.9,-122.6 39.9))'
occ_search(geometry = wkt, limit = 20)
# bounding box
occ_search(geometry = c(-122.6,37.9,-120.0,39.9), limit = 20)</pre>
```

257 Python

```
from pygbif import occurrences as occ
# well known text
occ.search(geometry = 'POLYGON((30.1 10.1, 10 20, 20 40, 40 40, 30.1 10.1))', limit = 20)
# bounding box
occ.search(geometry = '-125.0,38.4,-121.8,40.9', limit = 20)
```

258 Ruby

```
occ = Gbif::Occurrences
# well known text
occ.search(geometry: 'POLYGON((30.1 10.1, 10 20, 20 40, 40 40, 30.1 10.1))', limit: 20)
# bounding box
occ.search(geometry: '-125.0,38.4,-121.8,40.9', limit: 20)
```

Get only occurrences with lat/long data using the hasCoordinate parameter

 260 \mathbf{R}

```
occ_search(hasCoordinate = TRUE, limit = 5)
```

261 Python

```
from pygbif import occurrences as occ
occ.search(hasCoordinate = True, limit = 5)
```

262 Ruby

```
occ = Gbif::Occurrences
occ.search(hasCoordinate: true, limit: 5)
```

Get only those occurrences with spatial issues. Spatial issues are a set of issues that are returned in the issues field. They each indicate something different about that record. For example, the issue COUNTRY_COORDINATE_MISMATCH indicates that the interpreted occurrence coordinates fall outside of the indicated country. You can see how that might be useful when it comes to cleaning your data prior to analysis/visualization.

\mathbf{R}

```
occ_search(hasGeospatialIssue = TRUE, limit = 5)
```

Python Python

```
from pygbif import occurrences as occ
occ.search(hasGeospatialIssue = True, limit = 5)
```

270 Ruby

```
occ = Gbif::Occurrences
occ.search(hasGeospatialIssue: true, limit: 5)
```

Data cleaning. GBIF provides optional data issues with each occurrence record. These issues fall into many different pre-defined classes, covering issues with taxonomic names, geographic data, and more (see rgbif::occ_issues_lookup() to find out more information on GBIF issues; and the same data on GBIF's development site).

rgbif::occ_issues() provides a way to easily filter data downloaded via rgbif::occ_search() based on GBIF issues.

```
out <- occ_search(issue = 'DEPTH_UNLIKELY', limit = 500)
NROW(out)
#> [1] 5
out %>% occ_issues(-cudc) %>% .$data %>% NROW
#> [1] 0
```

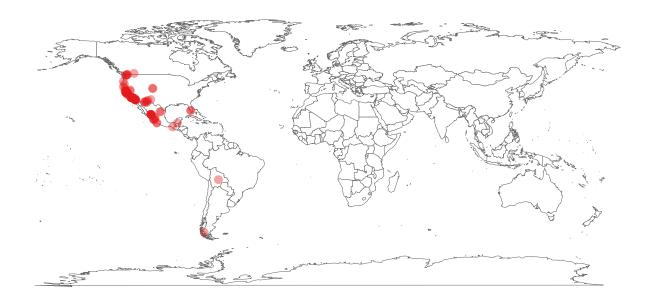
There's no equivalent interface in pygbif or gbifrb yet.

278 Mapping

283

An obvious downstream use case for species occurrence data is to map the data. rgbif per se is largely not concerned with making this easier, although we do have a simple wrapper around ggplot2 to make it easy to get a quick plot of occurrence data. For example, here we plot 100 occurrences for *Puma* concolor.

```
key <- name_backbone(name='Puma concolor')$speciesKey
dat <- occ_search(taxonKey = key, limit = 100, hasCoordinate = TRUE)
gbifmap(dat$data)</pre>
```



Another package, mapr, is the perfect mapping companion to rgbif. It has convenient functions for handling input data from rgbif, spocc, or arbitrary data.frame's, and output plots for base plots,

- ggplot2, ggmap (ggplot2 with map layers underneath), and interactive maps on GitHub gists or with
 Leaflet.is.
- 288 There's no equivalent interface in pygbif or gbifrb.
- 289 GBIF data in other R packages
- We discuss usage of GBIF data in other R packages throughout the manuscript, but provide a synopsis here for clarity.
- 292 taxize
- Some of the GBIF taxonomic services are also available in taxize, an R package that focuses on getting
 data from taxonomic data sources on the web. For example, with get_gbifid() one can get GBIF IDs
 used for a set of taxonomic names then use those IDs in other functions in taxize to get additional
 information, like taxonomically downstream children.
- 297 *spocc*
- GBIF occurrence data is available in the R package spocc via rgbif. spocc is a unified interface for fetching species occurrence data from many sources on the web. For example, a user can collect occurrence data from GBIF, iDigBio, and iNaturalist, and easily combine them, then use other packages to clean and visualize the data.

302 R vs. Python vs. Ruby

- Both R and Python are commonly used in science, and can be used for similar tasks. Python, however,
- is a more general programming lanuage, and can be used in more contexts than R can be used in.
- Ruby is used very little in science; but, like Python, Ruby is very widely used as a general purpose
- programming language, with heavy use in web development and web services.
- The three clients can do a lot of the same tasks. We envision rgbif being more common in workflows
- of academics asking research questions, whereas pygbif and gbifrb can do that as well, but may be
- more easily used in a website.
- The R client rgbif has had much more development time than pygbif and gbifrb, but with time pygbif and gbifrb will become equally mature.

Use cases

- The following are three use cases for the R library rgbif: niche modeling, spatial change in biodiversity,
- 314 and distribution mapping.
- 315 Ecological niche modeling
- In this example, we plot actual occurrence data for *Bradypus* species against a single predictor variable,
- BIO1 (annual mean temperature). This is only ont step in a species distribution modelling nworkflow.
- This example can be done using BISON data as well with our rbison package.
- 319 Load libraries

```
library("sp")
library("rgbif")
library("dismo")
library("maptools")
library("dplyr")
```

- 320 Raster files
- Make a list of files that are installed with the dismo package, then create a rasterStack from these

Get world boundaries

```
data(wrld_simpl)
```

Get GBIF data using the rOpenSci package rgbif

```
nn <- name_lookup("bradypus*", rank = "species")
nn <- na.omit(unique(nn$data$nubKey))
df <- occ_search(taxonKey = nn, hasCoordinate = TRUE, limit = 500)</pre>
```

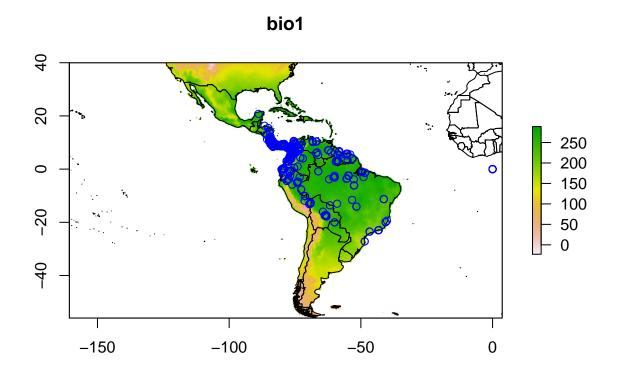
```
df_data <- df[ sapply(df, function(x) any(class(x$data) %in% "tbl_df")) ]
df_data <- dplyr::bind_rows(lapply(df_data, "[[", "data"))
df2 <- df_data %>% dplyr::select(decimalLongitude, decimalLatitude)
```

Plot

325

(1) Add raster data, (2) Add political boundaries, (3) Add the points (occurrences)

```
plot(predictors, 1)
plot(wrld_simpl, add = TRUE)
points(df2, col = "blue")
```



327 Biodiversity in big cities

In this example, we collect specimen records across different cities using GBIF data from the rgbif package.

Load libraries

326

```
library("rgbif")
library("ggplot2")
library("plyr")
library("httr")
library("RColorBrewer")
library("wicket")
```

- 331 Get bounding boxes for some cites
- Bounding lat/long data is from https://raw.github.com/amyxzhang/boundingbox-cities/master/
- boundbox.txt.

```
url <- 'https://raw.githubusercontent.com/amyxzhang/
boundingbox-cities/master/boundbox.txt'
rawdat <- content(GET(sub("\n", "", url)), as = "text")
dat <- read.table(
   text = rawdat, header = FALSE,
   sep = "\t", col.names = c("city", "minlat", "maxlon", "maxlat", "minlon"),
   stringsAsFactors = FALSE)
dat <- data.frame(
   city = dat$city, minlon = dat$minlon,
   minlat = dat$minlat, maxlon = dat$maxlon,
   maxlat = dat$maxlat,
   stringsAsFactors = FALSE
)</pre>
```

A helper function to get count data. GBIF has a count API, but we can't use that with a geometry search as that API doesn't support geospatial search. We can however use the search API via occ_search() and set limit = 1 so that we

```
getdata <- function(x){
  coords <- as.numeric(x[c('minlon','minlat','maxlon','maxlat')])
  wkt <- wicket::wkt_correct(wicket::bounding_wkt(values = coords))</pre>
```

```
num <- occ_search(geometry = wkt, limit = 1)$meta$count
data.frame(
    city = x['city'],
    richness = num,
    stringsAsFactors = FALSE
)</pre>
```

```
out <- apply(dat, 1, getdata)</pre>
```

337 Merge to original table

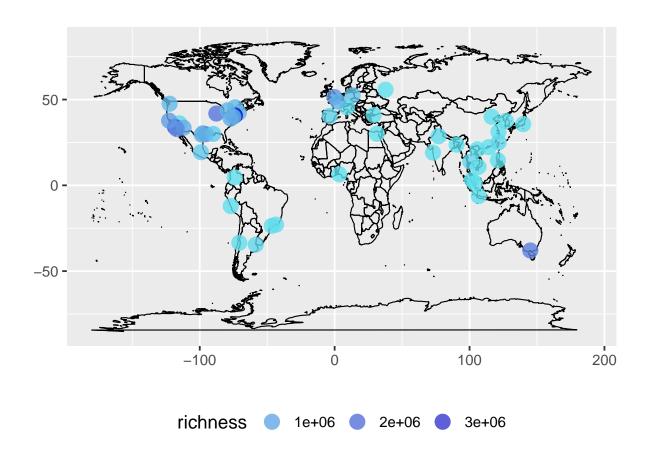
```
out <- merge(dat, ldply(out), by = "city")</pre>
```

Add centroids from bounding boxes

```
out <- transform(out, lat = (minlat + maxlat)/2, lon = (minlon + maxlon)/2)
```

Plot data

```
mapp <- map_data('world')
ggplot(mapp, aes(long, lat)) +
  geom_polygon(aes(group=group), fill="white", alpha=0, color="black", size=0.4) +
  geom_point(data=out, aes(lon, lat, color=richness), size=5, alpha=0.8) +
  scale_color_continuous(low = "#60E1EE", high = "#0404C8") +
  labs(x="", y="") +
  theme_grey(base_size=14) +
  theme(legend.position = "bottom", legend.key = element_blank()) +
  guides(color = guide_legend(keywidth = 2))</pre>
```



Valley oak occurrence data comparison

This example comes from Antonio J. Perez-Luque who shared his plot on Twitter. Antonio compared the occurrences of Valley Oak (*Quercus lobata*) from GBIF to the distribution of the same species from the Atlas of US Trees.

Load libraries

340

```
library('rgbif')
library('raster')
library('sp')
library('sf')
library('rgeos')
library('scales')
```

346 Get GBIF Data for Quercus lobata

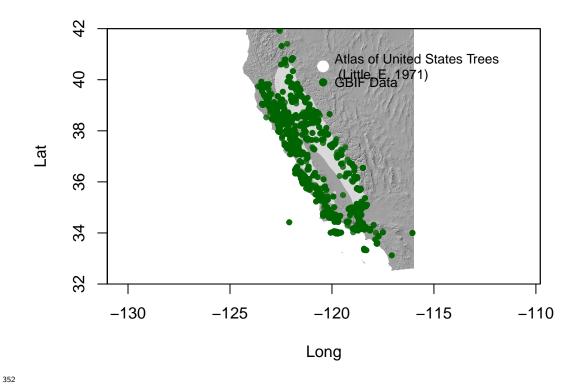
```
keyQl <- name_backbone(name='Quercus lobata', kingdom='plants')$speciesKey</pre>
dat.Ql <- occ_search(taxonKey=keyQl, return='data', limit=50000)</pre>
Get Distribution map of Q. lobata Atlas of US Trees (Little, E.)
From http://esp.cr.usgs.gov/data/little. And save shapefile in same directory
url <- 'https://github.com/sckott/gbifms/blob/gh-pages/querloba.zip?raw=true'</pre>
tmp <- tempdir()</pre>
download.file(url, destfile = "querloba.zip")
unzip("querloba.zip", exdir = tmp)
ql <- sf::read_sf(file.path(tmp, "querloba/querloba.shp"))</pre>
Get Elevation data of US
alt.USA <- getData('alt', country = 'USA')
Create Hillshade of US
alt.USA <- alt.USA[[1]]</pre>
slope.USA <- terrain(alt.USA, opt = 'slope')</pre>
aspect.USA <- terrain(alt.USA, opt = 'aspect')</pre>
hill.USA <- hillShade(slope.USA, aspect.USA, angle = 45, direction = 315)
Plot map
plot(hill.USA, col = grey(0:100/100), legend = FALSE,
      xlim = c(-125, -116), ylim = c(32, 42),
      main = 'Distribution of Quercus lobata',
      xlab = "Long", ylab = 'Lat')
# add shape from Atlas of US Trees
```

plot(q1, add = TRUE, col = alpha("white", 0.6),

border = FALSE)

add Gbif presence points

Distribution of Quercus lobata



Conclusions and future directions

The rgbif, pygbif, and gbibrb libraries provide programmatic interfaces to GBIF's application programming interface (API) - a powerful tool for working with species occurrence data, and facilitating reproducible research. In fact, the rgbif package has already been used in more than 20 scholarly publications (as of 2016-08-10), including (Amano, Lamming & Sutherland, 2016, Bartomeus et al. (2013), Barve (2014), Bone et al. (2015), Collins et al. (2015), Drozd & Šipoš (2013), Kong, Huang &

- Duan (2015), Richardson, Roux & Wilson (2015), Turner, Fréville & Rieseberg (2015), Verheijen et al.
- 360 (2015), Zizka & Antonelli (2015), Butterfield et al. (2016), Dellinger et al. (2015), Feitosa et al. (2015),
- 361 Malhado et al. (2015), Werner et al. (2015), Robertson, Visser & Hui (2016), Davison et al. (2015),
- 362 Janssens et al. (2016)).
- The rgbif package is relatively stable, and should not have many breaking changes unless necessitated
- due to changes in the GBIF API. However, it will gain function(s) to work with the maps API in the
- 365 near future.
- The pygbif and gbifrb libraries are in early development, and will greatly benefit from any feedback
- and use cases.
- One area of focus in the future is to attempt to solve many use cases that have been brought up with
- respect to GBIF data. For example, some specimens are included in GBIF that are located in botanical
- gardens. For many research questions, researchers are interested in "wild" type occurrences, not those
- in human curated scenarios. Making removal of these occurrences easy would be very useful, but is
- actually quite a hard problem. There are many other problems like this, for which these three libraries
- will help in making more efficient and reproducible.

374 Acknowledgments

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377 Data Accessibility

378 All scripts and data used in this paper can be found in the permanent data archive Zenodo under

the digital object identifier (DOI). This DOI corresponds to a snapshot of the GitHub repository

at https://github.com/sckott/gbifms. Software can be found at https://github.com/ropensci/rgbif,

https://github.com/sckott/pygbif, and https://github.com/sckott/gibfrb, all under MIT licenses. We

thank all the users that have used rgbif, pygbif, and gbifrb and have given feedback and reported

bugs. In addition, we greatly appreciate all the contributors to the three libraries, found at https://

github.com/ropensci/rgbif/graphs/contributors, https://github.com/sckott/pygbif/graphs/contributors,

and https://github.com/sckott/gbifrb/graphs/contributors.

386 References

- Amano T., Lamming JDL., Sutherland WJ. 2016. Spatial gaps in global biodiversity information and
- the role of citizen science. BioScience 66:393-400.
- Bartomeus I., Park MG., Gibbs J., Danforth BN., Lakso AN., Winfree R. 2013. Biodiversity ensures
- plant-pollinator phenological synchrony against climate change. Ecology Letters 16:1331–1338.
- Barve V. 2014. Discovering and developing primary biodiversity data from social networking sites: A
- novel approach. Ecological Informatics 24:194–199.
- Beck J., Ballesteros-Mejia L., Buchmann CM., Dengler J., Fritz SA., Gruber B., Hof C., Jansen
- F., Knapp S., Kreft H., Schneider A-K., Winter M., Dormann CF. 2012. Whats on the horizon for
- macroecology? *Ecography* 35:673–683.
- Bone RE., Smith JAC., Arrigo N., Buerki S. 2015. A macro-ecological perspective on crassulacean acid
- metabolism (CAM) photosynthesis evolution in afro-madagascan drylands: Eulophiinae orchids as a
- case study. New Phytologist 208:469–481.
- Brown JH. 1995. Macroecology. University of Chicago Press.
- 400 Brown KA., Parks KE., Bethell CA., Johnson SE., Mulligan M. 2015. Predicting plant diversity patterns
- 401 in madagascar: Understanding the effects of climate and land cover change in a biodiversity hotspot.
- 402 PLOS ONE 10:e0122721.
- Butterfield BJ., Copeland SM., Munson SM., Roybal CM., Wood TE. 2016. Prestoration: Using species
- in restoration that will persist now and into the future. Restor Ecol.
- 405 Ceballos G., Ehrlich PR., Barnosky AD., Garcia A., Pringle RM., Palmer TM. 2015. Accelerated
- modern human-induced species losses: Entering the sixth mass extinction. Science Advances 1:e1400253-
- 407 e1400253.
- 408 Chamberlain S., Ram K., Barve V., Mcglinn D. rgbif: An r interface to the global 'biodiversity'
- 409 information facility API.
- 410 Chamberlain S. pygbif: A python interface to the global biodiversity information facility API.
- 411 Chamberlain S. gbifrb: A ruby interface to the global biodiversity information facility API.
- ⁴¹² Collins R., Ribeiro ED., Machado VN., Hrbek T., Farias I. 2015. A preliminary inventory of the catfishes

- of the lower rio nhamundá, brazil (ostariophysi, siluriformes). BDJ 3:e4162.
- Davison J., Moora M., Opik M., Adholeya A., Ainsaar L., Ba A., Burla S., Diedhiou AG., Hiiesalu
- I., Jairus T., Johnson NC., Kane A., Koorem K., Kochar M., Ndiaye C., Partel M., Reier U., Saks
- 416 U., Singh R., Vasar M., Zobel M. 2015. Global assessment of arbuscular mycorrhizal fungus diversity
- reveals very low endemism. Science 349:970–973.
- Dellinger AS., Essl F., Hojsgaard D., Kirchheimer B., Klatt S., Dawson W., Pergl J., Pyšek P., Kleunen
- M van., Weber E., Winter M., Hörandl E., Dullinger S. 2015. Niche dynamics of alien species do not
- differ among sexual and apomictic flowering plants. New Phytologist 209:1313–1323.
- Drozd P., Šipoš J. 2013. R for all (i): Introduction to the new age of biological analyses. Casopis
- slezskeho zemskeho muzea (A) 62.
- Faulkner KT., Robertson MP., Rouget M., Wilson JR. 2014. A simple, rapid methodology for developing
- invasive species watch lists. Biological Conservation 179:25–32.
- Febbraro MD., Lurz PWW., Genovesi P., Maiorano L., Girardello M., Bertolino S. 2013. The use of
- climatic niches in screening procedures for introduced species to evaluate risk of spread: A case with
- the american eastern grey squirrel. *PLoS ONE* 8:e66559.
- 428 Feitosa YO., Absy ML., Latrubesse EM., Stevaux JC. 2015. Late quaternary vegetation dynamics from
- central parts of the madeira river in brazil. Acta Bot. Bras. 29:120-128.
- Ferretti F., Verd GM., Seret B., Šprem JS., Micheli F. 2015. Falling through the cracks: The fading
- history of a large iconic predator. Fish and Fisheries:n/a-n/a.
- 432 Ficetola GF., Rondinini C., Bonardi A., Baisero D., Padoa-Schioppa E. 2014. Habitat availability for
- amphibians and extinction threat: A global analysis. Diversity and Distributions 21:302–311.
- 434 Herring J. 2011. OpenGIS implementation standard for geographic information-simple feature access-
- part 1: Common architecture. OGC Document 4:122–127.
- Janssens SB., Vandelook F., Langhe ED., Verstraete B., Smets E., Vandenhouwe I., Swennen R. 2016.
- 437 Evolutionary dynamics and biogeography of musaceae reveal a correlation between the diversification
- of the banana family and the geological and climatic history of southeast asia. New Phytologist
- 439 210:1453-1465.
- 440 Kong X., Huang M., Duan R. 2015. SDMdata: A web-based software tool for collecting species

- occurrence records. PLOS ONE 10:e0128295.
- Malhado AC., Oliveira-Neto JA., Stropp J., Strona G., Dias LC., Pinto LB., Ladle RJ. 2015. Climato-
- logical correlates of seed size in amazonian forest trees. J Veg Sci 26:956–963.
- 444 María Mendoza., Ospina OE., Cárdenas-Henao H., García-R JC. 2015. A likelihood inference of
- historical biogeography in the world's most diverse terrestrial vertebrate genus: Diversification of
- direct-developing frogs (craugastoridae: Pristimantis) across the neotropics. Molecular Phylogenetics
- 447 and Evolution 85:50-58.
- Pimm SL., Jenkins CN., Abell R., Brooks TM., Gittleman JL., Joppa LN., Raven PH., Roberts CM.,
- Sexton JO. 2014. The biodiversity of species and their rates of extinction, distribution, and protection.
- 450 Science 344:1246752-1246752.
- R Core Team. 2014. R: A language and environment for statistical computing. Vienna, Austria: R
- 452 Foundation for Statistical Computing.
- Richardson DM., Roux JJL., Wilson JR. 2015. Australian acacias as invasive species: Lessons to be
- learnt from regions with long planting histories. Southern Forests: a Journal of Forest Science 77:31–39.
- Robertson MP., Visser V., Hui C. 2016. Biogeo: An r package for assessing and improving data quality
- of occurrence record datasets. Ecography 39:394–401.
- Turner KG., Fréville H., Rieseberg LH. 2015. Adaptive plasticity and niche expansion in an invasive
- thistle. Ecol Evol 5:3183-3197.
- Verheijen LM., Aerts R., Bönisch G., Kattge J., Bodegom PMV. 2015. Variation in trait trade-offs
- allows differentiation among predefined plant functional types: Implications for predictive ecology. New
- 461 Phytologist 209:563-575.
- Werner GDA., Cornwell WK., Cornelissen JHC., Kiers ET. 2015. Evolutionary signals of symbiotic
- persistence in the legumerhizobia mutualism. Proceedings of the National Academy of Sciences 112:10262-
- 464 10269.
- ⁴⁶⁵ Zizka A., Antonelli A. 2015. speciesgeocodeR: An r package for linking species occurrences, user-defined
- regions and phylogenetic trees for biogeography, ecology and evolution. Cold Spring Harbor Laboratory
- 467 Press.