

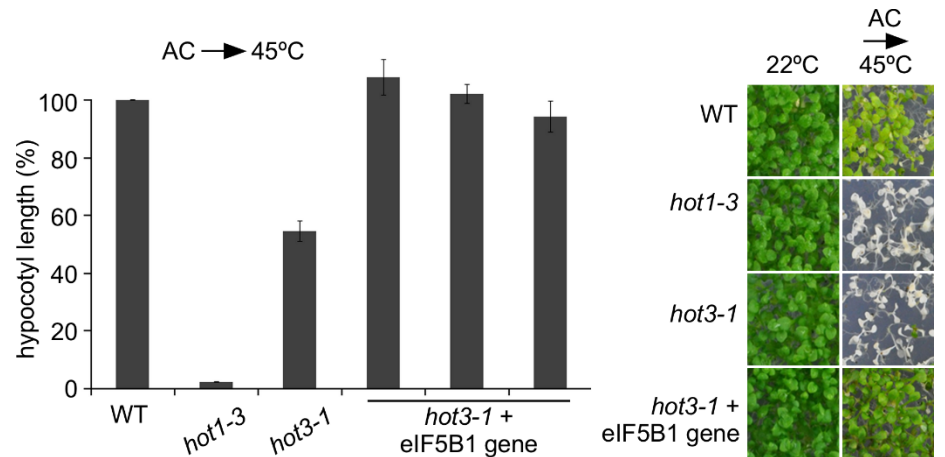
IN BRIEF

Some Like it HOT: Protein Translation and Heat Stress in Plants

The ability to acclimate to high temperatures that are normally lethal is common to virtually all organisms on the planet. A short exposure to milder heat stress informs organisms that they should ready themselves in case they experience even warmer conditions. Acquired thermo-tolerance in plants is important because plants must constantly balance growth and defense; any energy spent on heat stress responses will take away from plant growth and yield. Temperature stress is likely to become more prevalent in the coming decades due to climate change and the rise in extreme weather events. Understanding how plants respond to high temperature stress is therefore crucial in efforts to engineer or breed crop plants, for example, to tilt the balance between defense and growth to our advantage, at a minimal energetic cost for the plant. New work by Vierling, Peng and coauthors (Zhang, et al., 2017) reveals clues about how plants cope with and recover from heat stress.

A genetic analysis of thermotolerance is tricky, as desired mutants are likely to die since they would lack the ability to adapt to and/or recover from heat stress. The key is to design an assay with high survival rate and a clear phenotype. Hong and Vierling (2000) published the first genetic screen in a higher eukaryote for mutants with a loss in acquired thermotolerance, based on cell elongation in *Arabidopsis* seedling hypocotyls. Dark-grown seedlings kept at 72°F die when moved from 72°F to 115°F for only 2 hours, but survive if first pre-conditioned at 100°F before exposure to 115°F. A key sign of impending trouble is that the hypocotyl ceases to elongate, so the *hot* screen looked for mutants whose hypocotyls ceased to grow following pre-treatment at 100°F and treatment at 115°F. The first gene identified from this screen, *HOT1*, encodes the heat shock protein Hsp101, a molecular chaperone critical for acquired thermotolerance in yeast, flies, cyanobacteria and plants (Hong and Vierling, 2000). Heat shock proteins prevent the accumulation of protein denaturation and aggregation following temperature stress, and finding a mutant in *Hsp101* was a welcome validation of the screening conditions.

Zhang et al. (2017) reveal that *HOT3* encodes the translation initiation factor 5B



HOT3/eIF5B1 controls heat acclimation ability. Complementation of *hot3-1* with eIF5B1 rescues the heat sensitivity of *hot3-1* seedlings. Left-hand panel: hypocotyl length as a percentage of control seedlings maintained at 22°C. Error bars indicate SEM; n = 12. Right-hand panel: seedlings maintained at 22°C or subjected to heat stress. AC → 45°C: acclimation at 38°C for 1.5 h and 22°C for 2 h, followed by 45°C for 2.5 h. *hot1-3* carries a T-DNA insertion in the Hsp101 gene and is a highly heat sensitive mutant. [Adapted from Zhang et al. (2017), Figure 1].

(eIF5B). eIF5B plays several roles in initiation of translation, including the joining of ribosomal subunits at the beginning of messenger RNAs to kick-start translation. In a twist of fate, the initial *hot3* mutant, *hot3-1*, is temperature-sensitive, reducing protein stability or activity only at higher temperatures. Temperature-sensitive mutants are powerful tools to dissect the role of genes essential for growth and development, because the mutant phenotype is conditional. In fact, stronger *hot3* alleles have pronounced phenotypes even when grown at 72°F, and affect the translation potential of hundreds of genes even before being exposed to high temperatures.

How does protein translation fit into plant responses to heat stress? Ribosomes fall off mRNAs after exposure to heat stress, but begin to re-associate quickly to initiate new protein synthesis. This process is much slower in *hot3-1*, and as a consequence the translation of 2-3,000 genes is delayed. The mutant is therefore slower to correct heat-induced cellular damage, since it lacks the ability to replenish the necessary proteins in a timely manner. Translation already shows hints of breaking down under milder heat stress in *hot3-1*, suggesting that the mutant may also have problems acclimating to high temperatures.

The *hot3* mutants show delayed translation for only 10-15% of the roughly 20,000 genes expressed at this stage of

development, hinting at a new level of translation regulation governed by interaction between eIF5B and other ribosome components. A future challenge is to use our new knowledge about the role of eIF5B and translation in thermotolerance to understand how the activity of this translation initiation factor may be tuned to benefit plant fitness for agriculture.

Patrice A. Salomé
Department of Chemistry and
Biochemistry
University of California, Los Angeles
salome@chem.ucla.edu
ORCID: 0000-0003-4452-9064

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