- Comparison of large-scale citizen science data and
- long-term study data for phenology modeling
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#### Abstract

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Large-scale observational data from citizen science efforts are becoming increasingly common in ecology, and researchers often choose between these and data from intensive local-scale studies for their analyses. This choice has potential trade-offs related to spatial scale, observer variance, and inter-annual variability. Here we explored this issue with phenology by comparing models built using data from the large-scale, citizen science USA National Phenology Network (USA-NPN) effort with models built using data from more intensive studies at Long Term Ecological Research (LTER) sites. We built statistical and process based phenology models for species common to each dataset. From these models we compared parameter estimates, estimates of phenological events, and out-of-sample errors between models derived from both USA-NPN and LTER data. We found that model parameter estimates for the same species were most similar between the two datasets when using simple models, but parameter estimates varied widely as model complexity increased. Despite this, estimates for the date of phenological events and out-of-sample errors were similar, regardless of the model chosen. Predictions for USA-NPN data had the lowest error when using models built from the USA-NPN data, while LTER predictions were best made using LTER-derived models, confirming that models perform best when applied at the same scale they were built. This difference is likely due to variation in phenological requirements within species. Models using the USA-NPN dataset can integrate parameters over a large spatial scale while those using an LTER dataset can only estimate parameters for a single location. Accordingly, the choice of dataset depends on the research question. Inferences about species-specific phenological requirements are best made with LTER data, and if USA-NPN or similar data are all that is available, then analyses should be limited to simple models. Large-scale predictive modeling is best done with the larger-scale USA-NPN data, which has high spatial representation and a large regional species pool. LTER

datasets, on the other hand, have high site fidelity and thus characterize inter-annual variability
extremely well. Future research aimed at forecasting phenology events for particular species
over larger scales should develop models which integrate the strengths of both datasets.

Keywords: LTER, USA-NPN, forecasting, budburst, flowering, data integration, scale

## Introduction

role in ecological research extending from local to global scales (Cleland et al., 2007; Richardson et al., 2013; Tang et al., 2016). At large scales the timing of spring leaf out and fall senescence influence the carbon budget of earth system models, which has implications for correctly accounting for biosphere-atmosphere feedbacks in long-term climate forecasts (Richardson et al., 2012). At smaller scales, species-specific responses to temperature and precipitation can alter flower communities (Diez et al., 2012; CaraDonna et al., 2014; Theobald et al., 2017) and affect the abundance and richness of both pollinators (Ogilvie and Forrest, 2017; Ogilvie et al., 2017) and organisms at higher trophic levels (Tylianakis et al., 2008). Plant phenology models that are robust at multiple ecological scales, or deemed appropriate for a particular scale, are needed to better understand and 51 forecast the timing of key biological events. Many plant phenology studies use intensively collected datasets from a single location over a long time-period by a single research group (Cook et al., 2012; Wolkovich et al., 2012; Iler et al., 2013; Roberts et al., 2015). These datasets have regular sampling and large numbers of samples over long periods of time. As a result, the biological and climatic variability at that site is well represented. It is common for phenology models built with observations from a single site to not 57 transfer well to other sites (García-Mozo et al., 2008; Xu and Chen, 2013; Olsson and Jönsson, 2014; Basler, 2016). This lack of transferability can be driven by plasticity in phenology requirements, local adaptation, microclimates, or differences in plant age or population density (Kramer, 1995; Diez et al., 2012). For these reasons, data from a single location are not adequate for larger scale phenology modeling. Accurately forecasting phenology at larger scales will require models

Plant phenology, the timing of recurring biological events such as flowering, plays an important

that account for the full range of variation across a species' range (Richardson et al., 2013; Tang et al., 2016; Chuine and Régnière, 2017), which will necessitate the use of data sources beyond traditional single-site studies.

Data from citizen science projects are becoming increasingly important for ecological research (Kelling et al., 2009; Dickinson et al., 2010; Tulloch et al., 2013). Because these data are often collected by large numbers of volunteers, it is possible to gather data at much larger scales than with individual research teams. A relatively new citizen science project started in 2009, Nature's Notebook run by The USA National Phenology Network (USA-NPN), collects phenology observations from volunteers throughout the United States and makes the data openly available (Schwartz et al., 2012). Data from this project have already been used to study variation in oak phenology at a continental scale (Gerst et al., 2017), develop large-scale community phenology models (Melaas et al., 2016), and forecast long-term phenology trends (Jeong et al., 2013). Large-scale datasets from China and Europe have already contributed considerably to phenological research (Xu and Chen, 2013; Olsson and Jönsson, 2014; Basler, 2016; Zhang et al., 2017), and the USA-NPN dataset has the potential to meet these needs for North American plant species and communities. However, the features that allow citizen science projects to collect data at large scales can also introduce spatial biases toward cities and easily-accessible areas, and variation in sampling effort and observer skill (Dickinson et al., 2010). With thousands of participants, the potential for variation among observers in their determination of species identification and dating of phenological events is high. While volunteers have been shown to be accurate at distinguishing different leaf and flower stages for plants (Fuccillo et al., 2015) and can have high agreement on abundance estimates (Feldman et al., 2018), contributions to USA-NPN are sometimes made sporadically across seasons, years, and locations. This means that the quantity and quality of data at a specific site will typically be more variable for citizen science efforts than for intensive, long-term studies.

In order to accurately model and forecast phenology, it is important to understand how the strengths
and weaknesses of intensive local studies and large-scale citizen science projects influence both our
inferences about biological processes driving phenology (e.g. warming requirements for a specific
plant) and our ability to predict future phenology events (e.g. forecasting when flowering or leaf
out occurs). Here, we fit a suite of plant phenology models for the budburst and first-flowering
phenophases of 24 plant species to data from both the USA-NPN and a set of intensive long-term
studies from the Long Term Ecological Research (LTER) network. We compare the resulting models based on both inference about models and parameters and predictions for unobserved events.
We then use this comparison to assess the best methods for both local- and large-scale phenology
modeling and to point the way forward for integrating large-scale and local-scale data to determine
the best possible models across scales.

#### Methods

#### 99 Datasets

The USA National Phenology Network (USA-NPN) protocol uses status-based monitoring, where
via a phone app or web based interface observers answer 'yes,' 'no,' or 'unsure' when asked if
an individual plant has a specific phenophase present (Denny et al., 2014). Phenophases refer to
specific phases in the annual cycle of a plant, such as the presence of emerging leaves, flowers,

fruit, or senescing leaves. Sites in the USA-NPN datasets are located across the U.S. and generally clustered around populated areas (Fig. 1). To represent long-term, intensive phenology studies we used four datasets from North America representing three major ecosystem types (Table 1, Fig. 1).

All four long-term studies are located in the U.S. and are part of the Long Term Ecological Research network (LTER). The Harvard Forest and Hubbard Brook Long Term Experimental Forest are located in the northeastern U.S. and are dominated by deciduous broadleaf species. The H.J.

Andrews Experimental Forest is a coniferous forest in the coastal range of the western U.S. The Jornada Experimental Range is in the Chihuahua desert of the southwestern U.S.

We downloaded all USA-NPN observations from 2009, when collections began, to 2016 for the 112 following phenophases: Breaking Leaf Buds, Breaking Needle Buds, Emerging Needles, and Open 113 Flowers (USA National Phenology Network, 2017). The first three phenophases apply to the 114 'leaf out' phase for deciduous broadleafs, evergreen conifers, and pines, respectively. The 'Open 115 Flowers' phenophase refers to fully-open flowers and applies to all angiosperms. Hereafter, we 116 will refer to these as either 'Flowers' for the Open Flower phenophase, or 'Budburst' for all other 117 phenophases. We subset the USA-NPN observations similar to methods outlined in Crimmins et al. 118 (2017). First, 'yes' observations for individual plants were kept only if they were preceded by a 119 'no' observation within 30 days. Observations for 'Budburst' that were past day of year (DOY) 120 172, and for 'Flowers' that were past DOY 213 were dropped to minimize any influence from outliers. We inferred the observed DOY of each phenophase as the midpoint between each 'yes' 122 observation and the preceding 'no' observation. Finally, only species that had greater than 30 total observations were kept. Crimmins et al. (2017) only kept observations that were preceded by a 'no' within 15 days, and also grouped multiple individuals at single sites to a single observation. We used 30 days to allow for a greater number of species to be compared. We tested the sensitivity of
this choice by also performing the analysis using a 15 day cutoff. We chose not to group multiple
individuals at a single site to better incorporate intra-site variability.

In the LTER datasets observation metrics varied widely due to different protocols. To match the 129 USA-NPN data we converted all metrics to binary 'yes' and 'no' observations for each phenophase 130 (see supplementary methods). Three of the LTER datasets (Hubbard Brook, Harvard Forest, and 131 H.J. Andrews) had a sampling frequency of 3-7 days during the growing season. The Jornada 132 dataset had a sampling frequency of 30 days. As with the USA-NPN data, we inferred the date for 133 each phenophase as the midpoint between the first 'yes' observation and most recent 'no' observa-134 tion, and only kept species and phenophase combinations which had at least 30 total observations. 135 After data processing there were 38 species and phenophase combinations (with 24 unique species) 136 common to both the USA-NPN and LTER datasets to use in the analysis (Table 1 & S1). Using a 137 15 day cutoff in the USA-NPN dataset resulted in 35 unique combinations with 23 species. 138

#### 39 Models

It is common to fit multiple plant phenology models to find the one that best represents a specific species and phenophase (Chuine et al., 2013). For each of the 38 species and phenophase combinations in the five datasets (USA-NPN and four LTER datasets), we fit eight phenology models (Table 2). The *Naive model* uses the mean DOY from prior observations as the estimated DOY. The *Linear model* uses a regression with the mean spring (Jan. 1 - March 31) temperature as the independent variable and DOY as the response variable. For the six remaining models, the general form is based on the idea that a phenological event will occur once sufficient thermal forcing units,

 $F^*$ , accumulate from a particular start day of the year  $(t_1)$ . Forcing units are a transformation of the daily mean temperature and are calculated differently for each model (Table 2). The start day can either be estimated or fixed. For the *Growing Degree Day (GDD)* model, forcing units are the total degrees above the threshold  $T_{base}$  (Réaumur, 1735; Wang, 1960; Hunter and Lechowicz, 1992). The *Fixed GDD model* uses the same form but has fixed values for start day  $(t_1 = \text{Jan 1})$  and temperature threshold  $(T_{base} = 0^{\circ}\text{C})$ . The *Alternating model* has a variable number of required forcing units defined as a function of the total number of days below  $0^{\circ}\text{C}$  since Jan. 1 (number of chill days - NCD). The *Uniforc model* is like the *GDD model* but with the forcing units transformed via a sigmoid function (Chuine, 2000).

We also fit two models that attempt to capture spatial variation in phenological requirements. The
first spatial model, MI, is an extension of the GDD model which adds a correction in the required
forcing using the photoperiod (L) (Blümel and Chmielewski, 2012). The second, the *Macroscale*Species-specific Budburst model (MSB), uses the mean spring temperature as a linear correction
on the total forcing required in the *Alternating model* (Jeong et al., 2013). Since there is little to
no spatial variation in the LTER datasets, we fit the two spatial models to data from the USA-NPN
only. We compared the resulting parameters, estimates, and errors for the USA-NPN derived MIand MSB models to their non-spatial analogs (the GDD and Alternating models, respectively) for
each species and phenophase in the LTER data.

We extracted corresponding daily mean temperature for all USA-NPN and LTER observations from the gridded PRISM dataset using the latitude and longitude of the site associated with each observation (PRISM Climate Group, 2004). We parameterized all models using differential evolution to minimize the root mean square error (RMSE) of the estimated DOY of the phenological

event. Differential evolution is a global optimization algorithm which uses a population of randomly initialized models to find the set of parameters that minimize the RMSE (Storn and Price,
1997). Confidence intervals for parameters were obtained by bootstrapping, in which individual
models were re-fit 250 times using a random sample, with replacement, of the data. We made
predictions by taking the mean DOY estimated from the 250 bootstrapped iterations. A random
subset consisting of 20% of observations from each species and phenophase combination was held
out from model fitting for later evaluation.

### 76 Analysis

As described above, we fit two sets of models for each species and phenophase: one set of models parameterized using only USA-NPN data, and one set parameterized using only LTER data (with the exception of the *M1* and *MSB* models, see above). We performed three primary analyses from these model outputs by comparing: 1) the model parameters, 2) estimates from the models, and 3) out-of-sample errors from each model.

To compare the inferences about process made by the two datasets, we compared the distribution of each parameter between LTER and USA-NPN derived models for each species and phenophase combination. Using the mean value of each bootstrapped parameter, we also calculated the coefficient of determination ( $R^2$ ) between LTER and USA-NPN derived models among the 38 species-phenophases. In three cases where a species phenophase combination occurred in two LTER sites (Budburst for *Acer saccharum*, *Betula alleghaniensis*, and *Fagus grandifolia* in the Harvard and Hubbard Brook datasets) they were compared separately to the USA-NPN data.

Next we compared the estimates of phenological events between models. Models with different parameter values, and even entirely different structures, can produce similar estimates for the date of phenological events (Basler, 2016). Therefore, to compare the predictions and potential forecasts for models fit to the different datasets, we compared the estimated DOY predicted by the LTER and USA-NPN derived models for all held out observations. For each of the eight models, we calculated the coefficient of determination ( $R^2$ ) between LTER and USA-NPN derived estimates for estimates made at the four LTER sites and across all USA-NPN sites.

Finally, we directly evaluated performance using out-of-sample errors from the four combinations of models and observed data: A) LTER-derived models predicting LTER observations, B) USA-197 NPN derived models predicting LTER observations, C) LTER-derived models predicting USA-198 NPN observations, and D) USA-NPN derived models predicting USA-NPN observations. Using 199 the RMSE values from held out observations, we compared the performance of LTER and USA-200 NPN derived models on different data types in two different ways. First, we focused on local-scale 201 prediction by calculating the difference in the RMSE of LTER and USA-NPN derived models 202 solely with LTER observations. Secondly, to focus on large-scale prediction we calculated the 203 difference in RMSE using solely USA-NPN data. These differences were calculated for each of the 204 model types and 38 species-phenophase combinations. Negative values indicate that LTER-derived 205 models perform better, while positive values indicate that the USA-NPN derived model performed 206 better. We used a t-test to test the difference from zero in these values. In the three cases where the 207 same species and phenophase combination occurred in two LTER sites, we made the LTER-LTER 208 comparison within each site, not across sites, to focus on local scale prediction when LTER data are available. Absolute RMSE values as well as Pearson correlation coefficients are provided in the

- supplement for specific species (Fig. S5-S7) and with all observations aggregated together (Table S2).
- We performed all analysis using both the R and Python programming languages (R Core Team,
- 2017; Python Software Foundation, 2018). Primary R packages used in the analysis included
- dplyr (Wickham et al., 2017), tidyr (Wickham and Henry, 2018), ggplot2 (Wickham, 2016), lubri-
- date (Grolemund and Wickham, 2011), prism (Hart and Bell, 2015), raster (Hijmans, 2017), and
- sp (Pebesma and Bivand, 2005). Primary Python packages included SciPy (Jones et al., 2001),
- NumPy (Oliphant, 2006), Pandas (McKinney, 2010), and MPI for Python (Dalcin et al., 2011).
- <sup>219</sup> Code to fully reproduce this analysis is available on GitHub (https://github.com/sdtaylor/phenology\_dataset\_study
- and archived on Zenodo (https://doi.org/10.5281/zenodo.1256705)

## 221 Results

- Throughout the analysis there were no qualitative differences between a 30-day or 15-day threshold
- between the first 'yes' and most recent 'no' observation in the USA-NPN dataset. Results presented
- here reflect the 30 day cutoff, see the supplementary figures S2-S4 for matching figures using a 15
- 225 day cutoff.
- The best matches between parameter estimates based on USA-NPN and LTER data were the *Fixed*
- GDD model ( $R^2 = 0.49$ ) and the Linear model ( $R^2 = 0.39$  for  $\beta_1$  and -0.05 for  $\beta_2$ ). The parameters
- for all other models had  $R^2$  values <0 indicating that the relationship was worse than no relationship
- between the parameters (but with matching mean parameter values across the two sets of models)
- 230 (Fig. 2). The Naive model showed a distinct late bias in mean DOY estimates for phenological

events, likely resulting from the LTER datasets being mostly in the northern United States compared to the site locations of the USA-NPN dataset (Fig. 2). The large outlier for the *Fixed GDD*model is *Larrea tridentata*; this species' flower phenology is largely driven by precipitation, which
is not considered in the Fixed GDD model (Beatley, 1974). While the *Fixed GDD* and *Linear*models showed reasonable correspondence between parameter estimates, all parameters for individual species and phenophase combinations had different distributions between USA-NPN and
LTER-derived models (Fig. S10-S11).

When comparing estimates of phenological events between the two sets of models, many USA-NPN and LTER models produced similar estimates (Fig. 3). The *Fixed GDD model* had the highest correlation between the two models sets at USA-NPN sites ( $R^2 = 0.82$ ), while the *GDD*, MI, and *Uniforc* models had the highest correlation at LTER sites ( $R^2 = 0.51$ , 0.52, and 0.51, respectively). Comparing models with spatial corrections to the non-spatial alternatives, the *MSB* (an extension of the *Alternating model* with a spatial correction based on mean spring temperature, see Table 2 and Methods) improved the correlation between the two datasets over the *Alternating model*. The *MSB model* improved the  $R^2$  from 0.36 to 0.45 at LTER sites, and from -0.23 to -0.15 at USA-NPN sites. The *M1 model* (an extension of the *GDD model* with a spatial correction based on day length) improved the correlation over the *GDD model* only slightly at LTER sites (from 0.51 to 0.52) and did not improve the correlation at USA-NPN sites.

When comparing the predictive performance using out-of-sample errors, USA-NPN derived models made more accurate predictions for held-out USA-NPN observations, and LTER-derived models performed better on held-out LTER observations (all p < 0.001, Fig. 4). The *Naive* and *Lin*ear models had the largest differences between the two model sets, while the *Fixed GDD model*  had relatively similar errors when evaluated on both USA-NPN and LTER held-out observations.

Although the *Fixed GDD model* had the highest agreement in accuracy between USA-NPN and

LTER-derived models, it was not the best performing model overall. The *GDD* and *Uniforc* models

made the best out of sample predictions, having the lowest RMSE and Pearson coefficient when

aggregating all observations together (Table S2). One exception was that the Fixed GDD model

had a slightly higher Pearson value when using LTER-derived models to make predictions for

USA-NPN data. The best model for specific species and phenophases varied, but was commonly

the *Uniforc* and *GDD* models (Figs. S5-S6).

## Discussion

Data used to build phenology models typically falls into two categories: intensive long-term data 262 with long time-series at a small number of locations (e.g., LTER data in this study), and large-263 scale data with less intensive sampling at hundreds of locations (e.g., USA-NPN data) (Table 3). 264 This data scenario—a small amount of intensive data and a large amount of less intensive data—is 265 common in many areas of science and makes it necessary to understand how to choose between, 266 or combine, data sources (Hanks et al., 2011). We explored this issue for phenology modeling 267 in relation to making predictions and inferring process from models. For inference we found 268 that models based on different data sources resulted in different parameter estimates for all but 269 the simplest models. For prediction we found that models fit to different data sources tended to make similar predictions, but that models better predicted out-of-sample data from the data type to which they were fit. These results are consistent with other research showing that phenology model performance decreases when transferring single-site models to other locations (García-Mozo et al., 2008; Xu and Chen, 2013; Basler, 2016), and with the call for models that better incorporate spatial variation in phenology requirements (Richardson et al., 2013; Chuine and Régnière, 2017).

Understanding and making predictions for the phenology of a single location is best served by intensive local-scale data, when available, but large-scale datasets work better for extrapolating phenology predictions across a species range. Thus, the best choice of both data and models depends on the desired research goals.

In this study, parameter estimates differed widely within the same phenology model when fit to the two different types of data, except for the simplest process-oriented model: the Fixed GDD (Fig. 281 2). These differences may be caused by a variety of factors that have different implications for 282 interpreting process-oriented models and their parameters. First, the differences could result from 283 limitations in the sampling of the USA-NPN dataset, such as irregular sampling of the same loca-284 tion within or between seasons, leading to less accurate parameter estimates. If this is the case, it 285 would suggest that using LTER data is ideal for making inferences about plant physiology, and that 286 focusing on the Fixed GDD model is best for making inferences when USA-NPN data are all that 287 is available. Second, spatial variation (e.g. from local adaptation, acclimation, microclimates, or 288 plant age) in phenology requirements and drivers could contribute to these differences (Diez et al., 289 2012; Zhang et al., 2017). Models built using USA-NPN data integrate over that spatial variation, 290 while models built using LTER data only estimate the phenological requirements for a specific site. 291 In this case, USA-NPN data would provide a better estimate of the general phenological requirements of a species, but LTER data would provide a more accurate understanding for a single site. The best solution to this issue would be the development of models that accurately incorporate spatial variation, such as including genetic variation between different populations (Chuine and Régnière, 2017), although localized models could also be generated when large-scale predictions are unnecessary. Third, these differences could result from issues with model identifiability: since different parameter values can yield nearly identical estimates of phenological events, parameter estimates can differ between datasets even when the underlying processes generating the data are the same. Information about which of these issues may be causing the differences between datasets can be explored using the analyses in the current study, as will be explained below.

Despite substantial differences in parameter estimates, LTER and USA-NPN derived models pro-302 duced similar estimates for phenological events in most cases (Fig. 3). This greater correspondence 303 between predictions than parameters suggests that more complex models may have identifiability 304 issues. For example, two GDD models with parameters of  $t_1$ =1, F=10,  $T_{base}$ =0 and  $t_1$ =5, F=5, 305  $T_{base}$ =0 produce nearly identical estimates in many scenarios. This possibility is supported by the 306 fact that the highest correlation between parameter estimates is seen in models with only 1 or 2 307 parameters. In addition, bootstrap results for more complex models suggest a high degree of vari-308 ability in parameter estimates and potentially multiple local optima in fits to both USA-NPN and 309 LTER data (Fig. S10-S11). Finally, parameter estimates of more complex models are also not con-310 sistent among models for the same species when comparing multiple LTER datasets (Fig. S8-S9). 311 These results are consistent with research showing that models failed to estimate the starting day 312 of warming accumulation solely from budbreak time-series, thus producing parameter estimates 313 that were not biologically realistic (Chuine et al., 2016). Basler (2016) suggests that the key component in phenology models is the thermal forcing, with additional parameters being sensitive to over-fitting. Here, our simplest model, the Fixed GDD model which uses only a warming component, had the highest correlation among parameters between LTER and USA-NPN datasets. In combination with the aforementioned studies, our results indicate that caution is warranted in interpreting parameter estimates from complex phenology models regardless of the data source used for fitting the models.

While more complex phenology models appear to have identifiability issues, there is also evi-321 dence that they capture useful information, beyond the Fixed GDD model, based on their ability 322 to make out-of-sample predictions. Based on the RMSE, the GDD and Uniforc models produce 323 the best out-of-sample predictions for the majority of species and phenophases at both USA-NPN 324 and LTER datasets (Fig. S5 & S6). This demonstrates that the more complex models are capturing 325 additional information about phenology, and that some of the differences between datasets result 326 from differences in either the scales or the sampling of the data. Spatial variation in phenologi-327 cal requirements is known to exist in plants (Zhang et al., 2017). In combination with our results 328 showing observed differences in parameter estimates between LTER sites (Fig. S8-S9), this sug-329 gests that variation in phenological requirements across the range is likely important. However, the 330 models that attempted to address this by incorporating spatial variation did not yield improvements 331 over their base models in our analyses. Specifically, correspondence between parameter estimates 332 (Fig. 2), estimates of phenological events (Fig. 3), and out-of-sample error rates (Fig. 4) for the 333 MSB and M1 models were essentially the same as the Alternating and GDD models, respectively. 334 This lack of improvement from incorporating spatial variation could be caused either by models 335 not adequately capturing the process driving the spatial variation, the USA-NPN dataset having biases from variation in sampling effort and/or spatial auto-correlation, or some combination of these factors. Basler (2016) used the M1 model to predict budburst for six species across Europe

and found it was generally among the best models in terms of RMSE, albeit never by more than
a single day. Their result was strengthened by having a 40-year time-series across a large region.
Chuine and Régnière (2017) listed the incorporation of spatial variation in warming requirements
in models as a primary issue in future phenology research. Large-scale phenology datasets, like
USA-NPN, will be key in addressing this and other phenological research needs.

In addition to exploring differences between phenology datasets, our analyses provide guidance on
which models to use when making predictions at a local scale using models built from large-scale
data, or vice versa. Among the eight models tested, the *Uniforc* and *GDD* models performed the
best overall in the cross dataset comparison in terms of Pearson correlation and RMSE (Fig. S5-S6,
Table S2). The *GDD* model has one less parameter than the *Uniforc* model, thus the *GDD* model
is a suitable choice for making predictions when there is little to no information at the location
of interest (e.g. making phenology forecasts at a new location distant from any observed data).
This guidance can vary between species though and model testing should still be performed when
suitable data is available.

In conclusion, our results suggest that both LTER and USA-NPN data provide valuable information
on plant phenology. Models built using both data sources yield effective predictions for phenological events, but parameter estimates from the two data sources differ and models from each source
best predict that data source's phenology events. The primary difference in the datasets is spatial
scale, but due to trade-offs in data collection efforts, the larger scale USA-NPN data have shorter
time-series, less site fidelity and other differences from the intensively collected LTER data (Table
359
301. These differences can be strengths or potential limitations. Observers sampling opportunistically allows the USA-NPN dataset to have a large spatial scale, but also leads to low site fidelity,

which limits the ability to measure long-term trends at local scales (Gerst et al., 2016). Tracking long-term trends is the major strength of LTER data, but having a relatively small species pool limits their use in species-level predictive modeling. Due to these differences, the best data source for making predictions depends on the scale at which the predictions are being made. Identifying the most effective data sources for different types and scales of analysis is a useful first step, but 365 the ultimate solution to working with diverse data types is to focus on integrating all types of data 366 into analyses and forecasts (Hanks et al., 2011; Melaas et al., 2016). Our results suggest that meth-367 ods that can learn from the intensive information available in LTER data in regions where they 368 are available, and simultaneously use large-scale data to capture spatial variation in phenological 360 requirements will help improve our ability to understand and predict phenology. Data integration 370 efforts should also leverage data from remote sensing sources such as the PHENOCAM network or 371 satellite imagery, which have both a large spatial extent and high temporal resolution (Peng et al., 372 2017; Richardson et al., 2018a,b). Data integration provides the potential to use data from many 373 sources to produce the best opportunity for accurate inference about, and forecasting of, the timing 374 of biological events.

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## **References**

- Bailey, A. (2018). Hubbard brook experimental forest (us forest service): Routine seasonal phenology measurements, 1989 present. environmental data initiative.

  https://doi.org/10.6073/pasta/765084e2b4a5ec389403238c58784572.
- Basler, D. (2016). Evaluating phenological models for the prediction of leaf-out dates in six temperate tree species across central europe. *Agricultural and Forest Meteorology*, 217:10–21.
- Beatley, J. C. (1974). Effects of rainfall and temperature on the distribution and behavior of larrea tridentata (creosote-bush) in the mojave desert of nevada. *Ecology*, 55(2):245–261.
- Blümel, K. and Chmielewski, F. M. (2012). Shortcomings of classical phenological forcing models and a way to overcome them. *Agricultural and Forest Meteorology*, 164:10–19.
- Cannell, M. G. R. and Smith, R. I. (1983). Thermal time, chill days and prediction of budburst in picea sitchensis. *The Journal of Applied Ecology*, 20(3):951.

- CaraDonna, P. J., Iler, A. M., and Inouye, D. W. (2014). Shifts in flowering phenology reshape a
   subalpine plant community. *Proceedings of the National Academy of Sciences*, 111(13):4916–
   4921.
- Chuine, I. (2000). A unified model for budburst of trees. *Journal of Theoretical Biology*, 207(3):337–347.
- Chuine, I., Bonhomme, M., Legave, J. M., García de Cortázar-Atauri, I., Charrier, G., Lacointe,

  A., and Améglio, T. (2016). Can phenological models predict tree phenology accurately in the

  future? the unrevealed hurdle of endodormancy break. *Global Change Biology*, 22(10):3444–

  3460.
- Chuine, I., de Cortazar-Atauri, I. G., Kramer, K., and Hänninen, H. (2013). Plant development models. In Schwartz, M. D., editor, *Phenology: An Integrative Environmental Science*, pages 275–293. Springer Netherlands, Dordrecht.
- Chuine, I. and Régnière, J. (2017). Process-Based Models of Phenology for Plants and Animals.

  Annual Review of Ecology, Evolution, and Systematics, 48(1):159–182.
- Cleland, E., Chuine, I., Menzel, A., Mooney, H., and Schwartz, M. (2007). Shifting plant phenology in response to global change. *Trends in Ecology Evolution*, 22(7):357–365.
- Cook, B. I., Wolkovich, E. M., and Parmesan, C. (2012). Divergent responses to spring and winter warming drive community level flowering trends. *Proceedings of the National Academy of Sciences*, 109(23):9000–9005.
- Crimmins, T. M., Crimmins, M. A., Gerst, K. L., Rosemartin, A. H., and Weltzin, J. F. (2017).

- Usa national phenology network's volunteer-contributed observations yield predictive models of phenological transitions. *PLOS ONE*, 12(8):e0182919.
- Dalcin, L. D., Paz, R. R., Kler, P. A., and Cosimo, A. (2011). Parallel distributed computing using python. *Advances in Water Resources*, 34(9):1124–1139.
- Denny, E. G., Gerst, K. L., Miller-Rushing, A. J., Tierney, G. L., Crimmins, T. M., Enquist, C.
- A. F., Guertin, P., Rosemartin, A. H., Schwartz, M. D., Thomas, K. A., and Weltzin, J. F. (2014).
- Standardized phenology monitoring methods to track plant and animal activity for science and
- resource management applications. *International Journal of Biometeorology*, 58(4):591–601.
- Dickinson, J., Zuckerberg, B., and Bonter, D. (2010). Citizen science as an ecological research
- tool: Challenges and benefits. Annual Review of Ecology, Evolution and Systematics, 41(1):149–
- 430 172.
- 431 Diez, J. M., Ibáñez, I., Miller-Rushing, A. J., Mazer, S. J., Crimmins, T. M., Crimmins, M. A.,
- Bertelsen, C. D., and Inouye, D. W. (2012). Forecasting phenology: from species variability to
- community patterns. *Ecology Letters*, 15(6):545–553.
- Feldman, R. E., Žemaitė, I., and Miller-Rushing, A. J. (2018). How training citizen scientists af-
- fects the accuracy and precision of phenological data. *International Journal of Biometeorology*,
- 436 62(8):1421–1435.
- Fuccillo, K. K., Crimmins, T. M., de Rivera, C. E., and Elder, T. S. (2015). Assessing accuracy
- in citizen science-based plant phenology monitoring. *International Journal of Biometeorology*,
- 439 59(7):917–926.
- 440 García-Mozo, H., Chuine, I., Aira, M., Belmonte, J., Bermejo, D., Díaz de la Guardia, C., Elvira,

- B., Gutiérrez, M., Rodríguez-Rajo, J., Ruiz, L., Trigo, M., Tormo, R., Valencia, R., and Galán,
- 442 C. (2008). Regional phenological models for forecasting the start and peak of the quercus pollen
- season in spain. Agricultural and Forest Meteorology, 148(3):372–380.
- Gerst, K. L., Kellermann, J. L., Enquist, C. A. F., Rosemartin, A. H., and Denny, E. G. (2016).
- Estimating the onset of spring from a complex phenology database: trade-offs across geographic
- scales. *International Journal of Biometeorology*, 60(3):391–400.
- 447 Gerst, K. L., Rossington, N. L., and Mazer, S. J. (2017). Phenological responsiveness to climate
- differs among four species of quercus in north america. *Journal of Ecology*, 38(1):42–49.
- Grolemund, G. and Wickham, H. (2011). Dates and times made easy with lubridate. Journal of
- Statistical Software, 40(3):1–25.
- Hanks, E. M., Hooten, M. B., and Baker, F. A. (2011). Reconciling multiple data sources to
- improve accuracy of large-scale prediction of forest disease incidence. *Ecological Applications*,
- 453 21(4):1173–1188.
- 454 Hart, E. M. and Bell, K. (2015). prism: Download data from the oregon prism project.
- http://github.com/ropensci/prism.
- 456 Hijmans, R. J. (2017). raster: Geographic data analysis and modeling. r package version 2.6-7.
- https://CRAN.R-project.org/package=raster.
- Hunter, A. F. and Lechowicz, M. J. (1992). Predicting the Timing of Budburst in Temperate Trees.
- The Journal of Applied Ecology, 29(3):597.
- Iler, A. M., Høye, T. T., Inouye, D. W., and Schmidt, N. M. (2013). Nonlinear flowering re-

- sponses to climate: are species approaching their limits of phenological change? *Philosophical*
- *Transactions of the Royal Society of London*, 368(1624):20120489.
- Jeong, S.-J., Medvigy, D., Shevliakova, E., and Malyshev, S. (2013). Predicting changes in tem-
- perate forest budburst using continental-scale observations and models. Geophysical Research
- Letters, 40(2):359–364.
- Jones, E., Oliphant, T., Peterson, P., and Others (2001). Scipy: Open source scientific tools for
- python. http://www.scipy.org/.
- Kelling, S., Hochachka, W. M., Fink, D., Riedewald, M., Caruana, R., Ballard, G., and Hooker,
- G. (2009). Data-intensive science: A new paradigm for biodiversity studies. *BioScience*,
- 470 59(7):613–620.
- 471 Kramer, K. (1995). Phenotypic plasticity of the phenology of seven european tree species in
- relation to climatic warming. *Plant, Cell and Environment*, 18(2):93–104.
- 473 McKinney, W. (2010). Data structures for statistical computing in python. In *Proceedings of the*
- 9th Python in Science Conference, pages 51–56.
- Melaas, E. K., Friedl, M. A., and Richardson, A. D. (2016). Multiscale modeling of spring phenol-
- ogy across deciduous forests in the eastern united states. Global Change Biology, 22(2):792–
- 477 805.
- Ogilvie, J. E. and Forrest, J. R. (2017). Interactions between bee foraging and floral resource
- phenology shape bee populations and communities. Current Opinion in Insect Science, 21:75–
- 480 82.
- Ogilvie, J. E., Griffin, S. R., Gezon, Z. J., Inouye, B. D., Underwood, N., Inouye, D. W., and Irwin,

- R. E. (2017). Interannual bumble bee abundance is driven by indirect climate effects on floral resource phenology. *Ecology Letters*, 20(12):1507–1515.
- O'Keefe, J. (2015). Phenology of woody species at harvard forest since 1990. harvard forest data archive: Hf003.
- Oliphant, T. (2006). A guide to numpy. USA: Trelgol Publishing.
- Olsson, C. and Jönsson, A. M. (2014). Process-based models not always better than empirical
- models for simulating budburst of norway spruce and birch in europe. Global Change Biology,
- 20(11):3492–3507.
- Pebesma, E. J. and Bivand, R. S. (2005). Classes and methods for spatial data in R. *R News*, 5(2):9–13.
- Peng, D., Wu, C., Li, C., Zhang, X., Liu, Z., Ye, H., Luo, S., Liu, X., Hu, Y., and Fang, B. (2017).
- Spring green-up phenology products derived from MODIS NDVI and EVI: Intercomparison,
- interpretation and validation using National Phenology Network and AmeriFlux observations.
- 495 Ecological Indicators, 77:323–336.
- PRISM Climate Group (2004). Oregon state university. http://prism.oregonstate.edu.
- Python Software Foundation (2018). Python Language Reference Manual, version 3.6.
- http://www.python.org.
- <sup>499</sup> R Core Team (2017). R: a language and environment for statistical computing.
- Réaumur, R. (1735). Observations du thermomètres, faites a Paris pendant l'année 1735, com-

- parées avec celles qui ont été faites sous la ligne, a l'isle de France, a Alger et quelques unes de nos isles de l'Amérique. *Mem Paris Acad Sci*, 1735(545).
- Richardson, A. D., Anderson, R. S., Arain, M. A., Barr, A. G., Bohrer, G., Chen, G., Chen, J. M.,
- <sup>504</sup> Ciais, P., Davis, K. J., Desai, A. R., Dietze, M. C., Dragoni, D., Garrity, S. R., Gough, C. M.,
- Grant, R., Hollinger, D. Y., Margolis, H. A., McCaughey, H., Migliavacca, M., Monson, R. K.,
- Munger, J. W., Poulter, B., Raczka, B. M., Ricciuto, D. M., Sahoo, A. K., Schaefer, K., Tian,
- H., Vargas, R., Verbeeck, H., Xiao, J., and Xue, Y. (2012). Terrestrial biosphere models need
- better representation of vegetation phenology: results from the north american carbon program
- site synthesis. *Global Change Biology*, 18(2):566–584.
- Richardson, A. D., Hufkens, K., Milliman, T., Aubrecht, D. M., Chen, M., Gray, J. M., Johnston,
- M. R., Keenan, T. F., Klosterman, S. T., Kosmala, M., Melaas, E. K., Friedl, M. A., and Frol-
- king, S. (2018a). Tracking vegetation phenology across diverse north american biomes using
- phenocam imagery. Scientific Data, 5:180028.
- Richardson, A. D., Hufkens, K., Milliman, T., and Frolking, S. (2018b). Intercomparison of phe-
- nological transition dates derived from the PhenoCam Dataset V1.0 and MODIS satellite remote
- sensing. Scientific Reports, 8(1):5679.
- Richardson, A. D., Keenan, T. F., Migliavacca, M., Ryu, Y., Sonnentag, O., and Toomey, M.
- 518 (2013). Climate change, phenology, and phenological control of vegetation feedbacks to the
- climate system. Agricultural and Forest Meteorology, 169:156–173.
- Roberts, A. M. I., Tansey, C., Smithers, R. J., and Phillimore, A. B. (2015). Predicting a change in
- the order of spring phenology in temperate forests. *Global Change Biology*, 21(7):2603–2611.

- 522 Schulze, M. D. (2017). Vegetative phenology observations at the andrews experimental for-
- est, 2009 present. long-term ecological research. forest science data bank. corvallis, or.
- http://andlter.forestry.oregonstate.edu/data/abstract.aspx?dbcode=TV075.
- Schwartz, M. D., Betancourt, J. L., and Weltzin, J. F. (2012). From caprio's lilacs to the usa
- national phenology network. Frontiers in Ecology and the Environment, 10(6):324–327.
- Storn, R. and Price, K. (1997). Differential evolution a simple and efficient heuristic for global
- optimization over continuous spaces. *Journal of Global Optimization*, 11(4):341–359.
- Tang, J., Körner, C., Muraoka, H., Piao, S., Shen, M., Thackeray, S. J., and Yang, X. (2016).
- Emerging opportunities and challenges in phenology: a review. *Ecosphere*, 7(8):e01436.
- Theobald, E. J., Breckheimer, I., and HilleRisLambers, J. (2017). Climate drives phenological
- reassembly of a mountain wildflower meadow community. *Ecology*, 98(11):2799–2812.
- Tulloch, A. I. T., Possingham, H. P., Joseph, L. N., Szabo, J., and Martin, T. G. (2013). Realising
- the full potential of citizen science monitoring programs. *Biological Conservation*, 165:128–
- 138.
- Tylianakis, J. M., Didham, R. K., Bascompte, J., and Wardle, D. A. (2008). Global change and
- species interactions in terrestrial ecosystems. *Ecology Letters*, 11(12):1351–1363.
- 538 USA National Phenology Network (2017). Plant and animal phenology data. data type: Sta-
- tus and intensity. 01/01/2009-04/31/2017 for region: 49.9375, -66.4791667 (ur); 24.0625,
- -125.0208333 (II). USA-NPN, Tucson, Arizona, USA. Data set accessed 04/20/2017 at
- http://doi.org/10.5066/F78S4N1V.

- Wang, J. Y. (1960). A Critique of the Heat Unit Approach to Plant Response Studies. *Ecology*,
   41(4):785–790.
- Wickham, H. (2016). ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
- Wickham, H., Francois, R., Henry, L., and Müller, K. (2017). dplyr: A grammar of data manipulation.
- Wickham, H. and Henry, L. (2018). *tidyr: Easily Tidy Data with 'spread()' and 'gather()' Func-tions*.
- Wolkovich, E. M., Cook, B. I., Allen, J. M., Crimmins, T. M., Betancourt, J. L., Travers, S. E., Pau,
- 550 S., Regetz, J., Davies, T. J., Kraft, N. J. B., Ault, T. R., Bolmgren, K., Mazer, S. J., McCabe,
- G. J., McGill, B. J., Parmesan, C., Salamin, N., Schwartz, M. D., and Cleland, E. E. (2012).
- Warming experiments underpredict plant phenological responses to climate change. *Nature*,
- 485(7399):494–497.
- Xu, L. and Chen, X. (2013). Regional unified model-based leaf unfolding prediction from 1960 to
   2009 across northern china. *Global Change Biology*, 19(4):1275–1284.
- <sup>556</sup> Zhang, H., Liu, S., Regnier, P., and Yuan, W. (2017). New insights on plant phenological response
- to temperature revealed from long-term widespread observations in china. Global Change Biol-
- *ogy*, 12(10):3218–3221.

Dataset Name	Habitat	Phenological Event (Num. Species) Reference	Reference
Harvard Forest	N.E. Deciduous Forest	Budburst (17) Flowering (7)	(O'Keefe, 2015)
Jornada Experimental Range	Chihuahuan Desert	Flowering (2)	
H.J. Andrews Experimental Forest N.W. Wet Coniferous Forest Budburst (5) Flowering (4)	N.W. Wet Coniferous Forest	Budburst (5) Flowering (4)	(Schulze, 2017)
Hubbard Brook	N.E. Deciduous Forest	Budburst (3)	(Bailey, 2018)

 Table 1: LTER datasets used in the analysis

Name	DOY Estimator	Forcing Equations	Total	Reference
			Parameters	S
Naive	$\overline{DOY}$	ı	1	
Fixed GDD	$\sum_{i=0}^{DOY} R_f(T_i) \geq F^*$	$R_f(T_i) = max(T_i, 0)$	П	(Réaumur, 1735; Wang, 1960; Hunter and Lechowicz, 1992)
Linear	$DOY = eta_1 + eta_2 T_{mean}$	1	2	ı
GDD	$\sum_{i=t_1}^{DOY} R_f(T_i) \geq F^*$	$R_f(T_i) = max(T_i - T_{base}, 0)$	8	(Réaumur, 1735; Wang, 1960; Hunter and Lechowicz, 1992)
M1	$\sum_{t=t_1}^{DOY} R_f(T_i) \geq (rac{L_i}{24})^k F^*$	$R_f(T_i) = max(T_i - T_{base}, 5)$	4	(Blümel and Chmielewski, 2012)
Alternating	$\sum_{t=0}^{DOY} R_f(T_i) \geq a + be^{cNCD(t)}$	$R_f(T_i) = max(T_i - 5, 0)$	3	(Cannell and Smith, 1983)
N <b>SS</b> B	$\sum_{t=0}^{DOY} R_f(T_i) \ge a + be^{cNCD_i} + dT_{mean}$	$R_f(T_i) = max(T_i - 5, 0)$	4	(Jeong et al., 2013)
Uniforc	$\sum_{l=t_1}^{DOY} R_f(T_l) \geq F^*$	$R_f(T_i) = rac{1}{1+e^{b(T_i-c)}}$	4	(Chuine, 2000)

Table 2: Phenology models used in the analysis

		<u>LTER</u>	<u>USA-NPN</u>
	Time-series length	High	Low
	Spatial extent	Low	High
563	Local species representation	High	Low
	Regional/Continental species representation	Low	High

Number of observers Low High

Site fidelity High Low

Table 3: Attributes of the two datasets used in this study. Bold text indicates an attribute is expected

to increase over time.

Figure 1: Locations of National Phenology Network sites used (black points) and Long Term

Ecological Research sites (labeled circles), with greyscale showing elevation.

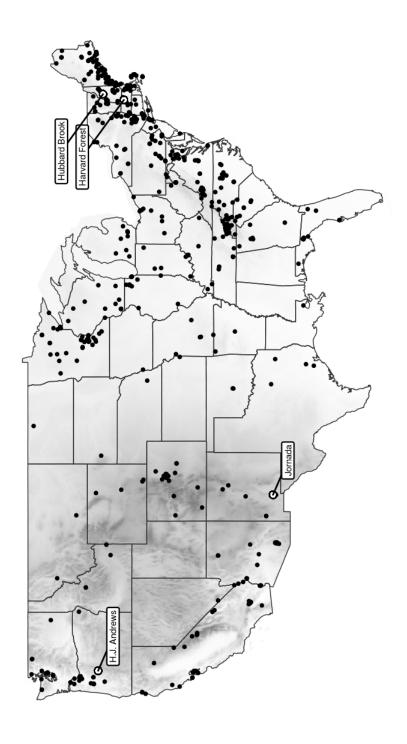


Figure 1: .

Figure 2: Comparisons of parameter estimates between USA-NPN and LTER derived models.

Each point represents a parameter value for a specific species and phenophase, and is the mean

value from 250 bootstrap iterations. The black line is the 1:1 line. The  $R^2$  is the coefficient of

determination, which can be negative if the relationship between the two parameter sets is worse

than no relationship but with the same mean values.

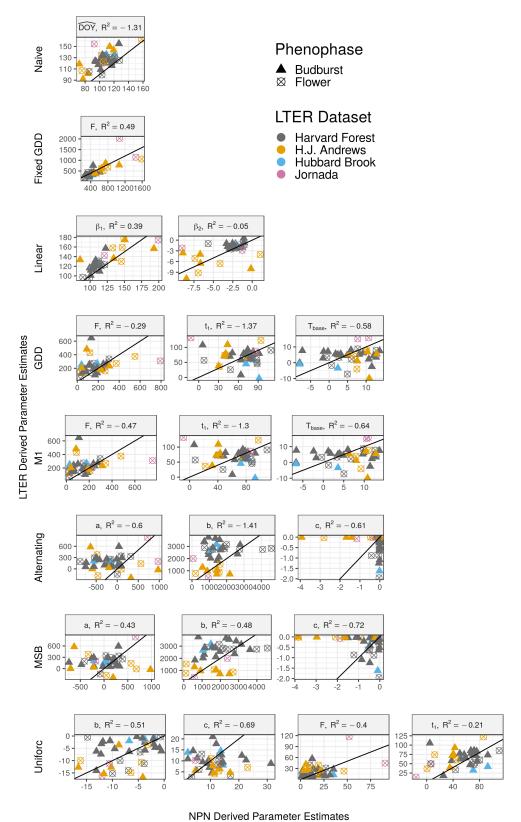


Figure 2: .

- Figure 3: Comparison of predicted day of year (DOY) of all phenological events between USA-
- NPN and LTER-derived models. Top panels show comparisons at LTER sites and bottom panels
- show comparisons at USA-NPN sites. Each point is an estimate for a single held-out observation.
- 576 Colors indicate observations for a single species and phenophase combination.

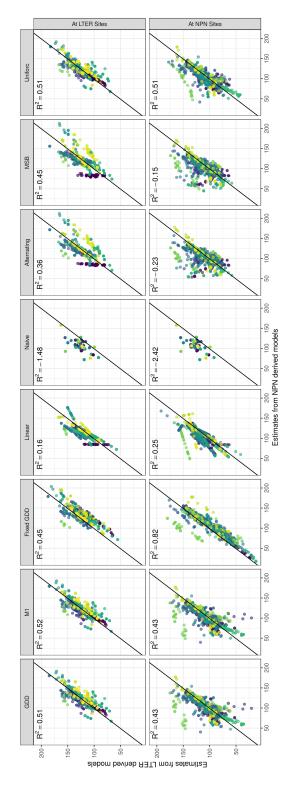


Figure 3: .

Figure 4: Differences in prediction error between USA-NPN and LTER-derived models. Density plots for comparisons of predictions on LTER data (top row) and USA-NPN data (bottom row). Each plot represents the difference between the RMSE for LTER-derived model and the USA-NPN derived model, meaning that values less than zero indicate more accurate prediction by LTER-derived models and values greater than zero indicate more accurate prediction by NPN-derived models. p < 0.001 for all t-tests. Differences are calculated pairwise for the 38 species/phenophase comparisons.

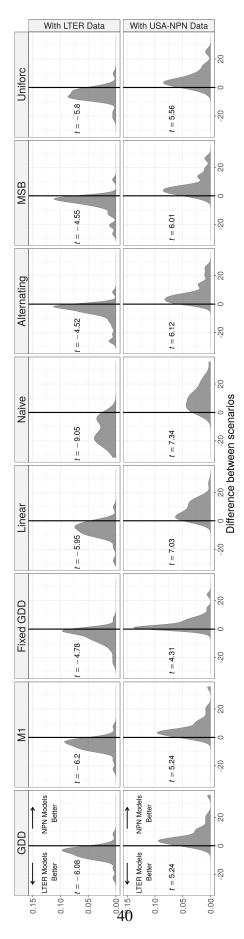


Figure 4: .