The following report addresses four questions from the exercise protocol and discusses the methods used. Section 1 provides a brief summary of the history of LiDAR applications in forestry science. In the following sections (2-4), details are discussed of the data and methods used to develop eight candidate regression models. In section 3, these models are evaluated using accuracy assessments and k-fold cross-validation tests.

1. LiDAR tools in forestry science

Since the 1980s, LiDAR tools have been applied to forest inventories (1–6). These have two approaches to remote sensing: area-based approaches and individual tree detection. Based on remote measurements of mean tree diameter and mean tree height, studies have used LiDAR data to estimate forest biomass (7–10), examine forest structure (11–13), and examine individual tree characteristics such as crown area and length, stem location (14–16), canopy fuel parameters (17), and even tree species (18,19). However, integrating remote sensing data into a multi-phase forest inventory sampling requires a statistical framework that is based on unbiased model-assisted estimators (20–22).

2. Building a LiDAR model for predicting above ground biomass (AGB)

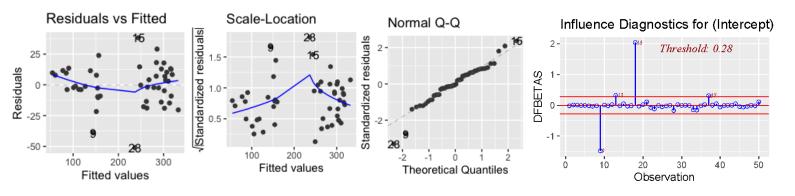
The aim of this exercise was to develop and evaluate regression models for predicting AGB of the Clocaenog forest site. Models were calibrated with data between field plot measurements and remote sensing statistics, which were linked using global navigation satellite systems (23). Field data was collected from among 50 randomly sampled concentric circular plots (10m radius), including measurements of tree diameters at breast height (dbh, cm). Using allometric equations, dbh measurements were used to estimate a response variable: plot-level above ground biomass (*AGB*, Mg/ha⁻¹). Remote sensing data was generated from an airborne laser scanning survey (ALS). From this survey, LiDAR returns (Light Detection and Ranging) were calculated to create three predictor variables. These included: mean tree height (MeanH) based on height returns (11,24–29), standard deviation (SD) of height returns, and fractional canopy height (Cover) based on ratio of height returns above 1.3m to total returns (30–35).

Table 1. Descriptive statistics of predictor variables.

Variable	Mean	SD	SE	Skewness	Kurtosis	SW
MeanH	13.05	5.77	0.82	-0.32	-1.40	***
Cover	0.92	0.08	0.01	-1.76	4.03	***
SD	5.41	2.84	0.40	-0.15	-1.47	***

Table 2. Phase 1 diagnostics; Hypothesis tests of 8 cross-validated regressions of simple linear (LM), stepwise linear models (LM.Stepwise), nonlinear least squared (NLS), stepwise nonlinear least squared regression (NLS.Stepwise), and robust regression (RR) using full dataset and cleaned dataset with influential outliers removed.

Model	SW	BP	DW	BF	Varia	Variance Inflation Factor			AIC
					MeanH	SD	Cover		
1.LM.Full	0.131	0.637	0.345	0.025	3.234	5.451	2.886	437.286	427.727
2.LM.Cleaned	0.949	0.187	0.303	0.001	3.743	4.951	9.493	413.282	403.926
3.LM.Stepwise	0.916	0.586	0.437	0.010	1.903	1.903	/	442.689	435.041
4.NLS.Full	0.968	0.417	0.344	0.049	28.554	12.128	2.959	439.239	427.766
5.NLS.Cleaned	0.961	0.110	0.287	0.002	34.967	13.391	5.221	415.310	404.083
6.RR.Huber	0.002	0.002	0.345	0.025	3.234	5.451	2.886	429.647	430.087
7.RR.Tukey	0.000	0.002	0.345	0.25	3.234	5.451	2.886	444.975	435.415
8.RR.Hampel	0.005	0.002	0.345	0.25	3.234	5.451	2.886	438.499	428.938



1. Accuracy assessments of LiDAR AGB models

Mean Biomass (full/cleaned): 220.628/223.965

Model	RMSE	RMSE%	MAE	MAE%	R^2	Theil's U	$G_4^{2 change}$
1.LM.Full	16.63	7.6%	13.67	6.2%	0.971	204.25	
2.LM.Cleaned	15.92	7.1%	13.15	5.9 %	0.970	41.64	
3.LM.Stepwise	16.44	7.5%	13.34	6.1%	0.973		
4.NLS.Full	16.34	7.4%	13.26	6.0%	0.974		
5.NLS.Cleaned	15.90	7.1%	13.91	6.2%	0.971		
6.RR.Huber	17.19	7.8%	14.08	6.4%	0.970		
7.RR.Tukey	17.31	7.9%	13.94	6.3%	0.967		
8.RR.Hampel	17.48	7.9%	14.08	6.4%	0.972		

Model estimates for final equation	MeanH	SD	Cover
1.LM.Full	9.75(0.73)***	15.56(1.93)***	157.69(51.38)**
2.LM.Cleaned	9.73(0.75)***	15.10(2.39)***	132.09(89.79)
3.LM.Stepwise	11.19(0.61)***	10.77(1.24)***	/
4.NLS.Full	7.03(2.16)**	12.72(2.86)***	146.78(51.59)
5.NLS.Cleaned	6.95(2.27)**	13.23(2.82)***	159.11(91.51)
6.RR.Huber	9.53(0.60)***	15.49(1.58)***	116.03(42.07)**
7.RR.Tukey	9.39(0.58)***	15.42(1.53)***	80.29(40.51)
8.RR.Hampel	9.59(0.66)***	15.60(1.73)***	129.46(46.00)**

Table 2: #Durban-Watson test for autocorrelation of disturbances. Shaprio-Wilks's p-values reporting signficant deviation from normal distribution at the levels of <0.05, 0.01, 0.001 as *, ** and **

From Tojal et al 2019: Shapiro–Wilk test to verify the normality of the residuals, Breusch–Pagan test to analyze the homoscedasticity of the residuals, Durbin–Watson test to detect possible dependencies between the data, Variance Inflation Factor (VIF) to detect collinearity problems in the model [37], Ramsey's RESET linearity test to verify linear relations, and, finally, Bonferroni's test to find statistically significant atypical values. All tests, except VIF, were calculated at the 95% level of significance. (GN: Gauss-Newton method is an iterative algorithm to solve non-linear least squares problems –inspired by kernel regression and Newton method but does not require computation of approximate Hessian, instead looks for a minimum sum of squared function values (36).

2. Limits to LiDAR biomass models

Based on observations in the literature that upper percentile heights typically correlate with plot-level tree height (e.g. Magnussen & Boudewyn, 1998; Næsset, 1997), all the major upper height percentiles from the median to max should be extracted and correlated to field height values. Using pearson's rank correlations, links to be made between plot-level field-measured mean tree height and all percentile heights should be performed and dated (Hopkinson)

Influence of Vegetation Structure on Lidar-derived Canopy Height and Fractional Cover in Forested Riparian Buffers During Leaf-Off and Leaf-On Conditions (Wasser et al)

The R^2 is a biased measure, inasmuch it increases the more terms are added into the model as predictors. The adjusted R^2 compensates for this overfitting by penalizing for the number of terms included as predictors

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```
R Syntax:
#### MSc Inventory Monitoring and Assessment ####
##### Assignment 3; LIDAR AGB Predict ########
#5 steps that follow:
#1. Explore predictors and run linear model using full dataset. Run diagnostics & check violations in residuals
#2.Clean dataset removing biggest outliers
#3.Run 7 alternative models using full & cleaned dataset. Run diagnostics & accuracy assessment
#4. Choose model based on pre/post-diagnostics and accuracy assessment
#5.Plot residuals using benchmark diagnostics
library(readxl)
ClocaenogField <- read excel("ClocaenogField.xlsx")
View(ClocaenogField)
#Explore data and normality of predictor variables:
describe(ClocaenogField$MEANH)
describe(ClocaenogField$COVER)
describe(ClocaenogField$SD)
shapiro.test(ClocaenogField$MEANH)
shapiro.test(ClocaenogField$COVER)
shapiro.test(ClocaenogField$SD)
#MODEL 1 (LM1): Linear regression
model lm1 <- lm(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField)
summary (model lm1)
#Run model diagnostics and check linear assumptions
ols test normality(model lm1)
ols test breusch pagan(model lm1)
dwtest(model lm1)
ols coll diag(model lm1)
outlierTest(model lm1, data=Duncan)
ols test outlier(model lm1)
ols vif tol(model lm1)
autoplot(model lm1)
ols plot diagnostics(model lm1)
ols plot cooksd bar(model lm1)
ols plot dfbetas(model lm1)
ols plot resid lev(model lm1)
ols plot comp plus resid(model lm1, print plot = TRUE)
#Bonferroni results significant and influential outliers found.
#Create new dataframe removing outliers and re-run linear model.
#Check again for violations and compare model 1 & 2 with non-linear models.
#remove outliers from rows 9 and 18
ClocaenogField cleaned <- ClocaenogField[-c(9,18),]
```

```
#MODEL 2 (LM Cleaned): Linear regression using cleaned dataset
model lm2 <- lm(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField cleaned)
summary (model lm2)
#run model diagnostics again
ols test normality(model lm2)
ols test breusch pagan(model lm2)
dwtest(model lm2)
ols coll diag(model lm2)
outlierTest(model lm2, data=Duncan)
ols test outlier(model lm2)
ols vif tol(model lm2)
autoplot(model lm2)
ols plot diagnostics(model lm2)
ols plot cooksd bar(model lm2)
ols plot dfbetas(model lm2)
ols plot resid lev(model lm2)
ols plot comp plus resid(model lm2, print plot = TRUE)
#same violations found from bonferroni results and variance inflation factors
#resort to stepwise, non-linear and robust regression and compare...
#MODEL 3 (LM-STEP-Full): Stepwise Linear regression using full dataset
# Step-wise backwards 10-fold cross-validation test of model accuracy based on RMSE
# Set seed for reproducibility
# Set up repeated k-fold cross-validation
train.control lm1 <- trainControl(method = "cv", number = 10)
# Train the model
step.model lm1 <- train(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField, method =
"leapBackward", tuneGrid = data.frame(nvmax = 1:3), trControl = train.control lm1)
step.model lm1$results
summary(step.model lm1$finalModel)
# Results suggest best 2-variable model contain MEANH and SD
# Step-wise model using most best 2-variables: MEANH and SD
model lm step full <- lm(BIOMASS ~ MEANH + SD, data = ClocaenogField)
summary(model lm step full)
#run diagnostics
ols test normality(model lm step full)
ols test breusch pagan(model lm step full)
dwtest(model lm step full)
ols coll diag(model lm step full)
outlierTest(model lm step full, data=Duncan)
ols test outlier(model lm step full)
ols vif tol(model lm step full)
autoplot(model lm step full)
ols plot diagnostics(model lm step full)
ols plot cooksd bar(model lm step full)
ols plot dfbetas(model lm step full)
ols plot resid lev(model lm step full)
ols plot comp plus resid(model lm step full, print plot = TRUE)
```

```
# MODEL 4 (NLS) Non-linear least squares regression model using full dataset
# Non-linear least squares regression
model nls1 <- lm(ClocaenogField$BIOMASS ~ ClocaenogField$MEANH + ClocaenogField$COVER +
ClocaenogField$SD + I((ClocaenogField$MEANH + ClocaenogField$COVER + ClocaenogField$SD)^2))
summary(model nls1)
# MODEL 5 (NLS) Non-linear least squares regression model using cleaned dataset
# Non-linear least squares regression
model nls1 cleaned <- lm(ClocaenogField cleaned$BIOMASS ~ ClocaenogField cleaned$MEANH +
ClocaenogField cleaned$COVER + ClocaenogField cleaned$SD + I((ClocaenogField cleaned$MEANH +
ClocaenogField cleaned$COVER + ClocaenogField cleaned$SD)^2))
summary(model nls1 cleaned)
# MODEL 6 (RLM.Huber) Robust regression using the Huber M-estimator to reduce outlier influence.
Though this does not reduce outliers in predictor variables, so Tukey needed below
rr.huber <- rlm(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField)
summary(rr.huber)
f.robftest(rr.huber,var = "MEANH")
f.robftest(rr.huber, var = "SD")
f.robftest(rr.huber, var = "COVER")
# MODEL 7 (RLM.Tukey) Robust regression using the Tukey M-estimator that assigns a weight of zero to
influential outliers
rr.bisquare <- rlm(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField, psi = psi.bisquare)
summary(rr.bisquare)
f.robftest(rr.bisquare,var = "MEANH")
f.robftest(rr.bisguare, var = "SD")
f.robftest(rr.bisquare, var = "COVER")
# MODEL 8 (RLM.Tukey) Robust regression using the Tukey M-estimator that assigns a weight of zero to
influential outliers
rr.hampel <- rlm(BIOMASS ~ MEANH + COVER + SD, data = ClocaenogField, psi = psi.hampel)
summary(rr.hampel)
f.robftest(rr.hampel,var = "MEANH")
f.robftest(rr.hampel, var = "SD")
f.robftest(rr.hampel, var = "COVER")
# Explore altherative accuracy assessments:
# sidak p value adjustment
ols test breusch pagan(model nls1, rhs = TRUE, multiple = TRUE, p.adj = 'sidak')
# holm's p value adjustment
ols test breusch pagan(model nls1, rhs = TRUE, multiple = TRUE, p.adj = 'holm')
# Global test of model assumptions
library(gvlma)
gvmodel <- gvlma(model lm1)
summary(gymodel)
gymodel del <- deletion.gylma(gymodel)
summary(gymodel del)
```

```
#compare models
ols mallows cp(model lm1, model_lm2)
ols fpe(model lm1)
ols hsp(model lm1)
ols mallows cp(model lm1, model lm2)
ols mallows cp(model lm1, model lm step full)
ols mallows cp(model lm1, model nls1)
ols mallows cp(model lm1, model nls1 cleaned)
ols mallows cp(model lm1, rr.huber)
ols mallows cp(model lm1, rr.bisquare)
ols mallows cp(model lm1, rr.hampel)
AIC(model lm1)
AIC(model lm2)
AIC(model lm step full)
AIC(model nls1)
AIC(model nls1 cleaned)
AIC(rr.huber)
AIC(rr.bisquare)
AIC(rr.hampel)
BIC(model lm1)
BIC(model lm2)
BIC(model lm step_full)
BIC(model nls1)
BIC(model nls1 cleaned)
BIC(rr.huber)
BIC(rr.bisquare)
BIC(rr.hampel)
glance(model lm1)
glance(model lm2)
glance(model lm step full)
glance(model nls1)
glance(model nls1 cleaned)
glance(rr.huber)
glance(rr.bisquare)
glance(rr.hampel)
#Explore residual diagnostics
ols plot resid box
ols plot resid fit
ols plot resid hist
ols plot resid qq
```

#big outliers so choose to run a robust regression using MASS pkg: #from https://stats.idre.ucla.edu/r/dae/robust-regression/ #Robust regression is iterated re-weighted least squares (IRLS). The #command is rlm in the MASS package. There are several

train control kfold repeat)

```
#weighting functions that can be used for IRLS. We
#first use the Huber weights and then bi-square weighting. We
#will then look at the final weights created by the IRLS process.
#robust methods due to variance of resids and measures of influence:
#from https://stats.idre.ucla.edu/r/dae/robust-regression/
#We can see that the weight given to Mississippi
#is dramatically lower using the bisquare weighting
#function than the Huber weighting function and
#the parameter estimates from these two different
#weighting methods differ. When comparing the
#results of a regular OLS regression and a robust
#regression, if the results are very different,
#you will most likely want to use the results
#from the robust regression. Large differences
#suggest that the model parameters are being
#highly influenced by outliers. Different functions
#have advantages and drawbacks. Huber weights can
#have difficulties with severe outliers, and
#bisquare weights can have difficulties converging
#or may yield multiple solutions.
#(robust) sandwich variance estimator for linear regression:
#from:
https://thestatsgeek.com/2014/02/14/the-robust-sandwich-variance-estimator-for-linear-regression-using-r/
#This method allowed us to estimate valid standard errors
#for our coefficients in linear regression, without requiring
#the usual assumption that the residual errors have constant variance.
# 10 k-fold cross validation
# define training control
train control kfold <- trainControl(method="cv", number=10)
# fix the parameters of the algorithm
grid <- expand.grid(.fL=c(0), .usekernel=c(FALSE))
# train the model
model lm1 kfold <- train(BIOMASS ~ MEANH + SD + COVER,
                                                                                     data=ClocaenogField,
trControl=train control kfold)
# summarize results
print(model lm1 kfold)
View(model lm1 kfold)
summary(model lm1 kfold)
# repeated 10 k-fold cross validation
# define training control
train control kfold repeat <- trainControl(method="repeatedcv", number=10, repeats=3)
# train the model
model lm1 kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField, trControl =
```

```
model lm2 kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField cleaned,
trControl=train control kfold repeat)
model lm step full kfold repeat <- train(BIOMASS ~ MEANH + SD, data = ClocaenogField,
trControl=train control kfold repeat)
model nls1 kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control kfold repeat)
model nls1 cleaned kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data =
ClocaenogField cleaned, trControl=train control kfold repeat)
rr.huber kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control kfold repeat)
rr.bisquare kfold repeat <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control kfold repeat)
rr.hampel kfold repeat <-train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control kfold repeat)
print(model lm1 kfold repeat)
print(model lm2 kfold repeat)
print(model lm step full kfold repeat)
print(model nls1 kfold repeat)
print(model nls1 cleaned kfold repeat)
print(rr.huber kfold repeat)
print(rr.bisquare kfold repeat)
print(rr.hampel kfold repeat)
# Leave-one-out cross validation
# define training control
train control LOOVC <- trainControl(method="LOOCV")
# train the model
model lm1 LOOVC <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control LOOVC)
model lm2 LOOVC <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField cleaned,
trControl=train control LOOVC)
model lm step full LOOVC <- train(BIOMASS ~ MEANH + SD, data = ClocaenogField,
trControl=train control LOOVC)
model nls1 LOOVC <- train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control LOOVC)
model nls1 cleaned LOOVC <- train(BIOMASS ~ MEANH + SD + COVER,
ClocaenogField cleaned, trControl=train control LOOVC)
rr.huber LOOVC <- train(BIOMASS ~ MEANH + SD + COVER, data =
trControl=train control LOOVC)
rr.bisquare LOOVC <- train(BIOMASS ~ MEANH + SD + COVER, data =
                                                                            ClocaenogField,
trControl=train control LOOVC)
rr.hampel LOOVC <-train(BIOMASS ~ MEANH + SD + COVER, data = ClocaenogField,
trControl=train control LOOVC)
print(model lm1 LOOVC)
summary(model lm1 LOOVC)
View(model lm1 LOOVC)
print(model lm2 LOOVC)
print(model lm step full LOOVC)
```

```
print(model nls1 LOOVC)
print(model nls1 cleaned LOOVC)
print(rr.huber LOOVC)
print(rr.bisquare LOOVC)
print(rr.hampel LOOVC)
#run diagnostics
shapiro.test(resid(model lm1 kfold repeat))
shapiro.test(resid(model lm2 kfold repeat))
shapiro.test(resid(model lm step full kfold repeat))
shapiro.test(resid(model nls1 kfold repeat))
shapiro.test(resid(model nls1 cleaned kfold repeat))
shapiro.test(resid(rr.huber kfold repeat))
shapiro.test(resid(rr.bisquare kfold repeat))
shapiro.test(resid(rr.hampel kfold repeat))
bptest(model lm1 kfold repeat)
bptest(model lm2 kfold repeat)
bptest(model lm step full kfold repeat)
bptest(model nls1 kfold repeat)
bptest(model nls1 cleaned kfold repeat)
bptest(rr.huber kfold repeat)
bptest(rr.bisquare kfold repeat)
bptest(rr.hampel kfold repeat)
dwtest(rr.huber)
dwtest(rr.bisquare)
dwtest(rr.hampel)
outlierTest(lm(rr.huber))
outlierTest(lm(rr.bisquare))
outlierTest(lm(rr.hampel))
vif(rr.huber)
vif(rr.bisquare)
vif(rr.hampel)
AIC(model lm1)
AIC(model lm2)
AIC(model lm step full)
AIC(model nls1)
AIC(model nls1 cleaned)
AIC(rr.huber)
AIC(rr.bisquare)
AIC(rr.hampel)
BIC(model lm1)
BIC(model lm2)
BIC(model lm step full)
BIC(model nls1)
BIC(model nls1 cleaned)
```

```
BIC(rr.huber)
BIC(rr.bisquare)
BIC(rr.hampel)
TheilU(ClocaenogField$BIOMASS, model lm1, type=1)
theil.wtd(model_lm1, weights = NULL)
bias training <- rnorm(40, 2, sd = 0.5)
bias(bias_training, ClocaenogField)
bias(samp, pop)
bias(samp, pop, type = 'relative')
bias(samp, pop, type = 'standardized')
dev <- samp - pop
bias(dev)
nom.uncertainty(model lm1)
lambda3(model nls1)
CronbachAlpha(model lm1)
guttman(model_lm1, missing = "complete", standardize = FALSE)
ols plot response(model lm1)
u theil train <- createDataPartition(ClocaenogField$BIOMASS, p=0.5, list = FALSE)
u theil trainingData <- ClocaenogField[u theil train]
u theil testData <- historical.df[-train,]
dim(trainingData)
```