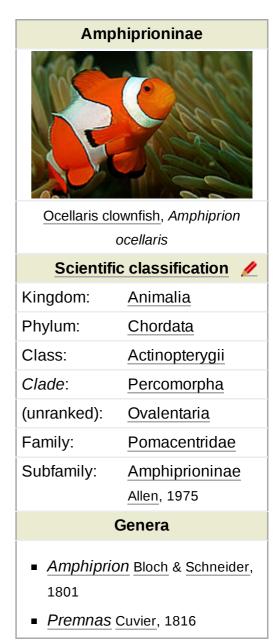
Amphiprioninae

Clownfish or **anemonefish** are <u>fishes</u> from the subfamily **Amphiprioninae** in the family <u>Pomacentridae</u>. Thirty <u>species</u> are recognized: one in the genus <u>Premnas</u>, while the remaining are in the genus <u>Amphiprion</u>. In the wild, they all form <u>symbiotic</u> <u>mutualisms</u> with <u>sea anemones</u>. Depending on species, anemonefish are overall yellow, orange, or a reddish or blackish color, and many show white bars or patches. The largest can reach a length of 17 cm (6.7 in), while the smallest barely achieve 7–8 cm (2.8–3.1 in).

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Distribution and habitats

Anemonefish are endemic to the warmer waters of the <u>Indian Ocean</u>, including the <u>Red Sea</u> and <u>Pacific Oceans</u>, the <u>Great Barrier Reef</u>, Southeast Asia, Japan, and the Indo-Malaysian region. While most species have restricted distributions, others are widespread. Anemonefish typically live at the bottom of shallow seas in sheltered reefs or in shallow <u>lagoons</u>. No anemonefish are found in the <u>Atlantic</u>. [1]

Diet

Anemonefish are <u>omnivorous</u> and can feed on undigested food from their host anemones, and the <u>fecal</u> matter from the anemonefish provides nutrients to the sea anemone. Anemonefish primarily feed on small zooplankton from the water column, such as copepods and tunicate larvae, with a small portion of their diet

coming from algae, with the exception of <u>Amphiprion perideration</u>, which primarily feeds on algae. They may also consume the tentacles of their host anemone. 4

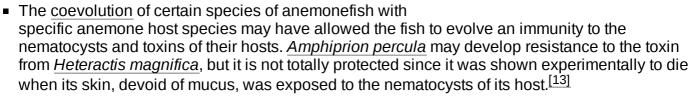
Symbiosis and mutualism

Anemonefish and sea anemones have a symbiotic, mutualistic relationship, each providing many benefits to the other. The individual species are generally highly host specific, and especially the genera Heteractis and Stichodactyla, and the species Entacmaea quadricolor are frequent anemonefish partners. The sea anemone protects the anemonefish from predators, as well as providing food through the scraps left from the anemone's meals and occasional dead anemone tentacles and functions as a safe nest site. In return, the anemonefish defends the anemone from its predators and parasites. [5][6] The anemone also picks up nutrients from the anemonefish's excrement. The nitrogen excreted from anemonefish increases the number of algae incorporated into the tissue of their hosts, which aids the anemone in tissue growth and regeneration. [3] The activity of the anemonefish results in greater water circulation around the sea anemone. [8] and it has been suggested that their bright coloring might lure small fish to the anemone, which then catches them. [9] Studies on anemonefish have found that they alter the flow of water around sea anemone tentacles by certain behaviors and movements such as "wedging" and "switching". Aeration of the host anemone tentacles allows for benefits to the metabolism of both partners, mainly by increasing anemone body size and both anemonefish and anemone respiration.[10]

Bleaching of the host anemone can occur when warm temperatures cause a reduction in algal symbionts within the anemone. Bleaching of the host can cause a short-term increase in the metabolic rate of resident anemonefish, probably as a result of acute stress. [11] Over time, however, there appears to be a down-regulation of metabolism and a reduced growth rate for fish associated with bleached anemones. These effects may stem from reduced food availability (e.g. anemone waste products, symbiotic algae) for the anemonefish. [12]

Several theories are given about how they can survive the sea anemone poison:

■ The mucus coating of the fish may be based on <u>sugars</u> rather than <u>proteins</u>. This would mean that anemones fail to recognize the fish as a potential food source and do not fire their <u>nematocysts</u>, or sting <u>organelles</u>.



Anemonefish are the best known example of fish that are able to live among the venomous sea anemone tentacles, but several others occur, including juvenile <u>threespot dascyllus</u>, certain <u>cardinalfish</u> (such as <u>Banggai</u> cardinalfish), incognito (or anemone) goby, and juvenile painted greenling. [14][15][16]



Ocellaris clownfish nestled in a magnificent sea anemone (Heteractis magnifica)



A pair of pink anemonefish (*Amphiprion perideraion*) in their anemone home



Clownfish swimming movements

Reproduction

In a group of anemonefish, a strict <u>dominance hierarchy</u> exists. The largest and most aggressive female is found at the top. Only two anemonefish, a male and a female, in a group reproduce – through <u>external fertilization</u>. Anemonefish are <u>sequential hermaphrodites</u>, meaning they develop into males first, and when they mature, they become females. If the female anemonefish is removed from the group, such as by death, one of the largest and most dominant males becomes a female. The remaining males move up a rank in the hierarchy.

Anemonefish lay eggs on any flat surface close to their host anemones. In the wild, anemonefish spawn around the time of the full moon. Depending on the species, they can lay hundreds or thousands of eggs. The male parent guards the eggs until they hatch about 6-10 days later, typically two hours after dusk. [17]

A clownfish swimming

Play media
Video of a cinnamon clownfish
swimming around an anemone

Parental investment

Anemonefish colonies usually consist of the reproductive male and female and a few male juveniles, which help tend the colony. [18] Although multiple males cohabit an environment with a single female, polygamy does not occur and only the adult pair exhibits reproductive

behavior. However, if the female dies, the social hierarchy shifts with the breeding male exhibiting protandrous sex reversal to become the breeding female. The largest juvenile then becomes the new breeding male after a period of rapid growth. The existence of protandry in anemone fish may rest on the case that nonbreeders modulate their phenotype in a way that causes breeders to tolerate them. This strategy prevents conflict by reducing competition between males for one female. For example, by purposefully modifying their growth rate to remain small and submissive, the juveniles in a colony present no threat to the fitness of the adult male, thereby protecting themselves from being evicted by the dominant fish. [20]

The reproductive cycle of anemonefish is often correlated with the lunar cycle. Rates of spawning for anemonefish peak around the first and third quarters of the moon. The timing of this <u>spawn</u> means that the eggs hatch around the full moon or new moon periods. One explanation for this lunar clock is that spring tides produce the highest tides during full or new moons. Nocturnal hatching during high tide may reduce predation by allowing for a greater capacity for escape. Namely, the stronger currents and greater water volume during high tide protect the hatchlings by effectively sweeping them to safety. Before spawning, anemonefish exhibit increased rates of anemone and substrate biting, which help prepare and clean the nest for the spawn. [19]

In terms of parental care, male anemonefish are often the caretakers of eggs. Before making the clutch, the parents often clear an oval-shaped clutch varying in diameter for the spawn. Fecundity, or reproductive rate, of the females, usually ranges from 600 to 1500 eggs depending on her size. In contrast to most animal species, the female-only occasionally takes responsibility for the eggs, with males expending most of the time and effort. Male anemonefish care for their eggs by fanning and guarding them for 6 to 10 days until they hatch. In general, eggs develop more rapidly in a clutch when males fan properly, and fanning represents a crucial mechanism of successfully developing eggs. This suggests that males can control the success of hatching an egg clutch by investing different amounts of time and energy towards the eggs. For example, a male could choose to fan less in times of scarcity or fan more in times of abundance. Furthermore, males display increased

alertness when guarding more valuable broods, or eggs in which paternity was guaranteed. Females, though, display generally less preference for parental behavior than males. All these suggest that males have increased parental investment towards the eggs compared to females. [21]

Taxonomy

Historically, anemonefish have been identified by morphological features and color pattern in the field, while in a laboratory, other features such as scalation of the head, tooth shape, and body proportions are used. [2] These features have been used to group species into six complexes, clownfish, tomato, skunk, clarkii, saddleback, and maroon. [22] As can be seen from the gallery, each of the fish in these complexes has a similar appearance. Genetic analysis has shown that these complexes are not monophyletic groups, particularly the 11 species in the *A. clarkii* group, where only *A. clarkii* and *A. tricintus* are in the same clade, with six species, *A. allardi A. bicinctus*, *A. chagosensis*, *A. chrosgaster*, *A. fuscocaudatus*, *A. latifasciatus*, and *A. omanensis* being in an Indian clade, *A. chrysopterus* having monospecific lineage, and *A. akindynos* in the Australian clade with *A. mccullochi*. [23] Other significant differences are that *A. latezonatus* also has monospecific lineage, and *A. nigripes* is in the Indian clade rather than with *A. akallopisos*, the skunk anemonefish. [24] *A. latezonatus* is more closely related to *A. percula* and *Premnas biaculeatus* than to the saddleback fish with which it was previously grouped. [25][24]

Obligate mutualism was thought to be the key innovation that allowed anemonefish to radiate rapidly, with rapid and convergent morphological changes correlated with the ecological niches offered by the host anemones. The complexity of mitochondrial DNA structure shown by genetic analysis of the Australian clade suggested evolutionary connectivity among samples of A. akindynos and A. mccullochi that the authors theorize was the result of historical <u>hybridization</u> and <u>introgression</u> in the evolutionary past. The two evolutionary groups had individuals of both species detected, thus the species lacked reciprocal monophyly. No shared haplotypes were found between species.

Phylogenetic relationships

Scientific name	Common name	Clade [23]	Complex	
Genus <i>Amphiprion</i> :[27]				
Amphiprion akallopisos	Skunk anemonefish	A. akallopisos	skunk	
A. akindynos	Barrier reef anemonefish	Australian	A. clarkii	
A. allardi	Allard's anemonefish	Indian	A. clarkii	
A. barberi	Barber's anemonefish	A. ephippium	A. ephippium	
A. bicinctus	Two-band anemonefish	Indian	A. clarkii	
A. chagosensis	Chagos anemonefish	Indian	A. clarkii	
A. chrysogaster	Mauritian anemonefish	Indian	A. clarkii	
A. chrysopterus	Orange-fin anemonefish	monospecific lineage	A. clarkii	
A. clarkii	Clark's anemonefish	A. clarkii	A. clarkii	
A. ephippium	Red saddleback anemonefish	A. ephippium	A. ephippium	
A. frenatus	Tomato anemonefish	A. ephippium	A. ephippium	
A. fuscocaudatus	Seychelles anemonefish	Indian ^[n 1]	clarkii	
A. latezonatus	Wide-band anemonefish	monospecific lineage	saddleback	
A. latifasciatus	Madagascar anemonefish	Indian	A. clarkii	
A. leucokranos	White-bonnet anemonefish	likely hybrid	skunk	
A. mccullochi	Whitesnout anemonefish	Australian	A. ephippium	
A. melanopus	Red and black anemonefish	A. ephippium	A. ephippium	
A. nigripes	Maldive anemonefish	Indian	skunk	
A. ocellaris	False clown anemonefish	percula	clownfish	
A. omanensis	Oman anemonefish	Indian	A. clarkii	
A. pacificus	Pacific anemonefish	A. akallopisos	skunk	
A. percula	Clown anemonefish	percula	clownfish	
A. perideraion	Pink skunk anemonefish	A. akallopisos	skunk	
A. polymnus	Saddleback anemonefish	A. polymnus	saddleback	
A. rubrocinctus	Australian anemonefish	A. ephippium	A. ephippium	
A. sandaracinos	Orange anemonefish	A. akallopisos	skunk	
A. sebae	Sebae anemonefish	A. polymnus	saddleback	
A. thiellei	Thielle's anemonefish	likely hybrid	skunk	
A. tricinctus	Three-band anemonefish	clarkii	clarkii	
Genus <i>Premnas</i> : ^[28]				
Premnas biaculeatus	Maroon anemonefish	percula	Maroon	

Morphological diversity by complex







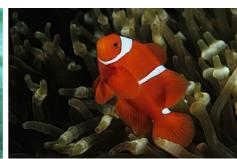
(clown percula anemonefish) in 'normal' orange and a melanistic blackish variant

clarkii a anemonefish)

(Clark's A. polymnus (saddleback clownfish) off Sulawesi, Indonesia







ephippium saddleback anemonefish)

anemonefish)

(red A. perideraion (pink skunk Male P. biaculeatus (maroon anemonefish) in Papua New Guinea

In the aquarium

Anemonefish make up 43% of the global marine ornamental trade, and 25% of the global trade comes from fish bred in captivity, while the majority is captured from the wild, [29][30] accounting for decreased densities in exploited areas. [31] Public aquaria and captive-breeding programs are essential to sustain their trade as marine ornamentals, and has recently become economically feasible. [32][33] It is one of a handful of marine ornamentals whose complete lifecycle has been in closed captivity. Members of some anemonefish species, such as the maroon clownfish, become aggressive in captivity; others, like the false percula clownfish, can be kept successfully with other individuals of the same species. [34]

When a sea anemone is not available in an aquarium, the anemonefish may settle in some varieties of soft corals, or large polyp stony corals.[35] Once an anemone or coral has been adopted, the anemonefish will defend it. Anemonefish, however, are not obligately tied to hosts, and can survive alone in captivity. [36][37]

In popular culture

In Disney/Pixar's 2003 film Finding Nemo and its 2016 sequel Finding Dory main characters Nemo and his dad Marlin are clownfish, probably the species A. ocellaris. [38] The popularity of anemonefish for aquaria increased following the film's release; it is the first film associated with an increase in the numbers of those captured in the wild. [39]

Notes

1. Exemplars of *A. fuscocaudatus* have never been sequenced. The authors hypothetically placed this species in the Indian clade because it is the most parsimonious solution regarding the biogeography of anemonefish species. [23]

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Further reading

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- Vargas-Abúndez, Arturo Jorge; Randazzo, Basilio; Foddai, Marco; Sanchini, Lorenzo; Truzzi, Cristina; Giorgini, Elisabetta; Gasco, Laura; Olivotto, Ike (January 2019). "Insect meal based diets for clownfish: Biometric, histological, spectroscopic, biochemical and molecular implications". *Aquaculture*. 498: 1–11. doi:10.1016/j.aquaculture.2018.08.018 (https://doi.org/10.1016%2Fj.aquaculture.2018.08.018). hdl:2318/1674109 (https://hdl.handle.net/2318%2F1674109).

External links

- Clownfish species profiles on AquariumDomain. (http://www.aquariumdomain.com/viewSpeciesList.php?species=Clownfish)
- (in German) Photo Gallery of *Amphiprion ocellaris* and their eggs (http://www.torsten-ernst.de/modules.php?name=Gallery&file=categories&cat_id=13)
- Monterey Bay Aquarium: Video and information (https://web.archive.org/web/20080724075222/ http://www.montereybayaquarium.org/efc/efc splash/splash animals clownfish.aspx)
- Clown Fish underwater photography gallery (https://web.archive.org/web/20100609112916/htt p://www.tommyschultz.com/component/searchimage/clown-fish-best/1.html)
- Aquaticcommunity.com (http://www.aquaticcommunity.com/clownfish/)
- Tolweb.org (http://tolweb.org/treehouses/?treehouse id=3390)

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