

# Diagnostic and therapy of severe *Clostridioides* difficile infections in the ICU

Benoit Guery<sup>a,b,c,f</sup>, Frédéric Barbut<sup>b,d,e,f</sup>, and Sarah Tschudin-Sutter<sup>f,g</sup>

### Purpose of review

The purpose of the review is to provide all the recent data focusing on the diagnostic and treatment of *Clostridioides difficile* infection in patients admitted in the ICU.

### **Recent findings**

In the ICU, diagnosis remains complicated with a large number of alternative diagnosis. The treatment classically relies on vancomycin but fidaxomicin and fecal microbiota transplantation are now potential solutions in selected indications.

#### Summary

Data on ICU-related CDI remain limited and conflicting. To date, there is no unique and simple way to obtain a diagnosis for CDI, the combination of clinical signs and a two-step testing algorithm remains the recommended gold-standard. Two molecules can be proposed for first line treatment: vancomycin and fidaxomicin. Although metronidazole may still be discussed as a treatment option for mild CDI in low-risk patients, its use for ICU-patients does not seem reasonable. Several reports suggest that fecal microbiota transplantation could be discussed, as it is well tolerated and associated with a high rate of clinical cure. CDI is a dynamic and active area of research with new diagnostic techniques, molecules, and management concepts likely changing our approach to this old disease in the near future.

#### **Keywords**

Clostridioides difficile, fecal microbiota transplantation, fidaxomicin, ICU, vancomycin

### INTRODUCTION

Over the last decade, *Clostridioides difficile* infection (CDI) has become one of the most important infections worldwide and the most common healthcare-associated infection in North America. CDI is associated with a wide spectrum of presentations ranging from mild infection to fulminant colitis. Prevention of recurrences is the key issue for CDI management, yet in ICU patients, clinical cure is the priority where, as for the most severe cases, the vital prognosis is at stake. Defining severity of CDI is challenging as several classification systems have been proposed, and severity provides the basis for the choice of treatment. In this review, we will focus on the epidemiology, the diagnosis, and the treatment of CDI in the ICU.

# EPIDEMIOLOGY OF CLOSTRIDIOIDES DIFFICILE IN THE ICU

CDI effects approximately 14.7/10000 persons in the United States [1] and a growing number of patients in Europe (4.1/10000 persons in 2008

and 7/10 000 persons in 2012–2013) [2,3]. In line, numbers of CDI are increasing in Asia and have, for instance been estimated at 7.4/10 000 persons in Japan [4]. In a recently published systematic review on the global burden of CDI, including 229 publications with data from 41 countries, the overall rate of healthcare facility-associated CDI was estimated at 2.24 [95% confidence interval (CI) 1.66–3.03] per

<sup>a</sup>Infectious Diseases Service, Department of Medicine, University Hospital and University of Lausanne, Lausanne, Switzerland, <sup>b</sup>French Group of Faecal Microbiota Transplantation (GFTF), <sup>c</sup>European Study Group on Host and Microbiota Interactions (ESGHAMI), <sup>d</sup>National Reference Laboratory for *Clostridium difficile*, <sup>e</sup>INSERM, U1139, Faculté de Pharmacie de Paris, Université Paris Descartes, UMR-S1139, Sorbonne Paris Cité, Paris, France, <sup>f</sup>ESCMID Study Group for *Clostridioides difficile* (ESGCD) and <sup>g</sup>Division of Infectious Diseases and Hospital Epidemiology and Department of Clinical Research, University Hospital Basel and University of Basel, Switzerland

Correspondence to Benoit Guery, Infectious Diseases Service, Department of Medicine, University Hospital and University of Lausanne, Rue du Bugnon 46, 1011 Lausanne, Switzerland. Tel: +41 21 314 1643; e-mail: benoit.guery@chuv.ch

**Curr Opin Crit Care** 2020, 26:450–458 DOI:10.1097/MCC.00000000000000753

### **KEY POINTS**

- No single currently available commercial test can serve as a stand-alone test for diagnosing CDI.
- To optimize the diagnosis of CDI, two-step algorithms are currently recommended.
- Metronidazole is not an option in the ICU but can be administered intravenously associated to vancomycin in fulminant episodes.
- FMT could be a reasonable option to treat severecomplicated CDI but further studies are needed.

1000 admission annually and 3.54 (95% CI 3.19-3.92) per 10 000 patient-days annually [5] with rates being generally higher in North America and among the elderly. Estimated rates for CDI with onset on the ICU were notably higher at 11.08 per 1000 admissions annually (95% CI 7.19–17.08). On the basis of a systematic review and meta-analysis including 80835 ICU patients from 22 studies, CDI may affect 2% (95% CI 1-2%) of all ICU patients and C. difficile was identified as the causative pathogen in 11% (95% CI 6–17%) of patients presenting with diarrhea [6]. Pooled prevalence was highest in North America (2%, 95% CI 1-4%), followed by Asia (3%, 95% CI 1–2%), and Europe (1%, 95% CI 1–2%). In conclusion, the literature points to a higher burden of CDI in ICU patients as compared with general cohorts of hospitalized patients. Data on mortality of CDI in ICU patients is conflicting, with some studies pointing to an increased risk of death and others rejecting it [7–9]. Yet, in a metaanalysis, overall pooled hospital mortality of patients with CDI differed significantly from mortality of ICU patients without CDI (32%, 95% CI 26– 39% as compared with 24%, 95% CI 14–36%) and was not explained by differences in the morbidity score (Acute Physiology and Chronic Health Evaluation II) on ICU admission, suggesting significant attributable mortality [6]. Conflicting data on attributable mortality may be related to varying distributions of different ribotypes of C. difficile [10]. On the basis of reports of a greater potential for toxin production and multidrug resistance profiles [11,12], most of the evidence regarding increased virulence of specific ribotypes derives from settings in which such strains emerged rapidly, often resulting in outbreaks. In endemic settings, the association of disease severity has not been confirmed [13].

ICU patients are at increased risk for developing CDI because of an accumulation of predisposing factors [14] and most importantly, antibiotics [15], with up to 60% of ICU patients having

concomitant infections [16]. Furthermore, colonization with toxigenic C. difficile on admission has been identified as an independent predictor of CDI in ICU patients [17]. This later association possibly points to the increasing importance of communityrelated, rather than healthcare-related acquisition of C. difficile [18]. Incidence of community-acquired CDI has almost doubled over the last decade [19] and has been estimated to account for 14.5% of all confirmed CDI cases among 5260 admitted to the ICU, the remainder being hospital-acquired (17.4%) or ICU-acquired [9]. Relating thereto, length of hospital and ICU stay have been associated with an increased risk for developing CDI [6]. Although surveillance studies commonly report categorization of CDI in respect to the onset and association of disease (i.e. community vs. healthcare) based on standardized criteria, such categorizations are of limited value to investigate sources of C. difficile acquisition. Both loss of colonization resistance and nosocomial transmission by direct or indirect contact or selection pressure exerted by prolonged administration of antimicrobials in the setting of community-acquired colonization may result in healthcare-associated CDI [20]. Thus, monitoring hospital-onset CDI cases followed by typing of strains in the context of the occurrence of clusters with an epidemiological link is needed to detect outbreaks and identifying gaps in infection prevention and control measures [21]. An ever-increasing body of evidence, however, supports antibiotic stewardship measures being the most important strategy to reduce CDI rates, impacting both loss of colonization resistance and selection pressure [22,23].

# **CLOSTRIDIOIDES DIFFICILE INFECTION DIAGNOSIS**

The diagnosis of *C. difficile* infection (CDI) has gained more attention since the emergence and spread of hypervirulent strains, such as ribotype 027, which have contributed to the increased morbidity and mortality of infection worldwide. Nevertheless, in 2012, the multicenter EUCLID study showed that 23% of CDI were still not diagnosed in Europe because of an absence of clinical suspicion [2].

According to European and North American guidelines, a CDI case is defined by the following criteria [24,25\*]:

(1) Clinical signs of infection (diarrhea, ileus, toxic megacolon) and a stool test positive for toxins or toxigenic *C. difficile*, without evidence of another cause of diarrhea

(2) Presence of pseudomembranes during rectosigmoid or colonoscopic examination or histopathologic findings revealing pseudomembranous colitis.

The endoscopic diagnosis of CDI is usually specific (although other pathogens can be sometimes implicated in pseudomembranous colitis [26]) but not very sensitive. A negative result does not exclude CDI as pseudomembranes can be absent at the early stage of the disease or during mild forms of antibiotic-associated diarrhea. Most of the time, the CDI diagnosis relies on laboratory tests.

Many tests detecting different targets are currently available for the diagnosis of CDI [27\*]. They usually fall into two major categories: those that detect free toxins (toxin A and/or toxin B) in fecal samples [enzyme immunoassays (EIAs) and stool cytotoxicity test (CTA)] and those that detect the organism [toxigenic culture, glutamate dehydrogenase (GDH), and nucleic acid amplification tests (NAATs)].

CTA is a reference method for detecting free toxins in the stools. This method consists in inoculating a stool filtrate on a cell culture and observing a specific cytopathic effect (CPE). CPE is characterized by cell rounding and results from the disruption of the actin cytoskeleton by toxins. Although this technique is highly sensitive (in the order of the toxin picogram), it has been abandoned progressively due the long turn-around time to get the result and the need of having cell culture facilities.

EIA tests include microwell immuno-enzymatic (ELISA) and immunochromatographic techniques. They detect both toxins A and B, with or without differentiation. They are easy-to-use and more rapid but their sensitivity ranges from 29 to 86% compared with CTA, which prevents their use as a standalone method for CDI diagnosis [24,25]. More recently, ultrasensitive assays detecting free toxins A and B has been developed by several companies using the single molecule array technology (SIMOA). The SIMOA tests have a higher sensitivity than currently available toxin EIA and have the potential in the future to improve and simplify the CDI diagnosis [28–30].

NAAT are based on real-time PCR, isothermal loop amplification or microarray technologies. They detect a wide variety of targets and have the advantage of being very fast and sensitive (average sensitivity of 96% compared with toxigenic culture) [31]. However, the presence of a toxigenic strain does not imply that the patient is infected as approximately 3% of the general population are asymptomatic carriers of a *C. difficile* toxigenic strain.

Toxigenic culture is a two-step method where *C*. *difficile* strains are first isolated on a selective

medium and then the isolates are tested for their ability to produce toxins *in vitro*. Although the time to obtain a result is too long for routine diagnosis (2 days or more), Toxigenic culture is very sensitive, and isolation of the strain is essential for determining its susceptibility to antimicrobials. This method is generally used as a gold standard when evaluating new NAAT assays.

GDH is a metabolic enzyme produced by all *C. difficile* strains (including toxigenic or nontoxigenic strains). It can be detected by ELISA or immunochromatographic tests. These tests display an excellent negative predictive value (NPV) [32] so that a negative result can rule out the diagnosis of CDI. However, a positive test must be confirmed by a more specific test detecting free toxins in stools.

No single currently available commercial test can serve as a stand-alone test for diagnosing CDI. Indeed, EIA methods detecting free toxins in stools are very specific but often lack sensitivity, which can lead to an underdiagnosis of CDI. Conversely, tests, such as NAAT and toxigenic culture, detecting the presence of toxigenic strains (irrespective of the presence of free toxins in stools) are more sensitive but less specific, which can lead to an overdiagnosis of CDI. Therefore, to optimize the diagnosis of CDI, two-step algorithms are currently recommended [24,25]. The first test must display a high NPV to reliably exclude patients without CDI. If positive, a second test with a high positive-predictive value (PPV) should be used. If positive, the patient is considered as truly infected. Feces samples without free toxins but with positive NAAT results need clinical evaluation to discern CDI from asymptomatic carriage.

Recent publications suggest that the bacterial load of *C. difficile* (evaluated by the cycle threshold of NAAT as a surrogate marker) is generally higher in patients with CDI compared with carriers and might serve as predictor of toxin presence [33,34].

To optimize the diagnosis of CDI, some general rules should be implemented to reduce inappropriate testing. First, it is important to take stool samples prior to initiation of CDI treatment. Any empirical treatment for CDI can lead to false negative test results [35]. Second, a test-of-cure should not be carried out at the end of the treatment. Actually, 30–40% of patients who are considered clinically cured are still positive for C. difficile toxins or culture at the end of the treatment [36]. Third, many guidelines recommend to implement criteria for stool selection to avoid detection of asymptomatic carriers. Only diarrheic stools (defined as stool taking the shape of the container) should be accepted by the laboratory. According to the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA) guidelines, if clinical and laboratory personnel agree to not submit stool specimens of patients receiving laxatives and to submit only stool samples from patients with an unexplained clinical diarrhea ( $\geq 3$  unformed stools in 24 h), then NAAT can be used as a stand-alone test.

### **DEFINITION OF SEVERITY**

The definition of severity of CDI is not yet consensual varying among the scientific societies [25,37,38] and clinical trials [39–41]. As treatment decisions are based on severity, these definitions are of great importance. A 3-year cohort study performed on 233 CDI patients showed that the frequency of severe CDI ranged from 11.6 to 59.2% depending on the definition [42]. In the latest IDSA/ SHEA guidelines, only leucocyte number and renal function define severity [25<sup>\*</sup>], the previous article showed that serum albumin could also be an important factor to define severity, although this factor has been shown to be independently linked to mortality in septic patients without CDI [43]. Finally, the characteristics of the patient are also important to correctly assess the severity, for example, we have shown that for hematological patients, IDSA/SHEAcriteria could underestimate the severity and the potential risk of unfavorable outcome [44]. Using leucocytes and renal definitions, as recommended by the IDSA/SHEA [25<sup>\*</sup>], seems a reasonable approach combining measures easy to obtain in all patients and supported by a large number of studies. In the next section of this manuscript, we will therefore consider severity according to this definition.

# TREATMENT OF CLOSTRIDIOIDES DIFFICILE INFECTION IN THE ICU

Treatment of CDI includes two distinct profiles of patients, CDI developing in the ICU and CDI requiring ICU admission. Although the entire spectrum of disease manifestations can be present in CDI developing during an ICU stay, severe and complicated forms lead to ICU admission. In 2018, the IDSA and SHEA provided an update of the clinical practice guidelines for CDI [25\*]. Episodes are classified as nonsevere, severe (defined by leukocytosis or serum creatinine), or fulminant (hypotension, shock, ileus, megacolon).

### ORAL ANTIBIOTIC TREATMENT IN THE ICU

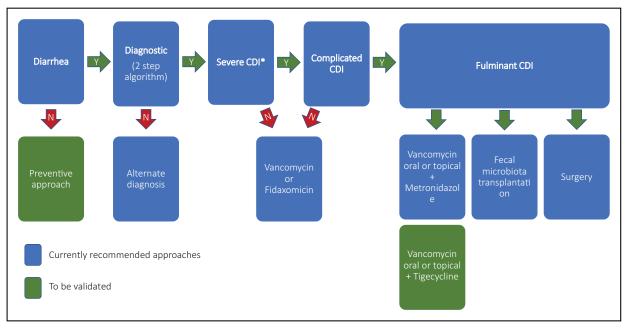
### Mild-to-severe episode

According to the IDSA/SHEA guidelines, initial nonsevere episodes can be treated with vancomycin or fidaxomicin; metronidazole is an alternate if these two agents are not available [25]. Metronidazole is not an option for severe episodes but can be administered intravenously associated to vancomycin in fulminant episodes. To shortly address the issue of vancomycin and metronidazole in the ICU, a limited number of studies are available. A meta-analysis of randomized controlled trials extracted only five studies, in the subgroup analysis on severity, the clinical effects of vancomycin were higher than in metronidazole in severe cases [45]. Interestingly, these data are related to only two of the five considered trials [39,46]. A retrospective, propensitymatched cohort study evaluated a total of 47 471 patients treated by vancomycin or metronidazole [47]. In the subcohort of 3130 patients with severe disease, vancomycin significantly reduced the risk of all-cause 30-day mortality.

Fidaxomicin was introduced for treatment of CDI and compared with vancomycin with two pivotal publications [40,41]. In both of these multicentric, double-blind, randomized noninferiority trials, no difference was observed in the subgroup of severe patients analyzing clinical cure as well as sustained clinical response, suggesting this molecule could be an alternative in severe patients. In proven CDI, a study compared 20 ICU patients with 30 patients treated in medical wards and showed that the response to fidaxomicin was comparable between these two groups [48]. A retrospective multicentric cohort study with propensity scorematched analysis in severe CDI compared 213 patients treated with fidaxomicin to 639 patients treated with oral vancomycin [49]. No difference was found on the primary outcome, which combined clinical failure and recurrence. A comparative efficacy of treatments analysis was performed using a network meta-analysis, including 24 trials with a total of 5361 patients [50]. In severe infections, vancomycin and fidaxomicin resulted in comparable outcomes. Thus, it seems reasonable to propose either vancomycin or fidaxomicin as first line drugs for ICU patients (Fig. 1).

### **Complicated episode**

The IDSA/SHEA guidelines recommend the administration of topical vancomycin with intravenous metronidazole. The evidence supporting this recommendation is not very high. Experimental data failed to show a difference [51], and a systematic review and meta-analysis also failed to show superiority of the combination treatment, but, because of the low number of studies performed, the combination treatment was not tested in each severity group [52]. A single-centre retrospective observational study compared critically ill patients receiving oral



**FIGURE 1.** Algorithm for the treatment and prevention of *Clostridioides difficile* in the ICU. Y, yes; N, no. \*Severe *Clostridioides difficile* (CDI) is defined according the 2018 Infectious Diseases Society of America (IDSA)/Society for Healthcare Epidemiology of America (SHEA) guideline [25\*].

vancomycin to patients receiving oral vancomycin combined with intravenous metronidazole [53]. Eighty-eight patients were included, mortality was 36.4 and 15.9% in the monotherapy and combination therapy groups, respectively (P = 0.03). Even though the patients were not clearly classified according to the IDSA/SHEA guidelines into severity groups, this study suggests that a combined treatment is associated with a better outcome in ICU patients. Moreover, although renal function was comparable between the two groups, leucocyte counts were significantly higher in the combination group (13.8 vs. 20.2 cells/ $\mu$ l, P = 0.004), suggesting more severe CDI, and thus reinforcing better outcome for combined treatment. We do not have currently any data on the use of fidaxomicin in complicated forms, and this drug is not approved in this indication. Intravenous immunoglobulin has also been proposed in CDI but the clinical data remain very limited [54–56]

# INTRACOLONIC ANTIBIOTIC TREATMENT IN THE ICU

In complicated forms of CDI, including the presence of ileus or megacolon, where oral drug administration is not possible, administration of adjunctive intracolonic vancomycin has largely been proposed [25\*] to be delivered by an endoscopic procedure [57]. A retrospective chart review of 696 patients showed that the group with intracolonic vancomycin had

higher rates of toxic megacolon, ICU admission and colectomy but maintained a similar mortality suggesting a potential protecting effect of locally administrated vancomycin [58]. A single-centre retrospective study included 47 patients treated with intracolonic vancomycin [59]. Overall, 70% (33/47) of the patients responded to adjunct of intracolonic vancomycin with complete resolution of symptoms without surgery. Another retrospective case-control study identified 24 ICU patients with adjunctive per rectum vancomycin [60] found different results and showed that this approach neither reduced the need for colectomy nor mortality. Overall, the level of evidence remains low and well conducted trials are still needed to clearly evaluate the potential advantage of locally delivering vancomycin in this subgroup of patients.

### ALTERNATE ANTIBIOTIC TREATMENT IN THE ICU

Tigecycline is broad spectrum protein synthesis inhibitor with a strong antimicrobial activity against *C. difficile* [61]. Several case report or small series have reported a potential effect of tigecycline in addition to the therapeutic regimen in severe and complicated CDI [62–66]. On a larger population, a single-centre retrospective cohort study compared 62 patients who received both tigecycline and vancomycin to 204 patients receiving only vancomycin [67]. In the propensity score-based analysis, the odds

ratio of favorable outcome was 0.92 (95% CI 0.60–1.44; P = 0.74) leading to the conclusion that adding tigecycline to standard therapy did not change rates of clinical cure. A well designed protocol is still needed to conclude on the potential effect of this molecule in severe and/or complicated forms of CDI but there is no strong rationale to propose this molecule as first-line treatment in complicated forms (Fig. 1).

# FECAL MICROBIOTA TRANSPLANTATION IN THE ICU

FMT has been largely proposed by international guidelines [25,37] for treatment of recurrent CDI following the pivotal trial of Van Nood et al. [68]. In the ICU, several reports suggest that FMT can also be a treatment option [69–72], even for severe-complicated CDI including toxic megacolon [73,74], or CDI in immunosuppressed patients [75]. A small series of 29 patients with severe and severe/complicated CDI showed that FMT reached an overall treatment response in, respectively, 100% (10/10) and 89% (17/19) for each group [76]. In the longterm, a multicenter follow-up study analyzed 17 patients with severe and/or complicated forms treated with FMT [77]. FMT was well tolerated in this cohort with a primary cure rate of 88.2%. A cohort of 57 patients with severe and severe complicated CDI was treated with FMT, 91% (52/57) experienced clinical cure at 1 month with 100% for severe (n=19) and 87% for severe complicated (n=33) [78]. There was no serious adverse event. These data suggest that FMT could be a reasonable option to treat severe-complicated CDI (Fig. 1).

### **SURGERY**

Colon-sparing diverting ileostomy with colonic lavage has been proposed as an alternative to colectomy in the treatment of severe complicated CDI [79]. In this study with a historical control group that underwent colectomy, 42 patients were treated using this surgical approach. This strategy resulted in reduced mortality compared with the historical population (19 vs. 50%) and preservation of the colon was achieved in 39/42 patients (93%). Interestingly, a retrospective review of patients treated surgically for severe complicated CDI compared 30day mortality, 1-year mortality, and colon preservation in 10 patients with loop ileostomy and colonic lavage vs. 13 patients with total abdominal colectomy [80]. There was no difference in mortality in both time points, but loop ileostomy allowed preservation of the colon in all six surviving patients

### PREVENTION OF CLOSTRIDIOIDES DIFFICILE INFECTION IN THE ICU

The ICU is associated with the exposition to large numbers of risks factors for CDI and it could be interesting to test a preventive approach for these patients exposed to large spectrum antibiotics, proton pump inhibitors, and other potential inductors of dysbiosis.

### PASSIVE AND ACTIVE IMMUNIZATION

- (1) Passive immunization: a randomized, double-blind, placebo-controlled study evaluated two monoclonal antibodies targeting *C. difficile* toxin A and B in 200 patients initially treated with metronidazole or vancomycin [81]. The rate of recurrence significantly decreased from 25 to 7% for patients treated with the antibodies. A phase 3 trial, involving 2655 patients with primary or recurrent CDI, showed that bezlotoxumab reduced the recurrence of infection from 28 to 17% [82].
- (2) Active immunization: three vaccines are currently in phase 2 and 3 clinical trials. A formalin-inactivated toxoid-based vaccine is currently the most advanced [83,84]. Several phase 3 trials have been initiated (NCT00772343, NCT03090191). VLA84 is a recombinant fusion protein with epitopes of toxin A and B [85]. A DNA vaccine [86,87] has been recently proposed with interesting results in animal models.

Although these preventive approaches are quite attractive, a cost–benefit analysis has to be performed for Bezlotoxumab in the ICU, and for vaccines, data on target populations benefiting most are lacking. Immunotherapy has also been proposed but remains at the experimental level [88].

#### **DRUGS**

Rifaximin and other microbiota-based drugs have been proposed to prevent CDI [89–91] but there is no data available for ICU patients.

### CONCLUSION

CDI in the ICU remains challenging at all levels, diarrhea is frequently encountered and can be multifactorial, CDI being one of the most important potential causes. The laboratory diagnosis does not rely on one test but must be combined with clinical data to optimize both positive and negative predictive values of the test performed. This approach remains the gold standard for diagnosis. In ICU patients, the use of metronidazole questionable

and both vancomycin and fidaxomicin should be proposed for first-line treatment. For severe-complicated forms, the combination of topical vancomycin and systemic metronidazole is still proposed, with many reports suggesting that FMT as an excellent, well tolerated, and efficient treatment option, potentially reducing the need for surgery. There is no data to promote an optimal preventive approach (i.e. immunization or preventive treatment) in the ICU. The figure proposes an algorithm for the management of CDI in the ICU.

### Acknowledgements

None.

### Financial support and sponsorship

None.

### **Conflicts of interest**

B.G. is a member of MSD Advisory Boards for C. difficile, of a Pfizer Antiinfectives Advisory Board. F.B. received Grants from Astellas, Anios, MSD, Biomérieux, Quidel, Cubist, Biosynex, GenePoc, Personal fees from Astellas, Pfizer, MSD, Danone, and Nonfinancial support from Astellas, Pfizer, Anios, MSD. S.T.S. is a member of the Astellas and MSD Advisory Boards for C. difficile, of the Pfizer Antiinfectives Advisory Board and the Menarini Scientific Advisory Board. She reports grants from the Swiss National Science Foundation NRP72 (407240\_167060), the Gottfried und Julia Bangerter-Rhyner Stiftung, the Fonds zur Förderung von Lehre und Forschung der Freiwilligen Akademischen Gesellschaft Basel, and the Jubiläumsstiftung from Swiss Life.

### REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- ■■ of outstanding interest
- Lessa FC, Mu Y, Bamberg WM, et al. Burden of Clostridium difficile infection in the United States. New Engl J Med 2015; 372:825-834.
- Davies KA, Longshaw CM, Davis GL, et al. Underdiagnosis of Clostridium difficile across Europe: the European, multicentre, prospective, biannual, point-prevalence study of Clostridium difficile infection in hospitalised patients with diarrhoea (EUCLID). Lancet Infect Dis 2014; 14: 1208-1219.
- Bauer MP, Notermans DW, van Benthem BH, et al. Clostridium difficile infection in Europe: a hospital-based survey. Lancet 2011; 377:63-73.
- Kato H, Senoh M, Honda H, et al. Clostridioides (Clostridium) difficile infection burden in Japan: a multicenter prospective study. Anaerobe 2019; 60:102011.
- Balsells E, Shi T, Leese C, et al. Global burden of Clostridium difficile infections: a systematic review and meta-analysis. J Glob Health 2018; 9:010407.
- Karanika S, Paudel S, Zervou FN, et al. Prevalence and clinical outcomes of Clostridium difficile infection in the intensive care unit: a systematic review and meta-analysis. Open Forum Infect Dis 2015; 3:ofv186.
- Dodek PM, Norena M, Ayas NT, et al. Length of stay and mortality due to Clostridium difficile infection acquired in the intensive care unit. J Crit Care 2013; 28: 335–340.

- Bouza E, Rodriguez-Creixems M, Alcalá L, et al. Is Clostridium difficile infection an increasingly common severe disease in adult intensive care units? A 10-year experience. J Crit Care 2015; 30:543-549.
- Zahar J-R, Schwebel C, Adrie C, et al., OUTCOMEREA study group. Outcome of ICU patients with Clostridium difficile infection. Crit Care 2012; 16:R215.
- Loo VG, Poirier L, Miller MA, et al. A predominantly clonal multiinstitutional outbreak of Clostridium difficile-associated diarrhea with high morbidity and mortality. New Engl J Med 2005; 353:2442–2449.
- Warny M, Pepin J, Fang A, et al. Toxin production by an emerging strain of Clostridium difficile associated with outbreaks of severe disease in North America and Europe. Lancet 2005; 366:1079–1084.
- 12. He M, Miyajima F, Roberts P, et al. Emergence and global spread of epidemic healthcare-associated *Clostridium difficile*. Nat Genet 2012; 45:109–113.
- Tschudin-Sutter S. Molecular epidemiology of Clostridium difficile for clinical practice. Swiss Med Wkly 2014; 144:w13995.
- Cao F, Chen CX, Wang M, et al. Updated meta-analysis of controlled observational studies: proton-pump inhibitors and risk of Clostridium difficile infection. J Hosp Infect 2017; 98:4-13.
- Antonelli M, Martin-Loeches I, Dimopoulos G, et al. Clostridioides difficile (formerly Clostridium difficile) infection in the critically ill: an expert statement. Intens Care Med 2020; 46:215–224.
- Marra AR, Edmond MB, Wenzel RP, Bearman GM. Hospital-acquired Clostridium difficile-associateddisease in the intensive care unit setting: epidemiology, clinical course and outcome. BMC Infect Dis 2007; 7:42.
- Tschudin-Sutter S, Carroll KC, Tamma PD, et al. Impact of toxigenic Clostridium difficile colonization on the risk of subsequent C. difficile infection in intensive care unit patients. Infect Control Hosp Epidemiol 2015; 36:1324-1329.
- Turner NA, Smith BA, Lewis SS. Novel and emerging sources of Clostridioides difficile infection. Plos Pathog 2019; 15:e1008125.
- Ofori E, Ramai D, Dhawan M, et al. Community-acquired Clostridium difficile: epidemiology, ribotype, risk factors, hospital and intensive care unit outcomes, and current and emerging therapies. J Hosp Infect 2018; 99:436–442.
- Freedberg DE, Zhou MJ, Cohen ME, et al. Pathogen colonization of the gastrointestinal microbiome at intensive care unit admission and risk for subsequent death or infection. Intens Care Med 2018; 44:1203-1211.
- Li C, Li Y, Huai Y, et al. Incidence and outbreak of healthcare-onset healthcare-associated Clostridioides difficile infections among intensive care patients in a large teaching hospital in China. Front Microbiol 2018; 9:566.
- 22. Lawes T, Lopez-Lozano J-M, Nebot CA, et al. Effects of national antibiotic stewardship and infection control strategies on hospital-associated and community-associated meticillin-resistant Staphylococcus aureus infections across a region of Scotland: a nonlinear time-series study. Lancet Infect Dis 2015; 15:1438–1449.
- Dingle KE, Didelot X, Quan TP, et al. Effects of control interventions on Clostridium difficile infection in England: an observational study. Lancet Infect Dis. 2017; 17:411–421.
- Crobach MJT, Planche T, Eckert C, et al. European Society of Clinical Microbiology and Infectious Diseases: update of the diagnostic guidance document for Clostridium difficile infection. Clin Microbiol Infec 2016; 22(Suppl 4):S63-S81.
- 25. McDonald LC, Gerding DN, Johnson S, et al. Clinical practice guidelines for Clostridium difficile infection in adults and children: 2017 Update by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). Clin Infect Dis 2018; 66:e1 e48.

Current recommendations for treatment of CDI. Extensive analysis of the literature.

- **26.** Krutova M, Wilcox MH, Kuijper EJ. The pitfalls of laboratory diagnostics of *Clostridium difficile* infection. Clin Microbiol Infect 2018; 24:682–683.
- 27. Gateau C, Couturier J, Coia J, Barbut F. How to: diagnose infection caused by
   Clostridium difficile. Clin Microbiol Infec 2018; 24:463468.

Very nice review of the pitfalls of diagnosis and the good practice to obtain a

- Banz A, Lantz A, Riou B, et al. Sensitivity of single-molecule array assays for detection of Clostridium difficile toxins in comparison to conventional laboratory testing algorithms. J Clin Microbiol 2018; 56:e00452–18.
- Pollock NR, Banz A, Chen X, et al. Comparison of Clostridioides difficile stool toxin concentrations in adults with symptomatic infection and asymptomatic carriage using an ultrasensitive quantitative immunoassay. Clin Infect Dis 2019; 68:78 – 86.
- 30. Sandlund J, Bartolome A, Almazan A, et al. Ultrasensitive detection of Clostridioides difficile toxins A and B by use of automated single-molecule counting technology. J Clin Microbiol 2018; 56:e00908-18.
- O'Horo JC, Jones A, Sternke M, et al. Molecular techniques for diagnosis of Clostridium difficile infection: systematic review and meta-analysis. Mayo Clin Proc 2012; 87:643–651.
- Shetty N, Wren MWD, Coen PG. The role of glutamate dehydrogenase for the detection of Clostridium difficile in faecal samples: a meta-analysis. J Hosp Infect 2011; 77:1–6.
- Crobach MJT, Duszenko N, Terveer EM, et al. Nucleic acid amplification test quantitation as predictor of toxin presence in Clostridium difficile infection. J Clin Microbiol 2018; 56:95.

- 34. Davies KA, Planche T, Wilcox MH. The predictive value of quantitative nucleic acid amplification detection of Clostridium difficile toxin gene for faecal sample toxin status and patient outcome. Plos One 2018; 13:e0205941.
- Sunkesula VCK, Kundrapu S, Muganda C, et al. Does empirical Clostridium difficile Infection (CDI) therapy result in false-negative CDI diagnostic test results? Clin Infect Dis 2013; 57:494–500.
- Sethi AK, Al-Nassir WN, Nerandzic MM, et al. Persistence of skin contamination and environmental shedding of Clostridium difficile during and after treatment of C. difficile infection. Infect Control Hosp Epidemiol 2010; 31:21-27.
- Debast SB, Bauer MP, Kuijper EJ; Committee. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for Clostridium difficile infection. Clin Microbiol Infec 2014; 20(Suppl 2):1–26.
- Surawicz CM, Brandt LJ, Binion DG, et al. Guidelines for diagnosis, treatment, and prevention of Clostridium difficile infections. Am J Gastroenterology 2013; 108:478-498.
- Zar FA, Bakkanagari SR, Moorthi KMLST, Davis MB. A comparison of vancomycin and metronidazole for the treatment of Clostridium difficileassociated diarrhea, stratified by disease severity. Clin Infect Dis 2007; 45:302-307.
- Cornely OA, Crook DW, Esposito R, et al. Fidaxomicin versus vancomycin for infection with Clostridium difficile in Europe, Canada, and the USA: a doubleblind, noninferiority, randomised controlled trial. Lancet Infect Dis 2012; 12:281 – 289.
- Louie TJ, Miller MA, Mullane KM, et al., OPT-80-003 Clinical Study Group. Fidaxomicin versus vancomycin for Clostridium difficile infection. New Engl J Med 2011; 364:422–431.
- Khanafer N, Barbut F, Eckert C, et al. Factors predictive of severe Clostridium difficile infection depend on the definition used. Anaerobe 2015; 37:43–48.
- 43. Furukawa M, Kinoshita K, Yamaguchi J, et al. Sepsis patients with complication of hypoglycemia and hypoalbuminemia are an early and easy identification of high mortality risk. Intern Emerg Med 2019; 14:539–548.
- **44.** Robin C, Héquette-Ruz R, Guery B, *et al.* Treating *Clostridium difficile* infection in patients presenting with hematological malignancies: are current guidelines applicable? Médecine Et Maladies Infect 2017; 47:532–539.
- Igarashi Y, Tashiro S, Enoki Y, et al. Oral vancomycin versus metronidazole for the treatment of Clostridioides difficile infection: meta-analysis of randomized controlled trials. J Infect Chemother 2018; 24:907–914.
- Johnson S, Louie TJ, Gerding DN, et al., Polymer Alternative for CDI Treatment (PACT) investigators. Vancomycin, metronidazole, or tolevamer for Clostridium difficile infection: results from two multinational, randomized, controlled trials. Clin Infect Dis 2014; 59:345–354.
- Stevens VW, Nelson RE, Schwab-Daugherty EM, et al. Comparative effectiveness of vancomycin and metronidazole for the prevention of recurrence and death in patients with Clostridium difficile infection. JAMA Intern Med 2017; 177:546–548.
- Penziner S, Dubrovskaya Y, Press R, Safdar A. Fidaxomicin therapy in critically ill patients with Clostridium difficile infection. Antimicrob Agents Chemother 2015; 59:1776–1781.
- Gentry CA, Nguyen PK, Thind S, et al. Fidaxomicin versus oral vancomycin for severe Clostridium difficile infection: a retrospective cohort study. Clin Microbiol Infec 2019; 25:987–993.
- Beinortas T, Burr NE, Wilcox MH, Subramanian V. Comparative efficacy of treatments for Clostridium difficile infection: a systematic review and network meta-analysis. Lancet Infect Dis 2018; 18:1035–1044.
- Erikstrup LT, Aarup M, Hagemann-Madsen R, et al. Treatment of Clostridium difficile infection in mice with vancomycin alone is as effective as treatment with vancomycin and metronidazole in combination. BMJ Open Gastroenterol 2015; 2:e000038.
- 52. Li R, Lu L, Lin Y, et al. Efficacy and safety of metronidazole monotherapy versus vancomycin monotherapy or combination therapy in patients with Clostridium difficile infection: a systematic review and meta-analysis. PloS One 2015; 10:e0137252.
- 53. Rokas KEE, Johnson JW, Beardsley JR, et al. The addition of intravenous metronidazole to oral vancomycin is associated with improved mortality in critically ill patients with Clostridium difficile infection. Clin Infect Dis 2015; 61:934–941.
- Shahani L, Koirala J. Use of intravenous immunoglobulin in severe Clostridium difficile -associated diarrhea. Hosp Pract 2015; 43:154–157.
- 55. Negm OH, MacKenzie B, Hamed MR, et al. Protective antibodies against Clostridium difficile are present in intravenous immunoglobulin and are retained in humans following its administration. Clin Exp Immunol 2017; 188:437-443.
- Abougergi MS, Kwon JH. Intravenous immunoglobulin for the treatment of Clostridium difficile infection: a review. Digest Dis Sci 2010; 56:19-26.
- Causey MW, Walker A, Cummings M, et al. Colonic decompression and direct intraluminal medical therapy for Clostridium difficile-associated megacolon using a tube placed endoscopically in the proximal colon. Colorectal Dis 2014: 16:071 – 074.
- Akamine CM, Ing MB, Jackson CS, Loo LK. The efficacy of intracolonic vancomycin for severe Clostridium difficile colitis: a case series. BMC Infect Dis 2016; 16:316.

- Kim PK, Huh HC, Cohen HW, et al. Intracolonic vancomycin for severe Clostridium difficile colitis. Surg Infect (Larchmt) 2013; 14:532–539.
- Malamood M, Nellis E, Ehrlich AC, Friedenberg FK. Vancomycin enemas as adjunctive therapy for Clostridium difficile infection. J Clin Med Res 2015; 7:422-427.
- Aldape MJ, Heeney DD, Bryant AE, Stevens DL. Tigecycline suppresses toxin A and B production and sporulation in Clostridium difficile. J Antimicrob Chemoth 2014; 70:153–159.
- Britt NS, Steed ME, Potter EM, Clough LA. Tigecycline for the treatment of severe and severe complicated Clostridium difficile infection. Infect Dis Ther 2014; 3:321–331.
- Thomas A, Khan F, Uddin N, Wallace MR. Tigecycline for severe Clostridium difficile infection. Int J Infect Dis 2014; 26:171–172.
- Knafl D, Winhofer Y, Lötsch F, et al. Tigecycline as last resort in severe refractory Clostridium difficile infection: a case report. J Hosp Infect 2016; 92:296-298.
- 65. Navalkele BD, Lerner SA. Intravenous Tigecycline Facilitates Cure of Severe Clostridium difficileInfection (CDI) After Failure of Standard Therapy: A Case Report and Literature Review of Tigecycline Use in CDI. Open Forum Infect Dis 2016; 3:ofw0943.
- 66. Bishop EJ, Tiruvoipati R, Metcalfe J, et al. The outcome of patients with severe and severe-complicated Clostridium difficile infection treated with tigecycline combination therapy: a retrospective observational study. Intern Med J 2018; 48:651–660.
- 67. Manea E, Sojo-Dorado J, Jipa RE, et al. The role of tigecycline in the management of Clostridium difficile infection: a retrospective cohort study. Clin Microbiol Infect 2018; 24:180–184.
- van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal infusion of donor feces for recurrent Clostridium difficile. New Engl J Med 2013; 368:407–415.
- Trubiano JA, Gardiner B, Kwong JC, et al. Faecal microbiota transplantation for severe Clostridium difficile infection in the intensive care unit. Eur J Gastroen Hepatol 2013; 25:255–257.
- Kim JE, Gweon T-G, Yeo CD, et al. A case of Clostridium difficile infection complicated by acute respiratory distress syndrome treated with fecal microbiota transplantation. World J Gastroenterol 2014; 20:12687–12690.
- Marcos LA, Gersh A, Blanchard K, et al. Fecal transplantation to treat initial severe Clostridium difficile infection with sepsis. J Miss State Med Ass 2015; 56:38-40.
- Schulz-Stübner S, Textor Z, Anetseder M. Fecal microbiota therapy as rescue therapy for life-threatening Clostridium difficile Infection in the critically ill: a small case series. Infect Control Hosp Epidemiol 2016; 37:1129-1131.
- Costello SP, Chung A, Andrews JM, Fraser RJ. Fecal microbiota transplant for Clostridium difficile colitis induced toxic megacolon. Am J Gastroenterol 2015; 110:775–776.
- 74. Alukal J, Dutta SK, Surapaneni BK, et al. Safety and efficacy of fecal microbiota transplant in 9 critically ill patients with severe and complicated Clostridium difficile infection with impending colectomy: a case series. J Digest Dis 2019; 20:301–307.
- Neemann K, Eichele DD, Smith PW, et al. Fecal microbiota transplantation for fulminant Clostridium difficileinfection in an allogeneic stem cell transplant patient. Transpl Infect Dis 2012; 14:E161–E165.
- Fischer M, Sipe BW, Rogers NA, et al. Faecal microbiota transplantation plus selected use of vancomycin for severe-complicated Clostridium difficileinfection: description of a protocol with high success rate. Aliment Pharm Therap 2015: 42:470–476.
- Aroniadis OC, Brandt LJ, Greenberg A, et al. Long-term follow-up study of fecal microbiota transplantation for severe and/or complicated Clostridium difficile infection: a multicenter experience. J Clin Gastroenterol 2016; 50:398-402.
- 78. Fischer M, Sipe B, Cheng Y-W, et al. Fecal microbiota transplant in severe and severe-complicated Clostridium difficile: a promising treatment approach. Gut Microbes 2016; 8:289-302.
- Neal MD, Alverdy JC, Hall DE, et al. Diverting loop ileostomy and colonic lavage: an alternative to total abdominal colectomy for the treatment of severe, complicated Clostridium difficile associated disease. Ann Surg 2011; 254-423-427.
- Fashandi AZ, Martin AN, Wang PT, et al. An institutional comparison of total abdominal colectomy and diverting loop ileostomy and colonic lavage in the treatment of severe, complicated Clostridium difficile infections. Am J Surg 2017; 213:507-511.
- Lowy I, Molrine DC, Leav BA, et al. Treatment with monoclonal antibodies against Clostridium difficile toxins. New Engl J Med 2010; 362:197-205.
- Wilcox MH, Gerding DN, Poxton IR, et al., MODIFY I and MODIFY II Investigators. Bezlotoxumab for prevention of recurrent Clostridium difficile infection. New Engl J Med 2017; 376:305–317.
- Kotloff KL, Wasserman SS, Losonsky GA, et al. Safety and immunogenicity of increasing doses of a Clostridium difficile toxoid vaccine administered to healthy adults. Infect Immun 2001; 69:988–995.
- 84. Bruyn G de. Saleh J, Workman D, et al., H-030-012 Clinical Investigator Study Team. Defining the optimal formulation and schedule of a candidate toxoid vaccine against Clostridium difficile infection: a randomized Phase 2 clinical trial. Vaccine 2016; 34:2170-2178.

- 85. Bézay N, Ayad A, Dubischar K, et al. Safety, immunogenicity and dose response of VLA84, a new vaccine candidate against Clostridium difficile, in healthy volunteers. Vaccine 2016; 34:2585-2592.
- **86.** Zhang B-Z, Cai J, Yu B, et al. A DNA vaccine targeting TcdA and TcdB induces protective immunity against Clostridium difficile. BMC Infect Dis 2016; 16:596.
- 87. Baliban SM, Michael A, Shammassian B, et al. An optimized, synthetic DNA vaccine encoding the toxin A and toxin B receptor binding domains of Clostridium difficile induces protective antibody responses in vivo. Infect Immun 2014; 82:4080-4091.
- Pizarro-Guajardo M, Chamorro-Veloso N, Vidal RM, Paredes-Sabja D. New insights for vaccine development against Clostridium difficile infections. Anaerobe 2019; 58:73-79.
- 89. Garey KW, Ghantoji SS, Shah DN, et al. A randomized, double-blind, placebo-controlled pilot study to assess the ability of rifaximin to prevent recurrent diarrhoea in patients with Clostridium difficile infection. J Antimicrob Chemoth 2011; 66:2850–2855.
- Major G, Bradshaw L, Boota N, et al., RAPID Collaboration Group. Follow-on RifAximin for the prevention of recurrence following standard treatment of Infection with Clostridium difficile (RAPID): a randomised placebo controlled trial. Gut 2018; 68:1224–1231.
- 91. Dubberke ER, Lee CH, Orenstein R, et al. Results from a randomized, placebo-controlled clinical trial of a RBX2660 a microbiota-based drug for the prevention of recurrent Clostridium difficile infection. Clin Infect Dis 2018; 67:1198–1204.