Package 'Rseb'

February 20, 2022

Type Package
Title An R-package for NGS data managing and visualization
Version 0.3.0
Author Sebastian Gregoricchio
${\bf Maintainer}\ {\bf Sebastian}\ {\bf Gregoricchio} < {\bf sebastian}. {\bf gregoricchio@gmail.com} >$
Description An R-package for daily tasks required to handle biological data as well as avoid recoding of small functions for quick but necessary data managing.
License GNU GENERAL PUBLIC LICENSE version 3
Depends R ($\xi = 4.0.0$)
Imports BiocManager, AnnotationFilter, Biostrings, biomaRt, diffloop, EnsDb.Hsapiens.v75, EnsDb.Hsapiens.v86, EnsDb.Mmusculus.v79, GenomicRanges, GO.db, rtracklayer, cowplot, data.table, ggplot2 (¿= 3.3.3), ggbio, ggforce, ggrepel, ggpubr, ggpmisc, matrixStats, plyr, dplyr, tidyr, purrr, readxl, robustbase, stringr, tools, devtools, rvcheck, curl, prettydoc, knitr, rmarkdown, stats, openssl
biocViews
Encoding UTF-8
LazyData true
RoxygenNote 7.1.2
VignetteBuilder knitr
<pre>URL https://sebastian-gregoricchio.github.io/Rseb/</pre>
https://github.com/sebastian-gregoricchio/Rseb/
https://sebastian-gregoricchio.github.io/
BugReports https://github.com/sebastian-gregoricchio/Rseb/issues
R topics documented:
actualize

actualize 3

actualize

 ${\tt Rseb}\ updates\ verification$

Description

It verifies if Rseb is up-to-date and installs it when required.

Usage

```
actualize(
  update = TRUE,
  verbose = TRUE,
  force = FALSE,
  build.manual = TRUE,
  build.vignettes = TRUE)
```

Arguments

update Logical value to define whether update the Rseb package. By default TRUE.

verbose Logical value to define whether print messages. By default TRUE.

force Logical value to define whether to force the installation of Rseb even

though already up-to-date. Parameter passed to devtools::install_github().

By default FALSE.

build.manual Logic value to define whether to build the manual. By default TRUE.

build.vignettes

Logic value to define whether to build the vignettes. By default TRUE.

Details

This function will check for internet availability.

Value

Warnings and/or messages. Installation of the latest version of Rseb if required.

4 build.bed

build.bed

 $Bed\ generator$

Description

Function that helps the building of a bed file providing the columns. It enables also the specification of the track line for software such as IGV in order to pre-define colors, track name, etc.

Usage

```
build.bed(
  chr,
  start,
  end,
  name = NULL,
  score = 0,
  strand = ".",
  thickStart = NULL,
  thickEnd = NULL,
  itemRgb = NULL,
  blockCount = NULL,
 blockSizes = NULL,
  blockStarts = NULL,
  track.name = NULL,
  display.mode = NULL,
  itemRgb.ON = T,
  useScore = F,
  colorByStrand = NULL,
  track.base.color = NULL,
  sort = T,
  bed.file.name = NULL,
  export.track.line = TRUE,
  return.data.frame = F,
  force.generation = F
)
```

Arguments

chr	String vector containing the name of the chromosome (e.g. chr3, chrY, chr2_random) or scaffold (e.g. scaffold10671).
start	Numeric vector indicating the starting position of the feature in the chromosome or scaffold. The first base in a chromosome is numbered 0 .
end	Numeric vector indicating the ending position of the feature in the chromosome or scaffold.

build.bed 5

name String vector defining the name of the BED line. This label is displayed to

the left of the BED line in the Genome Browser window when the track is open to full display mode or directly to the left of the item in pack mode. If set as NULL (default) and the column is required, the names will

correspond to the mid-point of the region.

A single value or a numeric vector with a score between 0 and 1000. If the track line useScore attribute is set as TRUE for this annotation data

set, the score value will determine the level of gray in which this feature is displayed (higher numbers = darker gray). By default 0.

A single character or a string vector defining the strand: either "." (=no strand) or "+" or "-". By default ".".

thickStart A numeric vector indicating the starting position at which the feature is drawn thickly (for example, the start codon in gene displays). When there is no thick part (default value, thickStart = NULL) it will be used

the start value.

thickEnd A numeric vector indicating the ending position at which the feature is drawn thickly (for example, the start codon in gene displays). When there is no thick part (default value, thickStart = NULL) it will be used the end

value.

A single value or a string vector containing the colors for each feature. It can be expressed as an RGB value of the form R,G,B (e.g. "255,0,0") or as any other R-supported color name (it will be converted automatically to RGB version). By default NULL. If the track line itemRgb.ON attribute is set as TRUE, this color value will determine the display color

of the data contained in this BED line. NOTE: It is recommended that a simple color scheme (eight colors or less) be used with this attribute to avoid overwhelming the color resources of the Genome Browser and your

Internet browser.

A single number or a numeric vector indicating the number of blocks

(exons) in the BED line. By default NULL.

blockSizes A vector containing a comma-separated list of the block sizes. The num-

ber of items in this list should correspond to blockCount. By default

NULL.

blockStarts A vector containing a comma-separated list of block starts. All of the blockStart positions should be calculated relative to start. The number

of items in this list should correspond to blockCount. By default NULL.

track.name A string defining the track label that will be displayed to the left of the

track in the Genome Browser window, and also the label of the track control at the bottom of the screen. The name can consist of up to 15 characters. It is recommended that the track-label be restricted to alphanumeric characters and spaces to avoid potential parsing problems. By

default NULL.

display.mode A string that defines the initial display mode of the annotation track. Values for display.mode include: "hide", "dense", "full", "pack", "squish".

By default NULL.

6 build.bed

itemRgb.ON

Logic value to define whether this attribute should be set to "On", the Genome Browser will use the RGB value shown in the itemRgb field in each data line of the associated BED track to determine the display color of the data on that line. If the itemRgb values are not provided, this parameter will be ignored. By default TRUE.

useScore

Logic value to define if the score field in each of the track's data lines should be used to determine the level of shading in which the data is displayed. By default FALSE.

colorByStrand

A vector composed by two strings for two colors, either in RGB comma separated format (eg. "0,250,30") or any R-supported color string (they will be converted automatically to RGB format). The order of color sets is c("strand +", "strand -"). Parameter ignored when itemRgb is active/provided. By default NULL.

track.base.color

A single string defining the main color for the annotation track. The track color consists of three comma-separated RGB values from 0-255 (eg. "0,250,30") or any R-supported color string (it will be converted automatically to RGB format). Parameter ignored when itemRgb or colorByStrand are active/provided. By default NULL.

sort

Logic value to define whether to sort the bed using the function sort.bed. By default TRUE.

bed.file.name

If a string with a full path to a bed_file is provided, the function will export the bed as a txt file. By default NULL.

export.track.line

Logic value to define if the track line should be exported. When bed.file.name = NULL this parameter is ignored. By default TRUE.

return.data.frame

Logic value to define if the to return the data frame corresponding to the bed (it will show the columns names). By default FALSE.

force.generation

Force the generation of bed even when certain errors occur (eg. score \not 1000, start \not end). By default FALSE.

Value

If required the function can export a bed file with or without the track line, return a data.frame (with column names) corresponding to the bed generated, or both. The bed file could be automatically sorted settin the parameter sort = TRUE.

References

- More information about bed format are available at the following link: https://genome.ucsc.edu/FAQ/FAQformat.html#format1.
- More information about track line parameters are available at the following link: https://genome.ucsc.edu/goldenPath/help/hgTracksHelp.html#lines.

calculate.mode 7

calculate.mode

 $Mode\ calculation$

Description

Calculate the mode value of a vector of numeric values.

Usage

```
calculate.mode(v)
```

Arguments

V

A vector of numeric numbers

Value

A single number corresponding to the mode of the list of numbers give as input

Examples

```
mode = calculate.mode(v = c(6, 8, 4, 845, 8, 5, 55, 84, 8, 84, 45, 5))
```

cmyk

 $CMYK\ color\ converter$

Description

Converts CMYK color values to hexadecimal color values

Usage

```
cmyk(C, M, Y, K)
```

Arguments

С	Value in the 0-100 range for the Cyan component.
М	Value in the 0-100 range for the Magenta component.
Υ	Value in the 0-100 range for the Yellow component.
K	Value in the 0-100 range for the Key component.

Value

The result is a string for the color in hexadecimal scale, eg. "#FFFFFF".

8 collapse.bed

Examples

```
color = cmyk(0, 0, 0, 0)
```

CNV.data

 $CNV\ data\ results\ example$

Description

Simulation of Copy Number Variation (CNV) analysis on a cohort of patients.

Usage

CNV.data

Format

A data frame with 25 rows and 9 variables:

```
geneName hypothetical gene symbols
patient_1 ... patient_N hypothetical patients ID
```

Source

Simulated data

collapse.bed

Merger of overlapping peaks in a provided .bed file.

Description

Merge overlapping peaks in a provided .bed file.

Usage

```
collapse.bed(
  bed,
  maximal.distance = 0,
  keep.strandness = FALSE,
  only.one.strand = NULL,
  score.operation = "mean",
  bed.header = FALSE,
  sep = "\t",
  return.bed = TRUE,
  export.file.name = NULL,
  export.header = FALSE
)
```

collapse.bed 9

Arguments

bed

Two options are possible:

- String with the path to a .bed file;

- data.frame corresponding to a bed file format (only the first 6 columns, BED6, will be kept).

maximal.distance

Maximal distance between regions allowed for regions to be merged. By default \emptyset .

keep.strandness

Logic value to indicate whether to force to only merge regions that are in the same strand. By default FALSE, disabled. Subordinated to not NULL value for 'only.one.strand' option.

only.one.strand

Atomic string to indicate whether to force merge for one specific strand only. It must be indicated the wished strand (e.g., '+', '-', '.'). Regions in the other strand/s will be kept without any modification. By default NULL.

score.operation

Applicable only if the regions contain scores. Atomic string to indicate the operation to apply to the scores of merged regions. Possible choices: 'mean', 'median', 'sum'. By default "mean".

bed.header

Logic value to define whether the .bed file contains an header or not. By default FALSE.

sep Si

String containing the separator character for a .bed file. By default " \t^* ".

return.bed

Logic value to define if to return the bed as a data.frame. By default TRUE. Only unique rows are kept.

export.file.name

Optional: string to define the path to the file to be exported, if required. By default NULL, not exported.

export.header

Logic value to define whether the header should be exported in the sorted bed file. By default FALSE.

Details

The function pre-sorts the bed and keeps only unique rows and only up to 6 columns (chr, start, end, name, score, strand).

The names of the regions (if available) of merged regions corresponds to the concatenation of all original region's name.

To get more information about the bed file format see the following page:

https://genome.ucsc.edu/FAQ/FAQformat.html#format1.

Value

If required, returns a data frame corresponding to the collapsed .bed file.

10 combine.lists

or.gradient Gradient colors generation and assignment

Description

Give a vector of colors generates a finite number of shadows that will be assigned to a numeric vector depending on the value of each element.

Usage

```
color.gradient(values, colors = c("blue", "white", "red"), bins = 100)
```

Arguments

values A numeric vector containing the values to which a color must be assigned

(NAs and NaN will be converted to 0).

colors A string vector with the colors, in the wished order, that have to be used

to generated the shadows. By default c("blue", "white", "red").

bins An atomic integer value to define the total number of bins/steps in which

the gradient should be dived.

Value

A vector containing the assigned colors corresponding to each element of values.

combine.lists	$List\ combiner$	
---------------	------------------	--

Description

Combines two or more lists in a single one keeping the element names.

Usage

```
combine.lists(list.of.lists)
```

Arguments

```
list.of.lists A list of lists.
```

Value

It returns a list that is a combination of the lists in the input list. If the list is not a nested list of list the original input is returned.

Examples

Description

This function runs a command line that uses deeptools to calculate scores per genome regions and to prepare an intermediate file that can be used with plot.density.profile and plot.density.summary. Typically, the genome regions are genes, but any other regions defined in a BED file can be used. computeMatrix accepts multiple score files (bigWig format) and multiple regions files (BED format). This tool can also be used to filter and sort regions according to their score.

Usage

```
computeMatrix.deeptools(
 mode,
  scoreFileName,
  regionsFileName,
  outFileName,
  outFileNameMatrix = NULL,
  outFileSortedRegions = NULL,
  referencePoint = "TSS",
  nanAfterEnd = FALSE,
  regionBodyLength = 1000,
  startLabel = "TSS",
  endLabel = "TES",
  unscaled5prime = 0,
  unscaled3prime = 0,
  upstream = 500,
  downstream = 500,
  binSize = 10,
  sortRegions = "keep",
  sortUsing = "mean",
  sortUsingSamples = NULL,
  averageTypeBins = "mean",
 missingDataAsZero = FALSE,
  skipZeros = FALSE,
 minThreshold = NULL,
 maxThreshold = NULL,
```

```
blackListFileName = NULL,
  samplesLabel = NULL,
  smartLabels = TRUE,
  scale = 1,
  numberOfProcessors = "max",
 metagene = FALSE,
  transcriptID = "transcript",
  exonID = "exon",
  transcript_id_designator = "transcript_id",
  srun = FALSE,
  computeMatrix.deeptools.command = paste0("/home/", Sys.getenv("USERNAME"),
    "/anaconda3/bin/computeMatrix"),
  return.command = FALSE,
  run.command = TRUE,
  quiet = FALSE,
  verbose = FALSE
)
```

Arguments

mode

The type of matrix computation. Allowed values are "reference-point" or "scale-region". No default.

• reference-point:

Reference-point refers to a position within a BED region (e.g., the starting point). In this mode, only those genomic positions before (upstream) and/or after (downstream) of the reference point will be plotted;

• scale-region:

In the scale-regions mode, all regions in the BED file are stretched or shrunken to the length (in bases) indicated by the user.

scoreFileName

String vector with the full paths to bigWig file(s) containing the scores to be plotted.

regionsFileName

String vector with the full paths to .BED or .GTF files containing the regions to plot. If multiple bed files are given, each one is considered a group that can be plotted separately. Also, adding a "#" symbol in the bed file causes all the regions until the previous "#" to be considered one group.

outFileName

String containing the full file name to save the gzipped matrix file (.gz) needed by plot.density.profile.

outFileNameMatrix

If this option is given, then the matrix of values underlying the heatmap will be saved using the indicated name, e.g. IndividualValues.tab. This matrix can easily be loaded into R or other programs. By default NULL.

outFileSortedRegions

File name in which the regions are saved after skiping zeros or min/max threshold values. The order of the regions in the file follows the sorting or-

der selected. This is useful, for example, to generate other heatmaps keeping the sorting of the first heatmap. Example: Heatmap1sortedRegions.bed. By default NULL.

referencePoint Possible choices: TSS, TES, center. The reference point for the plotting could be either the region start (TSS), the region end (TES) or the center of the region. Note that regardless of what you specify, plotHeatmap/plotProfile will default to using "TSS" as the label. By default TSS.

nanAfterEnd

Logic value. If set (TRUE), any values after the region end are discarded. This is useful to visualize the region end when not using the scale-regions mode and when the reference-point is set to the TSS. By default FALSE.

regionBodyLength

Distance in bases to which all regions will be fit. (Default: 1000).

startLabel

Label shown in the plot for the start of the region. Default is TSS (transcription start site), but could be changed to anything, e.g. "peak start". Note that this is only useful if you plan to plot the results yourself and not, for example, with plotHeatmap, which will override this. (Default: "TSS").

endLabel

Label shown in the plot for the region end. Default is TES (transcription end site). See the -startLabel option for more information. (Default: "TES").

unscaled5prime

Number of bases at the 5-prime end of the region to exclude from scaling. By default, each region is scaled to a given length (see the -regionBodyLength option). In some cases it is useful to look at unscaled signals around region boundaries, so this setting specifies the number of unscaled bases on the 5-prime end of each boundary. (Default: 0).

unscaled3prime

Number of bases at the 3-prime end of the region to exclude from scaling. By default, each region is scaled to a given length (see the -regionBodyLength option). In some cases it is useful to look at unscaled signals around region boundaries, so this setting specifies the number of unscaled bases on the 3-prime end of each boundary. (Default: 0).

upstream

Distance upstream of the reference-point selected. (Default: 500).

downstream

Distance downstream of the reference-point selected. (Default: 500).

binSize

Length, in bases, of the non-overlapping bins for averaging the score over the regions length. (Default: 10).

sortRegions

Possible choices: "descend", "ascend", "no", "keep". Whether the output file should present the regions sorted. The default is to not sort the regions. Note that this is only useful if you plan to plot the results yourself and not, for example, with plotHeatmap, which will override this. Note also that unsorted output will be in whatever order the regions happen to be processed in and not match the order in the input files. If you require the output order to match that of the input regions, then either specify "keep" or use computeMatrixOperations to resort the results file. (Default: "keep").

sortUsing

Possible choices: "mean", "median", "max", "min", "sum", "region_length". Indicate which method should be used for sorting. The value is computed for each row. Note that the region_length option will lead to a dotted line within the heatmap that indicates the end of the regions. (Default: "mean").

sortUsingSamples

List of sample numbers (order as in matrix), that are used for sorting by -sortUsing, no value uses all samples, example: -sortUsingSamples 1 3. By default NULL.

averageTypeBins

Possible choices: "mean", "median", "min", "max", "std", "sum". Define the type of statistic that should be used over the bin size range. (Default: "mean").

missingDataAsZero

Logic value to define if set, missing data (NAs) will be treated as zeros. The default is to ignore such cases (NULL). If not included, this parameter can be changed later in the function plot.density.profile.

Logic value to understand whether regions with only scores of zero should skipZeros be included or not. Default is to include them (FALSE).

Numeric value. Any region containing a value that is less than or equal to this will be skipped. This is useful to skip, for example, genes where the read count is zero for any of the bins. This could be the result of unmappable areas and can bias the overall results. (Default: NULL).

maxThreshold Numeric value. Any region containing a value greater than or equal to this will be skipped. The maxThreshold is useful to skip those few regions with very high read counts (e.g. micro satellites) that may bias the average values. (Default: NULL).

blackListFileName

A BED file containing regions that should be excluded from all analyses. Currently this works by rejecting genomic chunks that happen to overlap an entry. Consequently, for BAM files, if a read partially overlaps a blacklisted region or a fragment spans over it, then the read/fragment might still be considered. (Default: NULL).

Labels for the samples. This will then be passed to plot.density.profile samplesLabel function. The default is to use the file name of the sample. The sample labels should be separated by spaces and quoted if a label itself contains a space E.g. -samplesLabel label-1 "label 2".

> Instead of manually specifying labels for the input bigWig and BED/GTF files, this causes deepTools to use the file name after removing the path and extension. (Default: TRUE).

scale If set, all values are multiplied by this number. (Default: 1).

numberOfProcessors

Number of processors to use. Type "max/2" to use half the maximum number of processors or "max" to use all available processors. (Default: "max").

minThreshold

smartLabels

metagene When either a BED12 or GTF file are used to provide regions, perform

the computation on the merged exons, rather than using the genomic interval defined by the 5-prime and 3-prime most transcript bound (i.e., columns 2 and 3 of a BED file). If a BED3 or BED6 file is used as input,

then columns 2 and 3 are used as an exon. (Default: FALSE).

transcriptID When a GTF file is used to provide regions, only entries with this value

as their feature (column 3) will be processed as transcripts. (Default:

"transcript").

exonID When a GTF file is used to provide regions, only entries with this value as their feature (column 3) will be processed as exons. CDS would be

another common value for this. (Default: "exon").

transcript_id_designator

Each region has an ID (e.g., ACTB) assigned to it, which for BED files is either column 4 (if it exists) or the interval bounds. For GTF files this is instead stored in the last column as a key:value pair (e.g., as 'transcript_id "ACTB"', for a key of transcript_id and a value of ACTB). In some cases it can be convenient to use a different identifier. To do so, set this to the

desired key. (Default: "transcript_id").

srun Logic value to define whether the command should be run in srun mode.

By default FALSE.

computeMatrix.deeptools.command

String to define the command to use to recall the computeMatrix function of deeptools. An example: "/home/user/anaconda3/bin/computeMatrix".

By default "/home/USERNAME/anaconda3/bin/computeMatrix".

return.command Logic value to define whether to return the string corresponding to the

command for deeptools. By default FALSE.

run.command Logic value to define whether to run the the command line on system

terminal and generate the score matrix by deeptools. By default TRUE.

quiet Logic value to define if to remove any warning or processing messages.

By default FALSE.

verbose Logic value to define if to be VERY verbose in the status messages. –quiet

will disable this. By default FALSE.

Details

To know more about the deeptools's computeMatrix function see the package manual at the following link:

https://deeptools.readthedocs.io/en/develop/content/tools/computeMatrix.html.

Value

The function generates the files indicated by the output parameters. The matrix.gz output file can be read by the function read.computeMatrix.file.

16 convert_sequence

Examples

```
computeMatrix.deeptools(
  mode = "reference-point",
  scoreFileName = c("path_to/signal_file1.bw", "path_to/signal_file2.bw"),
  regionsFileName = c("path.to/regions1.bed", "path.to/regions2.bed"),
  upstream = 1000,
  downstream = 1000,
  outFileName = "path_to/output_matrix.gz",
  computeMatrix.deeptools.command = "/home/user/anaconda3/bin/computeMatrix",
   referencePoint = "peakMax")
computeMatrix.deeptools(
  mode = "scale-regions",
  scoreFileName = c("path_to/signal_file1.bw", "path_to/signal_file2.bw"),
  regionsFileName = c("path.to/regions1.bed", "path.to/regions2.bed"),
  upstream = 1000,
  downstream = 1000,
  regionBodyLength = 300,
  startLabel = "geneStart",
  endLabel = "geneEnd",
  outFileName = "path_to/output_matrix.gz",
  computeMatrix.deeptools.command = "/home/user/anaconda3/bin/computeMatrix",
   referencePoint = "peakMax")
```

convert_sequence

Nucleic acid sequences converter.

Description

Obtains de complementary, reverse complementary or the reverse of a DNA/RNA sequence.

Usage

```
convert_sequence(sequence = NULL, mode = "not specified", nucleic.acid = "DNA")
```

Arguments

sequence A string containing the sequence to be converted. By default NULL, it

returns an help for the mode.

mode A string value to define the modality of convertion. Possible options:

- Reverse complement = revComp — RC — rc — reverseComplement

- Reverse = rev - R - r - reverse

- Complement = comp — C — c — complement.

By default "not specified", it returns an help for the mode.

nucleic.acid A string to define the type of nucleic acid to which the input sequence

belongs. Available options "DNA", default value, or "RNA".

data.frame.to.list

Value

It returns a string with the converted sequence.

Examples

data.frame.to.list

Data frame conversion to a list of columns.

Description

Converts each column of a data.frame in a element of a list with the corresponding name of the original column. Useful for further use in functions such as purrr::pmap().

Usage

```
data.frame.to.list(x)
```

Arguments

Х

A data.frame to be converted

Value

A list of vectors in which each element is a column of input the data.frame.

Examples

```
data.frame.to.list(mtcars)
```

data.summary

Statistical data summary generator

Description

Produces a table with a summary of the statistics for a specific column of an input data.frame by a group of values defined by a group defined by another column.

Usage

```
data.summary(data, variable, group.names)
```

DE.status

Arguments

data Input data.frame to be analyzed.

variable A string with the name of the column to be analyzed.

group.names A string with the name of the column indicating the groups.

Value

It returns a list that is a combination of the lists in the input list. If the list is not a nested list of list the original input is returned.

Examples

```
data.summary(data = mtcars, variable = "mpg", group.names = "disp")
```

DE.status

Differential Expression status calculator for RNA-seq data

Description

Defines the differential expression status of genes from RNA-seq data depending on fold change expression and adjusted p-value.

Usage

```
DE.status(
  log2FC,
  p.value.adjusted,
  FC_threshold = 1.5,
  FC_NoResp_left = 0.9,
  FC_NoResp_rigth = NULL,
  p.value_threshold = 0.05,
  low.FC.status.label = "DOWN",
  high.FC.status.label = "UP",
  unresponsive.label = "NoResp",
  null.label = "NULL"
)
```

Arguments

log2FC Numeric vector of log2(fold change expression) values.

p.value.adjusted

Numeric vector of p-values. Use of adjusted p-values is recommended.

FC_threshold Value of the threshold to use for the fold change expression to define

differentially expressed genes, expressed as linear value. By default 1.5

and by consequence 1/1.5.

deeptools.matrix 19

FC_NoResp_left Value of the threshold to use for the fold change expression to define unresponsive genes when FC < 1, expressed as linear value. By default 0.9. If NULL it will be calculated symmetrically from FC_NoResp_rigth as 1/FC_NoResp_rigth.

FC_NoResp_rigth Value of the threshold to use for the fold change expression to define unresponsive genes when FC > 1, expressed as linear value. By default 1.1. If NULL it will be calculated symmetrically from FC_NoResp_left as 1/FC_NoResp_left.

p.value_threshold

Value of the threshold to use for the p-values to define differentially expressed genes, expressed as linear value. By default 0.05.

low.FC.status.label

String to define the label indicating the differentially expressed genes with a FoldChange < FC_threshold.

high.FC.status.label

String to define the label indicating the differentially expressed genes with a FoldChange > FC_threshold.

unresponsive.label

String to define the label indicating the unresponsive genes identified as FC_NoResp_left < FoldChange < FC_NoResp_rigth and p.value > p.value.threshold.

null.label String to define the label indicating the null genes.

Value

It returns a vector containing the differential expression status for each original value in the same order used in the input.

deeptools.matrix

RNA-seq example

Description

List result of the function read.computeMatrix.file used to read a matrix.gz file generated by deepTools computeMatrix function.

Usage

deeptools.matrix

Format

A named list of 3 variables:

metadata data.frame with the information gotten from the matrix_file.gz matrix.data data.frame with the scores gotten from original.file.path with full path to the original matrix_file.gz 20 density.matrix

Source

```
http://path.to.paper/
```

density.matrix

Density matrix builder

Description

A function (completely in R) that generates a matrix given a list of regions (.bed files) and signals (.bigWig files) alternative (even though more time consuming) to computeMatrix.deeptools. The output can be passed as it is to the functions plot.density.profile, plot.density.summary and, plot.density.differences.

Usage

```
## S3 method for class 'matrix'
density(
 mode,
  regions.list,
  samples.list,
  region.names = NULL,
  sample.names = NULL,
  sort.regions.coordinates = FALSE,
  reference.point = "center",
  reference.point.label = NULL,
  upstream = 500,
  downstream = 500,
  body.length = 1000,
 missing.data.as.zero = FALSE,
 bin.size = 10,
 binning.operation = "mean",
  stranded = FALSE
)
```

Arguments

mode

A string indicating the method for the matrix computation:

- scale-regions all regions in the BED file are stretched or shrunken to the length (in bases) indicated by the user (body.length);
- reference-point the matrix will be performed on the range-upstream+downstream from the indicated reference point (center, TSS, TES).

regions.list

A string vector with a list of full paths to bed files or list of data.frames in at least BED3 format (eg. generated by build.bed).

samples.list

A string vector with a list of full paths to bigWig files.

density.matrix 21

region.names

A string vector with the names of the regions. If NULL or of length lower than the number of regions the names will be assigned using the basename of the file if a path is provided otherwise "region_jorder number.". By default NULL.

sample.names

A string vector with the names of the samples. If NULL or of length lower than the number of samples the names will be assigned using the basename of the file. By default NULL.

sort.regions.coordinates

Logical value to define whether the output matrix should contain the regions sorted by genomic location for each region group (sorted by sort.bed). By default FALSE.

reference.point

The reference point for the matrix generation could be either the region start ("TSS"), the region end ("TES") or the "center" of the region. By default "center".

reference.point.label

A single string with the label for the reference point that could be used for the plots.

upstream

Distance, in bases (bp), upstream of the reference-point, in "reference-point" mode, or the region start, in "scale-regions" mode. By default 500.

downstream

Distance, in bases (bp), downstream of the reference-point, in "reference-point" mode, or the region start, in "scale-regions" mode. By default 500.

body.length Dista missing.data.as.zero

Distance, in bases (bp), to which all regions will be fit. By default: 1000. s.zero

A logical value to define whether missing data (NAs) should be treated as zeros. By default FALSE.

bin.size

Length, in bases (bp), of the non-overlapping bins for averaging the score over the regions length. By default 10.

binning.operation

A single string to define the type of statistic that should be used over the bin size range. The options are: "mean", "median", "sum". By default "mean".

stranded

Logical value to indicate whether the strand of the region should be taken into account. When TRUE, the order of the bigWig score for the given region will be reversed. Default FALSE.

Value

The function returns a named list containing:

- metadata data.frame with the parameters used to build the matrix;
- \bullet $\mbox{matrix.data}$ data.frame with the computed scores;
- original.file.path with the string: "Matrix generated by Rseb::density.matrix()".

This list can be passed as it is to the functions plot.density.profile, plot.density.summary and, plot.density.differences.

22 density_plot

density_plot

Plot density signal of NGS data.

Description

Plots the density profile of NGS data (e.g. ChIP-seq, ATAC-seq, MeDIP-seq, etc.). Used by the function plot.density.profile.

Usage

```
density_plot(
  samples,
  scores,
  positions,
  variance_scores,
  xlab = "Distance from regions center [bp]",
  ylab = "Average density signal",
  line_type = "solid",
 y_{lim} = NULL,
  x_{lim} = NULL
  x_{intercept} = 0,
  colors = c("blue", "red", "purple", "orange", "green"),
  title = "Density profile",
  text_size = 12,
  variance = T,
  print_plot = F,
  line_width = 1,
  variance_opacity = 0.25
)
```

Arguments

samples A character vector containing the samples list.

scores A numeric vector containing the scores for the Y-axis.

Positions A numeric vector containing the position for the X-axis.

variance_scores

A numeric vector containing the variance/error value at each position.

xlab A string containing the label for the X-axis. By default "Distance from

regions center [bp]".

ylab A string containing the label for the Y-axis. By default "Average density

signal".

line_type Vector to define each line type. Both numeric and string codes are ac-

cepted. if only one element is given this will be applied to all the lines.

By default "solid".

Example 1: c("solid", "dashed").

Example 2: c(1,2)

doughnut23

y_lim	List of numeric vectors with two elements each to define the range of the Y-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default NULL, the range will be defined automatically. Example list($c(0,20)$, $c(NA,30)$, $c(NA,NA)$).
x_lim	List of numeric vectors with two elements each to define the range of the X-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default NULL, the range will be defined automatically. Example list($c(0,20)$, $c(NA,30)$, $c(NA,NA)$).
$x_intercept$	A vector indicating the X intercepts for the vertical lines. By default 0.
colors	Vector to define the line and error area colors. If only one value is provided or the number of values is lower than the required ones only the first value will be used. All standard R.colors values are accepted. By default $c("blue","red","purple","orange","green")$.
title	A string containing the label for the X-axis. By default "Density profile".
text_size	Numeric value to define the size of the text for the labels of all the plots. By default 12.
variance	Logic value to define whether to plot the error/variance around the signal. By default TRUE.
print_plot	Logic value to define whether to print the plot once generated or not. By default $FALSE.$
line_width	Numeric value to define the line width for all the plots. By default 1.,
variance_opacit	y
	Numeric value to define the alpha/transparency of the error/variance. By default 0.25. Parameter considered only when variance = TRUE)

default 0.25. Parameter considered only when variance = TRUE).

Value

Returns a plot in ggplot2 format.

doughnut	$Donut/Doughnut\ plot$	

Description

Generation of a donut/doughnut plot (equivalent of a pie chart)

Usage

```
doughnut(
 labels = as.character(x),
 edges = 200,
 outer.radius = 0.8,
```

24 doughnut

```
inner.radius = 0.4,
clockwise = FALSE,
init.angle = if (clockwise) 90 else 0,
density = NULL,
angle = 45,
col = NULL,
border = FALSE,
lty = NULL,
main = NULL,
...
)
```

Arguments

Χ

labels	A string vector for the labels of the different sectors. By default as $\operatorname{character}(x)$.
edges	Number of edges of the shape. By default 200.
outer.radius	Fraction of the area to dedicate to the outer circle. By default 0.8.
inner.radius	Fraction of the area to dedicate to the inner circle. By default 0.4.
clockwise	Logic value to define whether the values should be plotted in clockwise sense. By default FALSE.
init.angle	Numeric value to define the starting angle for the data. By default if $clockwise = TRUE 90$, otherwise 0.
density	A vector or single number to define de density of the lines in the filling color of each value plotted. By default NULL.
angle	A vector or single number to define de angle of the lines in the filling color of each value plotted. By default 45.
-	

A vector containing the values to be plotted.

A vector of R standard colors for each value to be plotted. By default NULL.

Logic value to define whether plot the border of the sectors. By default

FALSE.

1ty Numeric value to define the type of line for the borders. By default NULL.

main String to set the title of the plot. By default NULL.

References

border

```
https://magesblog.com/
```

Examples

```
doughnut(x = c(3,5,9,12), inner.radius=0.5, col=c("red", "blue", "green", "yellow"))
```

floating.ceiling 25

floating.ceiling	Ceiling to floating value.
Toucing. Colling	certify to floating carae.

Description

Computes the ceiling of the given value but with any number of digits (to the closest floating number of given digits).

Usage

```
floating.ceiling(num, digits = 1)
```

Arguments

num A single number or a numeric vector.

digits A single integer indicating the maximum number of digits required.

Value

A floored number or numeric vector.

	floating.floor	Flooring to floating values	
--	----------------	-----------------------------	--

Description

Computes the floor of the given value but with any number of digits (to the closest floating number of given digits).

Usage

```
floating.floor(num, digits = 1)
```

Arguments

num A single number or a numeric vector.

digits A single integer indicating the maximum number of digits required.

Value

A floored number or numeric vector.

26 genomic.tracks

genomic.tracks

Genomic tracks plotter

Description

The functions allows to plot different types of genomic data (bigWig, bed, bedpe) at a specific genomic region. It is possible to highlight specific regions and the gene annotations are plotted automatically at the bottom of all the tracks.

Usage

```
genomic.tracks(
  tracks,
  genomic.region,
 genome,
  track.labels = NULL,
  track.labels.fontzise = 5,
  track.labels.position = c(-0.1, 0),
  track.colors = "#000000",
  grouping = NULL,
  gene.annotation.color = "darkblue",
  expand.bed = TRUE,
  arcs.direction = "down",
  fraction.arc.base = 0.025,
  highlight.bed = NULL,
  highlight.color = "yellow",
  highlight.transparency = 0.15,
 missing.data.as.zero.bw = FALSE,
  smooth.bigWig.signal = TRUE,
  smooth.bigWig.loess.span = 0.05,
  plot.bigWig.area = TRUE,
  bigWig.range.label.size = 2.5,
  score.bed.shadow = FALSE,
 height.ratios = NULL,
  width.ratios = c(1, 5)
)
```

Arguments

tracks A vector indicating the list of full paths of the files/tracks/signals to plot.

 $Supported \ formats: \ bed/bd/narrowPeak/broadPeak, bw/bigWig/bigwig,$

bedne.

genomic.region An atomic string indicating the genomic region into which restrict the

final plot in the format 'chr1:1234-5678'.

genome An atomic string indicating the genome to use for the annotations. Al-

lowed values are:

genomic.tracks 27

- hg19: loads an 'EnsDb' object from the library EnsDb. Hsapiens. v75;
- hg38: loads an 'EnsDb' object from the library EnsDb. Hsapiens. v86;
- mm10: loads an 'EnsDb' object from the library EnsDb. Mmusculus.v79;
- custom 'EnsDb' object: provide an 'EnsDb' object manually generated; visit the page https://bioconductor.org/packages/release/bioc/vignettes/ensembldb/inst/doc/ensembldb.html#102_building_annotation_packages for more information.

track.labels A vector indicating the labels to use for each track (genome annotation track excluded). By default NULL: the file base-name will be used.

track.labels.fontzise

A numerical value to indicate the font size of the track labels. Default value 5.

track.labels.position

A two-element numeric vector passed to xlim function for the definition of the frame size of the track labels. Default value c(-0.1,0).

track.colors A string vector indicating the color to use for each track (genome annotation track excluded). If only one value is provided it will be used for all the tracks. Default value "#000000" ("black").

A single numerical vector or a list of numeric vectors. Each list's element indicates the indexes corresponding to the tracks (1 = first track, 2 = second track, etc) for which the y-axes should be normalized. Each element will be taken into account in the order. Default value NULL.

gene.annotation.color

A string indicating the color to use for the genome annotation track.

expand.bed A logical value to define whether overlapping regions in a bed should be plotted on different levels. Default TRUE.

arcs.direction A string indicating the direction on which arcs should be plotted for bedpe files. Available options "up" or "down". Default value "down".

fraction.arc.base

A numerical value indicating the fraction of total plot height to be used as arc base thickness. By default 0.025 (2.5% of the track height).

highlight.bed Either a string indicating the full path to a bed file or a data.frame in BED3 format (chr, start, end) containing regions that should be highlighted in the plot. Regions included in the genomic range will be automatically selected. By default NULL.

highlight.color

A string indicating the color to use for the regions to highlight in the plot. By default 'yellow'.

highlight.transparency

A numerical value indicating the transparency (alpha) to use for the highlighted regions. Default value 0.15.

missing.data.as.zero.bw

A logical value to define where missing data in the bigWigs should be converted to zeros. Default FALSE.

28 get.gene.name

smooth.bigWig.signal

Logical value to indicate whether the bigWig signals should be smoothed (by loess x $\tilde{}$ y function). By default TRUE.

smooth.bigWig.loess.span

Numerical value to indicate the span value for the loess function used to smooth bigWig signals. By default 0.05.

plot.bigWig.area

Logical value to indicate whether the bigWig profile should be filled or not. If FALSE only the signal outline will be plotted. By default TRUE.

bigWig.range.label.size

A numerical value to indicate the font size of the bigWig signal range. Default value 2.5.

score.bed.shadow

Logical value to define whether the filling intensity of the bed segments should reflect the score of each signal. By default FALSE.

height.ratios

Numerical vector of relative track heights, passed to 'rel_heights' parameter of $cowplot::plot_grid()$. For example, in a two-row grid, rel_heights = c(2,1) would make the first column twice as wide as the second column. Value 1 indicates that all the tracks should have the same size. By default NULL, automatic ratios will be computed by this function.

width.ratios

Numerical vector of relative labels vs tracks widths, passed to 'rel_widths' parameter of cowplot::plot_grid(). For example, in a two-column grid, rel_widths = c(2, 1) would make the first column twice as wide as the second column. Value 1 indicates that all the tracks should have the same size. By default c(1,5) (1 label: 5 tracks).

Value

The function returns a named list containing:

- configuration: data.frame with the parameters used to build the plot(s);
- highlighted.region: data.frame with the regions used for the highlighting;
- single.track.list: a named list containing each single track plot used for the creation of the multi.track.plot;
- single.label.plot.list: a named list containing each single track label plot used for the creation of the multi.track.plot;
- multi.track.plot: the assembled multi.track labelled plot.

get.gene.name

Conversion of ENSEMBL gene IDs.

Description

Conversion of ENSEMBL gene IDs to gene symbols.

Usage

```
get.gene.name(ensembl.id, type = "gene", organism = "mmusculus")
```

Arguments

ensembl.id String vector of ENSEMBL genes IDs

type String to define the type of ENSEMBL inputs. By default gene to in-

dicate "ensembl_gene_id". If different from "gene" it will be set to "en-

sembl_transcript_id_version".

organism String to define de organism, e.g. mmusculus, hsapiens, etc. By default

mmusculus.

Value

A string vector with the corresponding gene_symbols.

Examples

```
gene_symbols =
get.gene.name(
  ensembl.id = c("ENSMUSG00000002111", "ENSMUSG00000027381"),
  type = "gene",
  organism = "mmusculus")
```

```
get.single.base.score.bw
```

Single base bigWig score selector

Description

Function to get the score from a bigWig for each base in a given genomic region.

Usage

```
get.single.base.score.bw(
  region,
  bigWig,
  missing.data.as.zero = TRUE,
  reverse.score = FALSE
)
```

30 grepl.data.frame

Arguments

region An atomic string indicating the genomic region into which restrict the

final plot in the format 'chr1:1234-5678'.

bigWig Full path to a bigWig file.

missing.data.as.zero

A logical value to define whether missing data (NAs) should be treated

as zeros. By default TRUE.

reverse.score A logical value to indicate whether the score order should be inverted.

Default TRUE.

Value

The output is a numeric vector containing the score for each base at a given position.

grepl.data.frame

Grep a pattern in a full data.frame.

Description

The function helps to define which rows of an input data.frame contain a specific patter.

Usage

```
grepl.data.frame(
  data.frame,
  pattern,
  ignore.case = FALSE,
  perl = FALSE,
  fixed = FALSE,
  useBytes = FALSE
)
```

Arguments

data.frame Input data.frame.

pattern Character string containing a regular expression (or character string for

fixed = TRUE) to be matched in the given character vector. Coerced by as.character to a character string if possible. If a character vector of length 2 or more is supplied, the first element is used with a warning.

Missing values are allowed except for regexpr and gregexpr.

ignore.case If FALSE, the pattern matching is case sensitive and if TRUE, case is ignored

during matching. By default FALSE.

perl Logical value to define if Perl-compatible regexps should be used. By

default FALSE.

GSEA.to.GOnumber 31

fixed Logical value to define if the pattern is a string to be matched as is.

Overrides all conflicting arguments. By default FALSE.

useBytes Logical value to define if the matching is done byte-by-byte rather than

character-by-character. By default FALSE.

Value

It will be return a logic vector with an element per each row of the data.frame. The value is TRUE when the patter is found at least once in the corresponding data.frame row.

Examples

```
iris = iris %>% filter(grepl.data.frame(iris, pattern = "setosa"))
```

GSEA.to.GOnumber

Conversion of GSEA terms into Gene Ontology numbers

Description

Helps to convert the terms of GSEA analyses into Gene Ontology (GO) ID numbers.

Usage

```
GSEA.to.GOnumber(
  input_terms,
  input_pvalue,
  return_table = T,
  export_table = F,
  output_file_name = paste(getwd(), "GO_numbers_table.tsv", sep = "/")
)
```

Arguments

input_terms A character vector containing the GSEA terms to be converted.

A numeric vector containing the p-values of the GSEA terms.

Logic value to define whether to return the resulting data.frame. By default TRUE.

export_table Logic value to define whether to export the resulting data.frame. By

default FALSE.

output_file_name

Path and file name of the output table if export is required. By default <working.directory>/GO_numbers_table.tsv.

Details

This functions requires the package GO.db.

If problems are encountered during the installation see https://www.biostars.org/p/50564/.

32 IGVsnap

Value

If required, returns a data.frame with 3 columns: GO_number, GO_annotation, p.value. This table could be directly exported.

Description

The function builds a script file that can be run on IGV to generate multiple screenshots at specific genomic regions.

Usage

```
IGVsnap(
 loci_vector,
  input_type,
 biomart = "ensembl",
 dataset = "mmusculus_gene_ensembl",
  reference_genome = NULL,
  fivePrime = 1000,
  threePrime = 1000,
  snap_names = NULL,
  IGV_batch_file = paste(getwd(), "/IGV_batch.txt", sep = ""),
  snap_image_format = "png",
  snap_directory = getwd(),
 maxPanelHeight = 1000,
 delay.interval = 10,
  session = NULL,
 exit = FALSE
)
```

Arguments

loci_vector	Either a gene name vector (e.g. c("Gapdh", "Spi1",)) or a regions vector (eg. c('chr1:253000-256503',). All IGV formats are allowed.
$input_type$	Define the input type. Allowed values are genes and regions.
biomart	Defines the biomart parameter for biomaRt package, by default ensembl.
dataset	$Defines \ the \ dataset\ parameter\ for\ biomaRt\ package, by\ default\ mmusculus_gene_ensembl.$
reference_genom	e
	[optional] Defines the genome to use, e.g. "mm9", "mm10", "hg19", "hg38", By default NULL.
fivePrime	Numeric value to define of how many base-pairs (bp) expand from full

gene position at it's 5'-end, default 1000bp.

install.pkg.source 33

threePrime Numeric value to define of how many base-pairs (bp) expand from full

gene position at it's 3'-end, default 1000bp.

snap_names [optional] String vector to define the names of images (without extention),

by default uses loci_vector.

IGV_batch_file String for the batch_script_file_name/path, by default <working_directory>/IGV_batch.txt.

snap_image_format

String to define the format of the images, e.g. "png", "jpeg", "svg", \dots .

By default png.

snap_directory String for the output directory for the snapshoots. By default jwork-

ing_directory.

maxPanelHeight Numeric value to define the height in pixel of the IGV pannel that will

be captured on IGV. By default 1000.

delay.interval Sets a delay (sleep) time in milliseconds. The sleep interval is invoked

between successive commands. By default 10. helps to give the time to

IGV to adapt the view before the snap (such as the autoscale).

session [optional] FULL path to an IGV session file (session.xml) to use for the

images. By default NULL.

exit Logical value to indicate whether exit IGV after image capture ended. By

default FALSE.

Details

To run the script on IGV: Tools ¿ Run Batch Script... ¿ choose the .txt output file from this function.

For more info on how batch tasks work on IGV see:

https://software.broadinstitute.org/software/igv/PortCommands.

Value

Exports a .txt file ready-to-use on IGV.

install.pkg.source Packet

Package installer from source archive.

Description

Allows the installation of R packages using the source archive file.

Usage

install.pkg.source(pkg.path)

Arguments

pkg.path String to define the path for the archive file to be installed.

34 intersect.bedtools

Value

No returned value. The package required will be installed.

```
 \begin{array}{ll} \hbox{\it intersect.bedtools} & Intersect\ two\ or\ more\ bed\ files\ (by\ {\it bedtools\ intersect}\ function). \end{array}
```

Description

This function runs a command line that uses bedtools intersect to intersect one or more .bed files.

Usage

```
intersect.bedtools(
  a,
 outputFileName = paste(getwd(), "intersected.bed", sep = "/"),
  abam = FALSE,
 ubam = FALSE,
 bed = FALSE,
 wa = FALSE,
 wb = FALSE,
 loj = FALSE,
 wo = FALSE,
 wao = FALSE,
 u = FALSE,
  c = FALSE,
 C = FALSE,
  v = FALSE,
  f = NULL,
 F. = NULL,
  r = FALSE,
 e = FALSE,
  s = FALSE,
  S = FALSE,
  split = FALSE,
  sorted = FALSE,
  g = NULL,
  srun = FALSE,
  intersect.bedtools.command = paste0("/home/", Sys.getenv("USERNAME"),
    "/anaconda3/bin/intersectBed"),
  return.command = FALSE,
  return.bed = FALSE,
 delete.output = FALSE,
  run.command = TRUE
)
```

intersect.bedtools 35

Arguments

A single string defining the BAM/BED/GFF/VCF file "A". Each feature in A is compared to B in search of overlaps. Use "stdin" if passing A with a UNIX pipe. A character vector with one or more BAM/BED/GFF/VCF file(s) "B". b It could be also a single string containing wildcard (*) character(s). Full path to output file name. By default <working.directory>/intersected.bed. outputFileName abam Logic value to define if file A is a BAM. Each BAM alignment in A is compared to B in search of overlaps. By default FALSE. ubam Logic value to define if to write the output as uncompressed BAM. The default is to write compressed BAM output (ubam = FALSE). bed Logic value to define whether to write output as BED when using a BAM input abam = TRUE. The default is to write output in BAM (bed = FALSE). Logic value to define if to write the original entry in A for each overlap. wa By default FALSE. Logic value to define if to write the original entry in B for each overlap. wb Useful for knowing what A overlaps. Restricted by -f and -r. By default FALSE. Logic value to define if to perform a "left outer join". That is, for each loj feature in A report each overlap with B. If no overlaps are found, report a NULL feature for B. By default FALSE. Logic value to define if to write the original A and B entries plus the WO number of base pairs of overlap between the two features. Only A features with overlap are reported. Restricted by -f and -r. By default FALSE. Logic value to define if to write the original A and B entries plus the wao number of base pairs of overlap between the two features. However, A features w/o overlap are also reported with a NULL B feature and overlap = 0. Restricted by -f and -r. By default FALSE. Logic value to define if to write original A entry once if any overlaps found u in B. In other words, just report the fact at least one overlap was found in B. Restricted by -f and -r. By default FALSE. С Logic value to define if to for each entry in A, report the number of hits in B while restricting to -f. Reports 0 for A entries that have no overlap with B. Restricted -f, -F, -r, and -s. By default FALSE. Logic value to define if to for each entry in A, separately report the number С of overlaps with each B file on a distinct line. Reports 0 for A entries that have no overlap with B. Overlaps restricted by -f, -F, -r, and -s. By default FALSE. Logic value to define if to only report those entries in A that have no ٧ overlap in B. Restricted by -f and -r. f Numeric value defining the minimum overlap required as a fraction of A. Default is 1E-9 (i.e. 1bp). By default NULL. F. Numeric value defining the minimum overlap required as a fraction of B. Default is 1E-9 (i.e., 1bp). By default NULL.

36 intersect.bedtools

r	Logic value defining if the fraction (parameter f) is required to be reciprocal fraction of overlap for A and B. In other words, if -f is 0.90 and -r is used, this requires that B overlap at least 90% of A and that A also overlaps at least 90% of B. By default NULL.
е	Logic value defining if the fraction (parameter f) must be satisfied for A $_{\rm OR}$ B. In other words, if -e is used with -f 0.90 and -F 0.10 this requires that either 90% of A is covered OR 10% of B is covered. Without -e, both fractions would have to be satisfied. By default NULL.
S	Logic value to define if to force "strandedness". That is, only report hits in B that overlap A on the same strand. By default, overlaps are reported without respect to strand. By default FALSE.
S	Logic value to define if to require different strandedness. That is, only report hits in B that overlap A on the <code>_opposite_</code> strand. By default, overlaps are reported without respect to strand. By default <code>FALSE</code> .
split	Logic value to define if to treat "split" BAM (i.e., having an "N" CIGAR operation) or BED12 entries as distinct BED intervals. By default FALSE.
sorted	Logic value to define, for very large B files, if to invoke a "sweeping" algorithm that requires position-sorted input. When using -sorted, memory usage remains low even for very large files. By default FALSE. It is possible to sort a bed file on terminal by (sort -k1,1 -k2,2n unsorted.bed > sorted.bed) or by the function sort.bed.
g	Specify a genome file the defines the expected chromosome order in the input files for use with the -sorted option. By default NULL.
srun	Logic value to define whether the command should be run in ${\sf srun}$ mode. By default ${\sf FALSE}.$
<pre>intersect.bedtools.command</pre>	
	String to define the command to use to recall the bedtools intersect function. An example: "/home/user/anaconda3/bin/intersectBed". By default "/home/USERNAME/anaconda3/bin/intersectBed".
return.command	Logic value to define whether to return the string corresponding to the command for bedtools. By default ${\sf FALSE}.$
return.bed	Logic value to define whether to return the resulting bed as data.frame. By default FALSE. Parameter not active when inputs are bam files.
delete.output	Logic value to define whether to delete the exported intersected bed file. By default FALSE. Parameter active only when return.bed = TRUE. Useful when is sufficient to get the result as a data.frame without saving it.
run.command	Logic value to define whether to run the the command line on system terminal and generate the bed resulting from the intersection. By default TRUE.

Details

To know more about the bedtools intersect function see the package manual at the following link:

https://bedtools.readthedocs.io/en/latest/content/tools/intersect.html.

is.color 37

Value

The function generates the files indicated by the output parameters. If required the command line used and/or the resulting intersected bed file. If both outputs are required, the output will be a named list with two values: "command" and "intersected.bed".

Examples

is.color

is.color

Description

Function to define if each element of a string vector is an R-supported color string.

Usage

```
is.color(x)
```

Arguments

Х

A string vector.

Value

A logical vector of the same length of x.

38 mass.to.volume

is.nan_df

is.nan() applied to a data.frame

Description

Applies the function is.nan() to a full data.frame.

Usage

```
is.nan_df(data.frame)
```

Arguments

data.frame

Input data.frame.

Value

It returns a matrix/array containing logic values for each element of the input data.frame. When TRUE it means that the corresponding element is a NaN.

Examples

```
is.nan.df(mtcars)
```

 ${\tt mass.to.volume}$

Get solvent volume to make a solution with a given amount of a compound.

Description

Given a specific ammount of solute calculates the volume of solvent necessary to obtain a certain final molarity concentration.

Usage

```
mass.to.volume(
   final_concentration,
   final_concentration_unit = "M",
   mass,
   mass_unit = "g",
   MW
)
```

molarity.to.mass 39

Arguments

final_concentration

Numeric value for the final concentration wanted.

final_concentration_unit

String to define the unit of the final concentration wanted. Available units

are: "M", "mM", "uM", "nM", "pM", "fM". By default "M".

mass Numeric value for the solute mass ammount.

mass_unit String to define the unit of the mass. Available units are: "kg", "g",

"mg", "ug", "ng". By default "g".

MW Numeric value for the Molecular Weigth (MW) of the compound expressed

in g/mol.

Value

It returns a string with the volume of solvent to use.

Examples

```
mass.to.volume(final_concentration = 5, mass = 10, MW = 215)
```

molarity.to.mass

Get solvent volume to make a solution with a given amount of a compound.

Description

Given a specific volume of solution wanted calculates the mass of solute necessary to obtain a certain final molarity concentration.

Usage

```
molarity.to.mass(
   final_concentration,
   final_concentration_unit = "M",
   final_volume,
   final_volume_unit = "mL",
   MW
)
```

Arguments

final_concentration

Numeric value for the final concentration wanted.

final_concentration_unit

String to define the unit of the final concentration wanted. Available units are: "M", "mM", "uM", "nM", "pM", "fM". By default "M".

40 move.df.col

final_volume Numeric value for the final volume wanted.

final_volume_unit

String to define the unit of the volume. Available units are: "L", "mL",

"uL". By default "mL".

MW Numeric value for the Molecular Weigth (MW) of the compound expressed

in g/mol.

Value

It returns a string with the mass of compound to use.

Examples

```
molarity.to.mass(final_concentration = 5, final_volume = 10, MW = 215)
```

move.df.col

Function to change easily the order of specific columns in a data frame.

Description

Allows to change the position of a column in a data frame using other columns as reference.

Usage

```
move.df.col(data.frame, move.command)
```

Arguments

data.frame An input data.frame.

move.command A string containing the moving command. The command is formed as

follows: "columnA movingCommand columnB". The basic options are: "first", "last", "before", "after". Compounded moves must be separated

by a semicolon. Example: "g first; a last; e before c".

Value

It returns the original data frame but with the columns moved as demanded.

References

https://stackoverflow.com/questions/3369959/moving-columns-within-a-data-frame-without-retyping

pkg.check 41

Examples

```
new.mtcars = move.df.col(mtcars, "mpg last")
new.mtcars = move.df.col(mtcars, "wt before carb")
new.mtcars = move.df.col(mtcars, "am before carb; cyl first")
```

pkg.check

Check package installation.

Description

Function to check if a package is installed. It works with bioconductor or CRAN packages.

Usage

```
pkg.check(package, archive)
```

Arguments

package A single string indicating the name of the package to check.

archive A single string indicating the type of archive. Possible values "CRAN"

and "bioconductor" (not case sensitive). Parameter without default...

Value

If the pkg is not already installed it will be installed.

Examples

```
pkg.check("ggplot2", "cran")
pkg.check("biomaRt", "bioconductor")
```

pkg.version

Get session info and package versions.

Description

Retrieves the information of the current session and the version of the packages loaded.

Usage

```
pkg.version(return.session = F, print.versions = T, return.versions = F)
```

Arguments

```
return.session Logic value to define if to save the session info. By default FALSE.

print.versions Logic value to define if to print the session and version info. By default TRUE.

return.versions
```

Logic value to define if to save package versions info. By default FALSE.

Value

If return.session and/or return.versions TRUE a list with these informations is returned. Otherwise nothing is returned.

```
plot.density.differences
```

Plot the distribution of overall NGS density at specific regions from deepTools matrices.

Description

Computes the score of each element in a list of regions and generates violins plots with percentiles and the mean (optional) for each sample/region. It uses as input a score matrix computed by deeptools's computeMatrix function or by computeMatrix.deeptools and density.matrix functions from this package.

Usage

```
## S3 method for class 'density.differences'
plot(
 matrix.file,
 missing.data.as.zero = NULL,
  sample.names = NULL,
  region.names = NULL,
  signal.type = "mean",
  error.type = "sem",
  subset.range = NULL,
  inverted.comparisons = F,
  stat.method = "wilcox.test",
  stat.paired = T,
  stat.p.levels = list(cutpoints = c(0, 1e-04, 0.001, 0.01, 0.05, 1), symbols =
    c("****", "***", "**", "*", "ns")),
  area.line.width = 0.5,
  area.fill.area = T,
  area.plot.zero.line = T,
  area.y.identical.auto = T,
  area.y.ticks.interval = NULL,
  area.y.digits = 1,
```

```
correlation.log2 = T,
  correlation.plot.correlation = T,
  correlation.correlation.method = "lm",
  correlation.show.equation = T,
  correlation.correlation.line.width = 0.75,
  correlation.correlation.line.color = "purple",
  correlation.correlation.line.type = 1,
  correlation.correlation.line.SE = T,
  correlation.correlation.formula = "y ~ x",
  correlation.add.rug = T,
  correlation.x.identical.auto = T,
  correlation.y.identical.auto = T,
  correlation.x.ticks.interval = NULL,
  correlation.y.ticks.interval = NULL,
  correlation.x.digits = 1,
  correlation.y.digits = 1,
  points.size = 0.5,
  transparency = 0.25,
  axis.line.width = 0.5,
  text.size = 12,
  legend.position = c(0.2, 0.85),
 colors = c(Sample1 = "#F8766D", Sample2 = "#00A5CF", No difference = "#00BA38"),
  n.row.multiplot = 1,
  by.row = T
)
```

Arguments

matrix.file

A single string indicating a full path to a matrix.gz file generated by deepTools/computeMatrix or by computeMatrix.deeptools, or a list generated by the function read.computeMatrix.file or density.matrix.

missing.data.as.zero

Logical value to define whether treat missing data as 0. If set as FALSE missing data will be converted to NA and will be excluded from the computations of the signal. By default TRUE.

sample.names

Samples names could be defined by a string vector. If set as NULL sample names will be get automatically by the matrix file. By default NULL. Example: c("sample1", "sample2", "sample3")

region.names

Region names could be defined by a string vector. If set as NULL sample names will be get automatically by the matrix file. By default NULL. Example: c("regionA", "regionB")

signal.type

String indicating the signal to be computed and plotted/compared. Available parameters are "mean", "median" and "sum". By default "mean".

error.type

String indicating the type of error to be computed and that will be available in the output data.table. Available parameters are "sem" and "sd", standard error mean and standard deviation respectively. By default "sem". Parameter considered only when show.mean = TRUE).

subset.range

A numeric vector indicating the range to which restrict the analyses (eg. c(-150, 250)). In the case of "scale-region" mode, the range is represented by (-upstream — 0 — body_length — body_length+downstream).By default NULL: the whole region is considered.

inverted.comparisons

Logical value to indicate whether to invert the order of the pair-comparisons. By default FALSE.

stat.method

A single string defining the method to use for the statistical comparisons. By default "wilcox.test". Available options: "t.test" "wilcox.test".

stat.paired

Logical value to define if the statistical comparisons should be performed paired. By default TRUE. Notice that to allow a paired comparison the number of data should be the same in the two groups compared, so in the most of the cases non applicable to the comparisons between two regions. Used only in "t.test" and "wilcox.test" methods.

stat.p.levels

A list containing the p-values levels/thresholds in the following format (default): list(cutpoints = c(0,0.0001,0.001,0.01,0.05,1), symbols = c("****","**","**","*","ns")). In other words, we use the following convention for symbols indicating statistical significance:

- ns: p ¿ 0.05
- * p = 0.05
- ** p := 0.01
- *** p j= 0.001
- **** p := 0.0001

area.line.width

Numeric value to define width of the line connecting the points in the area.plots. By default 0.5.

area.fill.area Logical value to indicate whether to fill the area under the line in the area.plot. By default TRUE.

area.plot.zero.line

Logical value to define whether to plot a dashed gray vertical line in correspondence of the 0 of each area.plot. By default TRUE.

area.y.identical.auto

Logical value to define whether use the same Y-axis range for all the area.plots automatically depending on their values. By default TRUE.

area.y.ticks.interval

A number indicating the interval/bin spacing two ticks on the Y-axis of area.plots. By default NULL: ticks are assigned automatically.

area.y.digits Numeric value defining the number of digits to use for the Y-axis values of area.plots. By default 1 (eg. 1.5).

correlation.log2

Logical value to define whether the correlation.plots should show the log2 value of the score. By default TRUE.

correlation.plot.correlation

Local value to indicate whether to plot the correlation curve on the correlation.plot. By default TRUE.

correlation.correlation.method

Atomic string describing the method to use to compute the regression curve, eg. "lm", "glm", "gam", "loess", "rlm". By default 'lm'.

correlation.show.equation

= T

correlation.correlation.line.width

Numeric value to define correlation line width for all correlation. plots. By default 0.75.

correlation.correlation.line.color

Numeric value to define correlation line width for all correlation.plots. By default "purple".

correlation.correlation.line.type

A numeric or character value to define the correlation line type. Both numeric and string codes are accepted. By default "solid".

correlation.correlation.line.SE

Logical value to indicate whether to plot the standard error (SE) of the correlation curve in the correlation plot. By default TRUE.

correlation.correlation.formula

Atomic string indicating the formula to use to compute the correlation curve. By default " $y \sim x$ ".

correlation.add.rug

Logical value to indicate whether to add a rug representation (1-d plot) of the data to the correlation.plot. By default TRUE.

correlation.x.identical.auto

Logical value to define whether use the same X-axis range for all the correlation plots automatically depending on their values. By default TRUE.

correlation.y.identical.auto

Logical value to define whether use the same Y-axis range for all the correlation.plots automatically depending on their values. By default TRUE.

correlation.x.ticks.interval

A number indicating the interval/bin spacing two ticks on the X-axis of correlation.plots. By default NULL: ticks are assigned automatically.

correlation.y.ticks.interval

A number indicating the interval/bin spacing two ticks on the Y-axis of correlation.plots. By default NULL: ticks are assigned automatically.

correlation.x.digits

Numeric value defining the number of digits to use for the X-axis values of correlation.plots. By default 1 (eg. 1.5).

correlation.y.digits

Numeric value defining the number of digits to use for the Y-axis values of correlation.plots. By default 1 (eg. 1.5).

points.size A numeric value defining the size of the points in both area and correlation plot. By default 0.5.

A numeric value to define the fraction of transparency of the fill area in the area.plot and the SE in the correlation plot (0 = transparent, 1 = full). By default 0.25.

axis.line.width

Numeric value to define the axes and ticks line width for all plots. By default 0.5.

text.size

Numeric value to define the size of the text for the labels of all the plots. By default 12.

legend.position

Any ggplot supported value for the legend position (eg. "none, "top", "bottom", "left", "right", c(fraction.x, fraction.y)). By default c(0.2,0.85).

colors

Vector of 3 elements to define the points and area colors ('Sample1', 'Sample2' and, 'No difference' values respectively). If only one value is provided it will applied to all the samples. If the number of values is less then 3, the default color set will be used. All supported R.colors values are accepted. By default c("Sample1" = "#F8766D", "Sample2" = "#00A5CF", "No difference" = "#00BA38").

n.row.multiplot

Numeric value to define the number of rows in the final multiplot.

by.row

Logical value to define whether the plots should be arranged by row. By default TRUE.

Details

To know more about the deepTools's function computeMatrix see the package manual at the following link:

https://deeptools.readthedocs.io/en/develop/content/tools/computeMatrix.html.

Value

The function returns a list containing:

- data.table with the computed values with all groups and all samples;
- metadata table with the information obtained from the matrix_file.gz;
- comparison.table.list with a list of tables for each group with a table per each comparison containing the original data and the compared values (differences);
- comparison.statistics.table with a table with all the statistical comparisons;
- area.plot.byGroup.list with a list per group with a all the area.plots of each comparison;
- correlation.plot.byGroup.list with a list per group with a all the correlation.plots of each comparison;
- area.multiplot.list with an area.multiplot per each group;
- correlation.multiplot.list with an correlation.multiplot per each group.

plot.density.profile 47

 $\begin{array}{ll} {\it plot.} {\it density.profile} & {\it Plot~of~NGS~density~signal~at~specific~regions~from~deep Tools} \\ & {\it matrices.} \end{array}$

Description

Plots the density profile of NGS data signals, using as input a score matrix computed by deeptools's computeMatrix function or by computeMatrix.deeptools and density.matrix functions from this package.

Usage

```
## S3 method for class 'density.profile'
plot(
 matrix.file,
  plot.by.group = T,
  missing.data.as.zero = NULL,
  sample.names = NULL,
  region.names = NULL,
  signal.type = "mean",
  error.type = "sem",
  plot.error = T,
  error.transparency = 0.125,
  title = NULL,
  x.lab = NULL,
  y.lab = NULL,
  line.type = "solid",
  line.width = 0.5,
  x.lim = NULL,
  y.lim = NULL,
  y.identical.auto = T,
  y.ticks.interval = NULL,
  y.digits = 1,
  axis.line.width = 0.5,
  text.size = 12,
  legend.position = c(0.2, 0.85),
  plot.vertical.lines = T,
  write.reference.points = T,
  colors = c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00",
    "#FF61C9", "gray30"),
  n.row.multiplot = 1,
  multiplot.export.file = NULL,
  real.width.single.plot = 2.9,
  real.height.single.plot = 3.5,
  by.row = TRUE,
  print.multiplot = F
)
```

48 plot.density.profile

Arguments

matrix.file A single string indicating a full path to a matrix.gz file generated by deepTools/computeMatrix or by computeMatrix.deeptools, or a list gen-

erated by the function read.computeMatrix.file or density.matrix.

 $\verb|plot.by.group| Logical value to define whether plot by group of regions or by sample. By$

default TRUE.

missing.data.as.zero

Logical value to define whether treat missing data as 0. If set as FALSE missing data will be converted to NA and will be excluded from the com-

putations of the signal. By default TRUE.

sample.names Samples names could be defined by a string vector. If set as NULL sample

names will be get automatically by the matrix file. By default NULL.

Example: c("sample1", "sample2", "sample3")

region.names Region names could be defined by a string vector. If set as NULL sample

names will be get automatically by the matrix file. By default $\ensuremath{\mathsf{NULL}}$.

Example: c("regionA", "regionB")

signal.type String indicating the signal to be computed and plotted. Available pa-

rameters are "mean", "median" and "sum". By default "mean".

error.type String indicating the type of error to be computed and plotted. Available

parameters are "sem" and "sd", standard error mean and standard deviation respectively. By default "sem". Parameter considered only when

plot.error = TRUE).

plot.error Logical value to define whether to plot the error around the signal. By

default TRUE.

error.transparency

Numeric value to define the alpha/transparency of the error. By default

0.125. Parameter considered only when plot.error = TRUE).

title Title of each plot could be defined by a string vector. If set as NULL titles

will be generated automatically. By default NULL.

Example: c("Title1", "Title2")

x.lab Single string or string vector to define the X-axis label for all the plots.

By default NULL, the label will be defined automatically.

y.lab Single string or string vector to define the Y-axis label for all the plots.

By default NULL, the label will be defined automatically.

line.type Vector to define each line type. Both numeric and string codes are ac-

cepted. If only one element is given this will be applied to all the lines.

By default "solid".

Example 1: c("solid", "dashed").

Example 2: c(1,2)

line.width Numeric value to define the line width for all the plots. By default 0.5.

x.lim List of numeric vectors with two elements each to define the range of the

X-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default

NULL, the range will be defined automatically.

Example list(c(0,20),c(NA,30),c(0,NA),c(NA,NA)).

plot.density.profile 49

y.lim

List of numeric vectors with two elements each to define the range of the Y-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default NULL, the range will be defined automatically.

Example list(c(0,20),c(NA,30),c(0,NA),c(NA,NA)).

y.identical.auto

Logical value to define whether use the same Y-axis range for all the plots automatically depending on the values. Not used when y.lim is not NULL. By default TRUE.

y.ticks.interval

A number indicating the interval/bin spacing two ticks on the Y-axis. By default NULL: ticks are assigned automatically. Active only when y.identical.auto = TRUE and y.lim != NULL.

y.digits

A numeric value to define the number of digits to use for the y.axis values. By default 1 (eg. 1.5).

axis.line.width

Numeric value to define the axes and ticks line width for all plots. By default 0.5.

text.size

Numeric value to define the size of the text for the labels of all the plots. By default 12.

legend.position

Any ggplot supported value for the legend position (eg. "none", top", "bottom", "left", "right", c(fraction.x, fraction.y)). By default c(0.2,0.85).

plot.vertical.lines

Logical value to define whether to plot a dashed gray vertical line in correspondence of the reference points of each plot. By default TRUE.

write.reference.points

Logical value to define whether to indicate the reference points on each plot. Applied only when x.lim is NULL. By default TRUE.

colors

Vector to define the line and error area colors. If only one value is provided it will applied to all the samples/groups. If the number of values is lower than the the required one, a random set of colors will be generated. All standard R.colors values are accepted. By default c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00", "#FF61C9", "gray30").

n.row.multiplot

Numeric value to define the number of rows in the final multiplot.

multiplot.export.file

If a string with the name of a PDF file is provided the multiplot will be exported. By default NULL.

real.width.single.plot

Numeric value, in inches, to define the real width of each plot in the multiplot exported, if required. By default 2.9 inches.

real.height.single.plot

Numeric value, in inches, to define the real height of each plot in the multiplot exported, if required. By default 3.5 inches.

by.row Logical value to define whether the plots should be arranged by row. By default TRUE.

print.multiplot

Logical value to define whether to print the multiplot once created. By default FALSE.

Details

To know more about the deepTools's function computeMatrix see the package manual at the following link:

https://deeptools.readthedocs.io/en/develop/content/tools/computeMatrix.html.

Value

The function returns a list containing:

- data.table with the computed values used for the plot;
- metadata table with the information gotten from the matrix_file.gz;
- plot.list with a plot for each list element;
- multiplot with the image of all the plots together.

Examples

```
plot.density.profile(
   matrix.file = "/path.to/matrix.file.gz", plot.by.group = TRUE,
   missing.data.as.zero = NULL, sample.names = NULL, region.names = NULL,
   signal.type = "mean", error.type = "sem", plot.error = TRUE,
   error.transparency = 0.125, title = NULL, x.lab = NULL, y.lab = NULL,
   line.type = "solid", line.width = 0.5, x.lim = NULL, y.lim = NULL,
   y.identical.auto = TRUE, y.ticks.number = 5, text.size = 12,
   plot.vertical.lines = TRUE, colors = c("red", "blue", "#00BA38"),
   n.row.multiplot = 1, multiplot.export.file = "/path.to/multiplot.pdf",
   real.width.single.plot = 2.5, real.height.single.plot = 3,
   print.multiplot = FALSE)
```

plot.density.profile.smooth

Plot of NGS density signal at specific regions from deepTools matrices (signal smoothing version).

Description

Plots the density profile of NGS data signals, using as input a score matrix computed by deeptools's computeMatrix function or by computeMatrix.deeptools and density.matrix functions from this package (signal smoothing version). The error on the line cannot be plotted in this case. See also plot.density.profile.

Usage

```
## S3 method for class 'density.profile.smooth'
plot(
  matrix.file,
  plot.by.group = T,
  missing.data.as.zero = NULL,
  sample.names = NULL,
  region.names = NULL,
  signal.type = "mean",
  smooth.span = 0.1,
  title = NULL,
  x.lab = NULL,
  y.lab = NULL,
  line.type = "solid",
  line.width = 0.5,
  x.lim = NULL,
  y.lim = NULL,
  y.identical.auto = T,
  y.ticks.interval = NULL,
  y.digits = 1,
  axis.line.width = 0.5,
  text.size = 12,
  legend.position = c(0.2, 0.85),
  plot.vertical.lines = T,
  write.reference.points = T,
  colors = c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00",
    "#FF61C9", "gray30"),
  n.row.multiplot = 1,
  multiplot.export.file = NULL,
  real.width.single.plot = 2.9,
  real.height.single.plot = 3.5,
  by.row = TRUE,
  print.multiplot = F
)
```

Arguments

Matrix.file A single string indicating a full path to a matrix.gz file generated by deepTools/computeMatrix or by computeMatrix.deeptools, or a list generated by the function read.computeMatrix.file or density.matrix.

plot.by.group Logical value to define whether plot by group of regions or by sample. By default TRUE.

missing.data.as.zero

Logical value to define whether treat missing data as 0. If set as FALSE missing data will be converted to NA and will be excluded from the computations of the signal. By default TRUE.

sample.names Samples names could be defined by a string vector. If set as NULL sample names will be get automatically by the matrix file. By default NULL.

Example: c("sample1", "sample2", "sample3")

 $\textbf{region.names} \qquad \text{Region names could be defined by a string vector. If set as $NULL$ sample}$

names will be get automatically by the matrix file. By default NULL.

Example: c("regionA", "regionB")

signal.type String indicating the signal to be computed and plotted. Available pa-

rameters are "mean", "median" and "sum". By default "mean".

smooth.span Numerical value to indicate the span value for the loess function used to

smooth bigWig signals. By default 0.1.

title Title of each plot could be defined by a string vector. If set as NULL titles

will be generated automatically. By default NULL.

Example: c("Title1", "Title2")

x.lab Single string or string vector to define the X-axis label for all the plots.

By default NULL, the label will be defined automatically.

y.lab Single string or string vector to define the Y-axis label for all the plots.

By default NULL, the label will be defined automatically.

line.type Vector to define each line type. Both numeric and string codes are ac-

cepted. If only one element is given this will be applied to all the lines.

By default "solid".

Example 1: c("solid", "dashed").

Example 2: c(1,2)

line.width Numeric value to define the line width for all the plots. By default 0.5.

x.lim List of numeric vectors with two elements each to define the range of the

X-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default

NULL, the range will be defined automatically.

Example list(c(0,20),c(NA,30),c(0,NA),c(NA,NA)).

y.lim List of numeric vectors with two elements each to define the range of the

Y-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default

NULL, the range will be defined automatically.

Example list(c(0,20),c(NA,30),c(0,NA),c(NA,NA)).

y.identical.auto

Logical value to define whether use the same Y-axis range for all the plots automatically depending on the values. Not used when y.lim is not NULL.

By default TRUE.

y.ticks.interval

A number indicating the interval/bin spacing two ticks on the Y-axis. By default NULL: ticks are assigned automatically. Active only when

y.identical.auto = TRUE and y.lim != NULL.

y.digits A numeric value to define the number of digits to use for the y.axis values.

By default 1 (eg. 1.5).

axis.line.width

Numeric value to define the axes and ticks line width for all plots. By

default 0.5.

text.size

Numeric value to define the size of the text for the labels of all the plots. By default 12.

legend.position

Any ggplot supported value for the legend position (eg. "none", top", "bottom", "left", "right", c(fraction.x, fraction.y)). By default c(0.2,0.85).

plot.vertical.lines

Logical value to define whether to plot a dashed gray vertical line in correspondence of the reference points of each plot. By default TRUE.

write.reference.points

Logical value to define whether to indicate the reference points on each plot. Applied only when x.lim is NULL. By default TRUE.

colors

Vector to define the line and error area colors. If only one value is provided it will applied to all the samples/groups. If the number of values is lower than the the required one, a random set of colors will be generated. All standard R.colors values are accepted. By default c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00", "#FF61C9", "gray30").

n.row.multiplot

Numeric value to define the number of rows in the final multiplot.

multiplot.export.file

If a string with the name of a PDF file is provided the multiplot will be exported. By default NULL.

real.width.single.plot

Numeric value, in inches, to define the real width of each plot in the multiplot exported, if required. By default 2.9 inches.

real.height.single.plot

Numeric value, in inches, to define the real height of each plot in the multiplot exported, if required. By default 3.5 inches.

by.row Logical value to define whether the plots should be arranged by row. By default TRUE.

print.multiplot

Logical value to define whether to print the multiplot once created. By default FALSE.

Details

To know more about the deepTools's function computeMatrix see the package manual at the following link:

https://deeptools.readthedocs.io/en/develop/content/tools/computeMatrix.html.

Value

The function returns a list containing:

- data.table with the computed values used for the plot;
- metadata table with the information gotten from the matrix_file.gz;
- plot.list with a plot for each list element;
- multiplot with the image of all the plots together.

Examples

```
plot.density.profile.smooth(
   matrix.file = "/path.to/matrix.file.gz", plot.by.group = TRUE,
   missing.data.as.zero = NULL, sample.names = NULL, region.names = NULL,
   signal.type = "mean", error.type = "sem", plot.error = TRUE,
   error.transparency = 0.125, title = NULL, x.lab = NULL, y.lab = NULL,
   line.type = "solid", line.width = 0.5, x.lim = NULL, y.lim = NULL,
   y.identical.auto = TRUE, y.ticks.number = 5, text.size = 12,
   plot.vertical.lines = TRUE, colors = c("red", "blue", "#00BA38"),
   n.row.multiplot = 1, multiplot.export.file = "/path.to/multiplot.pdf",
   real.width.single.plot = 2.5, real.height.single.plot = 3,
   print.multiplot = FALSE)
```

plot.density.summary $Plot\ the\ distribution\ of\ overall\ NGS\ density\ at\ specific\ regions$ from deep Tools matrices.

Description

Computes the score of each element in a list of regions and generates violins plots with percentiles and the mean (optional) for each sample/region. It uses as input a score matrix computed by deeptools's computeMatrix function or by computeMatrix.deeptools and density.matrix functions from this package.

Usage

```
## S3 method for class 'density.summary'
plot(
 matrix.file,
 plot.by.group = T,
 missing.data.as.zero = NULL,
  sample.names = NULL,
  region.names = NULL,
  signal.type = "mean",
  linear = F,
  error.type = "sem",
  show.mean = T,
 mean.error.type = "se",
 mean.color = "blue",
 mean.symbol.shape = 20,
 mean.symbol.size = 1,
  show.stat.multiplot = T,
  stat.method = "wilcox.test",
  stat.paired = F,
  stat.labels.format = "p.signif",
  stat.hide.ns = T,
  stat.p.levels = list(cutpoints = c(0, 1e-04, 0.001, 0.01, 0.05, 1), symbols =
```

plot.density.summary

```
c("****", "***", "**", "s")),
  title = NULL,
  x.lab = NULL
 y.lab = NULL,
  x.labs.angle = 0,
  dodge.width = 1,
  border.width = 0.5,
  border.color = "#000000",
  transparency = 0.5,
  subset.range = NULL,
 y.lim = NULL,
 y.identical.auto = T,
 y.ticks.interval = NULL,
 y.digits = 1,
  axis.line.width = 0.5,
  text.size = 12,
  legend.position = c(0.2, 0.85),
  colors = c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00",
    "#FF61C9", "gray30"),
  n.row.multiplot = 1,
 multiplot.export.file = NULL,
  real.width.single.violinplot = 1,
  real.height.single.violinplot = 3.5,
  by.row = TRUE,
  print.multiplot = F
)
```

Arguments

matrix.file A single string indicating a full path to a matrix.gz file generated by

deepTools/computeMatrix or by computeMatrix.deeptools, or a list generated by the function read.computeMatrix.file or density.matrix.

plot.by.group Logical value to define whether plot by group of regions or by sample. By default TRUE.

missing.data.as.zero

Logical value to define whether treat missing data as 0. If set as FALSE missing data will be converted to NA and will be excluded from the computations of the signal. By default TRUE.

sample.names Samples names could be defined by a string vector. If set as NULL sample

names will be get automatically by the matrix file. By default NULL.

Example: c("sample1", "sample2", "sample3")

region.names Region names could be defined by a string vector. If set as NULL sample names will be get automatically by the matrix file. By default NULL.

Example: c("regionA", "regionB")

signal.type String indicating the signal to be computed and plotted. Available pa-

rameters are "mean", "median" and "sum". By default "mean".

linear Logical value to define whether the plots should show the score in linear

scale. By default FALSE.

error.type

String indicating the type of error to be computed and that will be available in the output data.table. Available parameters are "sem" and "sd", standard error mean and standard deviation respectively. By default "sem". Parameter considered only when show.mean = TRUE).

show.mean

Logical value to define whether the mean value should be shown as a symbol on the plots. By default TRUE.

mean.error.type

String indicating the type of error for the mean to be computed. Available parameters are "se", "sd" and, "none". Respectively standard error, standard deviation, and no error plotted. By default "se". Parameter considered only when show.mean = TRUE).

mean.color

A single string expressing an R-supported color for the mean symbol. By default "blue".

mean.symbol.shape

A numeric value or string defining the shape for the mean symbol. By default 20.

mean.symbol.size

A numeric value defining the size of the mean symbol. By default 1.

show.stat.multiplot

Logical value to define if to add to the plot the statistical comparisons of the means for the groups present in the multiplot. By default TRUE. All possibile comparisons will be performed.

stat.method

A single string defining the method to use for the statistical comparisons. By default "wilcox.test". Available options: "t.test" "wilcox.test".

stat.paired

Logical value to define if the statistical comparisons should be performed paired. By default "FALSE". Notice that to allow a paired comparison the number of data should be the same in the two groups compared, so in the most of the cases non applicable to the comparisons between two regions. Used only in "t.test" and "wilcox.test" methods.

stat.labels.format

A single string indicating the format of the p-value to show for the statistical comparisons. By default "p.signif". Available options: "p.format" (normal p-value), "p.signif" (significance stars), "p.adj" (p-value adjusted).

stat.hide.ns

Logical value indicating if the NS ("Not Significant") comparisons should be shown or not. By default TRUE.

stat.p.levels

A list containing the p-values levels/thresholds in the following format (default): list(cutpoints = c(0,0.0001,0.001,0.01,0.05,1), symbols = c("****","***","**","**","ns")). In other words, we use the following convention for symbols indicating statistical significance:

- ns: p ; 0.05
- * p = 0.05
- ** p := 0.01
- *** p := 0.001
- **** p = 0.0001

title	Title of each plot could be defined by a string vector. If set as NULL titles will be generated automatically. By default NULL. Example: c("Title1", "Title2")	
x.lab	Single string or string vector to define the X-axis label for all the plots. By default NULL, the label will be defined automatically.	
y.lab	Single string or string vector to define the Y-axis label for all the plots. By default NULL, the label will be defined automatically.	
x.labs.angle	A single numeric value indicating the degrees of rotation of the category labels in the X-axis. By default \emptyset , horizontal without rotation.	
dodge.width	Numeric value defining the width of each single violin plot. By default 1.	
border.width	Numeric value to define the border width for all the violin plots. By default $\emptyset.5$.	
border.color	A single string indicating the color to use for the border of the violin plots. By default "#000000" (full black).	
transparency	A numeric value to define the fraction of transparency of the plots fill (0 = transparent, 1 = full). By default 0.5 .	
subset.range	A numeric vector indicating the range to which restrict the analyses (eg. c(-150, 250)). In the case of "scale-region" mode, the range is represented by (-upstream — 0 — body_length — body_length+downstream). By default NULL: the whole region is considered.	
y.lim	List of numeric vectors with two elements each to define the range of the Y-axis. To set only one side use NA for the side to leave automatic. If only one range is given this one will be applied to all the plots. By default NULL, the range will be defined automatically. Example list($c(0,20)$, $c(NA,30)$, $c(0,NA)$, $c(NA,NA)$).	
y.identical.auto		
	Logical value to define whether use the same Y-axis range for all the plots automatically depending on the values. Not used when y.lim is not NULL. By default TRUE.	
v ticke interval		

y.ticks.interval

A number indicating the interval/bin spacing two ticks on the Y-axis. By default NULL: ticks are assigned automatically. Active only when y.identical.auto = TRUE and y.lim != NULL.

y.digits A numeric value to define the number of digits to use for the y.axis values. By default 1 (eg. 1.5).

axis.line.width

Numeric value to define the axes and ticks line width for all plots. By default 0.5.

text.size Numeric value to define the size of the text for the labels of all the plots. By default 12.

legend.position

Any ggplot supported value for the legend position (eg. "none, "top", "bottom", "left", "right", c(fraction.x, fraction.y)). By default c(0.2,0.85).

colors

Vector to define the line and error area colors. If only one value is provided it will applied to all the samples/groups. If the number of values is lower than the the required one, a random set of colors will be generated. All standard R.colors values are accepted. By default c("#00A5CF", "#F8766D", "#AC88FF", "#E08B00", "#00BA38", "#BB9D00", "#FF61C9", "gray30").

n.row.multiplot

Numeric value to define the number of rows in the final multiplot.

multiplot.export.file

If a string with the name of a PDF file is provided the multiplot will be exported. By default NULL.

real.width.single.violinplot

Numeric value, in inches, to define the real width (not precise) of each single violin plot in the multiplot exported, if required. By default 1 inch.

real.height.single.violinplot

Numeric value, in inches, to define the real height (not precise) of each single violin plot in the multiplot exported, if required. By default 3.5 inches

by.row Logical value to define whether the plots should be arranged by row. By default TRUE.

print.multiplot

Logical value to define whether to print the multiplot once generated. By default FALSE.

Details

To know more about the deepTools's function computeMatrix see the package manual at the following link:

https://deeptools.readthedocs.io/en/develop/content/tools/computeMatrix.html.

Value

The function returns a list containing:

- data.table with the computed values used for the plot;
- metadata table with the information obtained from the matrix_file.gz;
- plot.list with a plot for each list element;
- density.profile with the density profile of the mean signal generated by plot.density.profile corresponding to the regions/samples for which the summary multiplot have been generated;
- multiplot with the image of all the plots together;
- summary.plot.samples with a plot showing the scores of all regions per each sample;
- summary.plot.regions with a plot showing the scores of all samples per each region;
- means.comparisons table with the statistical means comparisons (when show.stat.multiplot = TRUE, otherwise a string is returned).

pStars 59

|--|

Description

Converts a p-value score in equivalent stars of significance.

Usage

```
pStars(p.value, one = 0.05, two = 0.01, three = 0.001, four = 1e-04)
```

Arguments

p.value	A single numeric value indicating the p-value to evaluate.
one	A numeric value to define the p-value threshold for the first level of significance (*). By default 0.05 .
two	A numeric value to define the p-value threshold for the second level of significance (**). By default 0.01 .
three	A numeric value to define the p-value threshold for the third level of significance (***). By default 0.001 .
four	A numeric value to define the p-value threshold for the fourth level of significance (****). By default 0.0001.

Value

It returns a string with the corresponding level of significance: NS, *, **, ***.

Examples

```
significance = pStars(0.002)

require(dplyr)
data.frame =
   data.frame %>%
   mutate(p.stars = sapply(data.frame$p.value.column, pStars))
```

qPCR.rna.exp

qPCR.rna.exp

Merger of overlapping peaks in a provided .bed file.

Description

Merge overlapping peaks in a provided .bed file.

Usage

```
qPCR.rna.exp(
  results.file,
  housekeeping.genes = NULL,
 max.delta.reps = 0.5,
  reference.sample = NULL,
  exlude.houskeeping.FC = TRUE,
  exclude.samples = NULL,
  fix.y.axis = FALSE,
  x.labels.rotation = 45,
  text.size = 3,
  results.sheet.position = 3,
  rows.to.skip = 44,
  file.header = TRUE,
  file.tail = TRUE,
  samples.order = NULL,
  ignore.reps.errors = FALSE
)
```

Arguments

results.file String indicating the full path to the results excel file or a data.frame containing at least the following columns: 'Sample Name', 'Target Name', 'CT'.

housekeeping.genes

String vector with the list of genes that have to be used as target genes. By default NULL: an error message is printed.

max.delta.reps Numeric value indicating the maximum difference among replicate Ct. Default value: 0.5.

reference.sample

Single string indicating the name of the sample to use as reference for the computation of the FoldChanges. By default NULL: the first sample in the order is used as reference.

exlude.houskeeping.FC

Logic value to indicate whether the housekeeping genes should be excluded in the FoldChanges plots. By default TRUE.

exclude.samples

String vector indicating the samples that should be exuded in the expression and FoldChange plots. By default NULL.

qPCR.rna.exp 61

fix.y.axis Logic value indicating whether the y-axis of the plots should be kept fixed among all the genes. By default FALSE.

x.labels.rotation

Numeric value indicating the degrees of x-axis's labels rotation. By default 45.

text.size Numeric value to indicate the size of the text for the number above the bars. Default 3.

results.sheet.position

Numeric value indicating the position of the results sheet in the excel file. by default 3.

rows.to.skip How many rows must be skipped before to read the excel file. By default 44.

file.header Logic value to indicate whether the results excel file contains an header. By default TRUE.

file.tail Logic value to indicate whether the results excel file contains extra rows at the end of the results. By default TRUE.

samples.order A string vector indicating all the samples in order. This order will be used to order the samples in the plots. By default NULL: the reference sample will be the first, the other will be kept in the order available in the results table.

ignore.reps.errors

Logic value to define whether the difference between the Ct in replicates should be ignored: all the values are kept.

Value

The function returns a list containing:

- original.table: a data.frame containing the original results table;
- reshaped.table: a data.frame with the original results reorganized for the analyses;
- reshaped.table.cleaned: the reshaped data.frame upon filtering of the CT values (if required);
- reps.validation.plot: a plot representing a table with the differences two-by-two of the technical replicates (facet_wrapped by gene) where the cells have a red background if the difference is greater than the 'max.delta.reps' value;
- analyzed.data: a named list of data.frames, one for each housekeeping gene and one for the foldChange mean of all housekeeping normalization, containing the normalized expression scores and the FoldChanges over the reference sample;
- expression.plots: a named list of plots, one for each housekeeping gene, showing the gene expression histograms (facet_wrapped by gene);
- foldChange.plots: a named list of plots, one for each housekeeping gene and one for the foldChange mean of all housekeeping normalization, showing the FoldChange expression over the reference Sample (facet_wrapped by gene).

62 restore_packages

```
read.computeMatrix.file
```

computeMatrix *.gz file reader

Description

The function reads a matrix.file.gz generated by deepTools/computeMatrix function or by computeMatrix.deeptools. The value can be passed to plot.density.profile function.

Usage

```
read.computeMatrix.file(matrix.file)
```

Arguments

matrix.file

A string indicating indicating the full path to the matrix.file.gz generated by deepTools/computeMatrix function or by computeMatrix.deeptools.

Value

The functions returns a named list containing:

- metadatadata.frame with the information gotten from the matrix_file.gz
- matrix.datadata.frame with the scores gotten from
- original.file.pathwith full path to the original matrix_file.gz.

This list can be passed as it is to the function plot.density.profile.

restore_packages

Restores packages installed from a .rda file.

Description

Installs the packages contained in a .rda file. This file can be generated by the store_packages function of this package.

Usage

```
restore_packages(rda_file)
```

Arguments

rda_file

Path to the .rda from which get the information for the packages to reinstall.

Value

If it was not possible to re-install al packages, the list of not restored packages will be returned.

restriction.sites.to.bed 63

```
restriction.sites.to.bed
```

Generator of a bed file for enzymatic restriction sites.

Description

The function allows to create a bed file that can be added on IGV both as regions and track. It will show the restriction sites of a sequences if starting from the cut positions depending on sequence length. Chromosome, start and end of the input sequence are required.

Usage

```
restriction.sites.to.bed(
   cut_positions,
   chromosome,
   genome_start,
   return_bed = TRUE,
   export_bed_file = FALSE,
   output_file_name = paste(getwd(), "restriction_positions.bed", sep = "/"),
   enzyme_cut_length = 4,
   include_region_description = TRUE,
   region_name = "site",
   append = FALSE
)
```

Arguments

cut_positions A numeric vector with the list of the restriction/cut positions.

chromosome Chromosome number of the region analyzed.

genome_start Start position on the genome of the region analyzed.

return_bed Logic value to define if to return the bed as data.frame. By default TRUE. export_bed_file Logic value to define if to export the resulting .bed file. By default FALSE.

output_file_name

String corresponding to the path to the exported .bed file. By default "<working.directory>/restriction_positions.bed".

enzyme_cut_length

Numeric value to define the length of cut of the restriction enzyme. By default 4.

include_region_description

Logic value to define whether to include a fourth column containing the region name define by the parameter region_description. By default TRUE

region_name Regions base name. Automatically it will be added a number to the base

name. By default "site", the resulting regions will be: site_1, site_2,

append Logic value to define if to append the result to the file. By default FALSE,

the file will be overwritten.

RNAseq

Details

To map the positions of restriction enzymes it is possible to use http://restrictionmapper.org/ with the option Map (version 3).

Value

If required, it will be returned a classic bed file (chr start end [name]) with the regions centered on the cut position in the genome.

Examples

RNAseq

RNA-seq example

Description

Dummy example of a DESeq2 result for differential expression analysis on RNA-seq data

Usage

RNAseq

Format

A data frame with 300 rows and 7 variables:

```
geneName genes symbols
```

baseMean The average of the normalized count values, dividing by size factors, taken over all samples

log2FC the log2 value of the Fold Change expression between two conditions

1fcSE log2 Fold Change standard error (SE)

stat Wald statistic

pvalue Wald test p-value

padj BH adjusted p-values

Source

Simulated data

sort.bed 65

sort.bed

Sorter function for .bed files.

Description

Sorts .bed files by chromosome and position.

Usage

```
## S3 method for class 'bed'
sort(
  bed,
  bed.header = FALSE,
  sep = "\t",
  return.bed = TRUE,
  export.file.name = NULL,
  export.header = FALSE,
  unique.regions = TRUE
)
```

Arguments

bed Two options are possible:

- String with the path to a .bed file;

- Data.frame corresponding to a bed file format (all the columns and their

names will be kept).

bed.header Logic value to define whether the .bed file contains an header or not. By

default FALSE.

sep String containing the separator character for a .bed file. By default "\t".

return.bed Logic value to define if to return the bed as a data.frame. By default

TRUE. Only unique rows are kept.

export.file.name

Optional: string to define the path to the file to be exported, if required.

By default NULL, not exported.

export.header Logic value to define whether the header should be exported in the sorted

bed file. By default FALSE.

unique.regions Logic value to indicate whether the output bed must contain unique re-

gions. By default TRUE.

Details

The function keeps only unique rows.

To get more information about the bed file format see the following page:

https://genome.ucsc.edu/FAQ/FAQformat.html#format1.

66 substract.bw

Value

If required, returns a data frame corresponding to the sorted .bed file.

store_packages

Stores the information of installed packages in a .rda file.

Description

Saves the list of all the installed packages in a .rda file. This file can be used to restore the packages from a computer to another or after installation of a new R version by the function restore_packages of this package.

Usage

```
store_packages(output_directory = getwd())
```

Arguments

output_directory

Path to the directory in which export the .rda file. By default <working.directory>.

Value

Nothing is returned. An .rda file will be exported at the output_directory indicated.

substract.bw

Combination of two or more list in a unique one.

Description

Combines two or more lists in a single one keeping the elements names

Usage

```
substract.bw(bw1, bw2, return.substracted.bw = T, substracted.bw.file = NULL)
```

Arguments

bw1 Full path to the first bigWig (the second one will be substracted to this

one).

bw2 Full path to the second bigWig (it will be substracted to the first one).

return.substracted.bw

Logic value to define whether return the resulting bigWig as GRanges

object. By default TRUE.

substracted.bw.file

String for the path of the resulting bigwig file to be exported.

By default NULL, any file will be exported.

uniform.x.axis 67

Value

If required a subtraction bigWig is returned as GRanges object. The resulting bigWig can be also directly exported.

uniform.x.axis

Plot X-axis uniforming

Description

Takes a list of ggplot2 plots, compares their X-axis ranges and applies the highest/lowest limits to each plot in order to uniform all the plots. It can be used also to set the ticks step (to just change the breaks set all parameters as FALSE).

Usage

```
uniform.x.axis(
  plot.list,
  x.min = TRUE,
  x.max = TRUE,
  ticks.each = NULL,
  digits = 1
)
```

Arguments

plot.list	A single plot or a list of plots.
x.min	Either a logical value to define whether uniform the lower limit or a numeric value defining the lower limit. By default $TRUE$.
x.max	Either a logical value to define whether uniform the upper limit or a numeric value defining the upper limit. By default TRUE.
ticks.each	Numeric value to define every how much should be placed a tick. By default NULL, ticks will be placed automatically.
digits	A single integer indicating the maximum number of digits required for the rounding of the axis values. By default 1.

Value

Returns a plot list (or a single plot when only one input plot is provided) equivalent to the input list provided by the user in which the X-axis of all the plots will be uniformed.

68 update_pkgs

uniform.y.axis

Plot Y-axis uniforming

Description

Takes a list of ggplot2 plots, compares their Y-axis ranges and applies the highest/lowest limits to each plot in order to uniform all the plots. It can be used also to set the ticks step (to just change the breaks set all parameters as FALSE).

Usage

```
uniform.y.axis(
  plot.list,
  y.min = TRUE,
  y.max = TRUE,
  ticks.each = NULL,
  digits = 1
)
```

Arguments

plot.list	A single plot or a list of plots.
y.min	Either a logical value to define whether uniform the lower limit or a numeric value defining the lower limit. By default TRUE.
y.max	Either a logical value to define whether uniform the upper limit or a numeric value defining the upper limit. By default TRUE.
ticks.each	Numeric value to define every how much should be placed a tick. By default NULL, ticks will be placed automatically.
digits	A single integer indicating the maximum number of digits required for the rounding of the axis values. By default 1.

Value

Returns a plot list (or a single plot when only one input plot is provided) equivalent to the input list provided by the user in which the Y-axis of all the plots will be uniformed.

update_pkgs

function to automatically update the R packages.

Description

Automatically updates the R packages from CRAN and BioConductor repositories.

Usage

```
update_pkgs(ask = FALSE)
```

Arguments

ask

Logical indicating whether to ask the user to select packages before they are downloaded and installed, or the character string "graphics", which brings up a widget to allow the user to (de-)select from the list of packages which could be updated. (The latter value only works on systems with a GUI version of select.list, and is otherwise equivalent to ask = TRUE). By default FALSE.

Value

Nothing. The packages will be updated.

Examples

```
update_pkgs()
```

volcano

Volcano plot generator for RNA-seq data.

Description

Generates a volcano plot in order to visualize the differentially expressed genes. The plot is highly customizable.

Usage

```
volcano(
  log2FC_data,
  padj_data,
 FC_t = 1.5,
  p_t = 0.05
  FC_unresponsive_rigth = 1.1,
  FC_unresponsive_left = 1/FC_unresponsive_rigth,
 x_{ends} = NULL,
 y_min = 0,
  y_max = NULL,
  left_label = "UP",
  right_label = "DOWN",
  unresponsive_label = "NoResp",
  null_label = "NULL",
  names = as.character(c(1:length(log2FC_data))),
  left_names = FALSE,
  right_names = FALSE,
  padding = FALSE,
  names_size = 10,
  print_plot = F,
```

```
left_color = "#00BA38"
  right_color = "#F8766D",
  unresponsive_color = "#00A5CF",
  null_color = "gray30",
  point_size = 0.5,
  legend = TRUE,
  legend_title = "Expression status",
  x_label = bquote("log"["2"] * "(Fold Change expression)"),
 y_label = bquote("-log"["10"] * "(p-value"["adjusted"] * ")"),
  title = "Volcano plot",
  sub_title = NULL,
  add_threshold_lines = T,
  threshold_line_color = "gray70",
  threshold_line_type = "dotted",
  font_family = "Helvetica",
  font_size = 12
)
```

Arguments

log2FC_data Numeric vector containing the log2(FoldChange) values of each gene.

padj_data Numeric vector of p-values. Use of adjusted p-values is recommended.

FC_t Value of the threshold to use for the fold change expression to define differentially expressed genes, expressed as linear value. By default 1.5

and by consequence 1/1.5.

p_t Value of the threshold to use for the p-values to define differentially ex-

pressed genes, expressed as linear value. By default 0.05.

FC_unresponsive_rigth

Value of the threshold to use for the fold change expression to define unresponsive genes when FC > 1, expressed as linear value. By default 1.1. If NULL it will be calculated symmetrically from FC_NoResp_left as 1/FC_NoResp_left.

FC_unresponsive_left

Value of the threshold to use for the fold change expression to define unresponsive genes when FC < 1, expressed as linear value. By default 1/FC_unresponsive_rigth. If NULL it will be calculated symmetrically from FC_NoResp_rigth as 1/FC_NoResp_rigth.

x_ends Numeric positive value to define manually the range of the X-axis: it

will be calculated as $c(-x_ends,x_ends)$, for this reason the plot will be symmetrical. By default NULL, the range is assigned automatically and

the plot can be asymmetrical.

y_min Numeric value for the minimum value of the Y-axis. By default 0. Set it

to NULL for automatic computation.

y_max Numeric value for the maximum value of the Y-axis. By default NULL.

left_label String to indicate the label to use for the set of genes in the left side of the graph (those with FoldChange < 1/FC_t and p.value < p_t. By default

"UP".

right_label String to indicate the label to use for the set of genes in the right side of the graph (those with FoldChange > FC_t and p.value < p_t. By default "DOWN". unresponsive_label String to indicate the label to use for the set of unresponsive genes (those with FC_unresponsive_left < FoldChange < FC_unresponsive_rigth and p.value > p_t. By default "NoResp". null_label String to indicate the label to use for the set of null genes (those with 1/FC_t < FoldChange < FC_t and p.value < p_t. By default "NULL". String vector with the names to be plotted if required, eg. gene names. names By default as.character(c(1:length(log2FC_data))). left_names Logic value to indicate if to print the set of differentially expressed genes in the left side of the graph (those with FoldChange < 1/FC_t and p.value $< p_t$. By default FALSE. right_names Logic value to indicate if to print the set of differentially expressed genes in the right side of the graph (those with FoldChange $> FC_t$ and p.value < p_t. By default FALSE. padding Logic value to indicate if to plot the padding around the names of genes. By default FALSE. Numeric value to define de size of the point names size. By default 10. names_size Logic value to define whether to print the volcano plot once created. By print_plot default FALSE. left_color String to indicate the color to use for the set of genes in the left side of the graph (those with FoldChange < 1/FC_t and p.value < p_t. By default "#00BA38", a green. right_color String to indicate the color to use for the set of genes in the right side of the graph (those with FoldChange $> FC_t$ and p.value $< p_t$. By default "#F8766D", a pink/red. unresponsive_color String to indicate the color to use for the set of unresponsive genes (those with FC_unresponsive_left < FoldChange < FC_unresponsive_rigth and $p.value > p_t$. By default "#00A5CF", a light blue. null_color String to indicate the color to use for the set of null genes (those with 1/FC_t < FoldChange < FC_t and p.value < p_t. By default "gray30", a dark gray. point_size Numeric value to define de size of the points. By default 0.5. Logic value to define if to print the legend. By default TRUE. legend legend_title A string to indicate the label of the legend title. By default "Expression status". x_label A string to indicate the X-axis label. By default "log2(fold change expression)". A string to indicate the Y-axis label. By default "-log10(p-value adjusted)". y_label title A string to indicate the title of the plot. By default "Volcano plot".

sub_title A string to indicate the subtitle of the plot. By default NULL, no subtitle is written.

add_threshold_lines

Logic value to define if lines for the thresholds, both FC and p.value, should be plotted. By default TRUE .

threshold_line_color

String to define the color of the threshold lines. By default $"{\tt gray70"}$

threshold_line_type

String or numeric value to define the threshold lines type. Both numeric and string standard R codes are accepted. By default "dotted", equiva-

lent to 2.

font_family String to define the font family to use in the plot writings. By default

"Helvetica".

font_size Numeric value to define the font size. By default 12.

Value

A plot in ggplot2 format.