

RECOMB 2011 Satellite Workshop on Massively Parallel Sequencing (RECOMB-seq)
26-27 March 2011, Vancouver, BC, Canada; Short talk: 2011-03-27 12:10-12:30 (presentation: 15 minutes, questions: 5 minutes)
Slides available online at http://boisvert.info/dropbox/recomb-seq-2011-talk.pdf, version: 2011-03-23-1

Constrained traversal of repeats with paired sequences

Sébastien Boisvert, Élénie Godzaridis, François Laviolette & Jacques Corbeil



Department of Molecular Medicine

Department of Computer Science and Software Engineering

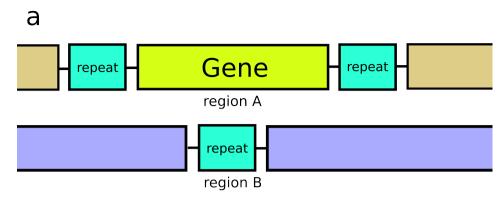


Biological usefulness:

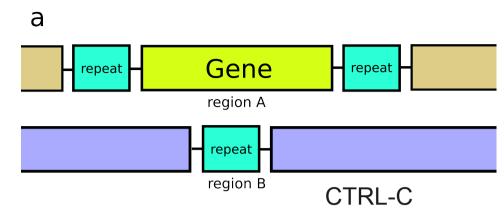
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- → ease copy-and-paste events in genomes

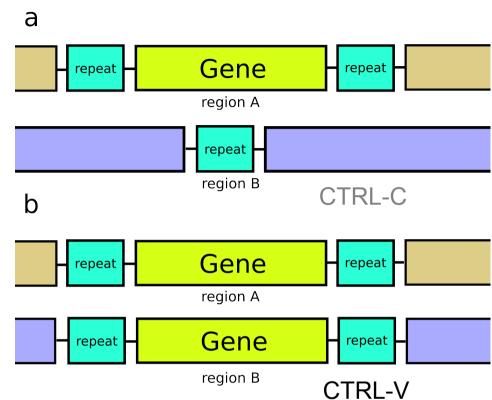
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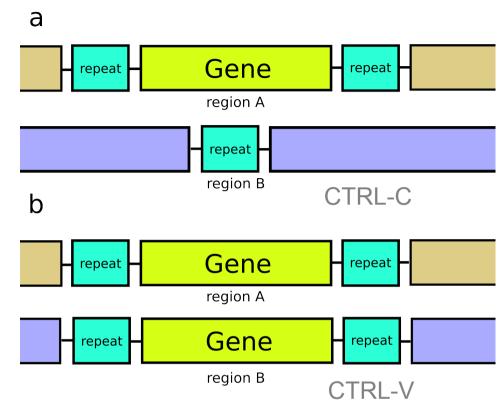


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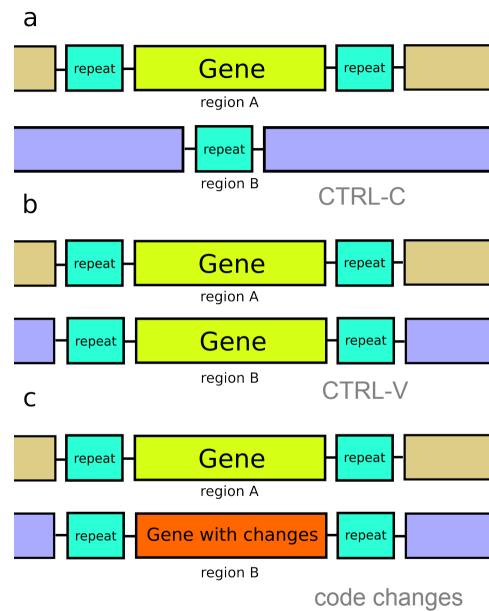
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code changes

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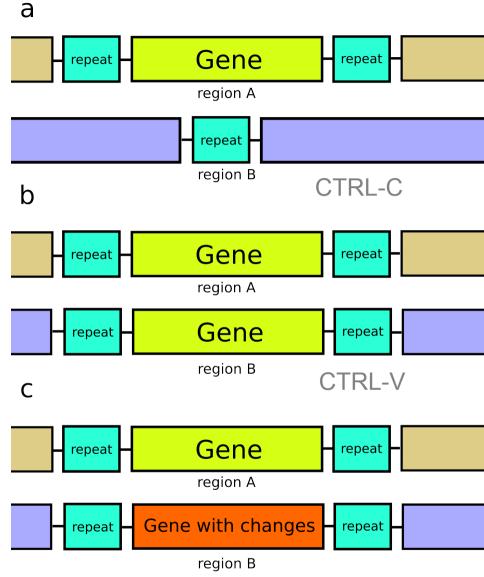


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- → role in 'gene innovation'
- → ease copy-and-paste events in genomes

Data analysis point-of-view:

→ often collapsed by assembly algorithms



code changes

Limitations of next-generation genome sequence assembly Alkan, Can and Sajjadian, Saba and Eichler, Evan E. *Nature Methods*, 2011

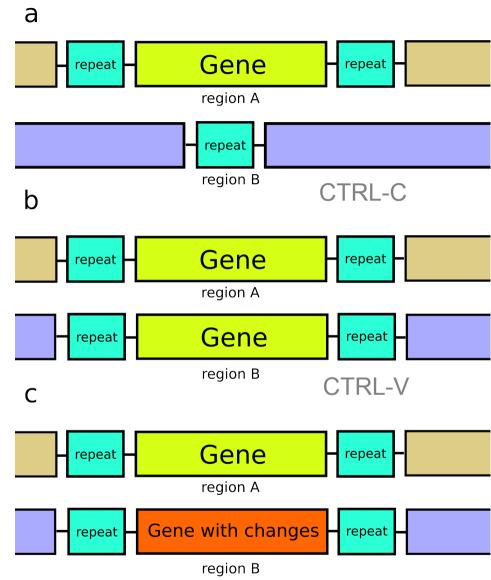
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Biological usefulness:

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- → ease copy-and-paste events in genomes

Data analysis point-of-view:

- → often collapsed by assembly algorithms
- → source of misassemblies



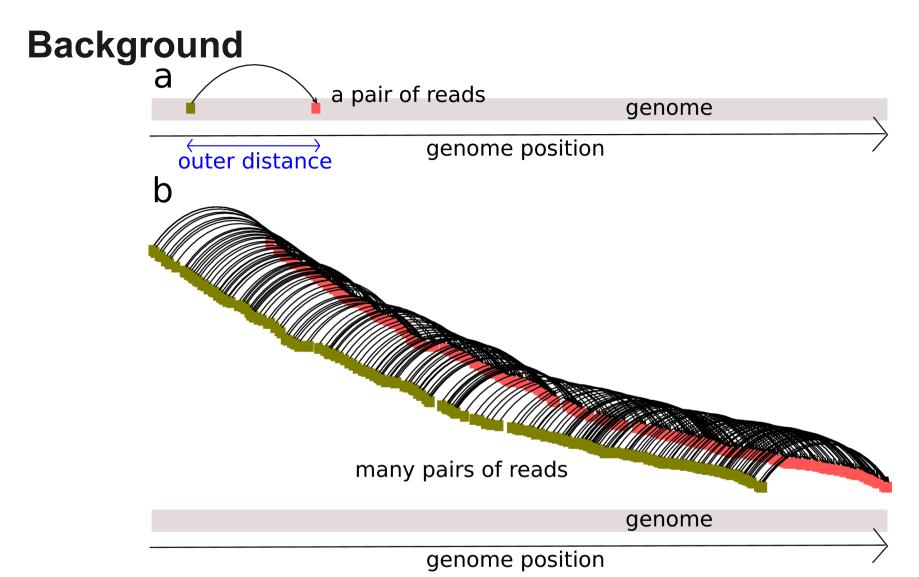
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Background

→ DNA sequence: contains blueprints of biological systems (genome)



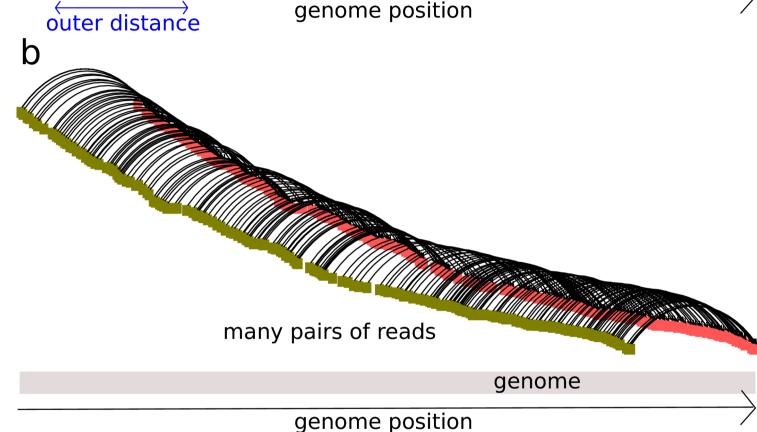
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- → Goal: get genome sequence from short (50-100 nt) paired reads

Background

a pair of reads

genome

genome position



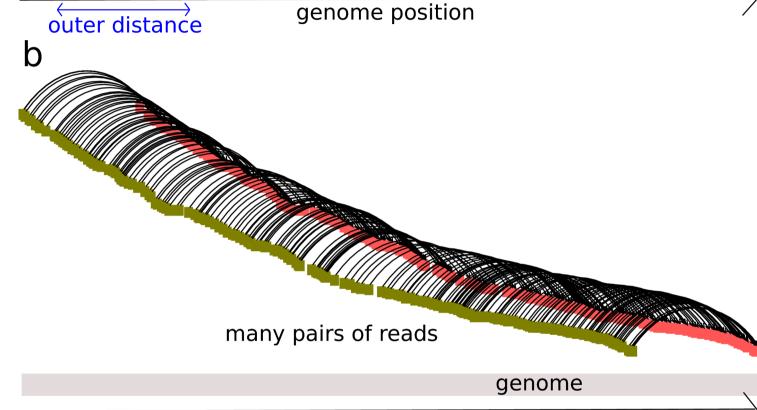
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- → DNA sequence: contains blueprints of biological systems (genome)
- → Goal: get genome sequence from short (50-100 nt) paired reads
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- → Outer distances of pairs: not constant, randomly distributed

Seminal work

→ Next-generation DNA sequencing deluge started ~2005

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- → How to assemble reads into genomes?

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"we cut the existing pieces of a puzzle into even smaller pieces of regular shape"

→ From the landmark paper by Pevzner et al.:

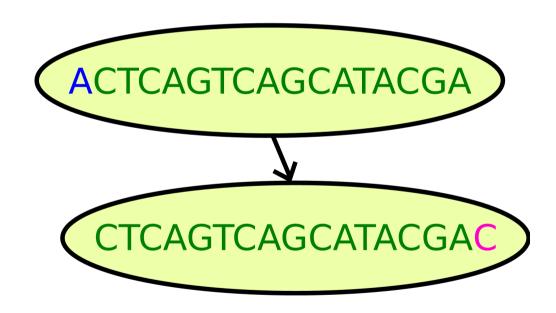
An Eulerian path approach to DNA fragment assembly Pevzner, Pavel A. and Tang, Haixu and Waterman, Michael S. *Proceedings of the National Academy of Sciences*, 2001 http://dx.doi.org/doi:10.1073/pnas.171285098

→ A graph G: vertices V(G) & arcs $E(G) \subseteq V(G) \times V(G)$

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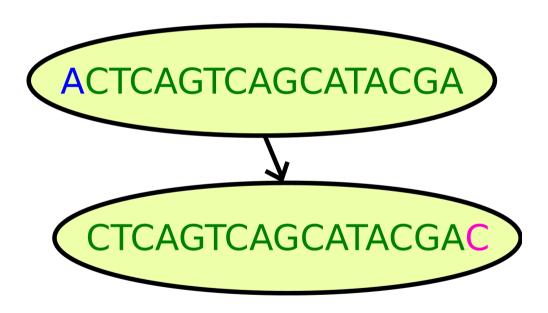
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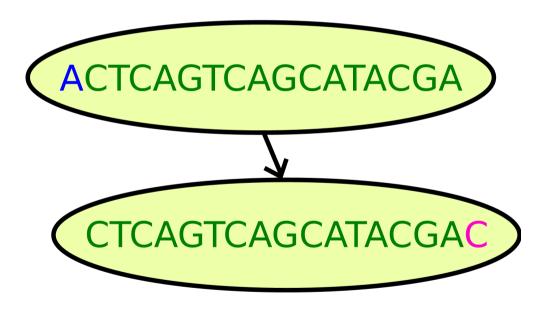
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→ Subgraph of the de Bruijn graph

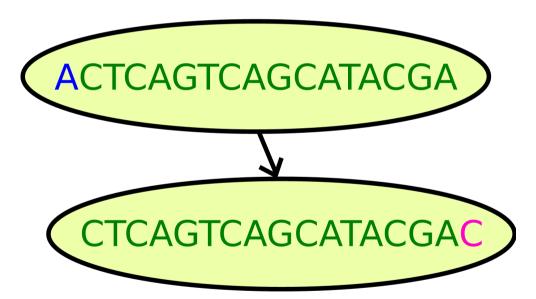


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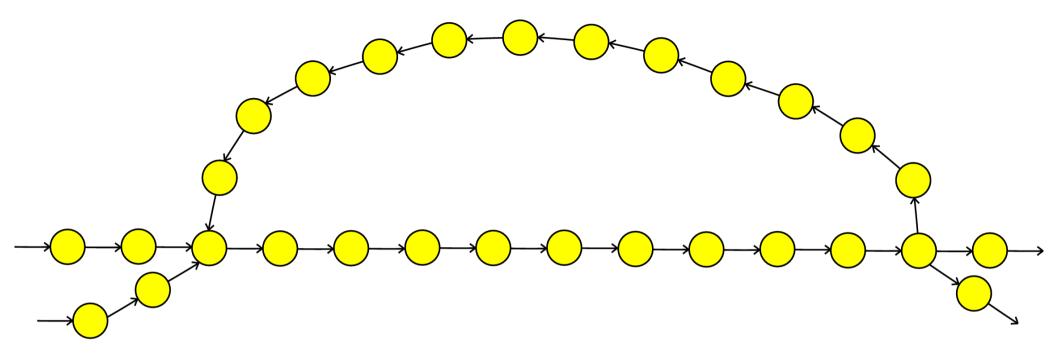


- → Subgraph of the de Bruijn graph
- → Genome: path in this graph
- → Owing to repeats, genome is a set of paths

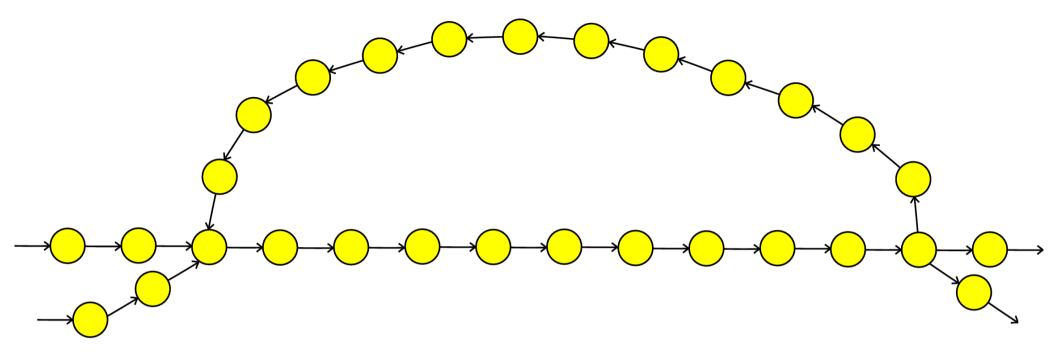
Read paths

GCTACGGAATAAAACCAGGAACAACAGACCCAGCAC **GCTACGGAATAAAACCAGGAA CTACGGAATAAAACCAGGAAC** TACGGAATAAAACCAGGAACA **ACGGAATAAAACCAGGAACAA CGGAATAAAACCAGGAACAAC GGAATAAAACCAGGAACAACA GAATAAAACCAGGAACAACAG AATAAAACCAGGAACAACAGA ATAAAACCAGGAACAACAGAC** TAAAACCAGGAACAACAGACC AAAACCAGGAACAACAGACCC **AAACCAGGAACAACAGACCCA AACCAGGAACAACAGACCCAG ACCAGGAACAACAGACCCAGC CCAGGAACAACAGACCCAGCA** CAGGAACAACAGACCCAGCAC

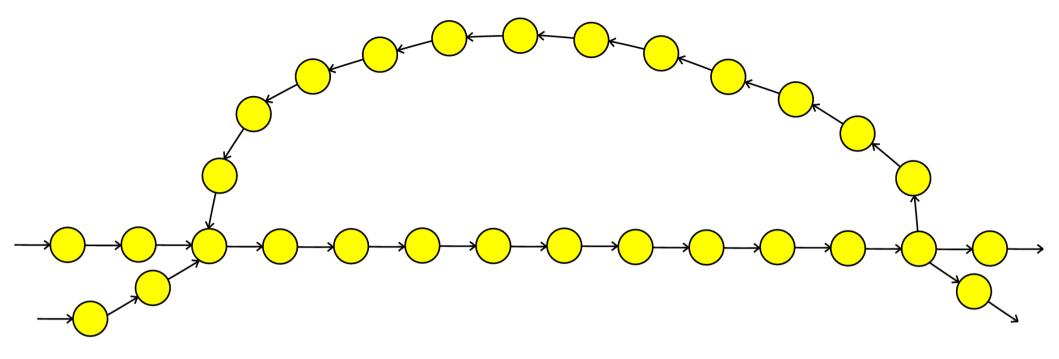
[→] A read is a path



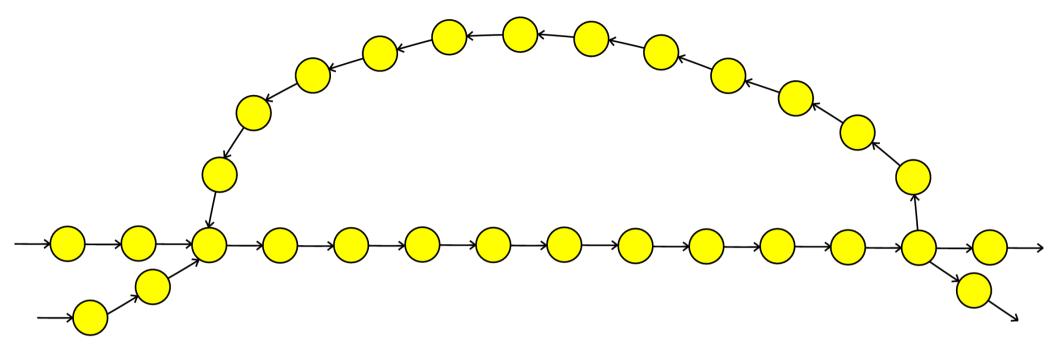
→ 2 entry points & 2 exit points



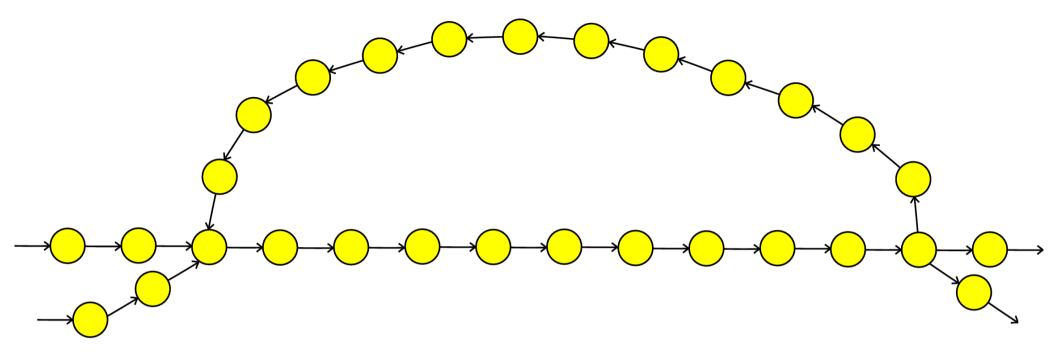
- → 2 entry points & 2 exit points
- → Matching entry & exit points



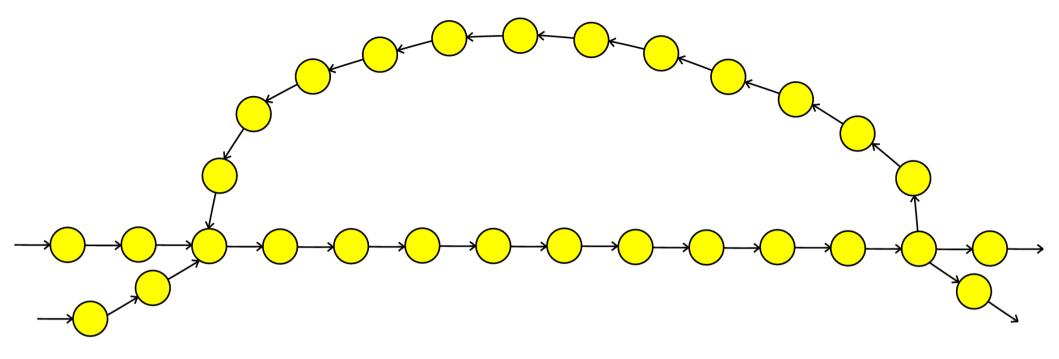
- → 2 entry points & 2 exit points
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- → How many copies ?



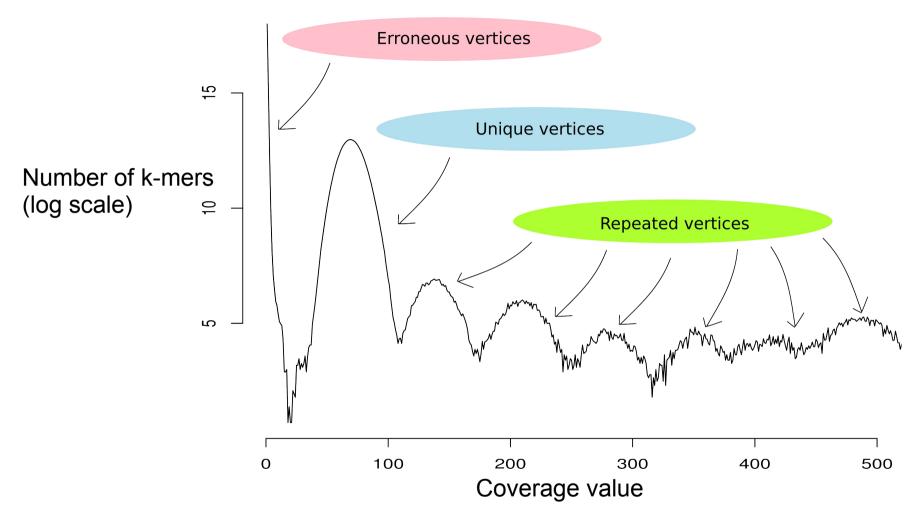
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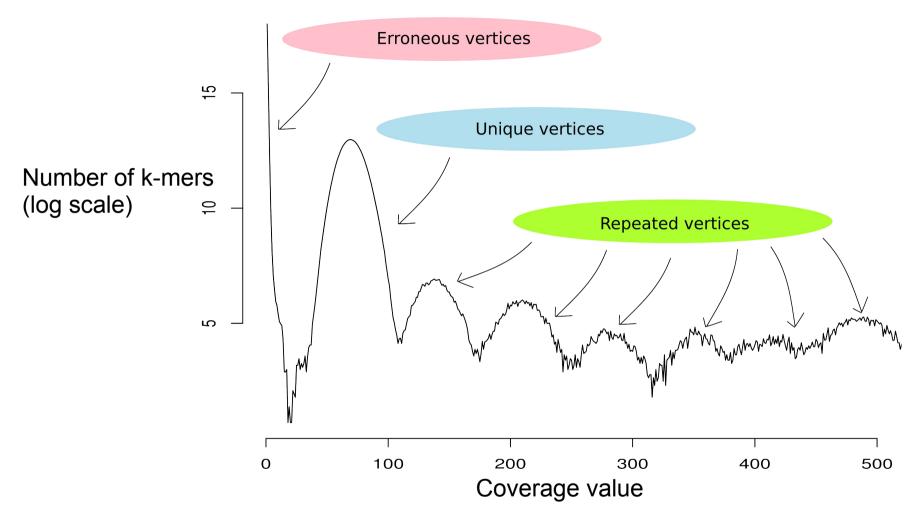


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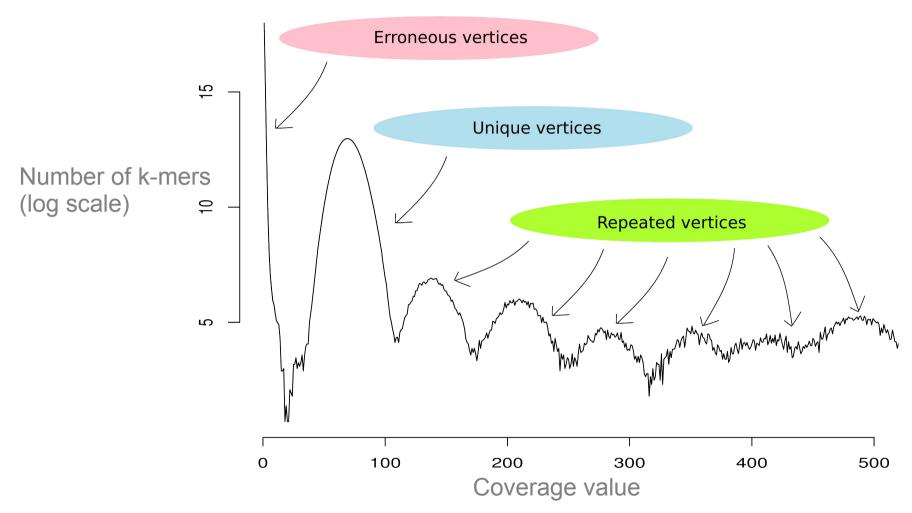


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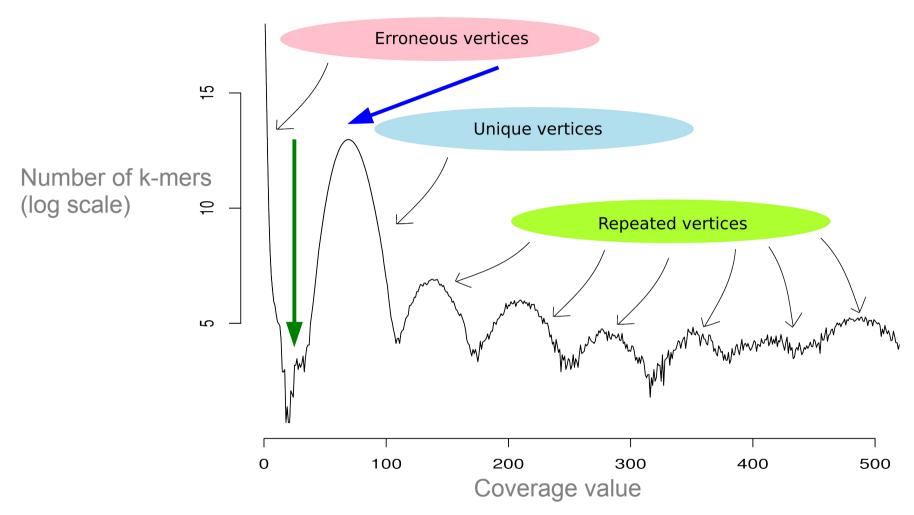




→ Numerous errors, not redundant; many are there only once



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- → Redundant genome k-mers



- → Numerous errors, not redundant; many are there only once
- → Redundant genome k-mers
- → From example: minimum coverage= 15, peak coverage= 69

Initial aim

→ Develop a powerful, parallel, *de novo* assembler able to handle reads from a mix of technologies

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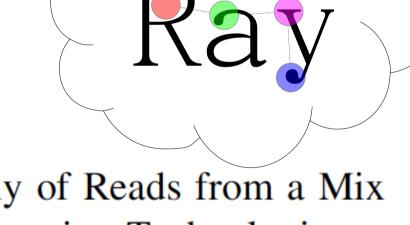
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Volume 17, Number 11, 2010

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Pp. 1519-1533

DOI: 10.1089/cmb.2009.0238



Ray: Simultaneous Assembly of Reads from a Mix of High-Throughput Sequencing Technologies

SÉBASTIEN BOISVERT,1,2 FRANÇOIS LAVIOLETTE,3 and JACQUES CORBEIL1,2

http://dx.doi.org/doi:10.1089/cmb.2009.0238

Other aims

→ Improve our assembler to traverse repeated regions more efficiently



Other aims

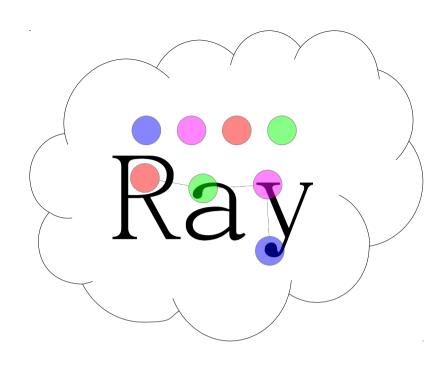
- → Improve our assembler to traverse repeated regions more efficiently
- → Assemble large genomes: <u>de novo assembly of Illumina CEO genome in 11.5 h on 512 processors with Ray</u> (Supplementary slides)



→ Computes seeds in the graph



- → Computes seeds in the graph
- → Extends them using pairs of sequences



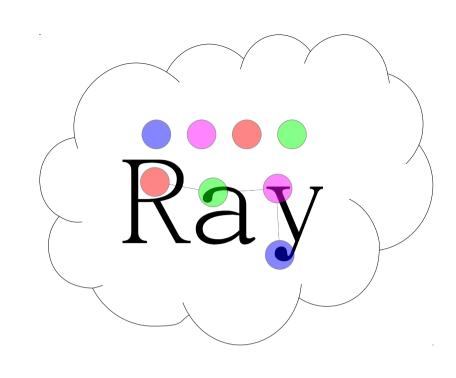
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- → Max. 4 choices for extension: A, T, C, or G
- → Selection is heuristic-based



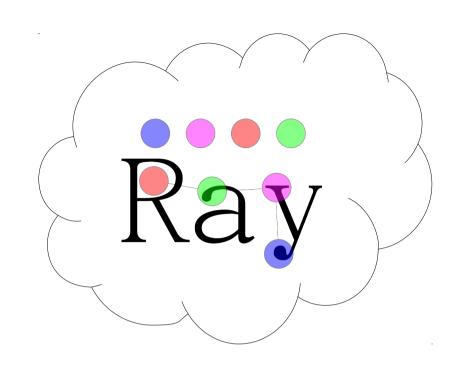
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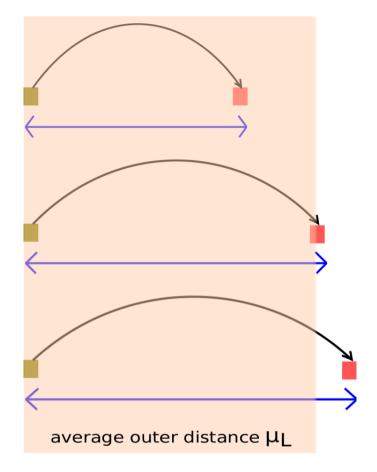
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- → Assembly result: no Ns only As, Ts, Cs and Gs
- → Message-Passing Interface (MPI); open system http://tiny.cc/ray-assembler
- → Cloud-ready (needs MPI + fast interconnect)



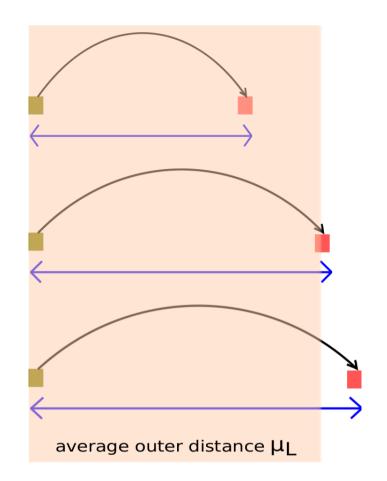
Paired reads



- → For any paired library L:
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Paired reads

→ Chaisson et al. (2009) transform a pair in meta-read



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De novo fragment assembly with short mate-paired reads: Does the read length matter? Chaisson, Mark J. and Brinza, Dumitru and Pevzner, Pavel A.

Genome Research, 2009

http://dx.doi.org/doi:10.1101/gr.079053.108

Paired reads

average outer distance LI

- → Chaisson et al. (2009) transform a pair in meta-read
- → in Ray: a pair is untransformed, used in synergy with other pairs
- → For any paired library L:
- \rightarrow average outer distance μ_L
- $ilde{m{\sigma}}$ standard deviation $m{\sigma}_L$

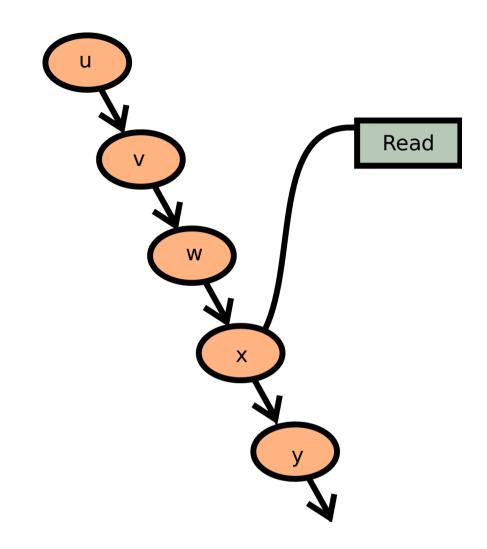
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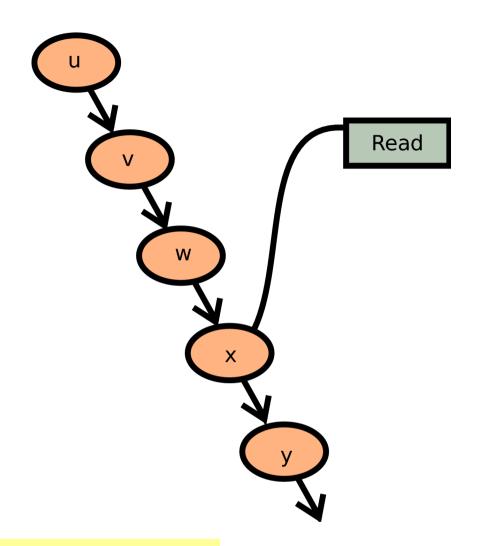
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11 / 21

→ List of reads for a k-mer



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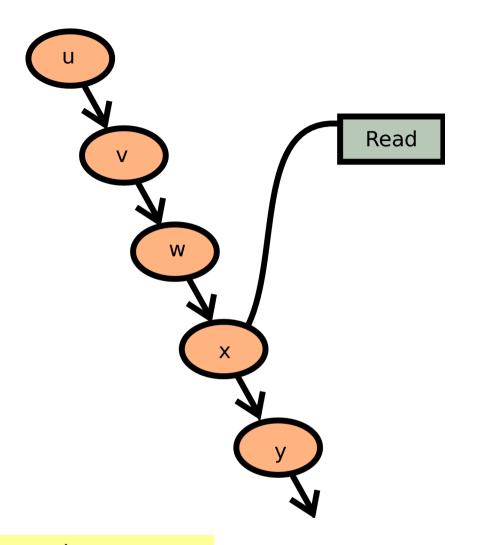


Genome assembly and comparison using de Bruijn graphs Zerbino, Daniel R.

PhD thesis, University of Cambridge, 2009 http://www.ebi.ac.uk/training/ftp/PhDtheses/Daniel_Zerbino.pdf

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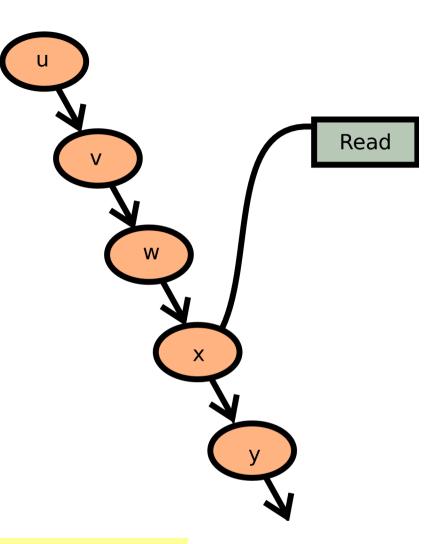


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- → List of reads for a k-mer
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- → In Velvet, read offset is 0 (k-mer position in read)
- → Not optimal, often a read starts on a repeated vertex
- → but may also contains unique vertices



Genome assembly and comparison using de Bruijn graphs Zerbino, Daniel R.

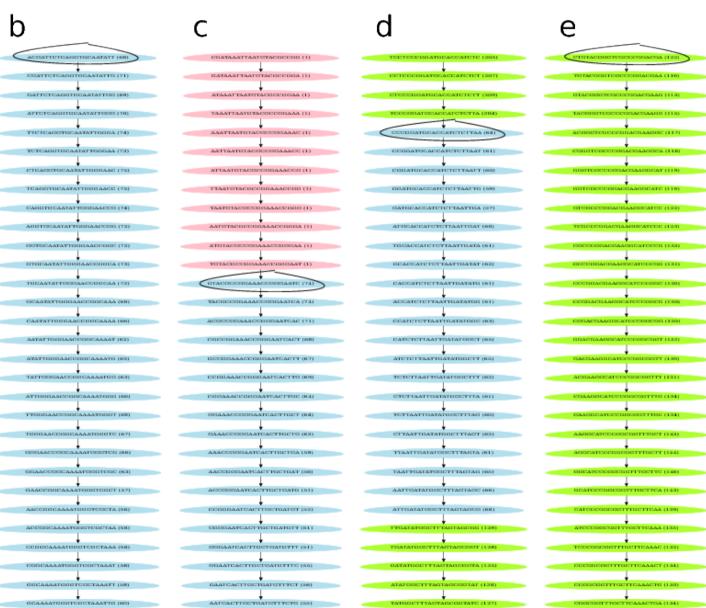
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→ 4 read path examples



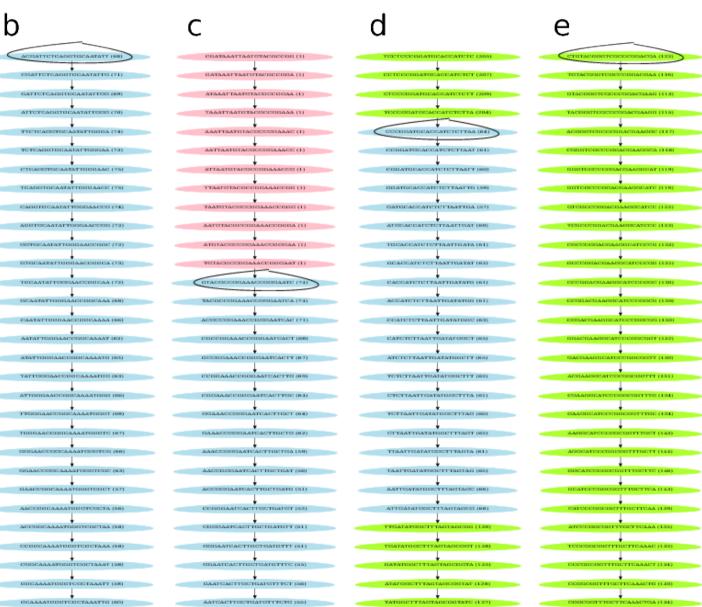
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→ 4 read path examples

→ Blue: unique

→ Pink: erroneous

→ Green: repeated



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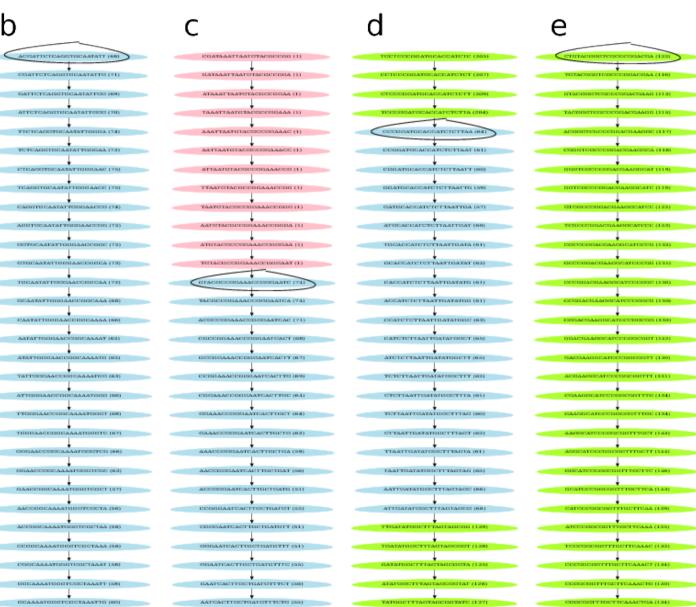


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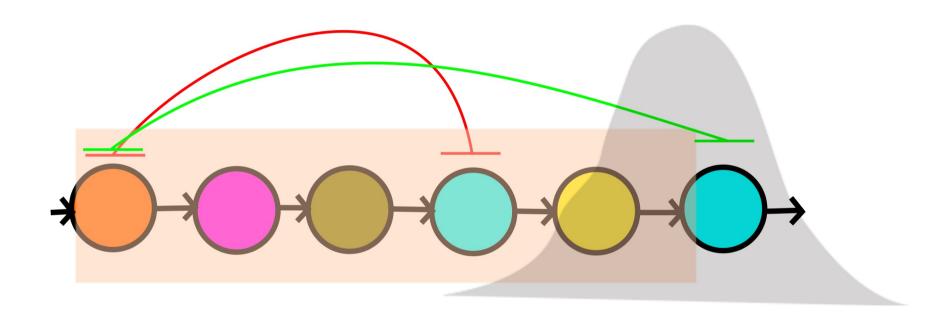
→ Green: repeated

→ Circle: read markers



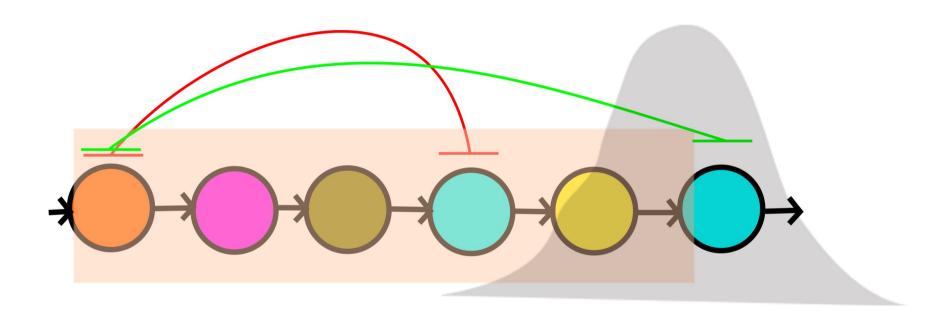
Constraint 1 -- acceptable outer distances

→ Outer distance must be within 3 standard deviations from the average



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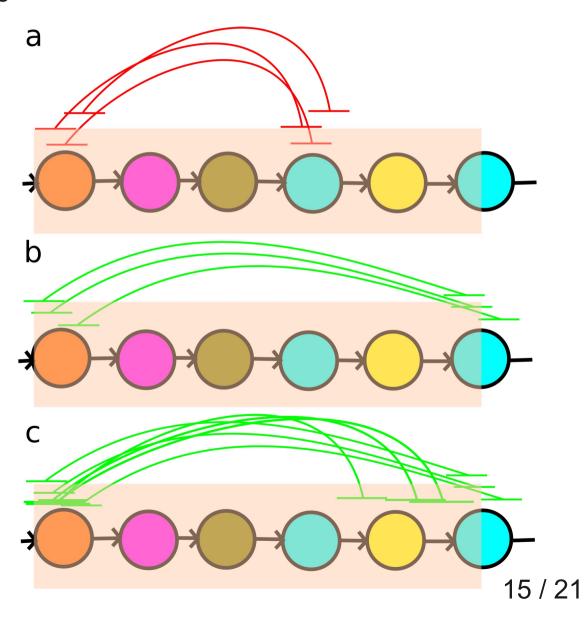
→ Outer distance must be within 3 standard deviations from the average



→ Indicates course of events

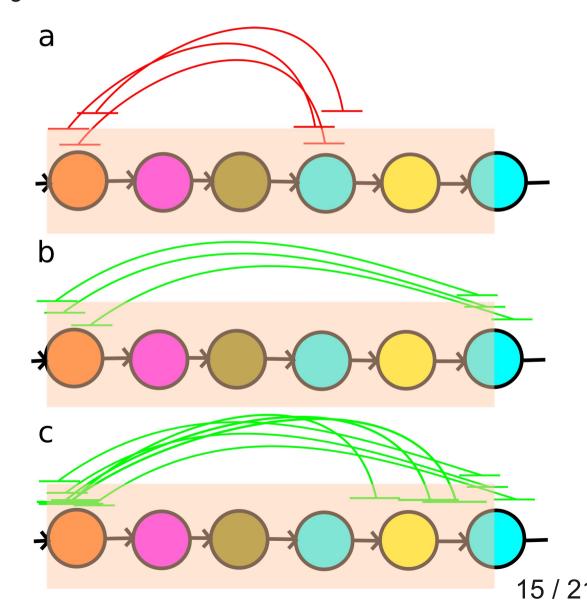
Constraint 2 -- local pair population v. genome-wide pair population

→ The average outer distance of local pair population must be within 1 standard deviations from the genome-wide average outer distance



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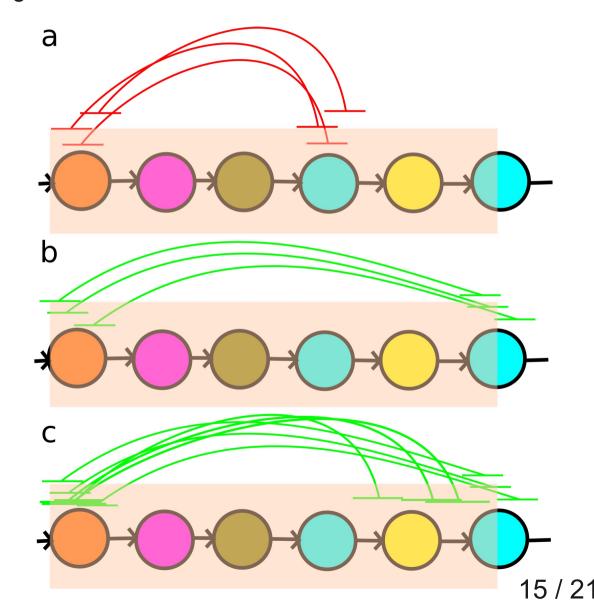
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→ Constraint on many pairs

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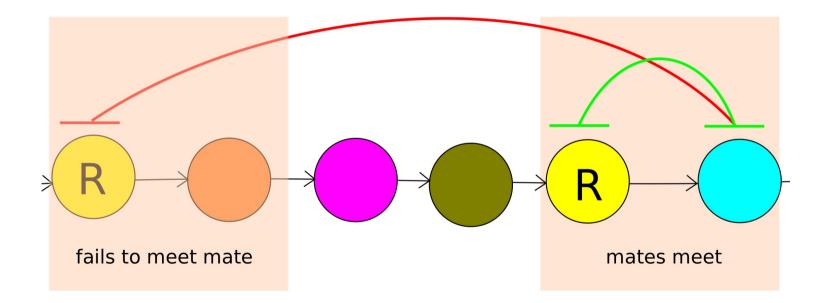
→ The average outer distance of local pair population must be within 1 standard deviations from the genome-wide average outer distance



- → Constraint on many pairs
- → Avoids collapsing of repeats

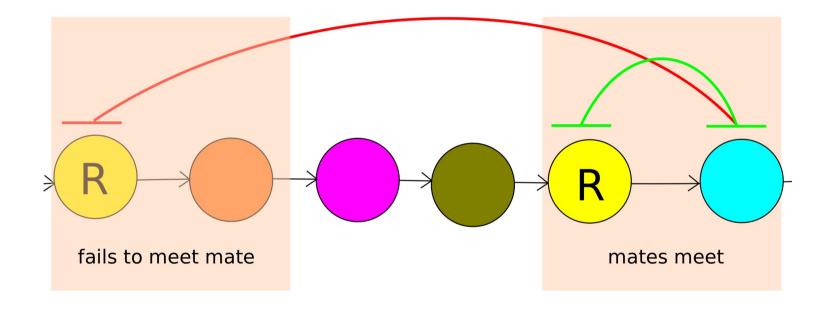
Constraint 3 -- mates meet

→ A read that fails to meet its mate within the average + 3 standard deviations is set as unused



Constraint 3 -- mates meet

→ A read that fails to meet its mate within the average + 3 standard deviations is set as unused



→ A read can meet its mate

Experiment

Table 1 - Description of libraries

Library	Average	Standard	Read	Substitution	Number
	outer	deviation	length	error rate	of pairs
	distance				
L1	200	20	50	0.5%	4000000
L2	1000	100	50	0.5%	4000000
L3	10000	1000	50	0.5%	4000000

^{→ 3} simulated paired libraries with *E. coli* genome: 200, 1000 & 10000

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- → 3 simulated paired libraries with *E. coli* genome: 200, 1000 & 10000
- → Standard deviation: 10 %; short read length; sequencing errors

Table 2 - Assembly validations

Libraries	Number	Number	Average	N50	Maximum	Genome	Large	Sub-	Small
	of	of	contig	contig	contig	breadth	indels	stitutions	indels
	contigs	nucleotides	length	length	length	coverage			
L1	108	4559977	42222	87242	269942	98.20%	0	0	1
L1-L2	82	4621035	56354	96588	232618	99.13%	0	5	0
L1-L3	43	4618437	107405	177402	409463	99.59%	0	1	0

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- → Maximum unique matches with MUMmer (Kurtz et al. 2004)
- → L1-L3 (L1+L2+L3) yields only 43 contigs; no misassembled contigs

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	of	of	contig	contig	contig	breadth	indels	stitutions	indels
	contigs	nucleotides	length	length	length	coverage			
L1	108	4559977	42222	87242	269942	98.20%	0	0	1
L1-L2	82	4621035	56354	96588	232618	99.13%	0	5	0
L1-L3	43	4618437	107405	177402	409463	99.59%	0	1	0





- → Maximum unique matches with MUMmer (Kurtz et al. 2004)
- → L1-L3 (L1+L2+L3) yields only 43 contigs; no misassembled contigs
- → Small increase of breadth of coverage -> sizable impact on number of contigs

Table 2 - Assembly validations

Libraries	Number	Number	Average	N50	Maximum	Genome	Large	Sub-	Small
	of	of	contig	contig	contig	breadth	indels	stitutions	indels
	contigs	nucleotides	length	length	length	coverage			
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- → Maximum unique matches with MUMmer (Kurtz et al. 2004)
- → L1-L3 (L1+L2+L3) yields only 43 contigs; no misassembled contigs
- → Small increase of breadth of coverage -> sizable impact on number of contigs
- → Resources necessary: 30 processors, 9 GiB of memory, 11 minutes for L1-L3

Comparison with Velvet

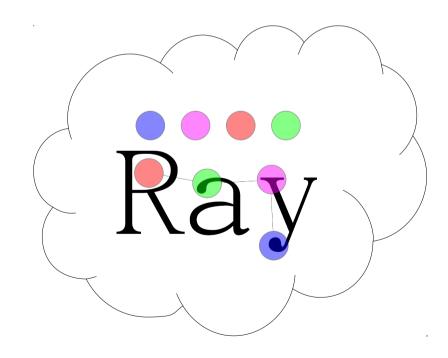
- → Comparison of *E. coli* genome assemblies with Velvet and Ray
- → with 3 paired libraries (200, 1000 & 10000)

	Velvet	Ray
Contigs	65	43
Misassembled contigs	36	0
Substitution errors	503	1
Small indels	829	0

→ Hypothesis: Velvet's bubble merging is unsuitable for repeats

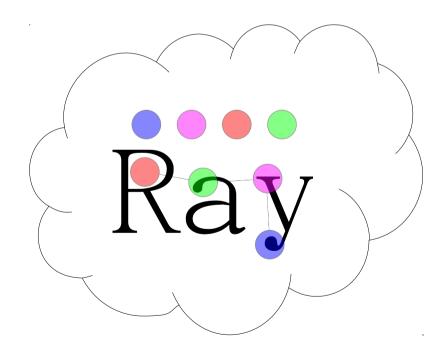
Conclusion

→ Optimal read markers place reads on unique vertices if possible



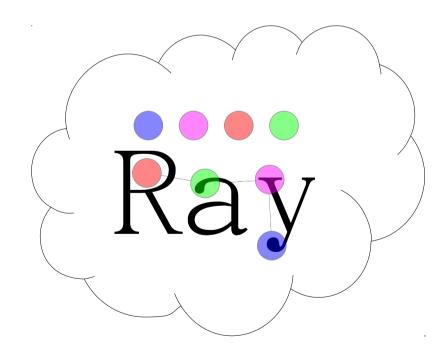
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- → Optimal read markers place reads on unique vertices if possible
- → Statistical constraints allow optimal pair placements



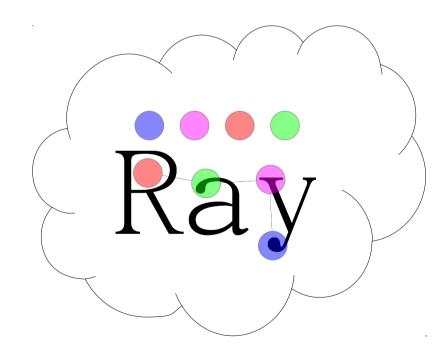
Conclusion

- → Optimal read markers place reads on unique vertices if possible
- → Statistical constraints allow optimal pair placements
- → Treat repeats precisely = large contiguous sequences without misassemblies



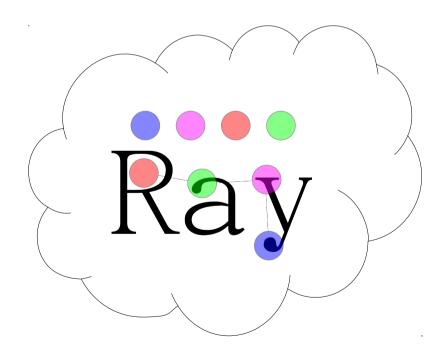
Perspective

→ Cloud-ready (needs MPI + fast interconnect)



Perspective

- → Cloud-ready (needs MPI + fast interconnect)
- → Assemble large genomes: <u>de novo assembly of Illumina CEO genome in 11.5 h with Ray</u> (Supplementary slides)

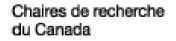


Acknowledgments

- → RECOMB-seq committees for organising this great workshop
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- → Natural Sciences and Engineering Research Council of Canada for funding to F.L.
- → Canadian Foundation for Innovation for infrastructure funding
- → Compute Canada (CLUMEQ) for compute resources
- → Free/libre software people

Canada Research



















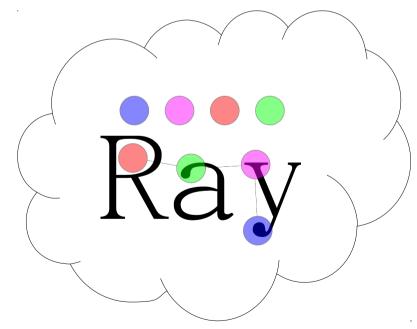


Questions?

Merci à vous pour votre attention!

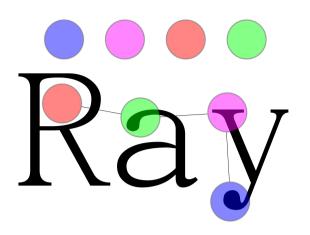
→ Cloud-ready (needs MPI + fast interconnect)

Example: Amazon EC2 Cluster Compute



→http://tiny.cc/ray-assembler

Supplementary: Ray command



\$ mpirun -np 30 /software/ray-1.2.4/bin/Ray -k 21 -o L1-L3-assembly \ -p L1_1.fasta L1_2.fasta -p L2_1.fasta L2_2.fasta -p L3_1.fasta L3_2.fasta

Supplementary: Velvet commands

Commands to convert files and run Velvet with automatic calculation of insert lengths and coverage values (3 categories):

```
$ /software/velvet_1.0.19/shuffleSequences_fasta.pl L1_1.fasta L1_2.fasta L1.fasta $ /software/velvet_1.0.19/shuffleSequences_fasta.pl L2_1.fasta L2_2.fasta L2.fasta $ /software/velvet_1.0.19/shuffleSequences_fasta.pl L3_1.fasta L3_2.fasta L3.fasta
```

- \$ /software/velvet_1.0.19/velveth velvetAssembly 21 -fasta \
 -shortPaired L1.fasta -shortPaired2 L2.fasta -shortPaired3 L3.fasta
- \$ /software/velvet_1.0.19/velvetg velvetAssembly -exp_cov auto

Supplementary: a Velvet incorrect contig

Example of Velvet incorrect contig (# 134; length: 4334 bp)

Reference segment Contig segment

2116786-2116856 1-71

2296192-2299712 51-3571 2309689-2310402 3582-4295

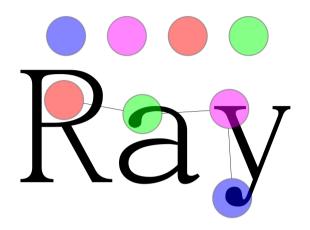
→ Velvet seems to jump over repeats, hence skipping important regions

Supplementary: Human genome chromosome 1

```
[1,0]<stdout>: Beginning of computation: 1 seconds
[1,0]<stdout>: Distribution of sequence reads: 1 minutes, 0 seconds
[1,0]<stdout>: Distribution of vertices & edges: 17 minutes, 58 seconds
[1,0]<stdout>: Calculation of coverage distribution: 8 seconds
[1,0]<stdout>: Indexing of sequence reads: 24 minutes, 38 seconds
[1,0]<stdout>: Computation of seeds: 18 minutes, 19 seconds
[1,0]<stdout>: Computation of library sizes: 5 minutes, 9 seconds
[1,0]<stdout>: Extension of seeds: 2 hours, 37 minutes, 12 seconds
[1,0]<stdout>: Computation of fusions: 43 minutes, 52 seconds
[1,0]<stdout>: Collection of fusions: 9 seconds
[1,0]<stdout>: Completion of the assembly: 4 hours, 28 minutes, 26 seconds

Memory usage: ~ 56 GiB
```

Peak coverage: 19



Supplementary: resource on Compute Canada's colosse

- → Ray 1.3.0 on 64 computers
- → Linked with InfiniBand QDR (40 Gigabits per second)
- → 24 GiB of memory per computer
- → 2 Intel Xeon Nehalem-EP (x86_64) processors per computer
- → 4 compute cores per processor
- → Total: 512 compute cores, 1535 GiB of memory





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InfiniBand is a trademark of the InfiniBand Trade Association.

Supplementary: SRA010766, Illumina CEO genome data

- → Illumina CEO
- → Jay T. Flatley
- → Illumina Genome Analyzer II
- → Input: 6 372 129 288 reads (477 909 696 600 nucleotides)

illumına^{*}

http://illumina.com/humangenome

Supplementary: de novo assembly of Illumina CEO genome (2011-03-21)

→ k=21

→ Running time: 11.5 h

→ Outer distances: 190 +/- 30; read length: 75

→ Peak coverage: 22, minimum coverage: 6

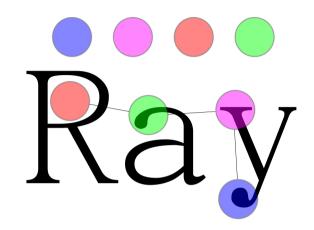
→ Peak probably too low

→ Output: 1 803 534 contiguous sequences

→ 1 772 120 417 nucleotides (haploid human genome is 3 Gb)

→ N50: 1341, average length: 982, longuest: 14584

→ Job identifier: 2814556, job code name: Nitro



Supplementary: next steps

→Try with a higher k-mer length (k=31)

Other dataset:

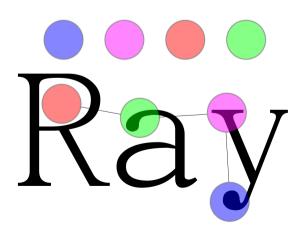
- → Test on the African Genome (SRA000271)
- → Yoruban male (NA18507)
- → Illumina Genome Analyzer platform

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Supplementary: Ray software systems

- → 143 MPI tag types, 28 master modes, 24 slave modes
- → Max. 250 paired libraries, max. coverage 65535, max processors: 1000000
- → With n processors: n slave rank + 1 master rank (which is also a slave)
- → Last revision: 4462 (1.3.0-dev)
- → 22792 lines of code (C++; .h, cpp)
- → 65 classes, 74 .cpp files, 68 .h files



Illumina is a registered trademark of Illumina, Inc.

Supplementary: Ray message transit systems

- → Manual message aggregation in some steps
- → Virtual communicator on top of MPI_COMM_WORLD (MPI default communicator)
- → Allows transparent message aggregation
- → Array of workers on each MPI ranks (max. 30000)
- → Workers see only 3 methods: pushMessage, isMessageProcessed, getResponseElements
- → All the aggregation logic is done by the virtual communicator