Package 'workingfunctions'

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alpha_diagnostics

Item total correlation and r drop

Description

Item total correlation and r drop

Usage

```
alpha_diagnostics(df)
```

Arguments

df

dataframe with one dimension

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
psych::alpha(df)
alpha_diagnostics(df=df)</pre>
```

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call_to_string

Model call to string

Description

Takes a call object and convert it to string

Usage

```
call_to_string(model)
```

Arguments

model

Model object

Examples

```
df<-generate_correlation_matrix()
model<-lm(df$X1~df$X2)
call_to_string(model)</pre>
```

cdf

Check dataframe

Description

dataframe summary

Usage

```
cdf(
   df,
   name_length = (getOption("width")/3),
   digits = 2,
   nuniques = 0,
   parralel = FALSE,
   file = NULL
)
```

Arguments

df dataframe
name_length number of characters to be displayed for names
digits number of rounding digits
nuniques number of unique items to display
parralel if TRUE it will run using multiple cores
file output filename

cfa_icc_index 7

Examples

cfa_icc_index

index of items to convert from lavaan to thurstonian order for analysis

Description

index of items to convert from lavaan to thurstonian order for analysis

Usage

```
cfa_icc_index(nitems, nfactors = 3)
```

Arguments

nitems number of items in the questionnaire

nfactors number of factors

Examples

```
cfa_icc_index(nitems=18,nfactors=3)
```

change_data_type

dataframe data type transformations

Description

dataframe data type transformations

Usage

```
change_data_type(df, type)
```

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Arguments

df dataframe

type "character" "numeric" "factor" "factor_character" "character_factor"

For "factor_character" if factors are found, are converted to characters For "character_factor" if characters are found, are converted to factors

Examples

```
cdf(df=change_data_type(df=mtcars, "character"))
cdf(df=change_data_type(df=mtcars, "numeric"))
cdf(df=change_data_type(df=mtcars, "factor"))
df<-change_data_type(df=mtcars, "factor")
cdf(df=change_data_type(df=df, "factor_character"))</pre>
```

clear_stopwords

Remove stopwods

Description

Remove stopwods

Usage

```
clear_stopwords(text, stopwords = stopwords::stopwords("english"))
```

Arguments

text character vector

stopwords character words to remove

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition, injected humour, or non-characteristic words etc."
stopwords<-stopwords::stopwords("english")
text<-c(text1,text2,text3,text4,text5)
clear_stopwords(text,stopwords=stopwords)</pre>
```

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clear_text

Clear text

Description

Clear text

Usage

```
clear_text(text)
```

Arguments

text

character vector

Examples

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition, injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
clear_text(text)
```

comparison_combinations

Produce combinations for comparisons from dataframe names

Description

Produce combinations for comparisons from dataframe names

Usage

```
comparison_combinations(df, all_orders = TRUE)
```

Arguments

df dataframe

all_orders if TRUE the order of combination is considered i.e. the combination X1 X2 also

appears as X2 X1 if FALSE it is assumed that X1 X2 and X2 X1 are the same

and only one of them appears

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Examples

```
comparison_combinations(generate_correlation_matrix(n=10)[,1:4])
```

compute_ability

Compute subject ability for thurstonian models

Description

Computes person ability for binary thurstonian coded items for a single dimension

Usage

```
compute_ability(
  response,
  eta,
  gamma,
  lambda,
  psi,
  plot = FALSE,
  map = compute_map(eta = eta, mean = 0, sd = 1)
)
```

Arguments

```
response item responses
eta eta or ability
gamma gamma or threshold
lambda lambda or loading
psi psi or error
plot if TRUE plots icc curves using the plot_icc_thurstonian function
map vector from compute_map
```

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
eta<-seq(-6,6,by=0.1)
response1<-c(0,0,0,0,0,0,0)
response2<-c(1,1,1,1,1,1,1,1)
response3<-c(1,0,1,0,1,0,1,0)
response4<-c(0,1,0,1,0,1,0,1,0)
map<-compute_map(eta=eta,mean=0,sd=1)
compute_ability(response1,eta,gamma,lambda,psi,map=map,plot=FALSE)</pre>
```

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```
compute_ability(response2,eta,gamma,lambda,psi,map=map,plot=FALSE)
compute_ability(response3,eta,gamma,lambda,psi,map=map,plot=FALSE)
compute_ability(response4,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

compute_adjustment

Compute adjustments

Description

Compute adjustments

Usage

```
compute_adjustment(a, ntests)
```

Arguments

a alpha criterion ntests number of tests

Examples

```
compute_adjustment(0.05,100)
```

compute_aggregate

Descriptive statistics

Description

uses plyr

Usage

```
compute_aggregate(df, iv, file = NULL)
```

Arguments

df dataframe

iv index of independent variables

file output filename

Details

returns xlsx

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Examples

```
compute_aggregate(df=mtcars,iv=9)
compute_aggregate(df=mtcars,iv=9:10)
compute_aggregate(df=mtcars,iv=9:11)
compute_aggregate(df=mtcars,iv=9:11,file="descriptives")
```

compute_aov_es

Compute eta and omega

Description

Computes omega using aov object. Based on http://stats.stackexchange.com/a/126520

Usage

```
compute_aov_es(model, ss = "I")
```

Arguments

model object aov

ss Character type of sums of squares "I" "III" "III"

```
form<-formula(uptake~Treatment)
one_way_between<-aov(form,CO2)
factorial_between<-aov(uptake~Treatment*Type,CO2)
compute_aov_es(model=one_way_between,ss="I")
sjstats::anova_stats(one_way_between,digits=10)
compute_aov_es(model=one_way_between,digits=10)
compute_aov_es(model=one_way_between,digits=10)
compute_aov_es(model=one_way_between,digits=10)
compute_aov_es(model=factorial_between,ss="II")
sjstats::anova_stats(factorial_between,digits=10)
compute_aov_es(model=factorial_between,ss="II")
sjstats::anova_stats(factorial_between,digits=10)
compute_aov_es(model=factorial_between,digits=10)
compute_aov_es(model=factorial_between,digits=10)
sjstats::anova_stats(factorial_between,ss="III")
sjstats::anova_stats(car::Anova(factorial_between,Type=3),digits=10)</pre>
```

```
compute_confidence_inteval
```

Compute confidence interval

Description

Compute confidence interval

Usage

```
compute_confidence_inteval(vector)
```

Arguments

vector vector

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_confidence_inteval(vector)</pre>
```

compute_crosstable

Compute crosstables

Description

Compute crosstables

Usage

```
compute_crosstable(df, factor_index = NULL, combinations = NULL)
```

Arguments

df dataframe factor_index index of factors combinations index of comparisons

```
combinations<-data.frame(index1=c("vs","am","gear"),index2=c("cyl","cyl","cyl"))
compute_crosstable(df=mtcars,combinations=combinations)
combinations<-data.frame(index1=c("vs","am"),index2=c("cyl","cyl"))
compute_crosstable(df=mtcars,combinations=combinations)
compute_crosstable(df=mtcars,factor_index=8:10)</pre>
```

compute_descriptives Descriptive statistics

Description

uses psych

Usage

```
compute_descriptives(df, dv, iv = NULL, file = NULL)
```

Arguments

df dataframe

dv index of dependent variablesiv index of independent variables

file output filename

Details

returns xlsx

Examples

```
compute_descriptives(df=mtcars,dv=1:5)
compute_descriptives(df=mtcars,dv=1:2,iv=9:10)
compute_descriptives(df=mtcars,dv=1:2,file="descriptives_no_factor")
compute_descriptives(df=mtcars,dv=1:2,iv=9:10,file="descriptives_factor")
```

compute_dissatenuation

Compute dissatenuation

Description

Compute dissatenuation

Usage

```
compute_dissatenuation(variable1, error1, variable2, error2)
```

Arguments

variable1 vector

error1 vector error measurement for variable1

variable2 vector

error2 vector error measurement for variable2

Examples

```
set.seed(1)
compute_dissatenuation(rnorm(10),rnorm(10),rnorm(10))
```

compute_dummy_comparisons

Compute number of dummy comparisons

Description

Compute number of dummy comparisons

Usage

```
compute_dummy_comparisons(items)
```

Arguments

items

number of items per block

Examples

```
compute_dummy_comparisons(1)
compute_dummy_comparisons(2)
compute_dummy_comparisons(3)
compute_dummy_comparisons(4)
compute_dummy_comparisons(5)
compute_dummy_comparisons(6)
```

 ${\tt compute_frequencies}$

Frequencies by levels

Description

returns frequency proportion percent

Usage

```
compute_frequencies(df, ordered = TRUE, file = NULL)
```

Arguments

df dataframe

ordered if TRUE it will output frequencies in descending order

file output filename

Details

returns xlsx

Examples

```
compute_frequencies(df=generate_missing(generate_factor(nrows=10,ncols=10),missing=5))
compute_frequencies(df=generate_factor())
compute_frequencies(df=generate_factor(),file="descriptives")
```

compute_icc_thurstonian

Compute item characteristic curves for thurstonian models

Description

Computes icc curves for binary thurstonian coded items for a single dimension

Usage

```
compute_icc_thurstonian(eta, gamma, lambda, psi, plot = FALSE)
```

Arguments

eta eta or ability

gamma or threshold
lambda lambda or loading

psi psi or error

plot if TRUE plots icc curves using the plot_icc_thurstonian function

```
 \begin{array}{l} {\rm gamma} < -c(\emptyset.556, -1.253, -1.729, \emptyset.618, \emptyset.937, \emptyset.295, -0.672, -1.127, -0.446, \emptyset.632, 1.147, \emptyset.498) \\ {\rm psi} < -c(2.172, 1.883, 2.055, 1.869, 2.231, 2.100, 1.762, 1.803, 1.565, 1.892, 1.794, 1.686) \\ {\rm lambda} < -c(1.082, 1.082, -1.297, -1.297, \emptyset.802, \emptyset.802, 1.083, 1.083) \\ {\rm gamma} < -{\rm gamma}[{\rm response\_dimension}(c(1:12), 3, c(1, 2))] \\ {\rm psi} < -{\rm psi}[{\rm response\_dimension}(c(1:12), 3, c(1, 2))] \\ {\rm eta} < -{\rm seq}(-6, 6, by=0.01) \\ {\rm compute\_icc\_thurstonian}({\rm eta} = {\rm eta}, {\rm gamma} = {\rm gamma}, {\rm lambda} = {\rm lambda}, {\rm psi} = {\rm psi}, {\rm plot} = {\rm FALSE}) \\ \end{array}
```

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compute_info_1pl

Compute item information for 1PL model

Description

Compute item information for 1PL model

Usage

```
compute_info_1pl(b, theta)
```

Arguments

b numeric difficulty parameter

theta numeric theta

Examples

```
compute_info_1pl(b=1,theta=-3)
compute_info_1pl(b=1,theta=-2)
compute_info_1pl(b=1,theta=-1)
compute_info_1pl(b=1,theta=0)
compute_info_1pl(b=1,theta=1)
compute_info_1pl(b=1,theta=2)
compute_info_1pl(b=1,theta=3)
ti<-compute_info_1pl(b=1,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))</pre>
```

compute_info_2pl

Compute item information for 2PL model

Description

Compute item information for 2PL model

Usage

```
compute_info_2pl(a, b, theta)
```

Arguments

a numeric discrimination parameter
b numeric difficulty parameter

theta numeric theta

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Examples

```
compute_info_2pl(a=1.5,b=1,theta=-3)
compute_info_2pl(a=1.5,b=1,theta=-2)
compute_info_2pl(a=1.5,b=1,theta=-1)
compute_info_2pl(a=1.5,b=1,theta=0)
compute_info_2pl(a=1.5,b=1,theta=1)
compute_info_2pl(a=1.5,b=1,theta=2)
compute_info_2pl(a=1.5,b=1,theta=3)
ti<-compute_info_2pl(a=1.5,b=1,theta=3)
ti<-compute_info_2pl(a=1,b=-2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))
ti<-compute_info_2pl(a=2,b=0,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))
ti<-compute_info_2pl(a=3,b=2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))</pre>
```

compute_info_3pl

Compute item information for 3PL model

Description

Compute item information for 3PL model

Usage

```
compute_info_3pl(a, b, g, theta)
```

Arguments

a	numeric discrimination parameter
b	numeric difficulty parameter
g	numeric guessing parameter
theta	numeric theta

```
compute_info_3pl(a=1.5,b=1,g=.2,theta=-3)
compute_info_3pl(a=1.5,b=1,g=.2,theta=-2)
compute_info_3pl(a=1.5,b=1,g=.2,theta=-1)
compute_info_3pl(a=1.5,b=1,g=.2,theta=0)
compute_info_3pl(a=1.5,b=1,g=.2,theta=1)
compute_info_3pl(a=1.5,b=1,g=.2,theta=2)
compute_info_3pl(a=1.5,b=1,g=.2,theta=3)
ti<-compute_info_3pl(a=1.5,b=1,g=.2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))</pre>
```

Description

Kruskal Wallis test

Usage

```
compute_kruskal_wallis_test(formula, df)
```

Arguments

formula one way formula in form of y~x. It will ignore more complex formulas

df dataframe

Note

```
eta squared ranges between 0 and 1 epsilon squared ranges between 0 and 1 eta squared multiplied by 100 indicates the percentage of variance in the dependent variable explained by the independent variable
```

Examples

compute_kurtosis

Compute kurtosis

Description

Compute kurtosis

Usage

```
compute_kurtosis(vector)
```

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Arguments

vector vector

Note

```
b_2 = m_4 / s^4 - 3 = (g_2 + 3) (1 - 1/n)^2 - 3. Used in MINITAB and BMDP.
```

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_kurtosis(vector)
e1071::kurtosis(vector)</pre>
```

compute_map

Simulate prior distribution

Description

Simulate prior distribution

Usage

```
compute_map(eta, mean = 0, sd = 1)
```

Arguments

eta vector
mean numeric
sd numeric

```
eta<-seq(-6,6,by=0.1)
compute_map(eta=eta,mean=0,sd=1)</pre>
```

compute_moving_average

Moving Average

Description

compute moving average

Usage

```
compute_moving_average(df, w)
```

Arguments

df dataframe w window

Examples

```
compute_moving_average(df=mtcars,w=5)
```

```
compute_one_way_test one way test
```

Description

one way test

Usage

```
compute_one_way_test(formula, df, var.equal = TRUE)
```

Arguments

formula one way formula in form of y~x. It will ignore more complex formulas

df dataframe

var.equal if TRUE it assumes equal variances

Note

eta and omega for welch statistics are not adequatelly tested and they should not be consulted

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Examples

```
form<-formula(qsec~cyl)
compute_one_way_test(formula=form,df=mtcars,var.equal=TRUE)
compute_one_way_test(formula=form,df=mtcars,var.equal=FALSE)
oneway.test(formula=form,data=mtcars,var.equal=TRUE)
oneway.test(formula=form,data=mtcars,var.equal=FALSE)
car::Anova(aov(form,data=mtcars),type=2)
model<-lm(form,data=mtcars)
lsr::etaSquared(aov(form,data=mtcars),type=3,anova=TRUE)
sjstats::anova_stats(model,digits=22)</pre>
```

compute_posthoc

Games Howell Tukey post hoc tests

Description

Based on http://www.psych.yorku.ca/cribbie/6130/games_howell.R

Usage

```
compute_posthoc(y, x, method = c("games-howell", "tukey"))
```

Arguments

y Vector continous variable

x Vector factor

method Character takes two values "games-howell" or "tukey" or c("games-howell", "tukey")

Examples

```
result<-compute_posthoc(mtcars[,6],mtcars[,10])</pre>
```

compute_power_r

Compute r power curve

Description

Compute r power curve

Usage

```
compute_power_r(
  n = 100,
  r = NULL,
  sig.level = 0.05,
  alternative = c("two.sided", "less", "greater"),
  title = "",
  base_size = 10
)
```

Arguments

n number of observations

sig.level alpha (type I error probability)

alternative a character string specifying the alternative hypothesis, must be one of "two.sided"

(default), "greater" or "less"

correlation coefficient

title plot title

base_size base font size

Examples

```
compute_power_r(n=100,r=.5,sig.level=.05,alternative=c("two.sided"))
```

compute_power_r_matrix

Compute correlation matrix

Description

Compute correlation matrix

Usage

```
compute_power_r_matrix(m, ...)
```

Arguments

m correlation matrix

. . . arguments passed to compute_power_r

```
compute_power_r_matrix(m=stats::cor(mtcars,use="pairwise.complete.obs"),n=100)
```

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```
compute_residual_stats
```

Residuals for matrices

Description

Root Mean Squared Residual Number of absolute residuals > 0.05 Proportion of absolute residuals > 0.05. It can either accept a psych EFA model or it can compare two correlation or covariance matrices

Usage

```
compute_residual_stats(model, data = NULL)
```

Arguments

model psych EFA model. It has to be a correlation or covariance matrix if data is not

NULL

data correlation or covariance matrix

Examples

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
compute_residual_stats(model)</pre>
```

compute_scores

Compute subject ability for thurstonian models

Description

Computes person ability for binary thurstonian coded items for a single dimension

Usage

```
compute_scores(mydata, ...)
```

Arguments

```
mydata item responses
```

... arguments passed to compute_ability

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Examples

```
 \begin{array}{l} \operatorname{gamma} < -\operatorname{c}(0.556, -1.253, -1.729, 0.618, 0.937, 0.295, -0.672, -1.127, -0.446, 0.632, 1.147, 0.498) \\ \operatorname{psi} < -\operatorname{c}(2.172, 1.883, 2.055, 1.869, 2.231, 2.100, 1.762, 1.803, 1.565, 1.892, 1.794, 1.686) \\ \operatorname{lambda} < -\operatorname{c}(1.082, 1.082, -1.297, -1.297, 0.802, 0.802, 1.083, 1.083) \\ \operatorname{gamma} < -\operatorname{gamma} [\operatorname{response\_dimension}(\operatorname{c}(1:12), 3, \operatorname{c}(1, 2))] \\ \operatorname{psi} < -\operatorname{psi} [\operatorname{response\_dimension}(\operatorname{c}(1:12), 3, \operatorname{c}(1, 2))] \\ \operatorname{eta} < -\operatorname{seq}(-6, 6, \operatorname{by} = 0.1) \\ \operatorname{map} < -\operatorname{compute\_map}(\operatorname{eta} = \operatorname{eta}, \operatorname{mean} = 0, \operatorname{sd} = 1) \\ \operatorname{response\_df} < -\operatorname{data.frame}(\operatorname{matrix}(\operatorname{nrow} = 0, \operatorname{ncol} = 8)) \\ \operatorname{response\_df} [1, ] < -\operatorname{c}(0, 0, 0, 0, 0, 0, 0, 0) \\ \operatorname{response\_df} [2, ] < -\operatorname{c}(1, 1, 1, 1, 1, 1, 1, 1) \\ \operatorname{response\_df} [3, ] < -\operatorname{c}(1, 0, 1, 0, 1, 0, 1, 0, 1) \\ \operatorname{compute\_scores}(\operatorname{response\_df}, \operatorname{eta}, \operatorname{gamma}, \operatorname{lambda}, \operatorname{psi}, \operatorname{map} = \operatorname{map}, \operatorname{plot} = \operatorname{FALSE}) \\ \end{array}
```

compute_se_theta

Compute the SE of theta

Description

Compute the SE of theta

Usage

```
compute_se_theta(info)
```

Arguments

info

numeric information

Examples

```
compute_se_theta(1)
ti<-compute_info_2pl(a=10,b=0,theta=seq(-3,3,by=.01)) # test information
plot(compute_se_theta(ti),x=seq(-3,3,by=.01))</pre>
```

 ${\tt compute_skewness}$

Compute skewness

Description

Compute skewness

Usage

```
compute_skewness(vector)
```

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Arguments

vector vector

Note

```
b_1 = m_3 / s^3 = g_1 ((n-1)/n)^3. Used in MINITAB and BMDP.
```

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_skewness(vector)
e1071::skewness(vector)</pre>
```

compute_standard

compute standard scores

Description

compute standard scores

Usage

```
compute_standard(vector, mean = 0, sd = 1, type = "z", input = "non_standard")
```

Arguments

vector vector

mean numeric applicable to "uz" sd numeric applicable to "uz"

type "z" "uz" "sten" "t" "stanine" "center" "center_reversed" "percent" "scale_zero_one"

"normal_density" "cumulative_density" "all"

input "standard" "non_standard" standard inputs are z scores and non standard are raw

scores

```
vector<-c(rnorm(10),NA,rnorm(10))
compute_standard(vector,type="z")
compute_standard(vector,mean=0,sd=1,type="uz")
compute_standard(vector,type="sten")
compute_standard(vector,type="t")
compute_standard(vector,type="stanine")
compute_standard(vector,type="center")
compute_standard(vector,type="center_reversed")
compute_standard(vector,type="percent")
compute_standard(vector,type="scale_zero_one")</pre>
```

```
ndf<-compute_standard(seq(-6,6,.01),mean=0,sd=1,type="normal_density")
plot(ndf)
cdf<-compute_standard(ndf,mean=0,sd=1,type="cumulative_density")
plot(cdf)
compute_standard(vector,type="all")
compute_standard(seq(-6,6,.1),type="all",input="standard")</pre>
```

compute_standard_error

Compute standard error

Description

Compute standard error

Usage

```
compute_standard_error(vector)
```

Arguments

vector

vector

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_standard_error(vector)</pre>
```

compute_unidimensional_ability

Compute theta for unidimensional models

Description

Compute theta for unidimensional models

Usage

```
compute_unidimensional_ability(
   a,
   b,
   g = NULL,
   d = 1.702,
   u,
   lim_theta = c(-6, 6)
)
```

Arguments

а	numeric vector discrimination parameters
b	numeric vector difficulty parameters
g	numeric vector guessing parameters
d	numeric scaling constant usually it is a value that approximating 1.749
u	numeric vector responses
lim_theta	vector minimum and maximum value of theta

Examples

```
 \begin{array}{l} \mathsf{a} < -\mathsf{c}(0.39, 0.45, 0.52, 0.3, 0.35, 0.43, 0.42, 0.44, 0.34, 0.42) \\ \mathsf{b} < -\mathsf{c}(-1.96, -1.9, -1.38, -0.58, 0.48, -0.81, -0.35, 1.59, 1.33, 2.93) \\ \mathsf{u} < -\mathsf{c}(1, 1, 1, 1, 0, 0, 1, 0, 1, 0) \\ \# \ \mathsf{SHOULD} \ \mathsf{RETURN} \ 0.48402574251176 \\ \mathsf{compute} \_ \mathsf{unidimensional}\_ \mathsf{ability}(\mathsf{a} = \mathsf{a}, \mathsf{b} = \mathsf{b}, \mathsf{u} = \mathsf{u}, \mathsf{d} = 1.7, \mathsf{g} = \mathsf{NULL}) \\ \mathsf{a} < -\mathsf{c}(1.27, 0.9, 0.94, 0.95, 0.55, 0.6, 0.44, 0.4) \\ \mathsf{b} < -\mathsf{c}(-0.54, 0.18, 0.21, 1.26, 1.73, -0.87, 1.72, 2.67) \\ \mathsf{u} < -\mathsf{c}(1, 1, 1, 1, 0, 0, 0, 0) \\ \# \ \mathsf{SHOULD} \ \mathsf{RETURN} \ 1.04621621510192 \\ \mathsf{compute} \_ \mathsf{unidimensional}\_ \mathsf{ability}(\mathsf{a} = \mathsf{a}, \mathsf{b} = \mathsf{b}, \mathsf{u} = \mathsf{u}, \mathsf{d} = 1.7, \mathsf{g} = \mathsf{NULL}) \\ \mathsf{a} < -\mathsf{c}(0.41, 0.32, 0.33, 1.2, 0.63, 0.62, 0.7, 0.61, 0.38, 0.53, 0.6, 1.16) \\ \mathsf{b} < -\mathsf{c}(-1.4, -1.3, -1.17, 0.2, 0.71, 0.86, -0.12, 0.12, 2.06, 1.38, 1.18, -0.33) \\ \mathsf{u} < -\mathsf{c}(1, 0, 1, 1, 0, 0, 0, 1, 1, 0, 1, 0) \\ \# \ \mathsf{SHOULD} \ \mathsf{RETURN} \ 0.0860506282671103 \\ \mathsf{compute} \_ \mathsf{unidimensional}\_ \mathsf{ability}(\mathsf{a} = \mathsf{a}, \mathsf{b} = \mathsf{b}, \mathsf{u} = \mathsf{u}, \mathsf{d} = 1.7, \mathsf{g} = \mathsf{NULL}) \\ \end{aligned}
```

compute_unidimensional_theta

Compute theta for unidimensional models

Description

Compute theta for unidimensional models

Usage

```
compute_unidimensional_theta(a, b = 0, g = 0, i = 1, d = 1.702, theta = 0)
```

Arguments

а	numeric discrimination parameter
b	numeric difficulty parameter
g	numeric guessing parameter
i	numeric innatentiveness parameter
d	numeric scaling constant usually a value 1.749 or 1.702
theta	numeric or vector theta

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Note

```
when scaling constant=1 it has no effect in equation when innatentiveness=1 and guessing=0 function computes a 2PL score when innatentiveness=1 and guessing!=0 function computes a 3PL score when innatentiveness!=1 and guessing!=0 function computes a 4PL score
```

Examples

```
compute_unidimensional_theta(a=10,b=0)
x<-seq(-3,3,by=.01)
plot(compute_unidimensional_theta(a=5,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=5,b=1,theta=x),x=x)
plot(compute_unidimensional_theta(a=5,b=1,theta=x),x=x)
plot(compute_unidimensional_theta(a=.1,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=1,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=.1,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=.5,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=1,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=.9,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=.9,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=.6,theta=x),x=x)</pre>
```

compute_y_logistic

Compute y for logistic function

Description

This function requires x range to produce a vector with y values

Usage

```
compute_y_logistic(intercept, coefficient, x)
```

Arguments

```
intercept Numeric
coefficient Numeric
x Numeric
```

```
x<--10:10
compute_y_logistic(0,1,x)
compute_y_logistic(0,1,1)
plot(x,compute_y_logistic(0,1,x),type="l");grid();abline(b=0,a=.5)</pre>
```

confusion

Create a confusion matrix from observed and expected vectors

Description

Create a confusion matrix from observed and expected vectors

Usage

```
confusion(observed, predicted)
```

Arguments

observed vector of observed variables
predicted vector of predicted variables

Examples

```
confusion(observed=c(1,2,3,4,5,10)),predicted=c(1,2,3,4,5,11)) confusion(observed=c(1,2,2,2,2)),predicted=c(1,1,2,2,2))
```

```
confusion_matrix_percent
```

Confusion matrix with row and column percent

Description

Confusion matrix with row and column percent

Usage

```
confusion_matrix_percent(observed, predicted)
```

Arguments

observed vector of observed variables
predicted vector of predicted variables

Note

Total measures - Accuracy: (TP+TN)/total
Total measures - Prevalence: (TP+FN)/total
Total measures - Proportion Incorrectly Classified: (FN+FP)/total
Horizontal measures - True Positive Rate - Sensitivity: TP/(TP+FN)
Horizontal measures - True Negative Rate - Specificity: TN/(FP+TN)
Horizontal measures - False Negative Rate - Miss Rate: FN/(TP+FN)
Horizontal measures - False Positive Rate - Fall-out: FP/(FP+TN)
Vertical measures - Positive Predictive value - Precision: TP/(TP+FP)

Vertical measures - Negative Predictive value: TN/(FN+TN) Vertical measures - False Omission Rate: FN/(FN+TN) Vertical measures - False Discovery Rate: FP/(TP+FP)

Examples

```
observed<-factor(round(rnorm(10000,m=10,sd=1)))
predicted<-factor(round(rnorm(10000,m=10,sd=1)))
confusion_matrix_percent(observed,predicted)</pre>
```

 ${\tt convert_excel_unix_timestamp}$

Convert UNIX EXCEL timestamp

Description

Convert UNIX EXCEL timestamp

Usage

```
convert_excel_unix_timestamp(timestamp)
```

Arguments

timestamp unix or excel timestamp

```
convert_excel_unix_timestamp(1)
```

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c_bind

cbind dataframes with unequal lengths or row lengths

Description

cbind dataframes with unequal lengths or row lengths

Usage

```
c_bind(..., first = TRUE)
```

Arguments

... dataframes or vectors to bind

first Logical

Author(s)

Ananda Mahto

Examples

```
c_bind(rnorm(10),rnorm(11),rnorm(12),rnorm(13))
```

data_frame_index

dataframe index

Description

dataframe index

Usage

```
data_frame_index(nrow, ncol)
```

Arguments

nrow number of rows
ncol number of collumns

```
data_frame_index(5,5)
```

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 $decompose_datetime$

Decompose datetime objects to dataframe collumns

Description

Decompose datetime objects to dataframe collumns

Usage

```
decompose_datetime(
    x,
    format = "",
    origin = "1970-01-01",
    tz = "GMT",
    extended = FALSE,
    breaks = c(-1, 5, 13, 16, 20, 23),
    ...
)
```

Arguments

x datetime object

format date time format

origin Starting date. The default is the unix time origin "1970-01-01"

tz Timezone

extended if TRUE it will display additional day time categories

WEEKDAY MONTH JULIAN QUARTER DAY_PERIOD

breaks Numeric vector Breaks define hour of day for classifiying into

"Night", "Morning", "Noon", "Afternoon", "Evening".

... arguments passed to as POSIXct This argument is used if extended=TRUE

```
timestamp1<-as.numeric(as.POSIXct(Sys.Date()))
timestamp2<-as.numeric(as.POSIXct(Sys.time()))
d1<-Sys.Date()
d2<-Sys.time()
decompose_datetime(x=d1)
decompose_datetime(x=d2)
decompose_datetime(x=d1,extended=TRUE)
decompose_datetime(x=d2,extended=TRUE)
decompose_datetime(x="01/15/1900",format="%m/%e/%Y")
decompose_datetime(x="01/15/1900",format="%m/%e/%Y",extended=TRUE)
decompose_datetime(x=as.Date(as.POSIXct(10000,origin="1970-01-01")))
decompose_datetime(x=as.Date(as.POSIXct(timestamp1,origin="1970-01-01")))</pre>
```

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```
format="\%m/\%e/\%Y") \\ decompose_datetime(x=as.Date(as.POSIXct(timestamp2,origin="1970-01-01")), \\ format="\%m/\%e/\%Y")
```

 ${\sf deg2rad}$

Convert degrees to radians

Description

Convert degrees to radians

Usage

deg2rad(degrees)

Arguments

degrees

degrees

Examples

deg2rad(180)

detach_package

Unload library

Description

Unload library

Usage

detach_package(package)

Arguments

package

Package name

```
display_upper_lower_triangle
```

Return upper diagonal from one matrix and lower diagonal from another matrix

Description

Return upper diagonal from one matrix and lower diagonal from another matrix

Usage

```
display_upper_lower_triangle(m_upper, m_lower, diagonal = NA)
```

Arguments

m_upper matrix m_lower matrix

diagonal if "upper" it returns upper diagonal if "lower" it returns lower diagonal if NA

returns NA in diagonal otherwise it returns any value spesified

Examples

```
m1<-matrix(1:9,nrow=3,ncol=3)
m2<-matrix(11:19,nrow=3,ncol=3)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal="upper")
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal="lower")
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=NA)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=1)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=c("X1","X2","X3"))
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=c(1,2,3))
display_upper_lower_triangle(m_upper=m1,m2)</pre>
```

dotnames

Get the names of objects in the arguments

Description

Get the names of objects in the arguments

Usage

```
dotnames(...)
```

Arguments

... objects

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Author(s)

Ananda Mahto

drop_levels

Drops unused factor levels

Description

Drops unused factor levels

Usage

```
drop_levels(df, factor_index = NULL, minimum_frequency = 5)
```

Arguments

df dataframe

factor_index numeric index of factors. If NULL the function uses is.factor() to discriminate

factors

minimum_frequency

the minimum frequency each factor will have, levels with frequency bellow or

equal to the defined frequency will be renamed "Other"

Examples

```
factor1<-factor(c(rep("A",10),rep("B",10)),levels=c("A","B","C","D"))
factor2<-factor(c(rep("A",10),rep("B",10)),levels=c("A","B","C","D"))
numeric1<-c(1:20)
df<-data.frame(numeric1,factor1,factor2)
df$factor1
drop_levels(df=df,minimum_frequency=9)
drop_levels(df=df,minimum_frequency=10)</pre>
```

dummy_arrange

Takes a vector with multiple responses and dummy arranges it in a dataframe

Description

Takes a vector with multiple responses and dummy arranges it in a dataframe

Usage

```
dummy_arrange(vector)
```

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Arguments

vector Vector

Examples

environment_options

Load environment options

Description

Load environment options

Usage

```
environment_options()
```

Examples

```
environment_options()
```

```
excel_confusion_matrix
```

Write matrix or dataframe to excel sheet

Description

Usefull for correlation matrices since it uses conditional formatting for matrices

Usage

```
excel_confusion_matrix(
   df,
   workbook,
   title = "Rows: Expected Collumns: Observed"
)
```

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Arguments

df dataframe or matrix

workbook workbook title comment

Examples

```
filename<-"excel_confusion_matrix.xlsx"
if (file.exists(filename)) file.remove(filename)
observed<-factor(round(rnorm(10000,m=10,sd=1)))
predicted<-factor(round(rnorm(10000,m=10,sd=1)))
confusion(observed,predicted)
cm<-confusion_matrix_percent(observed,predicted)
wb<-openxlsx::createWorkbook()
excel_confusion_matrix(cm,wb)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)</pre>
```

excel_critical_value

Write matrix or dataframe to excel sheet

Description

Usefull for generic data where conditional formating of a spesific collumn is required

Usage

```
excel_critical_value(
   df,
   workbook,
   sheet = "output",
   title = NULL,
   comment = NULL,
   numFmt = "#0.00",
   critical = NULL
)
```

Arguments

df dataframe or matrix

workbook workbook sheet sheet title title comment comment

numFmt number formatting

critical list in the form of (collumn1=critical_value1,collumn2=critical_value2...)

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Examples

```
comment<-list(mpg="Miles/(US) gallon",</pre>
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio",
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1=manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")
filename<-"excel_critical_value.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
df<-generate_missing(generate_correlation_matrix())</pre>
critical<-list(X1="<0.05",X5="<0")</pre>
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=list(X1="test"),
                     numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
filename<-"excel_critical_value_comment.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
df<-generate_missing(mtcars)</pre>
critical<-list(mpg=">20",am="=0")
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=comment,
                     numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
filename<-"excel_critical_value_comment_min_max.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
df<-generate_missing(mtcars)</pre>
critical<-list(mpg=c(">20","<11"),am="=0")</pre>
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=comment,
                     numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

excel_generic_format Generic function for creating workbooks and worksheets

Description

This function is used by excel_matrix and excel_critical_value functions

Usage

```
excel_generic_format(
```

```
df,
workbook,
sheet = "output",
title = NULL,
comment = NULL,
numFmt = "#0.00"
)
```

Arguments

df dataframe or matrix
workbook workbook
sheet sheet
title title
comment comment
numFmt number formatting

```
comment<-list(mpg="Miles/(US) gallon",</pre>
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio".
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1=manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")
mtcor<-data.frame(cor(mtcars))</pre>
filename<-"excel_generic.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()</pre>
openxlsx::addWorksheet(wb, "sheet")
openxlsx::addWorksheet(wb, "correlation")
openxlsx:: \verb|writeData| (wb, sheet="sheet", x=mtcars, colNames=TRUE, rowNames=TRUE)|
openxlsx::writeData(wb,sheet="correlation",x=mtcor,colNames=TRUE,rowNames=TRUE)
excel_generic_format(df=mtcars,workbook=wb,sheet="sheet",title="test",
                     comment=comment,numFmt="#0.00")
excel_generic_format(df=mtcor,workbook=wb,sheet="correlation",title="correlation",
                     comment=comment,numFmt="#0.00")
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

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excel_matrix

Write matrix or dataframe to excel sheet

Description

Usefull for corellation matrices. It uses conditional formatting for matrices, which outlines high and low values using background color

Usage

```
excel_matrix(
   df,
   workbook,
   sheet = "output",
   title = NULL,
   comment = NULL,
   numFmt = "#0.00",
   conditional_formatting = FALSE,
   diagonal = FALSE,
   diagonal_length = nrow(df)
)
```

Arguments

```
df
                  dataframe or matrix
                  workbook
workbook
sheet
                  sheet
title
                  title
comment
                 comment
numFmt
                 number formatting
conditional_formatting
                 if TRUE it will use conditional formatting
                 if TRUE it will add background fill to diagonal
diagonal
diagonal_length
                 length of diagonal for background fill
```

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```
vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1=manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")
mtcor<-data.frame(cor(mtcars))</pre>
filename<-"excel_matrix.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()</pre>
excel_matrix(mtcars,wb,sheet="matrix",comment=comment,
             conditional_formatting=TRUE,diagonal=FALSE)
excel_matrix(mtcars,wb,sheet="diagonal_non_square",comment=comment,
             conditional_formatting=FALSE,diagonal=TRUE)
excel_matrix(mtcars[1:10,1:10],wb,sheet="diagonal_square",comment=comment[1:10],
             conditional_formatting=FALSE,diagonal=TRUE)
excel_matrix(mtcars,wb,sheet="matrix_diagonal_non_square",comment=comment,
             conditional_formatting=TRUE,diagonal=TRUE)
excel_matrix(mtcars[1:10,1:10],wb,sheet="matrix_diagonal_square",comment=comment[1:10],
             conditional_formatting=TRUE,diagonal=TRUE)
excel_matrix(mtcor,wb,sheet="r",comment=comment,
             conditional_formatting=FALSE,diagonal=FALSE)
excel_matrix(mtcor,wb,sheet="conditional_formatting_r",comment=comment,
             conditional_formatting=TRUE,diagonal=TRUE)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

extract_components

Extract variance components from model

Description

Extract variance components from model

Usage

```
extract_components(model, title = "")
```

Arguments

model model containing variance components title plot title

```
design<-expand.grid(time=1:3,item=1:3,person=1:10)
design<-change_data_type(design,type="factor")
design$response<-rowSums(change_data_type(design[,1:2],type="numeric"))+rnorm(90,0,0.1)
model<-mixlm::lm(response~r(time)*r(person)+r(item)*r(person),data=design)
extract_components(model)</pre>
```

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flatten_list

Flatten two dimensional list

Description

Flatten two dimensional list

Usage

```
flatten_list(mydata)
```

Arguments

mydata

list with two dimensions

```
generate_comparisons_matrix
```

Generate comparisons matrix

Description

Generate comparisons matrix

Usage

```
generate_comparisons_matrix(items)
```

Arguments

items

number of items

```
generate_comparisons_matrix(2)
generate_comparisons_matrix(3)
generate_comparisons_matrix(4)
generate_comparisons_matrix(5)
generate_comparisons_matrix(6)
```

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```
generate_correlation_matrix
```

Generate dataframe which outputs a predetermined correlation matrix

Description

Generate dataframe which outputs a predetermined correlation matrix

Usage

```
generate_correlation_matrix(correlation_martix, nrows = 10)
```

Arguments

Examples

```
df<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-generate_correlation_matrix(correlation_martix,nrows=100)
stats::cor(df)</pre>
```

generate_data

Generate dataframe with random numbers

Description

Generate dataframe with random numbers

Usage

```
generate_data(
  nrows = 10,
  ncols = 5,
  mean = 0,
  sd = 1,
  min = 1,
  max = 5,
  type = "normal"
)
```

generate_factor 45

Arguments

nrows	number of rows to generate
ncols	number of collumns to generate
mean	mean of generated vectors
sd	standard deviation of generated vectors
min	minimum value in generated vector
max	maximum value in generated vector
type	character "normal" "uniform"

Examples

```
generate_data(nrows=10,ncols=5,mean=0,sd=1,type="normal")
generate_data(nrows=10,ncols=5,min=1,max=5,type="uniform")
```

generate_factor

Generate dataframe of factors

Description

Generate dataframe of factors

Usage

```
generate_factor(vector = LETTERS[1:5], nrows = 2, ncols = 10, type = "random")
```

Arguments

vector	factor pool
nrows	number of rows to generate
ncols	number of collumns to generate
type	"balanced" or "random" "balanced" generates balanced factor vectrors, "random" generates random factor vectors

```
generate_factor(vector=LETTERS[1:5],ncols=5,nrows=10,type="random")
generate_factor(vector=LETTERS[1:5],ncols=5,nrows=10,type="balanced")
generate_factor(vector=LETTERS[1:5],ncols=1,nrows=10,type="balanced")
generate_factor(vector=LETTERS[1:5],ncols=1,nrows=10,type="random")
```

generate_matrix_A

Generate Matrix A

Description

Generate Matrix A

Usage

```
generate_matrix_A(blocks = 3, items = 3)
```

Arguments

blocks number of blocks

items number of items per block

Examples

```
generate_matrix_A(blocks=3,items=3)
```

```
generate_matrix_lambda_hat
```

Generate matrix lambda for spesified number of comparisons

Description

Generate matrix lambda for spesified number of comparisons

Usage

```
generate_matrix_lambda_hat(blocks = 3, items = 3)
```

Arguments

blocks number of blocks

items number of items per block

```
generate_matrix_lambda_hat(blocks=3,items=4)
```

generate_missing 47

generate_missing

Generate missing data

Description

Generate missing data

Usage

```
generate_missing(df, missing = 5)
```

Arguments

df vector or dataframe

missing number of missing data per vector

Examples

```
generate_missing(rnorm(10),missing=5)
generate_missing(generate_data(nrow=10,ncol=2),missing=5)
```

```
generate_multiple_responce_vector
```

Generate multiple responce vector

Description

Generate multiple responce vector

Usage

```
generate_multiple_responce_vector(
  responces = 1:4,
  responded = 1:4,
  length = 10
)
```

Arguments

responces unique categories allowed

responded number of categories observed in iteration

length length of returned vector

```
generate_multiple_responce_vector(responces=1:4,responded=1:4,length=10)
```

generate_string

Generate random strings

Description

Generate random strings

Usage

```
generate_string(
  vector = c(LETTERS, letters, 0:9),
  vector_length = 1,
  nchar = 5
)
```

Arguments

vector character pool

vector_length number of strings to generate nchar Length of generated strings

Examples

```
generate_string(nchar=10)
generate_string(nchar=10,vector_length=10)
```

generate_unique_comparisons_index

Generate index for unique comparisons

Description

Generate index for unique comparisons

Usage

```
generate_unique_comparisons_index(items)
```

Arguments

items

number of items

getfwp 49

Examples

```
generate_unique_comparisons_index(1)
generate_unique_comparisons_index(2)
generate_unique_comparisons_index(3)
generate_unique_comparisons_index(4)
generate_unique_comparisons_index(5)
generate_unique_comparisons_index(6)
```

getfwp

Get working file path

Description

Get working file path

Usage

getfwp()

Examples

#getfwp()

get_mplus_thu_3t

Simulate prior distribution

Description

Simulate prior distribution

Usage

```
get_mplus_thu_3t(model)
```

Arguments

model

mplus thurstonian cfa model with 3 traits

50 increase_index

icc_cfa

Select responses for each dimension

Description

Select responses for each dimension

Usage

```
icc_cfa(eta, gamma, lambda, psi)
```

Arguments

eta eta or ability

gamma gamma or threshold lambda or loading

psi psi or error

Examples

```
icc_cfa(seq(-6,6,.1),1,1,1)
```

increase_index

index dataframe picks

Description

index dataframe picks

Usage

```
increase_index(blocks, items)
```

Arguments

blocks number of blocks

items number of items per block

```
increase_index(3,3)
```

install_all_packages 51

Description

Install all packages available in CRAN

Usage

```
install_all_packages()
```

Details

Install all packages available in CRAN. Already installed packages are not downloaded or installed

install_load

Install and load multiple packages

Description

Install and load multiple packages. If packages exist, they are loaded, if packages don't exist, they are downloaded installed and loaded

Usage

```
install_load(package)
```

Arguments

package

Vector Package names

Author(s)

Steven Worthington

```
install_load("car")
install_load(c("car", "ggplot2"))
```

52 k_fold

key_to_cfa_model

Converts key to cfa model spesification

Description

This function uses the key spesification used in report_alpha function and converts the key to a cfa model spesification

Usage

```
key_to_cfa_model(key)
```

Arguments

key

index of trait names and items constituring a trait

Examples

k_fold

K-Fold train test sampling

Description

splits a dataframe in train and test dataframes for model evaluation. Prepared data include data objects for xgboost

Usage

```
k_{fold}(df, model_{formula}, k = 10)
```

Arguments

```
df dataframe
model_formula
k k-folds
```

k_sample 53

Examples

```
infert_formula<-as.formula(factor(case)~age+parity+education+spontaneous+induced)
result<-k_fold(infert,k=10,model_formula=infert_formula)
model_formula<-as.formula(mpg~cyl+disp+hp+drat+wt+qsec+vs+am+gear+carb)
result<-k_fold(mtcars,k=2,model_formula=model_formula)</pre>
```

k_sample

train test sampling

Description

splits a dataframe in train and test dataframes for model evaluation. Prepared data include data objects for xgboost

Usage

```
k_sample(df, model_formula, k = 1)
```

Arguments

```
df dataframe
model_formula
k k-folds
```

Examples

```
infert\_formula <-as.formula(factor(case) \sim age+parity+education+spontaneous+induced) \\ result <-k\_sample(df=infert,k=10,model\_formula=infert\_formula) \\ model\_formula <-as.formula(mpg \sim cyl+disp+hp+drat+wt+qsec+vs+am+gear+carb) \\ result <-k\_sample(df=mtcars,k=10,model\_formula=model\_formula) \\ \end{aligned}
```

matrix_triangle

Return upper or lower matrix triangle

Description

Return upper or lower matrix triangle

Usage

```
matrix_triangle(m, off_diagonal = NA, diagonal = NULL, type = "lower")
```

54 mean_sd_alpha

Arguments

m matrix

off_diagonal off diagonal value

diagonal value. If NULL it returns the diagonal of the input matrix

type "upper" displays upper triangle, "lower" displays lower triangle

Examples

```
m<-matrix(1:9,nrow=3,ncol=3)
matrix_triangle(m=m)
matrix_triangle(m=m,diagonal=NA,type="lower")
matrix_triangle(m=m,diagonal=NULL,type="lower")
matrix_triangle(m=m,diagonal=NA,type="upper")
matrix_triangle(m=m,diagonal=NULL,type="upper")</pre>
```

mean_sd_alpha

Mean and SD

Description

Mean and SD

Usage

```
mean_sd_alpha(df, divisor = NULL)
```

Arguments

df dataframe with one dimension

divisor number to use for dividing the rowsums

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
mean_sd_alpha(df)
mean_sd_alpha(df,divisor=100)</pre>
```

mgsub 55

 ${\tt mgsub}$

Sub for multiple patterns

Description

Sub for multiple patterns

Usage

```
mgsub(mydata, pattern, replacement, ...)
```

Arguments

mydata Character

pattern Character to search for replacement Replacement character ... arguments passed to gsub

Examples

```
\label{eq:mgsub} \verb|mgsub| (\verb|mydata="#$%^&*_+", \verb|pattern=c("%","*"), "REPLACE", fixed=TRUE) \\
```

min_max_index

Return the minimum and maximum index of a vector

Description

Return the minimum and maximum index of a vector

Usage

```
min_max_index(vector)
```

Arguments

vector

Vector

56 model_loadings

Examples

```
vector1<-c(1,2,3,4,5,4,3,2,1)
vector2<-c(1,2,3,4,5,5,3,2,1)
vector3<-c(1,2,3,5,5,4,3,2,1)
vector4<-c(1,2,3,4,6,4,3,2,1)
vector5<-c(1,6,3,4,6,4,3,2,1)
vector<-vector1
which(vector==max(vector),arr.ind=TRUE)
which(vector==min(vector),arr.ind=TRUE)
min_max_index(vector1)
min_max_index(vector2)
min_max_index(vector3)
min_max_index(vector4)
min_max_index(vector5)</pre>
```

model_loadings

Pattern and structure matrix

Description

Pattern and structure matrix

Usage

```
model_loadings(model, cut = NULL, matrix_type = "pattern", sort = TRUE, ...)
```

Arguments

```
model psych EFA model

cut cut point for loadings

matrix_type "pattern" "structure" "all"

sort if TRUE it will sort loadings

... arguments passed to psych::fa.sort
```

Note

Check to see if you have multicolinearity values above .8 in the matrix are problematic Structure matrix represents Loadings after rotation

Pattern matrix represents Loadings before rotation

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
model_loadings(model=model,cut=NULL,matrix_type="pattern")
model_loadings(model=model,cut=0.4,matrix_type="structure")
model_loadings(model=model,cut=0.4,matrix_type="all",sort=FALSE)</pre>
```

off_diagonal_index 57

off_diagonal_index

index of off diagonal

Description

index of off diagonal

Usage

```
off_diagonal_index(length)
```

Arguments

length

length of diagonal

Examples

```
off_diagonal_index(length=6)
```

outlier_summary

Percent of outliers in vector

Description

Percent of outliers in vector

Usage

```
outlier_summary(vector)
```

Arguments

vector

numeric vector

Details

returns dataframe

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
outlier_summary(vector)
data.frame(sapply(mtcars,outlier_summary))</pre>
```

58 output_separator

```
output\_compare\_model\_logistic\\ Compare\ logistic\ regression\ models\ models
```

Description

Compare logistic regression models models

Usage

```
output_compare_model_logistic(model1, model2)
```

Arguments

```
model1 object glm model
model2 object glm model
```

Examples

output_separator

Output separator

Description

Heading, main output, and instructions for output for the console environment

Usage

```
output_separator(
   string,
   output = NULL,
   instruction = NULL,
   length = getOption("width")/2
)
```

padNA 59

Arguments

string Title of output
output object to print

instruction Character provided instructions regarding the output

length Numeric Length of separator measured in number of characters

Examples

```
output_separator(string="TEST",output="TEST",instruction="TEST",length=100)
output_separator(string="TEST",instruction="TEST",length=100)
output_separator(string="TEST",output="TEST",length=100)
output_separator(string="TEST")
```

padNA

pad NA's to collumns in dataframe

Description

pad NA's to collumns in dataframe

Usage

```
padNA(df, rowsneeded, first = TRUE)
```

Arguments

df dataframe

rowsneeded Numeric number of rows needed

first Boolean

Author(s)

Ananda Mahto

plot_boxplot

plot_acf

Plot autocorrelation function of correlation covariance and partial correlation

Description

uses ggplot

Usage

```
plot_acf(df, lag.max = length(df), base_size = 10, title = "")
```

Arguments

df ts object

lag.max maximum lags to include

base_size base font size
title plot title

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)
plot_acf(df=ts_data,base_size=20)</pre>
```

plot_boxplot

Boxplot

Description

Boxplot

Usage

```
plot_boxplot(df, title = "", base_size = 10)
```

Arguments

df dataframe or vector with continous or ordinal data

title Plot title

base_size numeric base font size

plot_cfa 61

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_boxplot(df=vector)
plot_boxplot(df=generate_missing(vector))
plot_boxplot(df=df)</pre>
```

plot_cfa

Plot cfa model

Description

Plot cfa model

Usage

```
plot_cfa(model, ...)
```

Arguments

model lavaan object

... arguments passed to semPlot::semPaths

62 plot_corrplot

plot_confusion

Plot confusion matrix

Description

Plot confusion matrix

Usage

```
plot_confusion(observed, predicted, base_size = 10, title = "")
```

Arguments

observed vector of observed outcomes predicted vector of predicted outcomes

base_size base font size title plot title

Examples

```
plot_confusion(observed=c(1,2,3,1,2,3),predicted=c(1,2,3,1,2,3))
observed<-c(rep("male",10),rep("female",10),"male","male")
predicted<-c(rep("male",10),rep("female",10),"female","female")
plot_confusion(observed=observed,predicted=predicted)</pre>
```

plot_corrplot

Correlation matrix plots

Description

Correlation matrix plots

Usage

```
plot_corrplot(mydata, title = "", base_size = 10, fill_limits = c(-1, 0, 1))
```

Arguments

mydata correlation matrix

title plot title base_size base font size

fill_limits lower and upper limit for fill

```
plot_corrplot(stats::cor(mtcars),title="Correlation")
plot_corrplot(stats::cor(mtcars),base_size=20)
```

plot_crosstable 63

plot_crosstable

Plot crosstables

Description

Plot crosstables

Usage

```
plot_crosstable(
   df,
   factor_index,
   combinations = NULL,
   shape = 16,
   angle = 0,
   base_size = 10,
   title = ""
)
```

Arguments

```
df dataframe
factor_index index of factors
combinations index of comparisons
shape shape of points
angle angle of xaxis labels
base_size base font size
title plot title
```

Examples

```
combinations<-data.frame(index1=c("vs","am","gear"),index2=c("cyl","cyl","cyl"))
plot_crosstable(df=mtcars,factor_index=8:9)
plot_crosstable(df=mtcars,combinations=combinations)</pre>
```

plot_histogram

Histograms with density function

Description

Histograms with density function

64 plot_icc_thurstonian

Usage

```
plot_histogram(
  df,
  bins = 30,
  title = "",
  base_size = 10,
  xlims = NULL,
  fill = "gray25",
  color = "gray50",
  ylab = "Count"
)
```

Arguments

df dataframe or vector with continous or ordinal data bins number of bars to display title plot title base_size numeric base font size xlims x axis limits fill color of bar color of bar outline color ylab y label

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_histogram(df=vector)
plot_histogram(df=df,xlims=c(0,50))
plot_histogram(df=df)
plot_multiplot(plotlist=plot_histogram(df=mtcars),cols=4)</pre>
```

Description

Plot icc curves for binary thurstonian coded items for a single dimension using the compute_icc_thurstonian function

plot_interaction 65

Usage

```
plot_icc_thurstonian(mydata, title = "Item Characteristic Curve")
```

Arguments

mydata dataframe from compute_icc_thurstonian function title plot title

Examples

```
 \begin{array}{l} {\rm gamma} < -c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498) \\ {\rm psi} < -c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686) \\ {\rm lambda} < -c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083) \\ {\rm gamma} < -{\rm gamma} [{\rm response\_dimension}(c(1:12),3,c(1,2))] \\ {\rm psi} < -{\rm psi} [{\rm response\_dimension}(c(1:12),3,c(1,2))] \\ {\rm eta} < -{\rm seq}(-6,6,by=1) \\ {\rm result} < -{\rm compute\_icc\_thurstonian}({\rm eta=eta,gamma=gamma,lambda=lambda,psi=psi,plot=TRUE}) \\ {\rm plot\_icc\_thurstonian}({\rm result\$icc}) \end{array}
```

plot_interaction

Plot two way interaction graphs

Description

Plot two way interaction graphs

Usage

```
plot_interaction(
   df,
   dv,
   iv,
   base_size = 20,
   type = "se",
   order_factor = TRUE,
   title = "",
   note = ""
)
```

Arguments

df dataframe

dv index of continous variables

iv index of factors base_size base font size

66 plot_irt_onefactor

type error bar type to display (1) "se" for standard error (2) "ci" for confidence inter-

val (3) "sd" for standard deviation (4) "" for no error bar

order_factor if TRUE it will sort the categorical axis by the continous variable value

title plot title note footnote

Examples

plot_irt_onefactor

Return data for irt plots

Description

Return data for irt plots

Usage

```
plot_irt_onefactor(model, theta = seq(-6, 6, 0.1), title = "", base_size = 10)
```

Arguments

model object mirt
theta theta
title plot title
base_size base size

```
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-4,high=4,d=NULL,a=1,mu=0,sd=1)$items
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Normal Test")
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-6,high=-4,d=NULL,a=1,mu=0,sd=1)$items
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Easy Items")
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=4,high=6,d=NULL,a=1,mu=0,sd=1)$items</pre>
```

plot_loadings 67

plot_loadings

Plot loadings

Description

Plot loadings

Usage

```
plot_loadings(
  model,
  matrix_type = NULL,
  title = "",
  base_size = 10,
  color = c("#5E912C", "white", "#5F2C91"),
  sort = TRUE
)
```

Arguments

```
model psych EFA model
matrix_type "pattern" "structure"
title plot title
base_size base font size
color color ranges for heatmap
```

68 plot_logistic_model

```
df1<-generate_correlation_matrix(cm,nrows=10000)</pre>
model1<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)</pre>
plot_loadings(model=model1,matrix_type="pattern",base_size=30)
cm<-matrix(c(1,.1,.1,.1,.1,.1,</pre>
              .1,1,.1,.1,.1,.1,
              .1, .1, 1, .1, .1, .1,
              .1, .1, .1, 1, .8, .8,
              .1, .1, .1, .8, 1, .8,
              .1,.1,.1,.8,.8,1),
              ncol=6,nrow=6)
df1<-generate_correlation_matrix(cm,nrows=10000)</pre>
model2<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)</pre>
plot_loadings(model=model2,matrix_type="pattern",base_size=30)
cm<-matrix(c(1,.01,.01,.01,.01,.01,</pre>
              .01,1,.01,.01,.01,.01,
              .01,.01,1,.01,.01,.01,
              .01,.01,.01,1,.01,.01,
              .01,.01,.01,.01,1,.01,
              .01,.01,.01,.01,.01,1),
              ncol=6,nrow=6)
df1<-generate_correlation_matrix(cm,nrows=10000)</pre>
model3<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)</pre>
plot_loadings(model=model3,matrix_type="pattern",base_size=10)
```

Description

Logistic model plot

Usage

```
plot_logistic_model(df, outcome = "outcome", title = "", base_size = 10)
```

Arguments

df dataframe with predictor and outcome outcome should be last

outcome name of outcome variable
title Character plot title
base_size base font size

plot_mosaic 69

plot_mosaic	Plot mosaic plots
-------------	-------------------

Description

Plot mosaic plots

Usage

```
plot_mosaic(df, factor_index, base_size = 10, title = "")
```

Arguments

df dataframe
factor_index index of factors
base_size base font size
title plot title

Examples

```
plot_mosaic(df=mtcars,factor_index=8:9)
plot_mosaic(df=mtcars,factor_index=9:10)
```

plot_mtmm

Plot multitrait multimethod matrix

Description

Plot multitrait multimethod matrix

Usage

```
plot_mtmm(df, key, method, subject, title = "")
```

Arguments

df	dataframe
----	-----------

key List index of trait names and items constituring a trait

method name of dataframe collumn spesifying the method used for the row observed

subject name of dataframe collumn spesifying subject id

title plot title

70 plot_multiplot

Examples

```
\label{eq:continuous} $\operatorname{population\_model} < -'t1 = -x1 + .9 + x2 + .9 + x3 \\ t2 = -x4 + .9 + x5 + .9 + x6 \\ t3 = -x7 + .9 + x8 + .9 + x9' \\ \operatorname{model\_data} < -\operatorname{lavaan} : : simulateData(population\_model, sample.nobs=1000) \\ \operatorname{model\_data} < -\operatorname{model\_data}[sample(1:1000,1000,TRUE),] \\ \operatorname{model\_data} < -\operatorname{rbind}(\operatorname{model\_data},\operatorname{model\_data},\operatorname{model\_data}) \\ \operatorname{model\_data} < -\operatorname{c(rep("m1",1000),rep("m2",1000),rep("m3",1000))} \\ \operatorname{model\_data} < -\operatorname{rep}(1:1000,3) \\ \operatorname{key} < -\operatorname{list}(t1 = \operatorname{paste0}("x",1:3),t2 = \operatorname{paste0}("x",4:6),t3 = \operatorname{paste0}("x",7:9)) \\ \operatorname{plot\_mtmm}(\operatorname{df=model\_data},\operatorname{key=key},\operatorname{method="method",subject="id"}) \\
```

plot_multiplot

Multiple ggplot plots in one graph

Description

Multiple ggplot plots in one graph

Usage

```
plot_multiplot(..., plotlist = NULL, cols = 2, layout = NULL)
```

Arguments

plotlist plot objects
plotlist a list of plots

cols number of columns in layout

layout a matrix specifying the layout. If present, 'cols' is ignored

```
p1<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet,group=Chick))+
           geom_line()+
           ggtitle("Growth curve for individual chicks")+
           theme_bw()
p2<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet))+
           geom_point(alpha=.3)+
           geom_smooth(alpha=.2,size=1,method="loess",formula="y~x")+
           ggtitle("Fitted growth curve per diet")+
           theme_bw()
p3<-ggplot(subset(ChickWeight,Time==21),aes(x=weight,colour=Diet))+
           geom_density()+
           ggtitle("Final weight, by diet")+theme_bw()
p4<-ggplot(subset(ChickWeight,Time==21),aes(x=weight,fill=Diet))+
           geom_histogram(colour="black",binwidth=50)+facet_grid(Diet~.)+
           ggtitle("Final weight, by diet")+theme_bw()
cars_plot<-plot_histogram(mtcars)</pre>
```

```
plot_multiplot(p1,p2,p3,p4,cols=2)
plot_multiplot(plotlist=plot_histogram(mtcars[,1:4]),cols=2)
plot_multiplot(plotlist=plot_histogram(mtcars),layout=matrix(1:4,ncol=2,byrow=TRUE))
plot_multiplot(plotlist=plot_scatterplot(mtcars[,1:4]),cols=2)
plot_multiplot(plotlist=cars_plot,layout=matrix(1:4,ncol=2,byrow=TRUE))
plot_multiplot(plotlist=cars_plot,cols=3)
```

plot_normality_diagnostics

Normality plots

Description

plot histogram density boxplot qq plot

Usage

```
plot_normality_diagnostics(
   df,
   breaks = NULL,
   title = "",
   file = NULL,
   w = 10,
   h = 10
)
```

Arguments

```
df dataframe or vector with continous or ordinal data breaks number of bars to display title plot title plot title output filename w width of pdf file h height of pdf file
```

Details

uses plot base

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_normality_diagnostics(df=vector,title="",file="rnorm",breaks=30)
plot_normality_diagnostics(df=vector,title="")
plot_normality_diagnostics(df=df,title="mtcars")
plot_normality_diagnostics(df=df,title="mtcars",file="rnorm")</pre>
```

72 plot_oneway

plot_oneway

Plot means with standard error for every level in a dataframe

Description

Plot means with standard error for every level in a dataframe

Usage

```
plot_oneway(
   df,
   dv,
   iv,
   base_size = 20,
   type = "se",
   order_factor = TRUE,
   title = "",
   note = "",
   width = 60
)
```

Arguments

```
df
                   dataframe
dν
                   index of continous variables
                   index of factors
i٧
                   base font size
base_size
                   error bar type to display (1) "se" for standard error (2) "ci" for confidence inter-
type
                   val (3) "sd" for standard deviation (4) "" for no error bar
order_factor
                   if TRUE it will sort the categorical axis by the continous variable value
title
                   plot title
note
                   footnote
width
                   wrap width for x title
```

plot_oneway_diagnostics

```
plot_oneway(df=mtcars,dv=2:3,iv=9:10,type="ci")
plot_oneway(df=mtcars,dv=2:3,iv=9:10,type="sd")
plot_oneway(df=mtcars,dv=2:3,iv=9:10,type="",order_factor=FALSE)
plot_oneway(df=mtcars,dv=2:3,iv=9:10,type="",order_factor=TRUE)
```

```
plot_oneway_diagnostics
```

Plot one way diagnostics

Description

Plot one way diagnostics

Usage

```
plot_oneway_diagnostics(df, dv, iv, base_size = 10)
```

Arguments

df	dataframe

dv index of continous variables

iv index of factors base_size base font size

Note

Residuals vs Fitted should be equally spread horizontally otherwize the assumption of equality of variances is violated

Normal QQ should show values in the diagonal otherwise the assumption of normality is violated

74 plot_qq

plot_outlier

Outlier graph using mean median and boxplot algorythms

Description

Outlier graph using mean median and boxplot algorythms

Usage

```
plot_outlier(df, method = "mean", title = "", base_size = 10)
```

Arguments

df dataframe or vector with continous or ordinal data

method "mean" "median" "boxplot"

title plot title base_size base font size

Author(s)

unknown

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_outlier(df=vector,method="mean",title="random vector")
plot_outlier(df=vector,method="median")
plot_outlier(df=vector,method="boxplot")
plot_outlier(df=df,method="mean",title="random vector")
plot_outlier(df=df,method="median")
plot_outlier(df=df,method="boxplot")
plot_multiplot(plotlist=plot_outlier(df=mtcars[,2:5],method="mean"),cols=2)</pre>
```

plot_qq

qq plots

Description

qq plots

Usage

```
plot_qq(df, title = "", base_size = 10)
```

Arguments

df dataframe or vector with continous or ordinal data

title plot title

base_size numeric base font size

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_qq(df=vector)
plot_qq(df=df)
plot_multiplot(plotlist=plot_qq(df=mtcars),cols=4)</pre>
```

```
plot_response_frequencies
```

Plot response frequencies

Description

Plot response frequencies

Usage

```
plot_response_frequencies(
   df,
   factor_index,
   base_size = 10,
   title = "",
   width = 100
)
```

Arguments

```
df dataframe
factor_index index of factors
base_size base font size
title plot title
```

width wrap width for x title

```
plot_response_frequencies(df=mtcars,factor_index=1:10)
```

76 plot_scatterplot

plot_roc

Plot ROC curve

Description

Plot ROC curve

Usage

```
plot_roc(observed, predicted, base_size = 10, title = "")
```

Arguments

observed vector of observed outcomes

predicted vector of predicted outcome probability

base_size base font size title plot title

Examples

```
observed<-round(abs(rnorm(100,m=0,sd=.5)))
predicted<-abs(rnorm(100,m=0,sd=.5))
plot_roc(observed=observed,predicted=predicted)
df1<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_martix<-as.matrix(df1)
diag(correlation_martix)<-1
df1<-generate_correlation_matrix(correlation_martix,nrows=1000)
df1$X1<-ifelse(abs(df1$X1)<1,0,1)
df1$X2<-abs(df1$X2)
df1$X2<-(df1$X2-min(df1$X2))/(max(df1$X2)-min(df1$X2))
plot_roc(observed=round(abs(df1$X1),0),predicted=abs(df1$X2))</pre>
```

plot_scatterplot

Plot plot_scatterplot

Description

Plot plot_scatterplot

plot_scatterplot 77

Usage

```
plot_scatterplot(
    df,
    method = lm,
    formula = y ~ x,
    base_size = 10,
    coord_equal = FALSE,
    all_orders = FALSE,
    title = "",
    combinations = NULL,
    string_aes = TRUE
)
```

Arguments

df dataframe if dataframe consists of 2 collumns the second collumn is the outcome

and the first collumn is the predictor

method smoothing method, "auto", "lm", "glm", "gam", "loess" or a function, e.g. MASS::rlm

or mgcv::gam, stats::lm, or stats::loess

formula formula used in smoothing function for geom_smooth

base_size base font size

coord_equal if TRUE axes maintain equal scale

all_orders if TRUE the order of combination is considered

title Plot title

combinations dataframe if not NULL user can provide a dataframe for variable combinations

for x and y axis. First collumn represents x and second collumn represents y

string_aes if TRUE string_aes function is used for names

```
result<-plot_scatterplot(df=mtcars,title="",coord_equal=TRUE,base_size=10)
plot_multiplot(plotlist=result[1:12],cols=4)
plot_scatterplot(df=mtcars[,1:2],base_size=10,coord_equal=TRUE,all_orders=FALSE)
plot_scatterplot(df=mtcars[,1:2],base_size=10,coord_equal=FALSE,all_orders=FALSE)
plot_scatterplot(df=mtcars,base_size=10,coord_equal=TRUE,all_orders=FALSE,
                 combinations=data.frame(x=c("mpg", "mpg", "mpg"),
                                          y=c("cyl","hp","mpg")))
plot_scatterplot(df=mtcars,base_size=10,coord_equal=TRUE,all_orders=TRUE,
                 combinations=data.frame(x=c("mpg"),y=c("cyl")))
x<-rnorm(1000)
y < -x + rnorm(x, sd = .1)
plot_scatterplot(df=data.frame(x,y),title="Random Simulation",coord_equal=TRUE)
df<-data.frame(matrix(-.999,ncol=2,nrow=2))</pre>
correlation_martix<-as.matrix(df)</pre>
diag(correlation_martix)<-1
df<-generate_correlation_matrix(correlation_martix,nrows=1000)</pre>
plot_scatterplot(df,title="Simulation of -.999 Correlation",coord_equal=TRUE,base_size=20)
```

78 plot_separability

plot_scree	Scree plot displaying the Kaiser and Jolife criteria for factor extraction

Description

Scree plot displaying the Kaiser and Jolife criteria for factor extraction

Usage

```
plot_scree(df, base_size = 15, title = "", color = c("#5F2C91", "#5E912C"))
```

Arguments

df dataframe
base_size base font size
title plot title

color color of line and point outline

Examples

```
plot_scree(df=mtcars,title="",base_size=15)
```

plot_separability
Plot separability

Description

Plot separability

Usage

```
plot_separability(observed, predicted, base_size = 10, title = "")
```

Arguments

observed vector of observed outcomes

predicted vector of predicted outcome probability

base_size base font size title plot title

plot_trees_xgboost 79

Examples

```
df1<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_martix<-as.matrix(df1)
diag(correlation_martix)<-1
df1<-generate_correlation_matrix(correlation_martix,nrows=1000)
df1$X1<-ifelse(abs(df1$X1)<1,0,1)
df1$X2<-abs(df1$X2)
df1$X2<-(df1$X2-min(df1$X2))/(max(df1$X2)-min(df1$X2))
plot_separability(observed=round(abs(df1$X1),0),predicted=abs(df1$X2))</pre>
```

plot_trees_xgboost

Plot trees for xgboost::xgb.train

Description

Plot trees for xgboost::xgb.train

Usage

```
plot_trees_xgboost(model, train, file = "xgboost")
```

Arguments

model object from xgboost::xgb.train

train Train dataset file output filename

```
infert_formula<-as.formula(factor(case)~education+spontaneous+induced)</pre>
boston\_formula <-as.formula(c("medv^",paste(names(MASS::Boston)[1:13],collapse="+")))
train_test_classification<-workingfunctions::k_fold(df=infert,model_formula=infert_formula)
train_test_regression<-workingfunctions::k_fold(df=MASS::Boston,model_formula=boston_formula)</pre>
xgb_classification<-xgboost::xgb.train(data=train_test_classification$xgb$f1$train,
                                   watchlist=train_test_classification$xgb$f1$watchlist,
                                       nthread=8,
                                       nround=20,
                                       objective="binary:logistic")
xgb_regression<-xgboost::xgb.train(data=train_test_regression$xgb$f1$train,
                                   watchlist=train_test_regression$xgb$f1$watchlist,
                                   eta=.3,
                                   nthread=8,
                                   nround=20)
xgboost::xgb.plot.multi.trees(model = xgb_classification, features_keep = 2)
plot_trees_xgboost(model=xgb_classification,train=train_test_classification$xgb$f1,file="Classification")
plot_trees_xgboost(model=xgb_regression,train=train_test_regression$xbg$f1,file="Regression")
```

proper proper

plot_ts

Plot timeseries

Description

Plot timeseries

Usage

```
plot_ts(df, base_size = 10, ylab = "Count", title = "")
```

Arguments

df ts object
base_size base font size
ylab y label
title plot title

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)
result<-plot_ts(ts_data,title="UK driver deaths")
for(i in 1969:1984)
    result<-result+geom_vline(xintercept=i,color="blue",size=1,alpha=.5)
result
autoplot(stl(ts_data,s.window='periodic'))+
    theme_bw(base_size=10)+
    labs(title="UK driver deaths")
forecast::gglagplot(data.frame(ts_data),do.lines=FALSE,lags=100)+
    theme_bw(base_size=10)+labs(title="UK driver deaths",y="count")</pre>
```

proper

Capitalize first character and lowercase the rest

Description

Capitalize first character and lowercase the rest

Usage

```
proper(x)
```

proportion_accurate 81

Arguments

x Character

Examples

```
x<-generate_string(nchar=10,vector=LETTERS,vector_length=10)
proper(x)</pre>
```

proportion_accurate

Proportion overall accuracy of a confusion matrix

Description

Proportion overall accuracy of a confusion matrix

Usage

```
proportion_accurate(observed, predicted)
```

Arguments

observed vector of observed variables predicted vector of predicted variables

Examples

```
proportion_accurate(observed=c(1,2,3,4,5,10),predicted=c(1,2,3,4,5,11))
```

questions_by_keys

Convert key to index list

Description

Convert key to index list

Usage

```
questions_by_keys(key)
```

Arguments

key

a vector indicating the dimension of each question. The order of the elements in the key represents the order of the questions, the numeric values represent the dimension the question belongs to

```
key<-c(1,2,3,4,5,1,2,3,4,5)
questions_by_keys(key)</pre>
```

82 rad2deg

```
{\it questions\_dimensions\_data} \\ {\it question~dimension~table}
```

Description

Return a dataframe with the order of the questions, their respective dimensions, and the description of the dimensions

Usage

```
questions_dimensions_dataframe(
  key,
  dimensions,
  elaborate_dimensions,
  questions
)
```

Arguments

key a vector indicating the dimension of each question. The order of the elements in

the key represents the order of the questions, the numeric values represent the

dimension the question belongs to

dimensions dimension names

elaborate_dimensions

full dimension names

questions question names

Examples

```
key<-c(1,2,3,4,5,1,2,3,4,5)
dimensions<-paste0("Dimension",1:10)
elaborate_dimensions<-paste0("Elaborated_Dimension",1:10)
questions<-paste0("Question",1:65)
questions_dimensions_dataframe(key,dimensions,elaborate_dimensions,questions)</pre>
```

rad2deg

Convert radians to degrees

Description

Convert radians to degrees

Usage

```
rad2deg(radians)
```

rank3_to_triplets 83

Arguments

radians radians

Examples

rad2deg(pi)

rank3_to_triplets

Convert thurstonian binary triplets to scale

Description

Convert thurstonian binary triplets to scale

Usage

```
rank3_to_triplets(mydata)
```

Arguments

mydata

dataframe

Examples

rank_df_to_binary

Convert scale to thurstonian binary with n items per block and n blocks

Description

Convert scale to thurstonian binary with n items per block and n blocks

Usage

```
rank_df_to_binary(mydata, items, reverse = TRUE)
```

84 rank_to_binary

Arguments

mydata dataframe

items number of items in block

reverse if TRUE assumes that the highest value is first item in rank if FALSE the lowest

value is the first item in rank

Examples

rank_to_binary

Convert scale to thurstonian binary with n items per ranking block

Description

Convert scale to thurstonian binary with n items per ranking block

Usage

```
rank_to_binary(mydata, items, reverse = TRUE)
```

Arguments

mydata dataframe

items number of items in block

reverse if TRUE assumes that the highest value is first item in rank if FALSE the lowest

value is the first item in rank

raw_alpha 85

raw_alpha

Raw alpha

Description

Raw alpha

Usage

```
raw_alpha(df)
```

Arguments

df

dataframe with one dimension

Examples

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
psych::alpha(df)
raw_alpha(df=df)</pre>
```

rbind_all

rbind dataframes or matrices with different lengths or collumn names

Description

rbind dataframes or matrices with different lengths or collumn names

Usage

```
rbind_all(df1, df2)
```

Arguments

df1 dataframe or matrix df2 dataframe or matrix

```
df1<-generate_correlation_matrix(n=10)
df2<-generate_correlation_matrix(n=10)
names(df2)[4]<-"X11"
rbind_all(df1=df1,df2=df2)
row.names(df1)<-21:30
rbind_all(df1=df1,df2=df2)</pre>
```

86 remove_nc

recode_scale_dummy

Scale and dummy code

Description

Scales numeric variables between 0 and 1 and creates dummy coding for character and vector variables

Usage

```
recode_scale_dummy(df, categories = 10)
```

Arguments

df dataframe

categories

Numeric Number of unique values a vector must have to perform dummy coding

Examples

```
recode_scale_dummy(infert)
```

remove_nc

Replace remove non computable values

Description

Replace remove non computable values

Usage

```
remove_nc(
   df,
   value = NA,
   remove_rows = FALSE,
   aggressive = FALSE,
   remove_cols = FALSE,
   remove_zero_variance = FALSE
)
```

remove_outliers 87

Arguments

df	dataframe
value	replacement
remove_rows	if TRUE it will remove rows with non computable values
aggressive	if TRUE it will remove entire row if a single non computable value exists if FALSE it will remove row if all values are non computable
remove_cols	if TRUE it will remove collumns with non computable values
remove_zero_variance	
	if TRUE it will remove collumns with no variance

Details

Non computable values are NA, NAN, inf and empty cells.

Note

This function internally replaces non computable values with the value choosen the default value is NA. Then it removes rows and collumns with NA values or zero variance

Examples

```
df<-mtcars
df[1,]<-as.numeric(NaN)</pre>
df[2,]<-as.numeric(Inf)</pre>
df[3,]<-as.numeric(-Inf)</pre>
df[4,]<-as.numeric(NA)</pre>
df[5,]<-""
remove_nc(df=df,value=NA)
cdf(remove_nc(df=df,value=NA))
df<-generate_missing(mtcars,missing=5)</pre>
remove_nc(df,remove_rows=TRUE,aggressive=FALSE)
remove_nc(df,remove_rows=TRUE,aggressive=TRUE)
df<-generate_missing(generate_correlation_matrix(nrows=5),missing=2)</pre>
df$X2<-NA
df$X3<-1
remove_nc(df,remove_cols=TRUE,remove_zero_variance=FALSE)
remove_nc(df,remove_cols=TRUE,remove_zero_variance=TRUE)
```

remove_outliers Remove outliers

Description

Remove outliers

Usage

```
remove_outliers(vector, probs = c(0.25, 0.75), na.rm = TRUE, ...)
```

Arguments

vector numeric

probs numeric vector with lowest and highest quantiles

na.rm if TRUE removes NA values
... arguments passed to quantile

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
remove_outliers(vector)
data.frame(sapply(df,remove_outliers))</pre>
```

Description

Remove all user packages

Usage

```
remove_user_packages()
```

```
replace_na_with_previous
```

Replace NA with the previous element in a vector

Description

Replace NA with the previous element in a vector

Usage

```
replace_na_with_previous(vector)
```

Arguments

vector Vector

report_alpha 89

Examples

```
df1<-generate_missing(rnorm(10), missing=5)
df2<-generate_missing(rnorm(10), missing=5)
df3<-generate_missing(rnorm(10), missing=5)
df4<-generate_missing(rnorm(10), missing=5)
df5<-generate_missing(rnorm(10), missing=5)
df<-data.frame(df1,df2,df3,df4,df5)
row.names(df)<-paste0("A",row.names(df))
replace_na_with_previous(df1)
df[]<-lapply(df,replace_na_with_previous)</pre>
```

report_alpha

Estimate alpha for several dimensions and export results to xlsx

Description

Uses an arbitrary input

Usage

```
report_alpha(
   df,
   key = NULL,
   questions = NULL,
   reverse = NULL,
   mini = NULL,
   maxi = NULL,
   file = NULL,
   ...
)
```

df	dataframe
key	index of trait names and items constituting a trait
questions	trait names and items constituting a trait
reverse	index of trait names and index for reversal
mini	minimum rating in scale if NULL reversal will be performed using the empirical minimum
maxi	maximum rating in scale if NULL reversal will be performed using the empirical maximum
file	output filename
	arguments passed to psych::alpha

90 report_cfa

Examples

report_cfa

Report

Description

Report

Usage

```
report_cfa(model, file = NULL, w = 10, h = 10)
```

Arguments

model	lavaan object
file	output filename
W	width of pdf file
h	height of pdf file

report_choric_serial 91

report_choric_serial Report polychoric tetrachoric polyserial biserial correlation

Description

Report polychoric tetrachoric polyserial biserial correlation

Usage

```
report_choric_serial(
    X,
    y = NULL,
    file = NULL,
    w = 10,
    h = 10,
    type = "tetrachoric",
    ...
)
```

Arguments

x The input may be in one of four forms:

- a) a data frame or matrix of dichotmous data (e.g., the lsat6 from the bock data set) or discrete numerical (i.e., not too many levels, e.g., the big 5 data set, bfi) for polychoric, or continuous for the case of biserial and polyserial
- b) a 2 x 2 table of cell counts or cell frequencies (for tetrachoric) or an n x m table of cell counts (for both tetrachoric and polychoric)
- c) a vector with elements corresponding to the four cell frequencies (for tetrachoric)
- d) a vector with elements of the two marginal frequencies (row and column) and the comorbidity (for tetrachoric)

y matrix or dataframe of discrete scores. In the case of tetrachoric, these should be dichotomous, for polychoric not too many levels, for biserial they should be

discrete (e.g., item responses) with not too many (<10?) categories

file output filename

w width of pdf file

h height of pdf file

type "tetrachoric" "polychoric" "polyserial" "biserial"

... arguments passed to psych::polychoric

92 report_correlation

 $report_correlation$

Report correlation matrix

Description

Report correlation matrix

Usage

```
report_correlation(
    x,
    y = NULL,
    use = "pairwise",
    method = "pearson",
    adjust = "holm",
    alpha = 0.05,
    ci = TRUE,
    file = NULL,
    w = 10,
    h = 10,
    base_size = 20,
    scatterplot = TRUE
)
```

X	matrix or dataframe
У	a second matrix or dataframe with the same number of rows as x
use	"pairwise" is the default value and will do pairwise deletion of cases. "complete" will select just complete cases
method	"pearson" "spearman" "kendall"
adjust	"holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none"
alpha	alpha level of confidence intervals
ci	By default, confidence intervals are found. However, this leads to a great slow-down of speed. So, for just the rs, ts and ps, set ci=FALSE
file	output filename
W	width of pdf file
h	height of pdf file
base_size	base font size
scatterplot	if TRUE it will outpu scatterplots

report_dataframe 93

Examples

report_dataframe

Write matrix or dataframe to excel sheet

Description

Usefull for generic data where conditional formating of a spesific collumn is required

Usage

```
report_dataframe(df, file = NULL, type = "critical_value", ...)
```

Arguments

```
df dataframe or matrix

file output filename of excel file

type "critical_value" "matrix"

... arguments passed to excel_critical_value or to excel_matrix
```

94 report_efa

report_efa

Output EFA model

Description

Output EFA model

Usage

```
report_efa(
  model,
  df,
  file = NULL,
  w = 10,
  h = 5,
  cut = 0,
  base_size = 10,
  scores = FALSE
)
```

Arguments

```
model psych EFA model

df dataframe

file output filename

w width of pdf file

h height of pdf file

cut cut point for loadings

base_size base font size

scores if TRUE it will output factor scores in excel file
```

Note

Orthogonal=varimax, Oblique=oblimin

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minres",oblique.scores=TRUE)
report_efa(model=model,df=mtcars,file="efa")
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="uls",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="ols",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="wls",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="gls",oblique.scores=TRUE)</pre>
```

report_factorial_anova 95

```
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="m1",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minchi",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minrank",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="old.min",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="alpha",oblique.scores=TRUE)
#report_efa(model=model,df=mtcars)
#report_efa(model=model,df=mtcars)</pre>
```

report_factorial_anova

Plot means with standard error for every level in a dataframe

Description

Plot means with standard error for every level in a dataframe

Usage

```
report_factorial_anova(
  df,
  dν,
 wid,
 within = NULL,
 within_full = NULL,
 between = NULL,
 within_covariates = NULL,
 between_covariates = NULL,
  observed = NULL,
  diff = NULL,
  reverse_diff = FALSE,
  type = 3,
  white.adjust = TRUE,
  detailed = TRUE,
  return_aov = TRUE,
  file = NULL,
  post_hoc_test = TRUE,
 base\_size = 15
)
```

Arguments

df

dataframe

dv names of dependent variables

wid names of

within names of within factors

within_full names of within factors after data are collapsed to means per condition

between names of between factors

within_covariates

names of within covariates

between_covariates

mames of between covariates

observed names in data that are already specified in either within or between that contain

predictor variables that are observed variables (not manipulated)

diff names of variables to collapse in a different score

reverse_diff If TRUE, triggers reversal of the difference collapse requested by diff

type sum of squares 1 2 3

white.adjust if TRUE corrects for heteroscedasticity detailed if TRUE returns detailed information

return_aov if TRUE returns aov object

file output filename

post_hoc_test if TRUE outputs post hoc in file

base_size base font size

```
set.seed(12345)
df<-data.frame(id=rep(seq(1,80),each=81,1),</pre>
                IV1=rep(LETTERS[1:3],each=1,2160),
                IV2=rep(LETTERS[4:6],each=3,720),
                IV3=rep(LETTERS[7:9],each=9,240),
               IV4=rep(LETTERS[10:12], each=27,80),
                stringsAsFactors=FALSE)
cdf<-data.frame(matrix(.01,ncol=4,nrow=4))</pre>
correlation_martix<-as.matrix(cdf)</pre>
diag(correlation_martix)<-1</pre>
cdf<-generate_correlation_matrix(correlation_martix,nrows=nrow(df))+10
names(cdf)<-paste0("DV",1:4)</pre>
df<-data.frame(df,cdf)</pre>
df$DV2<-df$DV2+10
df$DV3<-df$DV3+20
df$DV4<-df$DV4+30
df[df$IV1%in%"A",]$DV1<-df[df$IV1%in%"A",]$DV1+1
df[df$IV1%in%"B",]$DV1<-df[df$IV1%in%"B",]$DV1+2
df[df$IV1%in%"C",]$DV1<-df[df$IV1%in%"C",]$DV1+3
cdf(df)
r1<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                            within=c("IV1","IV2"),within_full=c("IV1","IV2"),
                            between=NULL,
```

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```
within_covariates=NULL, between_covariates=NULL,
                           file="anova_within",
                           post_hoc=TRUE)
r2<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=NULL, within_full=NULL,
                           between=c("IV1","IV2"),
                           within_covariates=NULL, between_covariates=NULL,
                           file="anova_between",
                           post_hoc=TRUE)
r3<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=c("IV3","IV4"),within_full=c("IV3","IV4"),
                           between=c("IV1","IV2"),
                           within_covariates=NULL, between_covariates=NULL,
                           file="anova_mixed",
                           post_hoc=FALSE)
r4<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=c("IV1","IV2"),within_full=c("IV1","IV2"),
                           between=NULL,
                           within_covariates=c("DV3","DV4"),between_covariates=NULL,
                           file="anova_within_cov",
                           post_hoc=TRUE)
```

report_hlr

Report HLR

Description

Report HLR

Usage

```
report_hlr(
   df,
   corlist,
   factorlist,
   predictor,
   random_effect,
   file = NULL,
   sheet = "report"
)
```

Arguments

df dataframe

corlist Numeric outcome index
factorlist Numeric predictor index
predictor Character predictor name
random_effect Character random effect name

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```
file Character file sheet Character sheet
```

Examples

report_irt

Output for irt model

Description

Output for irt model

Usage

```
report_irt(model, m2 = TRUE, file = NULL)
```

Arguments

model object mirt

m2 if TRUE report m2 statistics

file output filename

```
set.seed(12345)
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-4,high=4,d=NULL,a=1,mu=0,sd=1)$items
irt_onefactor<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
irt_twofactor<-mirt::mirt(cormatrix,2,empiricalhist=TRUE,calcNull=TRUE)
irt_threefactor<-mirt::mirt(cormatrix,3,empiricalhist=TRUE,calcNull=TRUE)
report_irt(model=irt_onefactor,file="one_factor")
report_irt(model=irt_twofactor,file="two_factors")
report_irt(model=irt_threefactor,file="three_factors")</pre>
```

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report_lda

Report for MASS::lda

Description

Report for MASS::lda

Usage

```
report_lda(model, file = NULL, w = 10, h = 10, base_size = 10, title = "")
```

Arguments

```
model object from MASS::lda
file output filename
w width of pdf file
h height of pdf file
base_size base font size
title plot title
```

Examples

```
model<-MASS::lda(case~.,data=infert)
result<-report_lda(model=model)
result<-report_lda(model=model,file="lda")
model<-MASS::lda(Species~.,data=iris)
result<-report_lda(model=model,file="lda")</pre>
```

report_logistic

Report logistic regression

Description

Report logistic regression

Usage

```
report_logistic(
  model,
  validation_data = NULL,
  file = NULL,
  title = "",
  w = 10,
  h = 10,
  base_size = 10,
  fast = FALSE
)
```

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Arguments

model object glm validation_data

validation data

file output filename

title plot title

w width of pdf file. Relevant only when file string is not emptyh height of pdf file. Relevant only when file string is not empty

base_size base font size

fast if TRUE it will not output individual scores and residuals

Note

(1) Problematic values for standardized residuals > +-1.96

Standardized residuals are residuals divided by an estimated standard deviation and they can be interpreted as z scores in that:

95 99 99.99 (2) Problematic values for dfBeta >=1

dfBeta estimates coefficients if the respective case is removed from the dataset

(3) Problematic values for Hat values (leverage) 2 or 3 times the average (k+1/n)

Hat values (leverage), gauge the influence of the observed value of the outcome variable over the predicted values

The average leverage value is defined as (k+1)/n, k=number of predictors, n=number of participants. Leverage values lie between 0 (no influence) and 1 (complete influence over prediction)

If no cases exert undue influence over the model then all leverage values should be close to (k+1)/n Hoaglin and Welsch (1978) recommends investigating cases with values greater than twice the average (2(k+1)/n)

Stevens (2002) recommends investigating cases with values greater than three times the average (3(k+1)/n)

(4) Problematic values for VIFs > 10

ASSUMPTIONS

- (1) Linearity between continous predictors and the logit (test wether the interaction term between the predictor and its log transformation is significant)
- (2) Independence of errors
- (3) No multicolinearity

```
modelcategoricalpredictor0<-glm(case~education,data=infert,family=binomial)
modelcategoricalpredictor1<-glm(case~education,data=infert,family=gaussian)
#modelcategoricalpredictor2<-glm(case~education,data=infert,family=Gamma)
#modelcategoricalpredictor3<-glm(case~education,data=infert,family=inverse.gaussian)
modelcategoricalpredictor4<-glm(case~education,data=infert,family=poisson)
modelcategoricalpredictor5<-glm(case~education,data=infert,family=quasi)
modelcategoricalpredictor6<-glm(case~education,data=infert,family=quasibinomial)
modelcategoricalpredictor7<-glm(case~education,data=infert,family=poisson)
modelcontinuouspredictor0<-glm(case~stratum,data=infert,family=binomial)</pre>
```

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```
modeltwopredictors0<-glm(case~education+stratum,data=infert,family=binomial)
modeltwopredictors1<-glm(case~education+stratum,data=infert,family=gaussian)
#modeltwopredictors2<-glm(case~education+stratum,data=infert,family=Gamma)</pre>
#modeltwopredictors3<-glm(case~education+stratum,data=infert,family=inverse.gaussian)</pre>
modeltwopredictors4<-glm(case~education+stratum,data=infert,family=poisson)</pre>
modeltwopredictors5<-glm(case~education+stratum,data=infert,family=quasi)</pre>
modeltwopredictors6<-glm(case~education+stratum,data=infert,family=quasibinomial)
modeltwopredictors7<-glm(case~education+stratum,data=infert,family=quasipoisson)
report_logistic(model=modelcategoricalpredictor0)
report_logistic(model=modelcategoricalpredictor1)
#report_logistic(model=modelcategoricalpredictor2)
#report_logistic(model=modelcategoricalpredictor3)
report_logistic(model=modelcategoricalpredictor4)
report_logistic(model=modelcategoricalpredictor5)
report_logistic(model=modelcategoricalpredictor6)
report_logistic(model=modelcategoricalpredictor7)
report_logistic(model=modelcontinuouspredictor0)
report_logistic(model=modeltwopredictors0)
report_logistic(model=modelcategoricalpredictor0,
                file="logistic_categorical_predictor",
                validation_data=infert)
report_logistic(model=modelcontinuouspredictor0,
                file="logistic_continuous_predictor",
                validation_data=infert)
report_logistic(model=modeltwopredictors0,
                file="logistic_two_predictors",
                validation_data=infert[1:10,])
```

report_manova

Manova result

Description

Manova result

Usage

```
report_manova(model, file = NULL)
```

Arguments

model object of manova model

file output filename

Note

Pillai-Bartlett trace (V): Represents the sum of the proportion of explained variance on the discriminant functions. As such, it is similar to the ratio of SS M /SS T, which is known as R 2.

Hotelling-s T 2: Represents the sum of the eigenvalues for each variate it compares directly to the

F-ratio in ANOVA

Wilks-s lambda (L): Represents the ratio of error variance to total variance (SS R /SS T) for each variate.

Roy-s largest root: Represents the proportion of explained variance to unexplained variance (SS M /SS R) for the first discriminant function.

ASSUMPTIONS

Independence: Observations should be statistically independent.

Random sampling: Data should be randomly sampled from the population of interest and measured at an interval level.

Multivariate normality: In ANOVA,we assume that our dependent variable is normally distributed within each group. In the case of MANOVA,we assume that the dependent variables (collectively) have multivariate normality within groups.

Homogeneity of covariance matrices: In ANOVA, it is assumed that the variances in each group are roughly equal (homogeneity of variance). In MANOVA we must assume that this is true for each dependent variable, but also that the correlation between any two dependent variables is the same in all groups. This assumption is examined by testing whether the population variance-covariance matrices of the different groups in the analysis are equal.

Examples

```
## Set orthogonal contrasts.
op<-options(contrasts=c("contr.helmert","contr.poly"))
model_mixed<-manova(cbind(yield,foo)~N*P*K,within(npk,foo<-rnorm(24)))
model_between<-manova(cbind(rnorm(24),rnorm(24))~round(rnorm(24),0)*round(rnorm(24),0))
report_manova(model=model_mixed)
report_manova(model=model_between)</pre>
```

report_normality_tests

Normality tests

Description

Shapiro-Wilk Anderson-Darling Cramer-von-Mises Shapiro-Francia Jarque-Bera Kolmogorov-Smirnov Lilliefors Pearson X2

Usage

```
report_normality_tests(df, file = NULL)
```

Arguments

df dataframe with continous or ordinal data

file output filename

Details

returns xlsx file

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Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
report_normality_tests(df=df)
report_normality_tests(df=vector,file="normality_tests")</pre>
```

report_oneway

One way

Description

One way

Usage

```
report_oneway(
   df,
   dv,
   iv,
   file = NULL,
   w = 10,
   h = 10,
   base_size = 10,
   note = "",
   title = "",
   type = "ci",
   plot_means = FALSE,
   plot_diagnostics = FALSE
)
```

```
df
                  dataframe
dν
                  index of continous variables
                  index of factors
iν
file
                  output filename
                  width of pdf file
                  height of pdf file
h
                  base font size
base_size
                  text for footnote
note
                  plot title
title
                  type of bar to display "se" "ci" "sd" ""
type
                  if TRUE it will output mean plots and descriptives for plots
plot_means
plot_diagnostics
                  if TRUE it will output ANOVA diagnostics plots
```

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Note

- (1) The Fisher procedure assumes heteroscedasticity
- (2) The Welch procedure does not assume heteroscedasticity
- (3) The Kruskal Wallis procedure does not assume normality but it is not an alternative for violations of heteroscedasticity
- (4) Posthoc Tuckey: not good for unequal sample sizes or heteroscedasticity
- (5) Posthoc Games Howell: good for unequal sample sizes and heteroscedasticity

Examples

report_pdf

Report pdf

Description

Report pdf

Usage

```
report_pdf(
    ...,
    plotlist = NULL,
    file = NULL,
    title = NULL,
    w = 10,
    h = 10,
    print_plot = TRUE
)
```

```
plotlist list of plot objects

file output filename

title output filename

w width of pdf file

h height of pdf file

print_plot if TRUE it prints plot on graphics device
```

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Examples

```
p1<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet,group=Chick))+</pre>
           geom_line()+
           ggtitle("Growth curve for individual chicks")+
           theme bw()
p2<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet))+</pre>
           geom_point(alpha=.3)+
           geom_smooth(alpha=.2,size=1,method="loess",formula="y~x")+
           ggtitle("Fitted growth curve per diet")+theme_bw()
cars_plot_multiplot<-plot_multiplot(plotlist=plot_histogram(mtcars[,1:4]),cols=2)</pre>
cars_plot_base<-plot_normality_diagnostics(mtcars)</pre>
report_pdf(p1,p2,print_plot=TRUE)
report_pdf(p1,p2,file="report",print_plot=FALSE)
report_pdf(plotlist=cars_plot_multiplot,print_plot=TRUE)
report_pdf(plotlist=cars_plot_multiplot,file="report",print_plot=FALSE)
report_pdf(plotlist=cars_plot_base,print_plot=TRUE)
report_pdf(plotlist=cars_plot_base,file="report",print_plot=FALSE)
```

report_regression

Regression

Description

Regression

Usage

```
report_regression(
  model,
  base_size = 10,
  title = "",
  file = NULL,
  w = 10,
  h = 10,
  plot_diagnostics = TRUE
)
```

```
model object ml
base_size base font size
title plot title
file output filename
w width of pdf file. Relevant only when file string is not empty
h height of pdf file. Relevant only when file string is not empty
plot_diagnostics
    if TRUE it will output linear model diagnostics plots
```

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Note

(1) Problematic values for standardized residuals > +-1.96

Standardized residuals are residuals divided by an estimated standard deviation and they can be interpreted as z scores in that:

95.00 99.00 99.99 (2) Studentized residuals indicate the the ability of the model to predict that case. They follow a t distribution

- (3) DFFits indicate the difference between the adjusted predicted value and the original predicted value. Adjusted predicted value for a case refers to the predicted value of that case, when that case is excluded from model fit.
- (4) Cook's distance indicates leverage. Problematic values for cook's distance > 1 Cook and Weisberg (1982).
- (5) Hat values indicate leverage. Problematic values for Hat values 2 or 3 times the average (k+1/n) The average leverage value is defined as (k+1)/n, k=number of predictors, n=number of participants. Leverage values lie between 0 (no influence) and 1 (complete influence over prediction)

Hoaglin and Welsch (1978) recommends investigating cases with values greater than twice the average (2(k+1)/n)

Stevens (2002) recommends investigating cases with values greater than three times the average (3(k+1)/n)

T-tests test the hypothesis that b's are different from 0

Multiple R^2: Variance Explained

Adjusted R^2: Indicates how much variance in Y would be accounted for if the model is derived from the population from which the sample was taken. Idealy, $R^2 = Adjusted R^2$

F-Statistic tests the null hypothesis is that the overall model has no effect

Covariance ratios critical values CVR>1+[3(k+1)/n] CRV<1-[3(k+1)/n]. In general we should obtain small values or we may have to remove cases ASSUMPTIONS

- (1) variable types: All predictors must be quantitative or categorical (with two levels), and the outcome variable must be quantitative (interval data), continuous and unbounded (no constraints on the variability of the outcome) (2) Non-zero variance
- (3) No perfect multicollinearity
- (4) Predictors are uncorrelated with -external variables-
- (5) Homoscedasticity: At each level of the predictor variable(s), the variance of the residual terms should be constant. Residuals at each level of the predictor(s) should have similar variance (homoscedasticity)
- (6) Independent errors: For any two observations the residual terms should be uncorrelated (or independent) This eventuality is sometimes described as a lack of autocorrelation. This assumption can be tested with the Durbin-Watson test, which tests for serial correlations between errors. Specifically, it tests whether adjacent residuals are correlated The size of the Durbin-Watson statistic depends upon the number of predictors in the model and the number of observations As a very conservative rule of thumb, values less than 1 or greater than 3 are definitely cause for concern; however, values closer to 2 may still be problematic depending on your sample and model R also provides a p-value of the autocorrelation. Be very careful with the Durbin-Watson test, though, as it depends on the order of the data: if you reorder your data, you-Il get a different value
- (7) Normally distributed errors: It is assumed that the residuals in the model are random, normally distributed variables with a mean of 0
- (8) Independence: It is assumed that all of the values of the outcome variable are independent (in other words, each value of the outcome variable comes from a separate entity)
- (9) Linearity: The mean values of the outcome variable for each increment of the predictor(s) lie along a straight line

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Examples

```
form<-formula(mpg~qsec)
regressionmodel<-lm(form,data=mtcars)
multipleregressionmodel<-lm(mpg~qsec*hp*wt*drat,data=mtcars)
res<-report_regression(model=regressionmodel,plot_diagnostics=TRUE)
res<-report_regression(model=multipleregressionmodel)
res<-report_regression(model=regressionmodel,file="regression")
res<-report_regression(model=multipleregressionmodel,file="regression",plot_diagnostics=TRUE)</pre>
```

report_ttests

T test

Description

T test

Usage

```
report_ttests(df, dv, iv, file = NULL, ...)
```

dataframe

Arguments df

dv	index of continous variables
iv	index of factors
file	output filename
	Arguments passed on to stats::t.test
	x a (non-empty) numeric vector of data values.

```
report_ttests(df=mtcars,dv=2,iv=9:10)
report_ttests(df=mtcars,dv=2:3,iv=9)
report_ttests(df=mtcars,dv=2:3,iv=9:10,alternative="two.sided")
report_ttests(df=mtcars,dv=2:7,iv=9:10,alternative="less")
report_ttests(df=mtcars,dv=2:7,iv=9:10,alternative="greater")
report_ttests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE)
report_ttests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE,file="ttest")
```

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report_wtests

Wilcoxon test

Description

Wilcoxon test

Usage

```
report_wtests(df, dv, iv, file = NULL, ...)
```

Arguments

df dataframe

dv index of continous variables

iv index of factorsfile output filename

... Arguments passed on to stats::wilcox.test

x numeric vector of data values. Non-finite (e.g., infinite or missing) values will be omitted.

Examples

```
report_wtests(df=mtcars,dv=2,iv=9)
report_wtests(df=mtcars,dv=2,iv=9:10)
report_wtests(df=mtcars,dv=2:3,iv=9)
report_wtests(df=mtcars,dv=2:3,iv=9:10,alternative="two.sided")
report_wtests(df=mtcars,dv=2:7,iv=9:10,alternative="less")
report_wtests(df=mtcars,dv=2:7,iv=9:10,alternative="greater")
report_wtests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE)
report_wtests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE,file="wilcoxontest")
```

report_xgboost

Report for xgboost::xgb.train

Description

Report for xgboost::xgb.train

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Usage

```
report_xgboost(
  model,
  validation_data = NULL,
  label = NULL,
  file = "xgboost",
  w = 10,
  h = 10,
  base_size = 10,
  title = "",
  fast = FALSE
)
```

Arguments

model object from xgboost::xgb.train
validation_data
validation data
label outcome variable name
file output filename
w width of pdf file
h height of pdf file
base_size base font size
title plot title

fast if TRUE error values are not saved in output

```
infert_formula<-as.formula(factor(case)~age+parity+education+spontaneous+induced)
boston\_formula <-as.formula (c("medv^",paste(names(MASS::Boston)[1:13],collapse="+")))
train_test_classification<-workingfunctions::k_fold(df=infert,model_formula=infert_formula)</pre>
train_test_regression<-workingfunctions::k_fold(df=MASS::Boston,model_formula=boston_formula)
xgb_classification<-xgboost::xgb.train(data=train_test_classification$xgb$f1$train,
                                   watchlist=train_test_classification$xgb$f1$watchlist,
                                        nthread=8,
                                        nround=20,
                                        objective="binary:logistic")
xgb_regression<-xgboost::xgb.train(data=train_test_regression$xgb$f1$train,</pre>
                                    watchlist=train_test_regression$xgb$f1$watchlist,
                                    eta=.3,
                                    nthread=8,
                                    nround=20)
report_xgboost(model=xgb_classification,
               validation_data=train_test_classification$f$test$f1,
               label=train_test_classification$outcome,
               file="Classification")
report_xgboost(model=xgb_regression,
```

response_frequency

```
validation_data=train_test_regression$f$test$f1,
label=train_test_regression$outcome,
file="Regression")
```

response_dimension

index parameter and items relative to their dimensions

Description

index parameter and items relative to their dimensions

Usage

```
response_dimension(response, dimensions, items)
```

Arguments

response vector one to number of items

dimensions number of dimensions items item comparisons

Examples

```
response_dimension(c(1:18),3,c(1,2))
response_dimension(c(1:18),3,c(1,3))
response_dimension(c(1:18),3,c(2,3))
```

response_frequency

Response frequencies

Description

returns count proportion percent

Usage

```
response_frequency(
  df,
  max = 10,
  uniqueitems = NULL,
  type = "percent",
  file = NULL
)
```

Arguments

df dataframe
max maximum score
uniqueitems number of unique items
type "frequency" "proportion" "percent" "all"
file output filename

Details

returns dataframe

Examples

result_confusion_performance

Plot performance of confusion matrix for different cut off points

Description

Plot performance of confusion matrix for different cut off points

Usage

```
result_confusion_performance(
  observed,
  predicted,
  step = 0.1,
  base_size = 10,
  title = ""
)
```

Arguments

observed vector of observed outcomes

predicted vector of predicted outcome probability

step stepping for tested cut values

base_size base font size title plot title

112 round_dataframe

Examples

round_dataframe

Round dataframe

Description

It only processes numeric values in a dataframe

Usage

```
round_dataframe(df, digits = 0, type = "round")
```

Arguments

df dataframe

digits decimal points to return. It works only with "round" type

type "round" "ceiling" "floor" "tenth"

```
round_dataframe(df=change_data_type(df=mtcars,type="factor"),digits=0)
round_dataframe(df=change_data_type(df=mtcars,type="character"),digits=0)
round_dataframe(df=mtcars,digits=0)
round_dataframe(df=mtcars,digits=0,type="ceiling")
round_dataframe(df=mtcars,digits=0,type="floor")
round_dataframe(df=mtcars*100,digits=2,type="tenth")
```

shrout 113

shrout	Shrout reliability
--------	--------------------

Description

Shrout reliability

Usage

```
shrout(sperson, spersonitem, stime, spersontime, serror, m, k)
```

Arguments

sperson variance component of participant
spersonitem variance component of participant by item interaction
stime variance component of time
spersontime variance component of participant by time interaction
serror variance component of error
m item reports

Examples

k

```
simulate_cfa_fit Simulate CFA from coefficients
```

k time points

Description

Simulates cfa from coefficients Simulates cfa from correlations of obeserved data Returns fit indices for predefined set of sample sizes

Usage

```
simulate_cfa_fit(
  model_sim = NULL,
  model = NULL,
  df = NULL,
  minnobs = 50,
  maxnobs = 1000,
  stepping = 10,
  file = NULL,
  w = 10,
  h = 10
)
```

Arguments

lavaan model spesification with defined coefficients model_sim lavaan model spesification with free coefficients model df dataframe minnobs start sample size end sample size maxnobs stepping stepping file output filename width of pdf file h height of pdf file

Examples

```
simulate_correlation_from_sample
```

Generate a dataframe that produces the same correlation matrix as the input dataframe

Description

Generate a dataframe that produces the same correlation matrix as the input dataframe

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Usage

```
simulate_correlation_from_sample(cordata, nrows = 10)
```

Arguments

cordata dataframe

nrows number of rows to generate

Examples

```
correlation_matrix<-generate_correlation_matrix()
stats::cor(correlation_matrix)
simulate_correlation_from_sample(correlation_matrix,nrows=1000)
stats::cor(simulate_correlation_from_sample(correlation_matrix,nrows=1000))</pre>
```

split_str

Split string to dataframe

Description

Split string to dataframe

Usage

```
split_str(vector, split = "/", include_original = FALSE)
```

Arguments

vector String

split Separation character

include_original

if TRUE it will return the input on a separate collumn

stat_word_char

a n 1		str	45
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Split string in dataframe

Description

Split string in dataframe

Usage

```
split_str_df(df, split = "/", type = "row", index, ...)
```

Arguments

df	dataframe
split	Separation character
type	"row" "collumn" if "row" it will split the string of row names and it will display it on seperate collumns if "collumn" it will split the string of a spesified collumn and it will display it on separate collumns
index	Numeric index of collumn to split. This is only relevant if type="collumn"
	arguments passed to split_str

Examples

stat_word_char

Text similarity measures

Description

Text similarity measures

Usage

```
stat_word_char(text)
```

string_aes 117

Arguments

text character vector

Examples

```
text<-"There are many variations of passages of Lorem Ipsum available,
but the majority have suffered alteration in some form, by injected humour,
or randomised words which don't look even slightly believable."
stat_word_char(text)
```

string_aes

Adjust string aesthetics

Description

Treats spesific characters such as ".", as separating characters and separates strings with space. Trims leading and trailing spaces and capitalizes the first letter of the string and lowers the rest.

Usage

Arguments

vector Vector

characterlist List the list of characters to treat as separating characters

proper Logical TRUE capitalizes the first letter in sentense format

```
vector<-c("TES.T","TES<p>T","TES&nbspT")
string_aes(vector=vector)
string_aes(vector=vector,proper=FALSE)
string_aes(vector=vector,proper=TRUE)
```

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 sub_str

Return n characters from left or right

Description

Return n characters from left or right

Usage

```
sub_str(x, n = 2, type)
```

Arguments

x Character

n Number of characters to return

type "right" "left"

Examples

```
sub_str("12345",n=2,type="right")
sub_str("12345",n=2,type="left")
```

swap

Reverse a numeric vector

Description

Reverse a numeric vector

Usage

```
swap(vector)
```

Arguments

vector

numeric

```
swap(c(1:10,1,2,3))
```

symmetric_matrix 119

symmetric_matrix

Symmetric Matrix

Description

Symmetric Matrix

Usage

```
symmetric_matrix(matrix, duplicate = "lower", diagonal = NULL)
```

Arguments

matrix matrix

duplicate "upper" duplicates upper triangle "lower" duplicates lower triangle

diagonal values

Examples

```
m_lower<-matrix_triangle(matrix(1:9,nrow=3,ncol=3),type="lower",diagonal=NA)
m_upper<-matrix_triangle(matrix(11:19,nrow=3,ncol=3),type="upper",diagonal=NA)
symmetric_matrix(matrix=m_lower,duplicate="lower",diagonal=NA)
symmetric_matrix(matrix=m_upper,duplicate="upper",diagonal=NA)</pre>
```

tag_pos

Part of speech tagging

Description

Part of speech tagging

Usage

```
tag_pos(text)
```

Arguments

text character vector

120 text_similarity

Examples

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
tag_pos(text)
```

text_similarity

Text similarity measures

Description

Text similarity measures

Usage

```
text_similarity(text1, text2)
```

Arguments

text1 character vector text2 character vector

```
text1<-"word_one word_two word_three"</pre>
text2<-"word_three word_four word_six"</pre>
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)</pre>
text<-unlist(strsplit(text,split=" "))</pre>
text1<-unlist(strsplit(text1,split=" "))</pre>
text2<-unlist(strsplit(text2,split=" "))</pre>
text3<-unlist(strsplit(text3,split=" "))</pre>
text4<-unlist(strsplit(text4,split=" "))</pre>
text5<-unlist(strsplit(text5,split=" "))</pre>
text_similarity(text1, text1)
text_similarity(text1,text2)
text_similarity(text1,text3)
text_similarity(text1, text4)
```

trim_df

trim_df

Trim whitespace in dataframe

Description

Trim whitespace in dataframe

Usage

```
trim_df(df)
```

Arguments

df

dataframe

Examples

ts_smoothing

Smoothing

Description

smoothing for timeseries. uses base plot

Usage

```
ts_smoothing(
   df,
   start = 0.01,
   stop = 2,
   step = 0.001,
   title = "",
   type = "kernel"
)
```

122 wrapper

Arguments

```
df ts object
start start value
stop stop value
step step
title plot title
type "default" "kernel" "lowess" "friedman" "splines" "polynomial" "linear"
```

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)
par(mfrow=c(2,2))
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="default")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="polynomial")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="linear")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="kernel")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="lowess")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="friedman")
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="splines")</pre>
```

wrapper

Wrap string

Description

Wrap string

Usage

```
wrapper(x, ...)
```

Arguments

```
x title
```

... arguments passed to strwrap

```
wrapper(rep("sting",50),30)
```

write_txt 123

write_txt

Log console in file

Description

Logs console in file and then displays log in console

Usage

```
write_txt(input, file = NULL)
```

Arguments

input Script to log in log file file Filename of log

```
write_txt(mtcars)
write_txt(mtcars,file="mtcars")
```

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