

Package ‘workingfunctions’

December 9, 2024

Title Statistical reporting and visualization for common methods

Version 0.1

Description Statistical reporting and visualization for common methodology used in psychology. Functions to minimize code and automate procedures using common R packages.

Depends R (>= 3.5), ggplot2

Imports MASS, future, future.apply, gtable, gtools, irr, openxlsx, pROC, plyr, psych, reshape2, stringr, xgboost, DescTools, Rmisc, ggfortify, ggrepel, car, dplyr, lmerTest, sjstats, doSNOW, foreach, ggpubr, gridExtra, scales, pwr, mirt, nlme, MplusAutomation, NLP, openNLP, tm, spelling, catR, lavaan, ggExtra, QuantPsyc, semPlot, R.utils, purrr, emmeans, ez

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Encoding UTF-8

LazyData true

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RoxygenNote 7.3.1

Contents

alpha_diagnostics	5
call_to_string	6
cdf	6
cfa_icc_index	7
change_data_type	7
clear_stopwords	8
clear_text	9
comparison_combinations	9
compute_ability	10
compute_adjustment	11
compute_aggregate	11
compute_aov_es	12
compute_confidence_inteval	13

compute_crosstable	13
compute_descriptives	14
compute_dissatenuation	14
compute_dummy_comparisons	15
compute_frequencies	15
compute_icc_thurstonian	16
compute_info_1pl	17
compute_info_2pl	17
compute_info_3pl	18
compute_kruskal_wallis_test	19
compute_kurtosis	19
compute_map	20
compute_moving_average	21
compute_one_way_test	21
compute_posthoc	22
compute_power_r	22
compute_power_r_matrix	23
compute_residual_stats	24
compute_scores	24
compute_scores_parallel	25
compute_se_theta	26
compute_skewness	26
compute_standard	27
compute_standard_error	28
compute_unidimensional_ability	28
compute_unidimensional_theta	29
compute_y_logistic	30
confusion	31
confusion_matrix_percent	31
convert_excel_unix_timestamp	32
c_bind	33
data_frame_index	33
decompose_datetime	34
deg2rad	35
detach_package	35
display_upper_lower_triangle	36
dotnames	36
drop_levels	37
dummy_arrange	37
duplicate_y_axis	38
environment_options	39
excel_confusion_matrix	39
excel_critical_value	40
excel_generic_format	41
excel_matrix	42
extract_components	44
flatten_list	44
generate_comparisons_matrix	45

generate_correlation_matrix	45
generate_data	46
generate_factor	47
generate_matrix_A	47
generate_matrix_lambda_hat	48
generate_missing	48
generate_multiple_responce_vector	49
generate_string	49
generate_unique_comparisons_index	50
getfwp	50
get_coding	51
get_mplus_thu_3t	51
get_questionnaire_reference	52
get_reversals	52
hinvert_title_grob	53
icc_cfa	53
increase_index	54
install_all_packages	54
install_load	55
json2dataframe	55
key_to_cfa_model	56
k_fold	56
k_sample	57
matrix_triangle	57
mean_sd_alpha	58
mgsb	59
min_max_index	59
model_loadings	60
off_diagonal_index	61
outlier_summary	61
output_compare_model_logistic	62
output_separator	62
padNA	63
plot_acf	64
plot_boxplot	64
plot_cfa	65
plot_confusion	66
plot_corrplot	66
plot_crosstable	67
plot_histogram	67
plot_icc_thurstonian	68
plot_interaction	69
plot_irt_onefactor	70
plot_loadings	71
plot_logistic_model	72
plot_mosaic	73
plot_mtmm	73
plot_multiplot	74

plot_normality_diagnostics	75
plot_oneway	76
plot_oneway_diagnostics	77
plot_outlier	78
plot_qq	78
plot_response_frequencies	79
plot_roc	80
plot_scatterplot	80
plot_scree	82
plot_separability	82
plot_ts	83
proper	84
proportion_accurate	84
questions_by_keys	85
questions_dimensions_dataframe	85
rad2deg	86
rank3_to_triplets	87
rank_df_to_binary	87
rank_to_binary	88
raw_alpha	89
rbind_all	89
recode_dataframe	90
recode_scale_dummy	90
remove_nc	91
remove_outliers	92
remove_user_packages	93
replace_na_with_previous	93
report_alpha	94
report_cfa	95
report_choric_serial	95
report_correlation	96
report_dataframe	97
report_efa	98
report_factorial_anova	99
report_hlr	101
report_irt	102
report_lda	103
report_logistic	104
report_manova	106
report_normality_tests	107
report_oneway	107
report_pdf	109
report_regression	110
report_ttests	111
report_wtests	112
report_xgboost	113
response_dimension	114
response_frequency	115

result_confusion_performance	116
round_dataframe	117
shrout	117
simulate_cfa_fit	118
simulate_correlation_from_sample	119
split_str	119
split_str_df	120
string_aes	121
sub_str	121
swap	122
symmetric_matrix	122
tag_pos	123
text_similarity	123
trim_df	124
ts_smoothing	125
wrapper	126
write_txt	126
Index	127

alpha_diagnostics	<i>Item total correlation and r drop</i>
-------------------	--

Description

Item total correlation and r drop

Usage

alpha_diagnostics(df)

Arguments

df dataframe with one dimension

Examples

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
psych::alpha(df)
alpha_diagnostics(df=df)
```

<code>call_to_string</code>	<i>Model call to string</i>
-----------------------------	-----------------------------

Description

Takes a call object and convert it to string

Usage

```
call_to_string(model)
```

Arguments

model Model object

Examples

```
df<-generate_correlation_matrix()
model<-lm(df$X1~df$X2)
call_to_string(model)
```

<code>cdf</code>	<i>Check dataframe</i>
------------------	------------------------

Description

dataframe summary

Usage

```
cdf(
  df,
  name_length = (getOption("width")/3),
  digits = 2,
  nuniqes = 0,
  parralel = FALSE,
  file = NULL
)
```

Arguments

df dataframe
name_length number of characters to be displayed for names
digits number of rounding digits
nuniqes number of unique items to display
parralel if TRUE it will run using multiple cores
file output filename

Examples

```

cdf(df=mtcars,parralel=TRUE)
cdf(df=change_data_type(mtcars,"factor"),nuniques=3)
cdf(df=data.frame(t(mtcars)),file="mtcars",nuniques=10)
cdf(df=mtcars)
cdf(df=generate_missing(mtcars))
cdf(df=infert,nuniques=10)
cdf(df=infert)
df<-data.frame(infert,
               date=seq(as.Date("2010-1-1"),
                        as.Date("2020-1-1"),
                        length.out=nrow(infert)))
cdf(df=df)

```

cfa_icc_index	<i>index of items to convert from lavaan to thurstonian order for analysis</i>
---------------	--

Description

index of items to convert from lavaan to thurstonian order for analysis

Usage

```
cfa_icc_index(nitems, nfactors = 3)
```

Arguments

nitems	number of items in the questionnaire
nfactors	number of factors

Examples

```
cfa_icc_index(nitems=18,nfactors=3)
```

change_data_type	<i>dataframe data type transformations</i>
------------------	--

Description

dataframe data type transformations

Usage

```
change_data_type(df, type)
```

Arguments

df	dataframe
type	"character" "numeric" "factor" "factor_character" "character_factor" For "factor_character" if factors are found, are converted to characters For "character_factor" if characters are found, are converted to factors

Examples

```
cdf(df=change_data_type(df=mtcars,"character"))
cdf(df=change_data_type(df=mtcars,"numeric"))
cdf(df=change_data_type(df=mtcars,"factor"))
df<-change_data_type(df=mtcars,"factor")
cdf(df=change_data_type(df=df,"factor_character"))
```

clear_stopwords	<i>Remove stopwods</i>
-----------------	------------------------

Description

Remove stopwods

Usage

```
clear_stopwords(
  text,
  stopwords = unique(c(tm::stopwords("SMART"), corpus::stopwords_en))
)
```

Arguments

text	character vector
stopwords	character words to remove

Examples

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
clear_stopwords(text)
```

clear_text

Clear text

Description

Clear text

Usage

```
clear_text(text)
```

Arguments

text character vector

Examples

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
clear_text(text)
```

comparison_combinations

Produce combinations for comparisons from dataframe names

Description

Produce combinations for comparisons from dataframe names

Usage

```
comparison_combinations(df, all_orders = TRUE)
```

Arguments

df dataframe

all_orders if TRUE the order of combination is considered i.e. the combination X1 X2 also appears as X2 X1 if FALSE it is assumed that X1 X2 and X2 X1 are the same and only one of them appears

Examples

```
comparison_combinations(generate_correlation_matrix(n=10)[,1:4])
```

compute_ability

Compute subject ability for thurstonian models

Description

Computes person ability for binary thurstonian coded items for a single dimension

Usage

```
compute_ability(
  response,
  eta,
  gamma,
  lambda,
  psi,
  plot = FALSE,
  map = compute_map(eta = eta, mean = 0, sd = 1)
)
```

Arguments

response	item responses
eta	eta or ability
gamma	gamma or threshold
lambda	lambda or loading
psi	psi or error
plot	if TRUE plots icc curves using the plot_icc_thurstonian function
map	vector from compute_map

Examples

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
eta<-seq(-6,6,by=0.1)
response1<-c(0,0,0,0,0,0,0,0,0)
response2<-c(1,1,1,1,1,1,1,1,1)
response3<-c(1,0,1,0,1,0,1,0,1)
response4<-c(0,1,0,1,0,1,0,1,0)
map<-compute_map(eta=eta,mean=0,sd=1)
compute_ability(response1,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

```
compute_ability(response2,eta,gamma,lambda,psi,map=map,plot=FALSE)
compute_ability(response3,eta,gamma,lambda,psi,map=map,plot=FALSE)
compute_ability(response4,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

compute_adjustment	<i>Compute adjustments</i>
--------------------	----------------------------

Description

Compute adjustments

Usage

```
compute_adjustment(a, ntests)
```

Arguments

a	alpha criterion
ntests	number of tests

Examples

```
compute_adjustment(0.05,100)
```

compute_aggregate	<i>Descriptive statistics</i>
-------------------	-------------------------------

Description

uses plyr

Usage

```
compute_aggregate(df, iv, file = NULL)
```

Arguments

df	dataframe
iv	index of independent variables
file	output filename

Details

returns xlsx

Examples

```
compute_aggregate(df=mtcars, iv=9)
compute_aggregate(df=mtcars, iv=9:10)
compute_aggregate(df=mtcars, iv=9:11)
compute_aggregate(df=mtcars, iv=9:11, file="descriptives")
```

compute_aov_es

Compute eta and omega

Description

Computes omega using aov object. Based on <http://stats.stackexchange.com/a/126520>

Usage

```
compute_aov_es(model, ss = "I")
```

Arguments

model	object aov
ss	Character type of sums of squares "I" "II" "III"

Examples

```
form<-formula(uptake~Treatment)
one_way_between<-aov(form,C02)
factorial_between<-aov(uptake~Treatment*Type,C02)
compute_aov_es(model=one_way_between,ss="I")
sjstats::anova_stats(one_way_between,digits=10)
compute_aov_es(model=one_way_between,ss="II")
sjstats::anova_stats(one_way_between,digits=10)
compute_aov_es(model=one_way_between,ss="III")
sjstats::anova_stats(one_way_between,digits=10)
compute_aov_es(model=factorial_between,ss="I")
sjstats::anova_stats(factorial_between,digits=10)
compute_aov_es(model=factorial_between,ss="II")
sjstats::anova_stats(factorial_between,digits=10)
compute_aov_es(model=factorial_between,ss="III")
sjstats::anova_stats(car::Anova(factorial_between,Type=3),digits=10)
```

compute_confidence_interval	
	<i>Compute confidence interval</i>

Description

Compute confidence interval

Usage

```
compute_confidence_interval(vector)
```

Arguments

vector	vector
--------	--------

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_confidence_interval(vector)
```

compute_crosstable	<i>Compute crosstables</i>
--------------------	----------------------------

Description

Compute crosstables

Usage

```
compute_crosstable(df, factor_index = NULL, combinations = NULL)
```

Arguments

df	dataframe
factor_index	index of factors
combinations	index of comparisons

Examples

```
combinations<-data.frame(index1=c("vs","am","gear"),index2=c("cyl","cyl","cyl"))
compute_crosstable(df=mtcars,combinations=combinations)
combinations<-data.frame(index1=c("vs","am"),index2=c("cyl","cyl"))
compute_crosstable(df=mtcars,combinations=combinations)
compute_crosstable(df=mtcars,factor_index=8:10)
```

`compute_descriptives` *Descriptive statistics*

Description

uses psych

Usage

```
compute_descriptives(df, dv, iv = NULL, file = NULL)
```

Arguments

<code>df</code>	dataframe
<code>dv</code>	index of dependent variables
<code>iv</code>	index of independent variables
<code>file</code>	output filename

Details

returns xlsx

Examples

```
compute_descriptives(df=mtcars,dv=1:5)
compute_descriptives(df=mtcars,dv=1:2,iv=9:10)
compute_descriptives(df=mtcars,dv=1:2,file="descriptives_no_factor")
compute_descriptives(df=mtcars,dv=1:2,iv=9:10,file="descriptives_factor")
```

`compute_dissatenuation`
 Compute dissatenuation

Description

Compute dissatenuation

Usage

```
compute_dissatenuation(variable1, error1, variable2, error2)
```

Arguments

<code>variable1</code>	vector
<code>error1</code>	vector error measurement for variable1
<code>variable2</code>	vector
<code>error2</code>	vector error measurement for variable2

Examples

```
set.seed(1)
compute_dissatenuation(rnorm(10),rnorm(10),rnorm(10),rnorm(10))
```

```
compute_dummy_comparisons
```

Compute number of dummy comparisons

Description

Compute number of dummy comparisons

Usage

```
compute_dummy_comparisons(items)
```

Arguments

items	number of items per block
-------	---------------------------

Examples

```
compute_dummy_comparisons(1)
compute_dummy_comparisons(2)
compute_dummy_comparisons(3)
compute_dummy_comparisons(4)
compute_dummy_comparisons(5)
compute_dummy_comparisons(6)
```

```
compute_frequencies
```

Frequencies by levels

Description

returns frequency proportion percent

Usage

```
compute_frequencies(df, ordered = TRUE, file = NULL)
```

Arguments

df	dataframe
ordered	if TRUE it will output frequencies in descending order
file	output filename

Details

returns xlsx

Examples

```
compute_frequencies(df=generate_missing(generate_factor(nrows=10,ncols=10),missing=5))
compute_frequencies(df=generate_factor())
compute_frequencies(df=generate_factor(),file="descriptives")
```

compute_icc_thurstonian

Compute item characteristic curves for thurstonian models

Description

Computes icc curves for binary thurstonian coded items for a single dimension

Usage

```
compute_icc_thurstonian(eta, gamma, lambda, psi, plot = FALSE)
```

Arguments

eta	eta or ability
gamma	gamma or threshold
lambda	lambda or loading
psi	psi or error
plot	if TRUE plots icc curves using the plot_icc_thurstonian function

Examples

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
eta<-seq(-6,6,by=0.01)
compute_icc_thurstonian(eta=eta,gamma=gamma,lambda=lambda,psi=psi,plot=FALSE)
```

compute_info_1pl	<i>Compute item information for 1PL model</i>
------------------	---

Description

Compute item information for 1PL model

Usage

```
compute_info_1pl(b, theta)
```

Arguments

b	numeric difficulty parameter
theta	numeric theta

Examples

```
compute_info_1pl(b=1, theta=-3)
compute_info_1pl(b=1, theta=-2)
compute_info_1pl(b=1, theta=-1)
compute_info_1pl(b=1, theta=0)
compute_info_1pl(b=1, theta=1)
compute_info_1pl(b=1, theta=2)
compute_info_1pl(b=1, theta=3)
ti<-compute_info_1pl(b=1, theta=seq(-6,6,by=.01)) # test information
plot(ti, x=seq(-6,6,by=.01))
```

compute_info_2pl	<i>Compute item information for 2PL model</i>
------------------	---

Description

Compute item information for 2PL model

Usage

```
compute_info_2pl(a, b, theta)
```

Arguments

a	numeric discrimination parameter
b	numeric difficulty parameter
theta	numeric theta

Examples

```

compute_info_2pl(a=1.5,b=1,theta=-3)
compute_info_2pl(a=1.5,b=1,theta=-2)
compute_info_2pl(a=1.5,b=1,theta=-1)
compute_info_2pl(a=1.5,b=1,theta=0)
compute_info_2pl(a=1.5,b=1,theta=1)
compute_info_2pl(a=1.5,b=1,theta=2)
compute_info_2pl(a=1.5,b=1,theta=3)
ti<-compute_info_2pl(a=1,b=-2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))
ti<-compute_info_2pl(a=2,b=0,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))
ti<-compute_info_2pl(a=3,b=2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))

```

compute_info_3pl	<i>Compute item information for 3PL model</i>
------------------	---

Description

Compute item information for 3PL model

Usage

```
compute_info_3pl(a, b, g, theta)
```

Arguments

a	numeric discrimination parameter
b	numeric difficulty parameter
g	numeric guessing parameter
theta	numeric theta

Examples

```

compute_info_3pl(a=1.5,b=1,g=.2,theta=-3)
compute_info_3pl(a=1.5,b=1,g=.2,theta=-2)
compute_info_3pl(a=1.5,b=1,g=.2,theta=-1)
compute_info_3pl(a=1.5,b=1,g=.2,theta=0)
compute_info_3pl(a=1.5,b=1,g=.2,theta=1)
compute_info_3pl(a=1.5,b=1,g=.2,theta=2)
compute_info_3pl(a=1.5,b=1,g=.2,theta=3)
ti<-compute_info_3pl(a=1.5,b=1,g=.2,theta=seq(-6,6,by=.01)) # test information
plot(ti,x=seq(-6,6,by=.01))

```

```
compute_kruskal_wallis_test
```

Kruskal Wallis test

Description

Kruskal Wallis test

Usage

```
compute_kruskal_wallis_test(formula, df)
```

Arguments

formula	one way formula in form of y~x. It will ignore more complex formulas
df	dataframe

Note

eta squared ranges between 0 and 1
 epsilon squared ranges between 0 and 1
 eta squared multiplied by 100 indicates the percentage of variance in the dependent variable explained by the independent variable

Examples

```
form<-formula(qsec~cyl)
kruskal.test(formula=form,data=mtcars)
rcompanion::epsilonSquared(x=mtcars$qsec,g=mtcars$cyl,group="row",ci=TRUE,conf=0.95,
                           type="perc",R=1000,digits=3)
rstatix::kruskal_effsize(mtcars,form,ci=TRUE,conf.level=0.95,ci.type="perc",nboot=100)
compute_kruskal_wallis_test(formula=form,df=mtcars)
```

```
compute_kurtosis
```

Compute kurtosis

Description

Compute kurtosis

Usage

```
compute_kurtosis(vector)
```

Arguments

vector vector

Note

$b_2 = m_4 / s^4 - 3 = (g_2 + 3) (1 - 1/n)^2 - 3$. Used in MINITAB and BMDP.

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_kurtosis(vector)
e1071::kurtosis(vector)
```

compute_map

Simulate prior distribution

Description

Simulate prior distribution

Usage

```
compute_map(eta, mean = 0, sd = 1)
```

Arguments

eta vector

mean numeric

sd numeric

Examples

```
eta<-seq(-6,6,by=0.1)
compute_map(eta=eta,mean=0,sd=1)
```

`compute_moving_average`*Moving Average*

Description

compute moving average

Usage

```
compute_moving_average(df, w)
```

Arguments

df	dataframe
w	window

Examples

```
compute_moving_average(df=mtcars,w=5)
```

`compute_one_way_test` *one way test*

Description

one way test

Usage

```
compute_one_way_test(formula, df, var.equal = TRUE)
```

Arguments

formula	one way formula in form of y~x. It will ignore more complex formulas
df	dataframe
var.equal	if TRUE it assumes equal variances

Note

eta and omega for welch statistics are not adequately tested and they should not be consulted

Examples

```

form<-formula(qsec~cyl)
compute_one_way_test(formula=form,df=mtcars,var.equal=TRUE)
compute_one_way_test(formula=form,df=mtcars,var.equal=FALSE)
oneway.test(formula=form,data=mtcars,var.equal=TRUE)
oneway.test(formula=form,data=mtcars,var.equal=FALSE)
car::Anova(aov(form,data=mtcars),type=2)
model<-lm(form,data=mtcars)
lsr::etaSquared(aov(form,data=mtcars),type=3,anova=TRUE)
sjstats::anova_stats(model,digits=22)

```

compute_posthoc

Games Howell Tukey post hoc tests

Description

Based on http://www.psych.yorku.ca/cribbie/6130/games_howell.R

Usage

```
compute_posthoc(y, x, method = c("games-howell", "tukey"))
```

Arguments

y	Vector continous variable
x	Vector factor
method	Character takes two values "games-howell" or "tukey" or c("games-howell","tukey")

Examples

```
result<-compute_posthoc(mtcars[,6],mtcars[,10])
```

compute_power_r

Compute r power curve

Description

Compute r power curve

Usage

```

compute_power_r(
  n = 100,
  r = NULL,
  sig.level = 0.05,
  alternative = c("two.sided", "less", "greater"),
  title = "",
  base_size = 10
)

```

Arguments

n	number of observations
r	correlation coefficient
sig.level	alpha (type I error probability)
alternative	a character string specifying the alternative hypothesis, must be one of "two.sided" (default), "greater" or "less"
title	plot title
base_size	base font size

Examples

```
compute_power_r(n=100,r=.5,sig.level=.05,alternative=c("two.sided"))
```

```
compute_power_r_matrix
```

Compute correlation matrix

Description

Compute correlation matrix

Usage

```
compute_power_r_matrix(m, ...)
```

Arguments

m	correlation matrix
...	arguments passed to compute_power_r

Examples

```
compute_power_r_matrix(m=stats::cor(mtcars,use="pairwise.complete.obs"),n=100)
```

compute_residual_stats

Residuals for matrices

Description

Root Mean Squared Residual Number of absolute residuals > 0.05 Proportion of absolute residuals > 0.05. It can either accept a psych EFA model or it can compare two correlation or covariance matrices

Usage

```
compute_residual_stats(model, data = NULL)
```

Arguments

model	psych EFA model. It has to be a correlation or covariance matrix if data is not NULL
data	correlation or covariance matrix

Examples

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
compute_residual_stats(model)
```

compute_scores

Compute subject ability for thurstonian models

Description

Computes person ability for binary thurstonian coded items for a single dimension

Usage

```
compute_scores(mydata, ...)
```

Arguments

mydata	item responses
...	arguments passed to compute_ability

Examples

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
```

```
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
```

```
eta<-seq(-6,6,by=0.1)
```

```
map<-compute_map(eta=eta,mean=0,sd=1)
```

```
response_df<-data.frame(matrix(nrow=0,ncol=8))
```

```
response_df[1,]<-c(0,0,0,0,0,0,0,0)
```

```
response_df[2,]<-c(1,1,1,1,1,1,1,1)
```

```
response_df[3,]<-c(1,0,1,0,1,0,1,0)
```

```
response_df[4,]<-c(0,1,0,1,0,1,0,1)
```

```
compute_scores(response_df,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

```
compute_scores_parallel
```

Compute subject ability for thurstonian models

Description

Computes person ability for binary thurstonian coded items for a single dimension

Usage

```
compute_scores_parallel(mydata, ...)
```

Arguments

mydata	dataframe
...	arguments passed to compute_ability

Examples

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
```

```
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
```

```
eta<-seq(-6,6,by=0.1)
```

```
map<-compute_map(eta=eta,mean=0,sd=1)
```

```
response_df<-data.frame(matrix(nrow=0,ncol=8))
```

```
response_df[1,]<-c(0,0,0,0,0,0,0,0)
```

```
response_df[2,]<-c(1,1,1,1,1,1,1,1)
```

```
response_df[3,]<-c(1,0,1,0,1,0,1,0)
```

```
response_df[4,]<-c(0,1,0,1,0,1,0,1)
```

```
compute_scores(response_df,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

```
compute_scores_parallel(response_df,eta,gamma,lambda,psi,map=map,plot=FALSE)
```

compute_se_theta	<i>Compute the SE of theta</i>
------------------	--------------------------------

Description

Compute the SE of theta

Usage

```
compute_se_theta(info)
```

Arguments

info	numeric information
------	---------------------

Examples

```
compute_se_theta(1)
ti<-compute_info_2pl(a=10,b=0,theta=seq(-3,3,by=.01)) # test information
plot(compute_se_theta(ti),x=seq(-3,3,by=.01))
```

compute_skewness	<i>Compute skewness</i>
------------------	-------------------------

Description

Compute skewness

Usage

```
compute_skewness(vector)
```

Arguments

vector	vector
--------	--------

Note

$b_1 = m_3 / s^3 = g_1 ((n-1)/n)^{3/2}$. Used in MINITAB and BMDP.

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_skewness(vector)
e1071::skewness(vector)
```

compute_standard	<i>compute standard scores</i>
------------------	--------------------------------

Description

compute standard scores

Usage

compute_standard(vector, mean = 0, sd = 1, type = "z", input = "non_standard")

Arguments

vector	vector
mean	numeric applicable to "uz"
sd	numeric applicable to "uz"
type	"z" "uz" "sten" "t" "stanine" "center" "center_reversed" "percent" "scale_zero_one" "normal_density" "cumulative_density" "all"
input	"standard" "non_standard" standard inputs are z scores and non standard are raw scores

Examples

```
vector<-c(rnorm(10),NA,rnorm(10))
compute_standard(vector,type="z")
compute_standard(vector,mean=0,sd=1,type="uz")
compute_standard(vector,type="sten")
compute_standard(vector,type="t")
compute_standard(vector,type="stanine")
compute_standard(vector,type="center")
compute_standard(vector,type="center_reversed")
compute_standard(vector,type="percent")
compute_standard(vector,type="scale_zero_one")
ndf<-compute_standard(seq(-6,6,.01),mean=0,sd=1,type="normal_density")
plot(ndf)
cdf<-compute_standard(ndf,mean=0,sd=1,type="cumulative_density")
plot(cdf)
compute_standard(vector,type="all")
compute_standard(seq(-6,6,.1),type="all",input="standard")
```

`compute_standard_error`*Compute standard error*

Description

Compute standard error

Usage

```
compute_standard_error(vector)
```

Arguments

vector	vector
--------	--------

Examples

```
set.seed(1)
vector<-rnorm(1000)
compute_standard_error(vector)
```

`compute_unidimensional_ability`*Compute theta for unidimensional models*

Description

Compute theta for unidimensional models

Usage

```
compute_unidimensional_ability(
  a,
  b,
  g = NULL,
  d = 1.702,
  u,
  lim_theta = c(-6, 6)
)
```

Arguments

a	numeric vector discrimination parameters
b	numeric vector difficulty parameters
g	numeric vector guessing parameters
d	numeric scaling constant usually it is a value that approximating 1.749
u	numeric vector responses
lim_theta	vector minimum and maximum value of theta

Examples

```

a<-c(0.39,0.45,0.52,0.3,0.35,0.43,0.42,0.44,0.34,0.42)
b<-c(-1.96,-1.9,-1.38,-0.58,0.48,-0.81,-0.35,1.59,1.33,2.93)
u<-c(1,1,1,1,0,0,1,0,1,0)
# SHOULD RETURN 0.48402574251176
compute_unidimensional_ability(a=a,b=b,u=u,d=1.7,g=NULL)
a<-c(1.27,0.9,0.94,0.95,0.55,0.6,0.44,0.4)
b<-c(-0.54,0.18,0.21,1.26,1.73,-0.87,1.72,2.67)
u<-c(1,1,1,1,0,0,0,0)
# SHOULD RETURN 1.04621621510192
compute_unidimensional_ability(a=a,b=b,u=u,d=1.7,g=NULL)
a<-c(0.41,0.32,0.33,1.2,0.63,0.62,0.7,0.61,0.38,0.53,0.6,1.16)
b<-c(-1.4,-1.3,-1.17,0.2,0.71,0.86,-0.12,0.12,2.06,1.38,1.18,-0.33)
u<-c(1,0,1,1,0,0,0,1,1,0,1,0)
# SHOULD RETURN 0.0860506282671103
compute_unidimensional_ability(a=a,b=b,u=u,d=1.7,g=NULL)

```

compute_unidimensional_theta

Compute theta for unidimensional models

Description

Compute theta for unidimensional models

Usage

```
compute_unidimensional_theta(a, b = 0, g = 0, i = 1, d = 1.702, theta = 0)
```

Arguments

a	numeric discrimination parameter
b	numeric difficulty parameter
g	numeric guessing parameter
i	numeric inattentiveness parameter
d	numeric scaling constant usually a value 1.749 or 1.702
theta	numeric or vector theta

Note

when scaling constant=1 it has no effect in equation
 when innatentiveness=1 and guessing=0 function computes a 2PL score
 when innatentiveness=1 and guessing!=0 function computes a 3PL score
 when innatentiveness!=1 and guessing!=0 function computes a 4PL score

Examples

```
compute_unidimensional_theta(a=10,b=0)
x<-seq(-3,3,by=.01)
plot(compute_unidimensional_theta(a=5,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=5,b=-1,theta=x),x=x)
plot(compute_unidimensional_theta(a=5,b=1,theta=x),x=x)
plot(compute_unidimensional_theta(a=.1,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=1,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=.1,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=.5,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=1,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=.9,theta=x),x=x)
plot(compute_unidimensional_theta(a=10,b=0,g=0,i=.6,theta=x),x=x)
```

compute_y_logistic	<i>Compute y for logistic function</i>
--------------------	--

Description

This function requires x range to produce a vector with y values

Usage

```
compute_y_logistic(intercept, coefficient, x)
```

Arguments

intercept	Numeric
coefficient	Numeric
x	Numeric

Examples

```
x<--10:10
compute_y_logistic(0,1,x)
compute_y_logistic(0,1,1)
plot(x,compute_y_logistic(0,1,x),type="l");grid();abline(b=0,a=.5)
```

confusion	<i>Create a confusion matrix from observed and expected vectors</i>
-----------	---

Description

Create a confusion matrix from observed and expected vectors

Usage

```
confusion(observed, predicted)
```

Arguments

observed	vector of observed variables
predicted	vector of predicted variables

Examples

```
confusion(observed=c(1,2,3,4,5,10),predicted=c(1,2,3,4,5,11))  
confusion(observed=c(1,2,2,2,2),predicted=c(1,1,2,2,2))
```

confusion_matrix_percent	<i>Confusion matrix with row and column percent</i>
--------------------------	---

Description

Confusion matrix with row and column percent

Usage

```
confusion_matrix_percent(observed, predicted)
```

Arguments

observed	vector of observed variables
predicted	vector of predicted variables

Note

Total measures - Accuracy: $(TP+TN)/total$
 Total measures - Prevalence: $(TP+FN)/total$
 Total measures - Proportion Incorrectly Classified: $(FN+FP)/total$
 Horizontal measures - True Positive Rate - Sensitivity: $TP/(TP+FN)$
 Horizontal measures - True Negative Rate - Specificity: $TN/(FP+TN)$
 Horizontal measures - False Negative Rate - Miss Rate: $FN/(TP+FN)$
 Horizontal measures - False Positive Rate - Fall-out: $FP/(FP+TN)$
 Vertical measures - Positive Predictive value - Precision: $TP/(TP+FP)$
 Vertical measures - Negative Predictive value: $TN/(FN+TN)$
 Vertical measures - False Omission Rate: $FN/(FN+TN)$
 Vertical measures - False Discovery Rate: $FP/(TP+FP)$

Examples

```

observed<-factor(round(rnorm(10000,m=10,sd=1)))
predicted<-factor(round(rnorm(10000,m=10,sd=1)))
confusion_matrix_percent(observed,predicted)

```

```
convert_excel_unix_timestamp
```

Convert UNIX EXCEL timestamp

Description

Convert UNIX EXCEL timestamp

Usage

```
convert_excel_unix_timestamp(timestamp)
```

Arguments

timestamp unix or excel timestamp

Examples

```
convert_excel_unix_timestamp(1)
```

c_bind	<i>cbind dataframes with unequal lengths or row lengths</i>
--------	---

Description

cbind dataframes with unequal lengths or row lengths

Usage

```
c_bind(..., first = TRUE)
```

Arguments

...	dataframes or vectors to bind
first	Logical

Author(s)

Ananda Mahto

Examples

```
c_bind(rnorm(10), rnorm(11), rnorm(12), rnorm(13))
```

data_frame_index	<i>dataframe index</i>
------------------	------------------------

Description

dataframe index

Usage

```
data_frame_index(nrow, ncol)
```

Arguments

nrow	number of rows
ncol	number of columns

Examples

```
data_frame_index(5, 5)
```

decompose_datetime	<i>Decompose datetime objects to dataframe columns</i>
--------------------	--

Description

Decompose datetime objects to dataframe columns

Usage

```
decompose_datetime(
  x,
  format = "",
  origin = "1970-01-01",
  tz = "GMT",
  extended = FALSE,
  breaks = c(-1, 5, 13, 16, 20, 23),
  ...
)
```

Arguments

x	datetime object
format	date time format
origin	Starting date. The default is the unix time origin "1970-01-01"
tz	Timezone
extended	if TRUE it will display additional day time categories WEEKDAY MONTH JULIAN QUARTER DAY_PERIOD
breaks	Numeric vector Breaks define hour of day for classifying into "Night", "Morning", "Noon", "Afternoon", "Evening".
...	arguments passed to as.POSIXct This argument is used if extended=TRUE

Examples

```
timestamp1<-as.numeric(as.POSIXct(Sys.Date()))
timestamp2<-as.numeric(as.POSIXct(Sys.time()))
d1<-Sys.Date()
d2<-Sys.time()
decompose_datetime(x=d1)
decompose_datetime(x=d2)
decompose_datetime(x=d1,extended=TRUE)
decompose_datetime(x=d2,extended=TRUE)
decompose_datetime(x="01/15/1900",format="%m/%e/%Y")
decompose_datetime(x="01/15/1900",format="%m/%e/%Y",extended=TRUE)
decompose_datetime(x=as.Date(as.POSIXct(10000,origin="1970-01-01")))
decompose_datetime(x=as.Date(as.POSIXct(timestamp1,origin="1970-01-01"))),
```

```
format="%m/%e/%Y")
decompose_datetime(x=as.Date(as.POSIXct(timestamp2,origin="1970-01-01")),
format="%m/%e/%Y")
```

deg2rad	<i>Convert degrees to radians</i>
---------	-----------------------------------

Description

Convert degrees to radians

Usage

deg2rad(degrees)

Arguments

degrees degrees

Examples

deg2rad(180)

detach_package	<i>Unload library</i>
----------------	-----------------------

Description

Unload library

Usage

detach_package(package)

Arguments

package Package name

`display_upper_lower_triangle`

Return upper diagonal from one matrix and lower diagonal from another matrix

Description

Return upper diagonal from one matrix and lower diagonal from another matrix

Usage

```
display_upper_lower_triangle(m_upper, m_lower, diagonal = NA)
```

Arguments

<code>m_upper</code>	matrix
<code>m_lower</code>	matrix
<code>diagonal</code>	if "upper" it returns upper diagonal if "lower" it returns lower diagonal if NA returns NA in diagonal otherwise it returns any value specified

Examples

```
m1<-matrix(1:9,nrow=3,ncol=3)
m2<-matrix(11:19,nrow=3,ncol=3)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal="upper")
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal="lower")
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=NA)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=1)
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=c("X1","X2","X3"))
display_upper_lower_triangle(m_upper=m1,m_lower=m2,diagonal=c(1,2,3))
display_upper_lower_triangle(m_upper=m1,m2)
```

`dotnames`

Get the names of objects in the arguments

Description

Get the names of objects in the arguments

Usage

```
dotnames(...)
```

Arguments

<code>...</code>	objects
------------------	---------

Author(s)

Ananda Mahto

drop_levels	<i>Drops unused factor levels</i>
-------------	-----------------------------------

Description

Drops unused factor levels

Usage

```
drop_levels(df, factor_index = NULL, minimum_frequency = 5)
```

Arguments

df	dataframe
factor_index	numeric index of factors. If NULL the function uses is.factor() to discriminate factors
minimum_frequency	the minimum frequency each factor will have, levels with frequency bellow or equal to the defined frequency will be renamed "Other"

Examples

```
factor1<-factor(c(rep("A",10),rep("B",10)),levels=c("A","B","C","D"))
factor2<-factor(c(rep("A",10),rep("B",10)),levels=c("A","B","C","D"))
numeric1<-c(1:20)
df<-data.frame(numeric1,factor1,factor2)
df$factor1
drop_levels(df=df,minimum_frequency=9)
drop_levels(df=df,minimum_frequency=10)
```

dummy_arrange	<i>Takes a vector with multiple responses and dummy arranges it in a dataframe</i>
---------------	--

Description

Takes a vector with multiple responses and dummy arranges it in a dataframe

Usage

```
dummy_arrange(vector)
```

Arguments

vector Vector

Examples

```
vector1<-gsub(" ", "",
              generate_multiple_responce_vector(responces=c("Agree", "Hi", "All"),
              responded=1:3, length=10), fixed=TRUE)
vector2<-gsub(" ", "",
              generate_multiple_responce_vector(responces=1:4, responded=1:4, length=10),
              fixed=TRUE)
vector3<-sample(1:4, 10, replace=TRUE)
vector4<-sample(LETTERS[1:3], 10, replace=TRUE)
dummy_arrange(vector1)
dummy_arrange(vector2)
dummy_arrange(vector3)
dummy_arrange(vector4)
```

duplicate_y_axis	<i>Duplicate y axis</i>
------------------	-------------------------

Description

Duplicate y axis

Usage

```
duplicate_y_axis(p1, p2)
```

Arguments

p1 Plot 1
p2 Plot 2

Examples

```
p1<-ggplot(ChickWeight, aes(x=Time, y=weight, colour=Diet, group=Chick))+
  geom_line()+
  ggtitle("Growth curve for individual chicks")
duplicate_y_axis(p1, p1)
```

environment_options	<i>Load environment options</i>
---------------------	---------------------------------

Description

Load environment options

Usage

```
environment_options()
```

Examples

```
environment_options()
```

excel_confusion_matrix	
------------------------	--

Write matrix or dataframe to excel sheet

Description

Usefull for correlation matrices since it uses conditional formatting for matrices

Usage

```
excel_confusion_matrix(
  df,
  workbook,
  title = "Rows: Expected Collumns: Observed"
)
```

Arguments

df	dataframe or matrix
workbook	workbook
title	comment

Examples

```
filename<-"excel_confusion_matrix.xlsx"
if (file.exists(filename)) file.remove(filename)
observed<-factor(round(rnorm(10000,m=10,sd=1)))
predicted<-factor(round(rnorm(10000,m=10,sd=1)))
confusion(observed,predicted)
cm<-confusion_matrix_percent(observed,predicted)
wb<-openxlsx::createWorkbook()
excel_confusion_matrix(cm,wb)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

excel_critical_value *Write matrix or dataframe to excel sheet*

Description

Usefull for generic data where conditional formating of a spesific collumn is required

Usage

```
excel_critical_value(
  df,
  workbook,
  sheet = "output",
  title = NULL,
  comment = NULL,
  numFmt = "#0.00",
  critical = NULL
)
```

Arguments

df	dataframe or matrix
workbook	workbook
sheet	sheet
title	title
comment	comment
numFmt	number formatting
critical	list in the form of (collumn1=critical_value1,collumn2=critical_value2...)

Examples

```
comment<-list(mpg="Miles/(US) gallon",
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio",
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1=manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")
filename<-"excel_critical_value.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
```



```

df<-generate_missing(generate_correlation_matrix())
critical<-list(X1="<0.05",X5="<0")
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=list(X1="test"),
                    numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
filename<-"excel_critical_value_comment.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
df<-generate_missing(mtcars)
critical<-list(mpg=">20",am="=0")
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=comment,
                    numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
filename<-"excel_critical_value_comment_min_max.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
df<-generate_missing(mtcars)
critical<-list(mpg=c(">20","<11"),am="=0")
excel_critical_value(df=df,workbook=wb,sheet="critical",comment=comment,
                    numFmt="#0.00",critical=critical)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)

```

excel_generic_format *Generic function for creating workbooks and worksheets*

Description

This function is used by excel_matrix and excel_critical_value functions

Usage

```

excel_generic_format(
  df,
  workbook,
  sheet = "output",
  title = NULL,
  comment = NULL,
  numFmt = "#0.00"
)

```

Arguments

df	dataframe or matrix
workbook	workbook
sheet	sheet
title	title
comment	comment
numFmt	number formatting

Examples

```
comment<-list(mpg="Miles/(US) gallon",
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio",
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1=manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")
mtcor<-data.frame(cor(mtcars))
filename<-"excel_generic.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
openxlsx::addWorksheet(wb,"sheet")
openxlsx::addWorksheet(wb,"correlation")
openxlsx::writeData(wb,sheet="sheet",x=mtcars,colNames=TRUE,rowNames=TRUE)
openxlsx::writeData(wb,sheet="correlation",x=mtcor,colNames=TRUE,rowNames=TRUE)
excel_generic_format(df=mtcars,workbook=wb,sheet="sheet",title="test",
                    comment=comment,numFmt="#0.00")
excel_generic_format(df=mtcor,workbook=wb,sheet="correlation",title="correlation",
                    comment=comment,numFmt="#0.00")
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

excel_matrix

Write matrix or dataframe to excel sheet

Description

Usefull for corellation matrices. It uses conditional formatting for matrices,which outlines high and low values using background color

Usage

```
excel_matrix(
  df,
  workbook,
  sheet = "output",
  title = NULL,
  comment = NULL,
  numFmt = "#0.00",
  conditional_formatting = FALSE,
  diagonal = FALSE,
  diagonal_length = nrow(df)
)
```

Arguments

df	dataframe or matrix
workbook	workbook
sheet	sheet
title	title
comment	comment
numFmt	number formatting
conditional_formatting	if TRUE it will use conditional formatting
diagonal	if TRUE it will add background fill to diagonal
diagonal_length	length of diagonal for background fill

Examples

```
comment<-list(mpg="Miles/(US) gallon",
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio",
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1>manual)",
              gear="Number of forward gears",
              carb="Number of carburetors",
              extra_comment1="test1",
              extra_comment2="test2")

mtcor<-data.frame(cor(mtcars))
filename<-"excel_matrix.xlsx"
if (file.exists(filename)) file.remove(filename)
wb<-openxlsx::createWorkbook()
excel_matrix(mtcars,wb,sheet="matrix",comment=comment,
             conditional_formatting=TRUE,diagonal=FALSE)
excel_matrix(mtcars,wb,sheet="diagonal_non_square",comment=comment,
             conditional_formatting=FALSE,diagonal=TRUE)
excel_matrix(mtcars[1:10,1:10],wb,sheet="diagonal_square",comment=comment[1:10],
             conditional_formatting=FALSE,diagonal=TRUE)
excel_matrix(mtcars,wb,sheet="matrix_diagonal_non_square",comment=comment,
             conditional_formatting=TRUE,diagonal=TRUE)
excel_matrix(mtcars[1:10,1:10],wb,sheet="matrix_diagonal_square",comment=comment[1:10],
             conditional_formatting=TRUE,diagonal=TRUE)
excel_matrix(mtcors,wb,sheet="r",comment=comment,
             conditional_formatting=FALSE,diagonal=FALSE)
excel_matrix(mtcors,wb,sheet="conditional_formatting_r",comment=comment,
             conditional_formatting=TRUE,diagonal=TRUE)
openxlsx::saveWorkbook(wb,invisible(paste(filename)),TRUE)
```

extract_components	<i>Extract variance components from model</i>
--------------------	---

Description

Extract variance components from model

Usage

```
extract_components(model, title = "")
```

Arguments

model	model containing variance components
title	plot title

Examples

```
design<-expand.grid(time=1:3,item=1:3,person=1:10)
design<-change_data_type(design,type="factor")
design$response<-rowSums(change_data_type(design[,1:2],type="numeric"))+rnorm(90,0,0.1)
model<-mixlm::lm(response~r(time)*r(person)+r(item)*r(person),data=design)
extract_components(model)
```

flatten_list	<i>Flatten two dimensional list</i>
--------------	-------------------------------------

Description

Flatten two dimensional list

Usage

```
flatten_list(mydata)
```

Arguments

mydata	list with two dimensions
--------	--------------------------

```
generate_comparisons_matrix
```

Generate comparisons matrix

Description

Generate comparisons matrix

Usage

```
generate_comparisons_matrix(items)
```

Arguments

items	number of items
-------	-----------------

Examples

```
generate_comparisons_matrix(2)
generate_comparisons_matrix(3)
generate_comparisons_matrix(4)
generate_comparisons_matrix(5)
generate_comparisons_matrix(6)
```

```
generate_correlation_matrix
```

Generate dataframe which outputs a predetermined correlation matrix

Description

Generate dataframe which outputs a predetermined correlation matrix

Usage

```
generate_correlation_matrix(correlation_matrix, n_rows = 10)
```

Arguments

correlation_matrix	correlation matrix of resulting dataframe
n_rows	number of rows to generate

Examples

```
df<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_matrix<-as.matrix(df)
diag(correlation_matrix)<-1
df<-generate_correlation_matrix(correlation_matrix,nrows=100)
stats::cor(df)
```

generate_data

*Generate dataframe with random numbers***Description**

Generate dataframe with random numbers

Usage

```
generate_data(
  nrows = 10,
  ncols = 5,
  mean = 0,
  sd = 1,
  min = 1,
  max = 5,
  type = "normal"
)
```

Arguments

nrows	number of rows to generate
ncols	number of columns to generate
mean	mean of generated vectors
sd	standard deviation of generated vectors
min	minimum value in generated vector
max	maximum value in generated vector
type	character "normal" "uniform"

Examples

```
generate_data(nrows=10,ncols=5,mean=0,sd=1,type="normal")
generate_data(nrows=10,ncols=5,min=1,max=5,type="uniform")
```

generate_factor	<i>Generate dataframe of factors</i>
-----------------	--------------------------------------

Description

Generate dataframe of factors

Usage

```
generate_factor(vector = LETTERS[1:5], nrows = 2, ncols = 10, type = "random")
```

Arguments

vector	factor pool
nrows	number of rows to generate
ncols	number of collumns to generate
type	"balanced" or "random" "balanced" generates balanced factor vectrors, "random" generates random factor vectors

Examples

```
generate_factor(vector=LETTERS[1:5],ncols=5,nrows=10,type="random")
generate_factor(vector=LETTERS[1:5],ncols=5,nrows=10,type="balanced")
generate_factor(vector=LETTERS[1:5],ncols=1,nrows=10,type="balanced")
generate_factor(vector=LETTERS[1:5],ncols=1,nrows=10,type="random")
```

generate_matrix_A	<i>Generate Matrix A</i>
-------------------	--------------------------

Description

Generate Matrix A

Usage

```
generate_matrix_A(blocks = 3, items = 3)
```

Arguments

blocks	number of blocks
items	number of items per block

Examples

```
generate_matrix_A(blocks=3,items=3)
```

`generate_matrix_lambda_hat`*Generate matrix lambda for spesified number of comparisons*

Description

Generate matrix lambda for spesified number of comparisons

Usage

```
generate_matrix_lambda_hat(blocks = 3, items = 3)
```

Arguments

blocks	number of blocks
items	number of items per block

Examples

```
generate_matrix_lambda_hat(blocks=3,items=4)
```

`generate_missing`*Generate missing data*

Description

Generate missing data

Usage

```
generate_missing(df, missing = 5)
```

Arguments

df	vector or dataframe
missing	number of missing data per vector

Examples

```
generate_missing(rnorm(10),missing=5)
generate_missing(generate_data(nrow=10,ncol=2),missing=5)
```

generate_multiple_responce_vector
Generate multiple response vector

Description

Generate multiple response vector

Usage

```
generate_multiple_responce_vector(  
  responce = 1:4,  
  responded = 1:4,  
  length = 10  
)
```

Arguments

responce	unique categories allowed
responded	number of categories observed in iteration
length	length of returned vector

Examples

```
generate_multiple_responce_vector(responce=1:4,responded=1:4,length=10)
```

generate_string *Generate random strings*

Description

Generate random strings

Usage

```
generate_string(  
  vector = c(LETTERS, letters, 0:9),  
  vector_length = 1,  
  nchar = 5  
)
```

Arguments

vector	character pool
vector_length	number of strings to generate
nchar	Length of generated strings

Examples

```
generate_string(nchar=10)
generate_string(nchar=10,vector_length=10)
```

generate_unique_comparisons_index
<i>Generate index for unique comparisons</i>

Description

Generate index for unique comparisons

Usage

```
generate_unique_comparisons_index(items)
```

Arguments

items	number of items
-------	-----------------

Examples

```
generate_unique_comparisons_index(1)
generate_unique_comparisons_index(2)
generate_unique_comparisons_index(3)
generate_unique_comparisons_index(4)
generate_unique_comparisons_index(5)
generate_unique_comparisons_index(6)
```

getfwp	<i>Get working file path</i>
--------	------------------------------

Description

Get working file path

Usage

```
getfwp()
```

Examples

```
#getfwp()
```

get_coding	<i>Takes question response options from a json object and returns a list of dataframes mapping response option number to response option text</i>
------------	---

Description

Takes question response options from a json object and returns a list of dataframes mapping response option number to response option text

Usage

```
get_coding(coding_list)
```

Arguments

coding_list Json object containing question options

Examples

```
file<-"/opt/repo/rworking/projects/pcentral/data/aristotle-accounttests-6-17.json"
coding_list<-rjson::fromJSON(file=file)$instr_descr_options$biodata
get_coding(coding_list)
```

get_mplus_thu_3t	<i>Simulate prior distribution</i>
------------------	------------------------------------

Description

Simulate prior distribution

Usage

```
get_mplus_thu_3t(model)
```

Arguments

model mplus thurstonian cfa model with 3 traits

get_questionnaire_reference	<i>Questionnaire reference</i>
-----------------------------	--------------------------------

Description

Questionnaire reference

Usage

```
get_questionnaire_reference(mydata, questionnaire, tag_remove = "")
```

Arguments

- mydata json output
- questionnaire name of questionnaire to extract reference from
- tag_remove character

Examples

```
stud<-rjson::fromJSON(file="/opt/repo/rworking/projects/jacob/stud.json")
iboSTUDPULS2<-json2dataframe(stud$`items-by-order`$STUDPULS2)
get_questionnaire_reference(stud,"STUDPULS2","DTL2-ST")
```

get_reversals	<i>Questionnaire reference</i>
---------------	--------------------------------

Description

Questionnaire reference

Usage

```
get_reversals(df)
```

Arguments

- df dataframe from get_questionnaire_reference

Examples

```
stud<-rjson::fromJSON(file="/opt/repo/rworking/projects/jacob/stud.json")
res<-get_questionnaire_reference(stud,"STUDPULS2","DTL2-ST")
ibo<-res$items_by_order[!res$items_by_order$instrdtl_value %in% "-9999.00",]
ibo$instrdtl_value<-as.numeric(ibo$instrdtl_value)
get_reversals(ibo)
```

hinvert_title_grob	<i>Invert title for duplicating y axis</i>
--------------------	--

Description

Invert title for duplicating y axis

Usage

hinvert_title_grob(grob)

Arguments

grob grob object

icc_cfa	<i>Select responses for each dimension</i>
---------	--

Description

Select responses for each dimension

Usage

icc_cfa(eta, gamma, lambda, psi)

Arguments

eta eta or ability
gamma gamma or threshold
lambda lambda or loading
psi psi or error

Examples

icc_cfa(seq(-6,6,.1),1,1,1)

increase_index	<i>index dataframe picks</i>
----------------	------------------------------

Description

index dataframe picks

Usage

```
increase_index(blocks, items)
```

Arguments

blocks	number of blocks
items	number of items per block

Examples

```
increase_index(3,3)
```

install_all_packages	<i>Install all packages available in CRAN</i>
----------------------	---

Description

Install all packages available in CRAN

Usage

```
install_all_packages()
```

Details

Install all packages available in CRAN. Already installed packages are not downloaded or installed

install_load	<i>Install and load multiple packages</i>
--------------	---

Description

Install and load multiple packages. If packages exist, they are loaded, if packages don't exist, they are downloaded installed and loaded

Usage

```
install_load(package)
```

Arguments

package	Vector Package names
---------	----------------------

Author(s)

Steven Worthington

Examples

```
install_load("car")
install_load(c("car", "ggplot2"))
```

json2dataframe	<i>Convert a json object to dataframe</i>
----------------	---

Description

Convert a json object to dataframe

Usage

```
json2dataframe(mydata, fast = FALSE, row_to_collumn = FALSE)
```

Arguments

mydata	json object
fast	if TRUE uses a faster algorithm
row_to_collumn	if TRUE it will return row in first collumn

Examples

```
file<-"/opt/repo/rworking/projects/pcentral/data/aristotle-accounttests-6-17.json"
pcentral<-rjson::fromJSON(file=file)
biodata<-json2dataframe(mydata=pcentral$itemresponses$biodata)
biodata<-json2dataframe(mydata=pcentral$itemresponses$biodata, fast=TRUE)
```

key_to_cfa_model	<i>Converts key to cfa model spesification</i>
------------------	--

Description

This function uses the key spesification used in report_alpha function and converts the key to a cfa model spesification

Usage

```
key_to_cfa_model(key)
```

Arguments

key index of trait names and items constituring a trait

Examples

```
population_model<- 't1=~x1+.5*x2+.5*x3
                    t2=~x4+.5*x5+.5*x6
                    t3=~x7+.5*x8+.5*x9'
model_data<-lavaan::simulateData(population_model,sample.nobs=1000)
key<-list(f1=paste0("x",1:3),f2=paste0("x",4:6),f3=paste0("x",7:9))
model<-key_to_cfa_model(key)
fit<-lavaan::cfa(model,model_data)
```

k_fold	<i>K-Fold train test sampling</i>
--------	-----------------------------------

Description

splits a dataframe in train and test dataframes for model evaluation. Prepared data include data objects for xgboost

Usage

```
k_fold(df, model_formula, k = 10)
```

Arguments

df dataframe
model_formula model formula
k k-folds

Examples

```
infert_formula<-as.formula(factor(case)~age+parity+education+spontaneous+induced)
result<-k_fold(infert,k=10,model_formula=infert_formula)
model_formula<-as.formula(mpg~cyl+disp+hp+drat+wt+qsec+vs+am+gear+carb)
result<-k_fold(mtcars,k=2,model_formula=model_formula)
```

k_sample	<i>train test sampling</i>
----------	----------------------------

Description

splits a dataframe in train and test dataframes for model evaluation. Prepared data include data objects for xgboost

Usage

```
k_sample(df, model_formula, k = 1)
```

Arguments

df	dataframe
model_formula	model formula
k	k-folds

Examples

```
infert_formula<-as.formula(factor(case)~age+parity+education+spontaneous+induced)
result<-k_sample(df=infert,k=10,model_formula=infert_formula)
model_formula<-as.formula(mpg~cyl+disp+hp+drat+wt+qsec+vs+am+gear+carb)
result<-k_sample(df=mtcars,k=10,model_formula=model_formula)
```

matrix_triangle	<i>Return upper or lower matrix triangle</i>
-----------------	--

Description

Return upper or lower matrix triangle

Usage

```
matrix_triangle(m, off_diagonal = NA, diagonal = NULL, type = "lower")
```

Arguments

m	matrix
off_diagonal	off diagonal value
diagonal	diagonal value. If NULL it returns the diagonal of the input matrix
type	"upper" displays upper triangle, "lower" displays lower triangle

Examples

```
m<-matrix(1:9,nrow=3,ncol=3)
matrix_triangle(m=m)
matrix_triangle(m=m,diagonal=NA,type="lower")
matrix_triangle(m=m,diagonal=NULL,type="lower")
matrix_triangle(m=m,diagonal=NA,type="upper")
matrix_triangle(m=m,diagonal=NULL,type="upper")
```

mean_sd_alpha	<i>Mean and SD</i>
---------------	--------------------

Description

Mean and SD

Usage

```
mean_sd_alpha(df, divisor = NULL)
```

Arguments

df	dataframe with one dimension
divisor	number to use for dividing the rowsums

Examples

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
mean_sd_alpha(df)
mean_sd_alpha(df,divisor=100)
```

mgsub	<i>Sub for multiple patterns</i>
-------	----------------------------------

Description

Sub for multiple patterns

Usage

```
mgsub(mydata, pattern, replacement, ...)
```

Arguments

mydata	Character
pattern	Character to search for
replacement	Replacement character
...	arguments passed to gsub

Examples

```
mgsub(mydata="#$%^&*_+",pattern=c("%","*"),"REPLACE",fixed=TRUE)
```

min_max_index	<i>Return the minimum and maximum index of a vector</i>
---------------	---

Description

Return the minimum and maximum index of a vector

Usage

```
min_max_index(vector)
```

Arguments

vector	Vector
--------	--------

Examples

```
vector1<-c(1,2,3,4,5,4,3,2,1)
vector2<-c(1,2,3,4,5,5,3,2,1)
vector3<-c(1,2,3,5,5,4,3,2,1)
vector4<-c(1,2,3,4,6,4,3,2,1)
vector5<-c(1,6,3,4,6,4,3,2,1)
vector<-vector1
which(vector==max(vector),arr.ind=TRUE)
which(vector==min(vector),arr.ind=TRUE)
min_max_index(vector1)
min_max_index(vector2)
min_max_index(vector3)
min_max_index(vector4)
min_max_index(vector5)
```

model_loadings	<i>Pattern and structure matrix</i>
----------------	-------------------------------------

Description

Pattern and structure matrix

Usage

```
model_loadings(model, cut = NULL, matrix_type = "pattern", sort = TRUE, ...)
```

Arguments

model	psych EFA model
cut	cut point for loadings
matrix_type	"pattern" "structure" "all"
sort	if TRUE it will sort loadings
...	arguments passed to psych::fa.sort

Note

Check to see if you have multicollinearity values above .8 in the matrix are problematic
Structure matrix represents Loadings after rotation
Pattern matrix represents Loadings before rotation

Examples

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
model_loadings(model=cut=NULL,matrix_type="pattern")
model_loadings(model=model,cut=0.4,matrix_type="structure")
model_loadings(model=model,cut=0.4,matrix_type="all",sort=FALSE)
```

off_diagonal_index	<i>index of off diagonal</i>
--------------------	------------------------------

Description

index of off diagonal

Usage

```
off_diagonal_index(length)
```

Arguments

length	length of diagonal
--------	--------------------

Examples

```
off_diagonal_index(length=6)
```

outlier_summary	<i>Percent of outliers in vector</i>
-----------------	--------------------------------------

Description

Percent of outliers in vector

Usage

```
outlier_summary(vector)
```

Arguments

vector	numeric vector
--------	----------------

Details

returns dataframe

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
outlier_summary(vector)
data.frame(sapply(mtcars,outlier_summary))
```

```
output_compare_model_logistic
    Compare logistic regression models models
```

Description

Compare logistic regression models models

Usage

```
output_compare_model_logistic(model1, model2)
```

Arguments

model1	object glm model
model2	object glm model

Examples

```
modelcategoricalpredictor<-glm(case~education,data=infert,family=binomial)
modelcontinuouspredictor<-glm(case~age,data=infert,family=binomial)
modeltwopredictors<-glm(case~education*age,data=infert,family=binomial)
modelmultiple<-glm(case~education*age*parity,data=infert,family=binomial)
anova(modelcategoricalpredictor,modelcontinuouspredictor)
output_compare_model_logistic(model1=modelcategoricalpredictor,
                              model2=modeltwopredictors)
output_compare_model_logistic(model1=modelcontinuouspredictor,
                              model2=modeltwopredictors)
output_compare_model_logistic(model1=modelcontinuouspredictor,
                              model2=modelcategoricalpredictor)
```

```
output_separator    Output separator
```

Description

Heading, main output, and instructions for output for the console environment

Usage

```
output_separator(
  string,
  output = NULL,
  instruction = NULL,
  length = getOption("width")/2
)
```

Arguments

string	Title of output
output	object to print
instruction	Character provided instructions regarding the output
length	Numeric Length of separator measured in number of characters

Examples

```
output_separator(string="TEST",output="TEST",instruction="TEST",length=100)
output_separator(string="TEST",instruction="TEST",length=100)
output_separator(string="TEST",output="TEST",length=100)
output_separator(string="TEST")
```

padNA	<i>pad NA's to collumns in dataframe</i>
-------	--

Description

pad NA's to collumns in dataframe

Usage

```
padNA(df, rowsneeded, first = TRUE)
```

Arguments

df	dataframe
rowsneeded	Numeric number of rows needed
first	Boolean

Author(s)

Ananda Mahto

plot_acf	<i>Plot autocorrelation function of correlation covariance and partial correlation</i>
----------	--

Description

uses ggplot

Usage

```
plot_acf(df, lag.max = length(df), base_size = 10, title = "")
```

Arguments

df	ts object
lag.max	maximum lags to include
base_size	base font size
title	plot title

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)
plot_acf(df=ts_data,base_size=20)
```

plot_boxplot	<i>Boxplot</i>
--------------	----------------

Description

Boxplot

Usage

```
plot_boxplot(df, title = "", base_size = 10)
```

Arguments

df	dataframe or vector with continous or ordinal data
title	Plot title
base_size	numeric base font size

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_boxplot(df=vector)
plot_boxplot(df=generate_missing(vector))
plot_boxplot(df=df)
```

plot_cfa

Plot cfa model

Description

Plot cfa model

Usage

```
plot_cfa(model, ...)
```

Arguments

model	lavaan object
...	arguments passed to semPlot::semPaths

Examples

```
model='LATENT1=~X1+X2+X3
      LATENT2=~X4+X5+X6'
df<-lavaan::simulateData(model=model,model.type="cfa",
                        return.type="data.frame",sample.nobs=100)
df<-generate_missing(df)
fit<-lavaan::cfa(model,data=df,missing="ML")
plot_cfa(fit)
model='LATENT1=~X1+X2+X3+X4+X5+X6
      LATENT2=~X1+X2+X3+X4+X5+X6'
```

plot_confusion	<i>Plot confusion matrix</i>
----------------	------------------------------

Description

Plot confusion matrix

Usage

```
plot_confusion(observed, predicted, base_size = 10, title = "")
```

Arguments

observed	vector of observed outcomes
predicted	vector of predicted outcomes
base_size	base font size
title	plot title

Examples

```
plot_confusion(observed=c(1,2,3,1,2,3),predicted=c(1,2,3,1,2,3))
observed<-c(rep("male",10),rep("female",10),"male","male")
predicted<-c(rep("male",10),rep("female",10),"female","female")
plot_confusion(observed=observed,predicted=predicted)
```

plot_corrplot	<i>Correlation matrix plots</i>
---------------	---------------------------------

Description

Correlation matrix plots

Usage

```
plot_corrplot(mydata, title = "", base_size = 10, fill_limits = c(-1, 0, 1))
```

Arguments

mydata	correlation matrix
title	plot title
base_size	base font size
fill_limits	lower and upper limit for fill

Examples

```
plot_corrplot(stats::cor(mtcars),title="Correlation")
plot_corrplot(stats::cor(mtcars),base_size=20)
```

plot_crosstable	<i>Plot crosstables</i>
-----------------	-------------------------

Description

Plot crosstables

Usage

```
plot_crosstable(  
  df,  
  factor_index,  
  combinations = NULL,  
  shape = 16,  
  angle = 0,  
  base_size = 10,  
  title = ""  
)
```

Arguments

df	dataframe
factor_index	index of factors
combinations	index of comparisons
shape	shape of points
angle	angle of xaxis labels
base_size	base font size
title	plot title

Examples

```
combinations<-data.frame(index1=c("vs","am","gear"),index2=c("cyl","cyl","cyl"))  
plot_crosstable(df=mtcars,factor_index=8:9)  
plot_crosstable(df=mtcars,combinations=combinations)
```

plot_histogram	<i>Histograms with density function</i>
----------------	---

Description

Histograms with density function

Usage

```
plot_histogram(
  df,
  bins = 30,
  title = "",
  base_size = 10,
  xlims = NULL,
  fill = "gray25",
  color = "gray50",
  ylab = "Count"
)
```

Arguments

df	dataframe or vector with continous or ordinal data
bins	number of bars to display
title	plot title
base_size	numeric base font size
xlims	x axis limits
fill	color of bar
color	color of bar outline
ylab	y label

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_histogram(df=vector)
plot_histogram(df=df,xlims=c(0,50))
plot_histogram(df=df)
plot_multiplot(plotlist=plot_histogram(df=mtcars),cols=4)
```

plot_icc_thurstonian *Plot thurstonian icc*

Description

Plot icc curves for binary thurstonian coded items for a single dimension using the compute_icc_thurstonian function

Usage

```
plot_icc_thurstonian(mydata, title = "Item Characteristic Curve")
```

Arguments

mydata	dataframe from compute_icc_thurstonian function
title	plot title

Examples

```
gamma<-c(0.556,-1.253,-1.729,0.618,0.937,0.295,-0.672,-1.127,-0.446,0.632,1.147,0.498)
psi<-c(2.172,1.883,2.055,1.869,2.231,2.100,1.762,1.803,1.565,1.892,1.794,1.686)
lambda<-c(1.082,1.082,-1.297,-1.297,0.802,0.802,1.083,1.083)
gamma<-gamma[response_dimension(c(1:12),3,c(1,2))]
```

```
psi<-psi[response_dimension(c(1:12),3,c(1,2))]
```

```
eta<-seq(-6,6,by=1)
```

```
result<-compute_icc_thurstonian(eta=eta,gamma=gamma,lambda=lambda,psi=psi,plot=TRUE)
```

```
plot_icc_thurstonian(result$icc)
```

plot_interaction	<i>Plot two way interaction graphs</i>
------------------	--

Description

Plot two way interaction graphs

Usage

```
plot_interaction(
  df,
  dv,
  iv,
  base_size = 20,
  type = "se",
  order_factor = TRUE,
  title = "",
  note = ""
)
```

Arguments

df	dataframe
dv	index of continous variables
iv	index of factors
base_size	base font size

type	error bar type to display (1) "se" for standard error (2) "ci" for confidence interval (3) "sd" for standard deviation (4) "" for no error bar
order_factor	if TRUE it will sort the categorical axis by the continous variable value
title	plot title
note	footnote

Examples

```
nrows=1000
df<-data.frame(generate_factor(vector=LETTERS[1:5],nrows=nrows,ncols=10,type="random"),
               generate_data(nrows=nrows,ncols=5,type="normal"))
#result<-plot_interaction(df=df,dv=11:15,iv=1:10)
plot_interaction(df=mtcars,dv=2:3,iv=8:9,base_size=20,title="",type="se")
plot_interaction(df=mtcars,dv=2,iv=8:9,base_size=20,title="",type="se")
plot_interaction(df=mtcars,dv=2:3,iv=8:9,base_size=20,title="",type="ci")
plot_interaction(df=mtcars,dv=2:3,iv=9:10,base_size=20,title="",type="ci")
plot_interaction(df=mtcars,dv=2:3,iv=9:10,base_size=20,title="",type="sd")
plot_interaction(df=mtcars,dv=2,iv=9:10,base_size=20,
               title="",type="",order_factor=FALSE)
```

plot_irt_onefactor	<i>Return data for irt plots</i>
--------------------	----------------------------------

Description

Return data for irt plots

Usage

```
plot_irt_onefactor(model, theta = seq(-6, 6, 0.1), title = "", base_size = 10)
```

Arguments

model	object mirt
theta	theta
title	plot title
base_size	base size

Examples

```
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-4,high=4,d=NULL,a=1,mu=0,sd=1)$items
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Normal Test")
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-6,high=-4,d=NULL,a=1,mu=0,sd=1)$items
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Easy Items")
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=4,high=6,d=NULL,a=1,mu=0,sd=1)$items
```

```
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Difficult Items")
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-4,high=-4,d=NULL,a=0.01,mu=0,sd=1)$items
model<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
plot_irt_onefactor(model=model,base_size=10,title="Low Discrimination")
cormatrix<-psych::sim.poly(nvar=5,n=50000,low=-4,high=4,a=1,c=0,z=1,d=NULL,
                           mu=0,sd=1,cat=5,mod="logistic",theta=NULL)$items
model<-mirt::mirt(cormatrix,1,itemtype="graded")
plot_irt_onefactor(model=model,base_size=10,title="graded response")
```

plot_loadings	<i>Plot loadings</i>
---------------	----------------------

Description

Plot loadings

Usage

```
plot_loadings(
  model,
  matrix_type = NULL,
  title = "",
  base_size = 10,
  color = c("#5E912C", "white", "#5F2C91"),
  sort = TRUE
)
```

Arguments

model	psych EFA model
matrix_type	"pattern" "structure"
title	plot title
base_size	base font size
color	color ranges for heatmap

Examples

```
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
plot_loadings(model=model,matrix_type="structure")
plot_loadings(model=model,matrix_type="pattern")
cm<-matrix(c(1,.8,.8,.1,.1,.1,
             .8,1,.8,.1,.1,.1,
             .8,.8,1,.1,.1,.1,
             .1,.1,.1,1,.8,.8,
             .1,.1,.1,.8,1,.8,
             .1,.1,.1,.8,.8,1),
           ncol=6,nrow=6)
```

```

df1<-generate_correlation_matrix(cm,nrows=10000)
model1<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
plot_loadings(model=model1,matrix_type="pattern",base_size=30)
cm<-matrix(c(1,.1,.1,.1,.1,.1,
             .1,1,.1,.1,.1,.1,
             .1,.1,1,.1,.1,.1,
             .1,.1,.1,1,.8,.8,
             .1,.1,.1,.8,1,.8,
             .1,.1,.1,.8,.8,1),
           ncol=6,nrow=6)
df1<-generate_correlation_matrix(cm,nrows=10000)
model2<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
plot_loadings(model=model2,matrix_type="pattern",base_size=30)
cm<-matrix(c(1,.01,.01,.01,.01,.01,
             .01,1,.01,.01,.01,.01,
             .01,.01,1,.01,.01,.01,
             .01,.01,.01,1,.01,.01,
             .01,.01,.01,.01,1,.01,
             .01,.01,.01,.01,.01,1),
           ncol=6,nrow=6)
df1<-generate_correlation_matrix(cm,nrows=10000)
model3<-psych::fa(df1,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
plot_loadings(model=model3,matrix_type="pattern",base_size=10)

```

plot_logistic_model *Logistic model plot*

Description

Logistic model plot

Usage

```
plot_logistic_model(df, outcome = "outcome", title = "", base_size = 10)
```

Arguments

df	dataframe with predictor and outcome outcome should be last
outcome	name of outcome variable
title	Character plot title
base_size	base font size

Examples

```

df<-data.frame(outcome=c(rep(1,10),rep(0,10)),
               pd1=c(rep(1,11),rep(0,9)),
               pd2=c(rep(1,9),rep(0,11)),
               pc1=c(rnorm(10,mean=5),rnorm(10,mean=10)),
               pc2=c(rnorm(10,mean=5),rnorm(10,mean=20)))
plot_logistic_model(df=df,base_size=15)

```

plot_mosaic	<i>Plot mosaic plots</i>
-------------	--------------------------

Description

Plot mosaic plots

Usage

```
plot_mosaic(df, factor_index, base_size = 10, title = "")
```

Arguments

df	dataframe
factor_index	index of factors
base_size	base font size
title	plot title

Examples

```
plot_mosaic(df=mtcars, factor_index=8:9)
plot_mosaic(df=mtcars, factor_index=9:10)
```

plot_mtmm	<i>Plot multitrait multimethod matrix</i>
-----------	---

Description

Plot multitrait multimethod matrix

Usage

```
plot_mtmm(df, key, method, subject, title = "")
```

Arguments

df	dataframe
key	List index of trait names and items constituring a trait
method	name of dataframe collumn spesifying the method used for the row observed
subject	name of dataframe collumn spesifying subject id
title	plot title

Examples

```

population_model<- 't1=~x1+.9*x2+.9*x3
                    t2=~x4+.9*x5+.9*x6
                    t3=~x7+.9*x8+.9*x9'

model_data<-lavaan::simulateData(population_model,sample.nobs=1000)
model_data<-model_data[sample(1:1000,1000,TRUE),]
model_data<-rbind(model_data,model_data,model_data)
model_data$method<-c(rep("m1",1000),rep("m2",1000),rep("m3",1000))
model_data$id<-rep(1:1000,3)
key<-list(t1=paste0("x",1:3),t2=paste0("x",4:6),t3=paste0("x",7:9))
plot_mtmm(df=model_data,key=key,method="method",subject="id")

```

plot_multiplot

*Multiple ggplot plots in one graph***Description**

Multiple ggplot plots in one graph

Usage

```
plot_multiplot(..., plotlist = NULL, cols = 2, layout = NULL)
```

Arguments

...	plot objects
plotlist	a list of plots
cols	number of columns in layout
layout	a matrix specifying the layout. If present, 'cols' is ignored

Examples

```

p1<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet,group=Chick))+
  geom_line()+
  ggtitle("Growth curve for individual chicks")+
  theme_bw()
p2<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet))+
  geom_point(alpha=.3)+
  geom_smooth(alpha=.2,size=1,method="loess",formula="y~x")+
  ggtitle("Fitted growth curve per diet")+
  theme_bw()
p3<-ggplot(subset(ChickWeight,Time==21),aes(x=weight,colour=Diet))+
  geom_density()+
  ggtitle("Final weight, by diet")+theme_bw()
p4<-ggplot(subset(ChickWeight,Time==21),aes(x=weight,fill=Diet))+
  geom_histogram(colour="black",binwidth=50)+facet_grid(Diet~.)+
  ggtitle("Final weight, by diet")+theme_bw()
cars_plot<-plot_histogram(mtcars)

```

```

plot_multiplot(p1,p2,p3,p4,cols=2)
plot_multiplot(plotlist=plot_histogram(mtcars[,1:4]),cols=2)
plot_multiplot(plotlist=plot_histogram(mtcars),layout=matrix(1:4,ncol=2,byrow=TRUE))
plot_multiplot(plotlist=plot_scatterplot(mtcars[,1:4]),cols=2)
plot_multiplot(plotlist=cars_plot,layout=matrix(1:4,ncol=2,byrow=TRUE))
plot_multiplot(plotlist=cars_plot,cols=3)

```

```

plot_normality_diagnostics
      Normality plots

```

Description

plot histogram density boxplot qq plot

Usage

```

plot_normality_diagnostics(
  df,
  breaks = NULL,
  title = "",
  file = NULL,
  w = 10,
  h = 10
)

```

Arguments

df	dataframe or vector with continous or ordinal data
breaks	number of bars to display
title	plot title
file	output filename
w	width of pdf file
h	height of pdf file

Details

uses plot base

Examples

```

vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_normality_diagnostics(df=vector,title="",file="rnorm",breaks=30)
plot_normality_diagnostics(df=vector,title="")
plot_normality_diagnostics(df=df,title="mtcars")
plot_normality_diagnostics(df=df,title="mtcars",file="rnorm")

```

plot_oneway

Plot means with standard error for every level in a dataframe

Description

Plot means with standard error for every level in a dataframe

Usage

```
plot_oneway(
  df,
  dv,
  iv,
  base_size = 20,
  type = "se",
  order_factor = TRUE,
  title = "",
  note = "",
  width = 60
)
```

Arguments

df	dataframe
dv	index of continous variables
iv	index of factors
base_size	base font size
type	error bar type to display (1) "se" for standard error (2) "ci" for confidence interval (3) "sd" for standard deviation (4) "" for no error bar
order_factor	if TRUE it will sort the categorical axis by the continous variable value
title	plot title
note	footnote
width	wrap width for x title

Examples

```
nrows=1000
df<-data.frame(generate_factor(vector=LETTERS[1:5],nrows=nrows,ncols=10,type="random"),
               generate_data(nrows=nrows,ncols=5,type="normal"))
result<-plot_oneway(df=df,dv=11:15,iv=1:10)
plot_oneway(df=mtcars,dv=2,iv=9)
plot_oneway(df=mtcars,dv=2,iv=9:10)
plot_oneway(df=mtcars,dv=2:3,iv=10)
plot_oneway(df=mtcars,dv=2:3,iv=9:10)
plot_oneway(df=mtcars,dv=2:3,iv=9:10,type="se")
```

```
plot_oneway(df=mtcars, dv=2:3, iv=9:10, type="ci")
plot_oneway(df=mtcars, dv=2:3, iv=9:10, type="sd")
plot_oneway(df=mtcars, dv=2:3, iv=9:10, type="", order_factor=FALSE)
plot_oneway(df=mtcars, dv=2:3, iv=9:10, type="", order_factor=TRUE)
```

plot_oneway_diagnostics

Plot one way diagnostics

Description

Plot one way diagnostics

Usage

```
plot_oneway_diagnostics(df, dv, iv, base_size = 10)
```

Arguments

df	dataframe
dv	index of continous variables
iv	index of factors
base_size	base font size

Note

Residuals vs Fitted should be equally spread horizontally otherwise the assumption of equality of variances is violated

Normal QQ should show values in the diagonal otherwise the assumption of normality is violated

Examples

```
nrows=1000
df<-data.frame(generate_factor(vector=LETTERS[1:5],nrows=nrows,ncols=10,type="random"),
               generate_data(nrows=nrows,ncols=5,type="normal"))
result<-plot_oneway_diagnostics(df=df, dv=11:15, iv=1:10)
plot_oneway_diagnostics(df=mtcars, dv=1:2, iv=9:10)
```

plot_outlier	<i>Outlier graph using mean median and boxplot algorythms</i>
--------------	---

Description

Outlier graph using mean median and boxplot algorythms

Usage

```
plot_outlier(df, method = "mean", title = "", base_size = 10)
```

Arguments

df	dataframe or vector with continous or ordinal data
method	"mean" "median" "boxplot"
title	plot title
base_size	base font size

Author(s)

unknown

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_outlier(df=vector,method="mean",title="random vector")
plot_outlier(df=vector,method="median")
plot_outlier(df=vector,method="boxplot")
plot_outlier(df=df,method="mean",title="random vector")
plot_outlier(df=df,method="median")
plot_outlier(df=df,method="boxplot")
plot_mplot(plotlist=plot_outlier(df=mtcars[,2:5],method="mean"),cols=2)
```

plot_qq	<i>qq plots</i>
---------	-----------------

Description

qq plots

Usage

```
plot_qq(df, title = "", base_size = 10)
```

Arguments

df	dataframe or vector with continous or ordinal data
title	plot title
base_size	numeric base font size

Details

uses ggplot

Examples

```
vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
plot_qq(df=vector)
plot_qq(df=df)
plot_mplot(plotlist=plot_qq(df=mtcars),cols=4)
```

plot_response_frequencies

Plot response frequencies

Description

Plot response frequencies

Usage

```
plot_response_frequencies(
  df,
  factor_index,
  base_size = 10,
  title = "",
  width = 100
)
```

Arguments

df	dataframe
factor_index	index of factors
base_size	base font size
title	plot title
width	wrap width for x title

Examples

```
plot_response_frequencies(df=mtcars,factor_index=1:10)
```

plot_roc	<i>Plot ROC curve</i>
----------	-----------------------

Description

Plot ROC curve

Usage

```
plot_roc(observed, predicted, base_size = 10, title = "")
```

Arguments

- observed vector of observed outcomes
- predicted vector of predicted outcome probability
- base_size base font size
- title plot title

Examples

```
observed<-round(abs(rnorm(100,m=0,sd=.5)))
predicted<-abs(rnorm(100,m=0,sd=.5))
plot_roc(observed=observed,predicted=predicted)
df1<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_martix<-as.matrix(df1)
diag(correlation_martix)<-1
df1<-generate_correlation_matrix(correlation_martix,nrows=1000)
df1$X1<-ifelse(abs(df1$X1)<1,0,1)
df1$X2<-abs(df1$X2)
df1$X2<-(df1$X2-min(df1$X2))/(max(df1$X2)-min(df1$X2))
plot_roc(observed=round(abs(df1$X1),0),predicted=abs(df1$X2))
```

plot_scatterplot	<i>Plot plot_scatterplot</i>
------------------	------------------------------

Description

Plot plot_scatterplot

Usage

```
plot_scatterplot(
  df,
  method = lm,
  formula = y ~ x,
  base_size = 10,
  coord_equal = FALSE,
  all_orders = FALSE,
  title = "",
  combinations = NULL,
  string_aes = TRUE
)
```

Arguments

df	dataframe if dataframe consists of 2 columns the second column is the outcome and the first column is the predictor
method	smoothing method, "auto", "lm", "glm", "gam", "loess" or a function, e.g. MASS::rlm or mgcv::gam, stats::lm, or stats::loess
formula	formula used in smoothing function for geom_smooth
base_size	base font size
coord_equal	if TRUE axes maintain equal scale
all_orders	if TRUE the order of combination is considered
title	Plot title
combinations	dataframe if not NULL user can provide a dataframe for variable combinations for x and y axis . First column represents x and second column represents y
string_aes	if TRUE string_aes function is used for names

Examples

```
result<-plot_scatterplot(df=mtcars,title="",coord_equal=TRUE,base_size=10)
plot_multiplot(plotlist=result[1:12],cols=4)
plot_scatterplot(df=mtcars[,1:2],base_size=10,coord_equal=TRUE,all_orders=FALSE)
plot_scatterplot(df=mtcars[,1:2],base_size=10,coord_equal=FALSE,all_orders=FALSE)
plot_scatterplot(df=mtcars,base_size=10,coord_equal=TRUE,all_orders=FALSE,
  combinations=data.frame(x=c("mpg","mpg","mpg"),
    y=c("cyl","hp","mpg")))
plot_scatterplot(df=mtcars,base_size=10,coord_equal=TRUE,all_orders=TRUE,
  combinations=data.frame(x=c("mpg"),y=c("cyl")))
x<-rnorm(1000)
y<-x+rnorm(x,sd=.1)
plot_scatterplot(df=data.frame(x,y),title="Random Simulation",coord_equal=TRUE)
df<-data.frame(matrix(-.999,ncol=2,nrow=2))
correlation_matrix<-as.matrix(df)
diag(correlation_matrix)<-1
df<-generate_correlation_matrix(correlation_matrix,nrows=1000)
plot_scatterplot(df,title="Simulation of -.999 Correlation",coord_equal=TRUE,base_size=20)
```

plot_scree	<i>Scree plot displaying the Kaiser and Jolife criteria for factor extraction</i>
------------	---

Description

Scree plot displaying the Kaiser and Jolife criteria for factor extraction

Usage

```
plot_scree(df, base_size = 15, title = "", color = c("#5F2C91", "#5E912C"))
```

Arguments

df	dataframe
base_size	base font size
title	plot title
color	color of line and point outline

Examples

```
plot_scree(df=mtcars,title="",base_size=15)
scree(mtcars)
```

plot_separability	<i>Plot separability</i>
-------------------	--------------------------

Description

Plot separability

Usage

```
plot_separability(observed, predicted, base_size = 10, title = "")
```

Arguments

observed	vector of observed outcomes
predicted	vector of predicted outcome probability
base_size	base font size
title	plot title

Examples

```
df1<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_matrix<-as.matrix(df1)
diag(correlation_matrix)<-1
df1<-generate_correlation_matrix(correlation_matrix,nrows=1000)
df1$X1<-ifelse(abs(df1$X1)<1,0,1)
df1$X2<-abs(df1$X2)
df1$X2<-(df1$X2-min(df1$X2))/(max(df1$X2)-min(df1$X2))
plot_separability(observed=round(abs(df1$X1),0),predicted=abs(df1$X2))
```

plot_ts

*Plot timeseries***Description**

Plot timeseries

Usage

```
plot_ts(df, base_size = 10, ylab = "Count", title = "")
```

Arguments

df	ts object
base_size	base font size
ylab	y label
title	plot title

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)
result<-plot_ts(ts_data,title="UK driver deaths")
for(i in 1969:1984)
  result<-result+geom_vline(xintercept=i,color="blue",size=1,alpha=.5)
result
autoplot(stl(ts_data,s.window='periodic'))+
  theme_bw(base_size=10)+
  labs(title="UK driver deaths")
forecast::gglagplot(data.frame(ts_data),do.lines=FALSE,lags=100)+
  theme_bw(base_size=10)+labs(title="UK driver deaths",y="count")
```

proper	<i>Capitalize first character and lowercase the rest</i>
--------	--

Description

Capitalize first character and lowercase the rest

Usage

proper(x)

Arguments

x Character

Examples

```
x<-generate_string(nchar=10,vector=LETTERS,vector_length=10)
proper(x)
```

proportion_accurate	<i>Proportion overall accuracy of a confusion matrix</i>
---------------------	--

Description

Proportion overall accuracy of a confusion matrix

Usage

proportion_accurate(observed, predicted)

Arguments

observed vector of observed variables
predicted vector of predicted variables

Examples

```
proportion_accurate(observed=c(1,2,3,4,5,10),predicted=c(1,2,3,4,5,11))
```

questions_by_keys	<i>Convert key to index list</i>
-------------------	----------------------------------

Description

Convert key to index list

Usage

questions_by_keys(key)

Arguments

key	a vector indicating the dimension of each question. The order of the elements in the key represents the order of the questions, the numeric values represent the dimension the question belongs to
-----	--

Examples

```
key<-c(1,2,3,4,5,1,2,3,4,5)
questions_by_keys(key)
```

questions_dimensions_dataframe	<i>Question dimension table</i>
--------------------------------	---------------------------------

Description

Return a dataframe with the order of the questions, their respective dimensions, and the description of the dimensions

Usage

```
questions_dimensions_dataframe(
  key,
  dimensions,
  elaborate_dimensions,
  questions
)
```

Arguments

key	a vector indicating the dimension of each question. The order of the elements in the key represents the order of the questions, the numeric values represent the dimension the question belongs to
dimensions	dimension names
elaborate_dimensions	full dimension names
questions	question names

Examples

```
key<-c(1,2,3,4,5,1,2,3,4,5)
dimensions<-paste0("Dimension",1:10)
elaborate_dimensions<-paste0("Elaborated_Dimension",1:10)
questions<-paste0("Question",1:65)
questions_dimensions_dataframe(key,dimensions,elaborate_dimensions,questions)
```

rad2deg	<i>Convert radians to degrees</i>
---------	-----------------------------------

Description

Convert radians to degrees

Usage

```
rad2deg(radians)
```

Arguments

radians	radians
---------	---------

Examples

```
rad2deg(pi)
```

rank3_to_triplets	<i>Convert thurstonian binary triplets to scale</i>
-------------------	---

Description

Convert thurstonian binary triplets to scale

Usage

```
rank3_to_triplets(mydata)
```

Arguments

mydata	dataframe
--------	-----------

Examples

```
set.seed(12345)
mydata<-data.frame(i1=rnorm(10,mean=2,sd=.5),
                   i2=rnorm(10,mean=2,sd=.5),
                   i3=rnorm(10,mean=2,sd=.5),
                   i4=rnorm(10,mean=2,sd=.5),
                   i5=rnorm(10,mean=2,sd=.5),
                   i6=rnorm(10,mean=2,sd=.5))
result<-rank_to_binary(mydata[,1:3])
rank3_to_triplets(result)
```

rank_df_to_binary	<i>Convert scale to thurstonian binary with n items per block and n blocks</i>
-------------------	--

Description

Convert scale to thurstonian binary with n items per block and n blocks

Usage

```
rank_df_to_binary(mydata, items, reverse = TRUE)
```

Arguments

mydata	dataframe
items	number of items in block
reverse	if TRUE assumes that the highest value is first item in rank if FALSE the lowest value is the first item in rank

Examples

```
set.seed(12345)
mydata<-data.frame(i1=rnorm(10,mean=2,sd=.5),
                   i2=rnorm(10,mean=2,sd=.5),
                   i3=rnorm(10,mean=2,sd=.5),
                   i4=rnorm(10,mean=2,sd=.5),
                   i5=rnorm(10,mean=2,sd=.5),
                   i6=rnorm(10,mean=2,sd=.5))
rank_df_to_binary(mydata[,c("i1","i2","i3","i4")],4)
rank_df_to_binary(mydata,3)
```

rank_to_binary	<i>Convert scale to thurstonian binary with n items per ranking block</i>
----------------	---

Description

Convert scale to thurstonian binary with n items per ranking block

Usage

```
rank_to_binary(mydata, items, reverse = TRUE)
```

Arguments

mydata	dataframe
items	number of items in block
reverse	if TRUE assumes that the highest value is first item in rank if FALSE the lowest value is the first item in rank

Examples

```
set.seed(12345)
mydata<-data.frame(i1=round(rnorm(10,mean=2,sd=1),2),
                   i2=round(rnorm(10,mean=2,sd=1),2),
                   i3=round(rnorm(10,mean=2,sd=1),2),
                   i4=round(rnorm(10,mean=2,sd=1),2),
                   i5=round(rnorm(10,mean=2,sd=1),2),
                   i6=round(rnorm(10,mean=2,sd=1),2))
rank_to_binary(mydata[,c("i1","i2","i3")],items=3)
rank_to_binary(mydata[,c("i1","i2","i3")],items=3,reverse=FALSE)
rank_to_binary(mydata,items=3)
```

`raw_alpha`*Raw alpha*

Description

Raw alpha

Usage`raw_alpha(df)`**Arguments**`df` dataframe with one dimension**Examples**

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
psych::alpha(df)
raw_alpha(df=df)
```

`rbind_all`*rbind dataframes or matrices with different lengths or collumn names*

Description

rbind dataframes or matrices with different lengths or collumn names

Usage`rbind_all(df1, df2)`**Arguments**`df1` dataframe or matrix
`df2` dataframe or matrix**Examples**

```
df1<-generate_correlation_matrix(n=10)
df2<-generate_correlation_matrix(n=10)
names(df2)[4]<-"X11"
rbind_all(df1=df1,df2=df2)
row.names(df1)<-21:30
rbind_all(df1=df1,df2=df2)
```

recode_dataframe	<i>Takes a dataframe mapping response option number to response option text and recodes a numeric response vector to text response option for the entire dataframe</i>
------------------	--

Description

Takes a dataframe mapping response option number to response option text and recodes a numeric response vector to text response option for the entire dataframe

Usage

```
recode_dataframe(df, coding_map)
```

Arguments

- df dataframe
- coding_map dataframe containg map between coding and text

Examples

```
file<-"/opt/repo/rworking/projects/pcentral/data/aristotle-accounttests-6-17.json"
pcentral<-rjson::fromJSON(file=file)
df<-json2dataframe(pcentral$itemresponses$biodata)
coding_map<-get_coding(pcentral$instr_descr_options$biodata)
names(df)[c(2,11,9,3,5,6,10,12,7)]<-names(coding_map)
recode_dataframe(df=df,coding_map=coding_map)
file<-"/opt/repo/rworking/projects/pcentral/data/aristotle-accounttests-6-17.json"
pcentral<-rjson::fromJSON(file=file)
coding_map<-get_coding(pcentral$instr_descr_options$biodata)$Education
vector<-json2dataframe(pcentral$itemresponses$biodata)$education[1:50]
recode_dataframe(df=vector,coding_map=coding_map)
```

recode_scale_dummy	<i>Scale and dummy code</i>
--------------------	-----------------------------

Description

Scales numeric variables between 0 and 1 and creates dummy coding for character and vector variables

Usage

```
recode_scale_dummy(df, categories = 10)
```

Arguments

df	dataframe
categories	Numeric Number of unique values a vector must have to perform dummy coding

Examples

```
recode_scale_dummy(infert)
```

remove_nc	<i>Replace remove non computable values</i>
-----------	---

Description

Replace remove non computable values

Usage

```
remove_nc(
  df,
  value = NA,
  remove_rows = FALSE,
  aggressive = FALSE,
  remove_cols = FALSE,
  remove_zero_variance = FALSE
)
```

Arguments

df	dataframe
value	replacement
remove_rows	if TRUE it will remove rows with non computable values
aggressive	if TRUE it will remove entire row if a single non computable value exists if FALSE it will remove row if all values are non computable
remove_cols	if TRUE it will remove collumns with non computable values
remove_zero_variance	if TRUE it will remove collumns with no variance

Details

Non computable values are NA, NAN, inf and empty cells.

Note

This function internally replaces non computable values with the value choosen the default value is NA. Then it removes rows and collumns with NA values or zero variance

Examples

```

df<-mtcars
df[1,]<-as.numeric(NaN)
df[2,]<-as.numeric(Inf)
df[3,]<-as.numeric(-Inf)
df[4,]<-as.numeric(NA)
df[5,]<-" "
remove_nc(df=df,value=NA)
cdf(remove_nc(df=df,value=NA))
df<-generate_missing(mtcars,missing=5)
remove_nc(df,remove_rows=TRUE,aggressive=FALSE)
remove_nc(df,remove_rows=TRUE,aggressive=TRUE)
df<-generate_missing(generate_correlation_matrix(nrows=5),missing=2)
df$X2<-NA
df$X3<-1
remove_nc(df,remove_cols=TRUE,remove_zero_variance=FALSE)
remove_nc(df,remove_cols=TRUE,remove_zero_variance=TRUE)

```

remove_outliers

*Remove outliers***Description**

Remove outliers

Usage

```
remove_outliers(vector, probs = c(0.25, 0.75), na.rm = TRUE, ...)
```

Arguments

vector	numeric
probs	numeric vector with lowest and highest quantiles
na.rm	if TRUE removes NA values
...	arguments passed to quantile

Examples

```

vector<-generate_missing(rnorm(1000))
df<-generate_missing(mtcars[,1:2])
remove_outliers(vector)
data.frame(sapply(df,remove_outliers))

```

remove_user_packages	<i>Remove all user packages</i>
----------------------	---------------------------------

Description

Remove all user packages

Usage

```
remove_user_packages()
```

replace_na_with_previous	<i>Replace NA with the previous element in a vector</i>
--------------------------	---

Description

Replace NA with the previous element in a vector

Usage

```
replace_na_with_previous(vector)
```

Arguments

vector	Vector
--------	--------

Examples

```
df1<-generate_missing(rnorm(10),missing=5)
df2<-generate_missing(rnorm(10),missing=5)
df3<-generate_missing(rnorm(10),missing=5)
df4<-generate_missing(rnorm(10),missing=5)
df5<-generate_missing(rnorm(10),missing=5)
df<-data.frame(df1,df2,df3,df4,df5)
row.names(df)<-paste0("A",row.names(df))
replace_na_with_previous(df1)
df[]<-lapply(df,replace_na_with_previous)
```

report_alpha

*Estimate alpha for several dimensions and export results to xlsx***Description**

Uses an arbitrary input

Usage

```
report_alpha(
  df,
  key = NULL,
  questions = NULL,
  reverse = NULL,
  mini = NULL,
  maxi = NULL,
  file = NULL,
  ...
)
```

Arguments

df	dataframe
key	index of trait names and items constituting a trait
questions	trait names and items constituting a trait
reverse	index of trait names and index for reversal
mini	minimum rating in scale if NULL reversal will be performed using the empirical minimum
maxi	maximum rating in scale if NULL reversal will be performed using the empirical maximum
file	output filename
...	arguments passed to psych::alpha

Examples

```
set.seed(12345)
df<-data.frame(matrix(.5,ncol=6,nrow=6))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-round(generate_correlation_matrix(correlation_martix,nrows=1000),0)+5
key<-list(f1=c("X1","X2","X3"),
          f2=c("X4","X5","X6"))
reverse<-list(f1=c(1,1,1),
              f2=c(1,1,1))
report_alpha(df=df,key=key,cumulative=TRUE,n.iter=1)
report_alpha(df=df,key=key,reverse=reverse,check.keys=FALSE,n.iter=2)
report_alpha(df=df,key=key,check.keys=FALSE,n.iter=2,file="alpha")
```

report_cfa	<i>Report</i>
------------	---------------

Description

Report

Usage

```
report_cfa(model, file = NULL, w = 10, h = 10)
```

Arguments

model	lavaan object
file	output filename
w	width of pdf file
h	height of pdf file

Examples

```
model='LATENT=~ITEM1+ITEM2+ITEM3+ITEM4+ITEM5'
df<-lavaan::simulateData(model=model,model.type="cfa",
                          return.type="data.frame",sample.nobs=100)
df<-generate_missing(df)
fit<-lavaan::cfa(model,data=df,missing="ML")
report_cfa(fit)
report_cfa(fit,file="cfa")
```

report_choric_serial	<i>Report polychoric tetrachoric polyserial biserial correlation</i>
----------------------	--

Description

Report polychoric tetrachoric polyserial biserial correlation

Usage

```
report_choric_serial(
  x,
  y = NULL,
  file = NULL,
  w = 10,
  h = 10,
  type = "tetrachoric",
  ...
)
```

Arguments

x	<p>The input may be in one of four forms:</p> <p>a) a data frame or matrix of dichotomous data (e.g., the lsat6 from the bock data set) or discrete numerical (i.e., not too many levels, e.g., the big 5 data set, bfi) for polychoric, or continuous for the case of biserial and polyserial</p> <p>b) a 2 x 2 table of cell counts or cell frequencies (for tetrachoric) or an n x m table of cell counts (for both tetrachoric and polychoric)</p> <p>c) a vector with elements corresponding to the four cell frequencies (for tetrachoric)</p> <p>d) a vector with elements of the two marginal frequencies (row and column) and the comorbidity (for tetrachoric)</p>
y	matrix or dataframe of discrete scores. In the case of tetrachoric, these should be dichotomous, for polychoric not too many levels, for biserial they should be discrete (e.g., item responses) with not too many (<10?) categories
file	output filename
w	width of pdf file
h	height of pdf file
type	"tetrachoric" "polychoric" "polyserial" "biserial"
...	arguments passed to psych::polychoric

Examples

```
report_choric_serial(generate_data(min=0,max=1,type="uniform"),
  type="tetrachoric",file="tetrachoric")
report_choric_serial(generate_data(min=1,max=5,type="uniform"),
  type="polychoric")
report_choric_serial(x=psych::lsat6,y=psych::lsat6,
  type="polyserial",file="polyserial")
report_choric_serial(x=psych::lsat6,y=psych::lsat6,
  type="biserial",file="biserial")
```

report_correlation	<i>Report correlation matrix</i>
--------------------	----------------------------------

Description

Report correlation matrix

Usage

```
report_correlation(
  x,
  y = NULL,
  use = "pairwise",
```



```

    method = "pearson",
    adjust = "holm",
    alpha = 0.05,
    ci = TRUE,
    file = NULL,
    w = 10,
    h = 10,
    base_size = 20,
    scatterplot = TRUE
  )

```

Arguments

x	matrix or dataframe
y	a second matrix or dataframe with the same number of rows as x
use	"pairwise" is the default value and will do pairwise deletion of cases. "complete" will select just complete cases
method	"pearson" "spearman" "kendall"
adjust	"holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none"
alpha	alpha level of confidence intervals
ci	By default, confidence intervals are found. However, this leads to a great slow-down of speed. So, for just the rs, ts and ps, set ci=FALSE
file	output filename
w	width of pdf file
h	height of pdf file
base_size	base font size
scatterplot	if TRUE it will output scatterplots

Examples

```

report_correlation(x=generate_missing(mtcars[,1:3],10))
report_correlation(x=generate_missing(mtcars[,1:3],10),
                   file="correlation",scatterplot=TRUE)
report_correlation(x=mtcars[,1:3],file="correlation")

```

report_dataframe	<i>Write matrix or dataframe to excel sheet</i>
------------------	---

Description

Usefull for generic data where conditional formating of a spesific collumn is required

Usage

```
report_dataframe(df, file = NULL, type = "critical_value", ...)
```

Arguments

df	dataframe or matrix
file	output filename of excel file
type	"critical_value" "matrix"
...	arguments passed to excel_critical_value or to excel_matrix

Examples

```
comment<-list(mpg="Miles/(US) gallon",
              cyl="Number of cylinders",
              disp="Displacement (cu.in.)",
              hp="Gross horsepower",
              drat="Rear axle ratio",
              wt="Weight (1000 lbs)",
              qsec="1/4 mile time",
              vs="Engine (0=V-shaped,1=straight)",
              am="Transmission (0=automatic,1>manual)",
              gear="Number of forward gears",
              carb="Number of carburetors")
report_dataframe(mtcars,sheet="report",file="mtcars",comment=comment,numFmt="#0.00",
                critical=list(am="<0.05"))
report_dataframe(mtcars,sheet="report",file=NULL,comment=comment,numFmt="#0.00",
                critical=list(am="<0.05"))
```

report_efa	<i>Output EFA model</i>
------------	-------------------------

Description

Output EFA model

Usage

```
report_efa(
  model,
  df,
  file = NULL,
  w = 10,
  h = 5,
  cut = 0,
  base_size = 10,
  scores = FALSE
)
```

Arguments

model	psych EFA model
df	dataframe
file	output filename
w	width of pdf file
h	height of pdf file
cut	cut point for loadings
base_size	base font size
scores	if TRUE it will output factor scores in excel file

Note

Orthogonal=varimax, Oblique=oblimin

Examples

```

model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minres",oblique.scores=TRUE)
report_efa(model=model,df=mtcars,file="efa")
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="uls",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="ols",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="wls",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="gls",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="pa",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="ml",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minchi",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="minrank",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="old.min",oblique.scores=TRUE)
report_efa(model=model,df=mtcars)
model<-psych::fa(mtcars,nfactors=2,rotate="oblimin",fm="alpha",oblique.scores=TRUE)
#report_efa(model=model,df=mtcars)

```

report_factorial_anova

Plot means with standard error for every level in a dataframe

Description

Plot means with standard error for every level in a dataframe

Usage

```
report_factorial_anova(
  df,
  dv,
  wid,
  within = NULL,
  within_full = NULL,
  between = NULL,
  within_covariates = NULL,
  between_covariates = NULL,
  observed = NULL,
  diff = NULL,
  reverse_diff = FALSE,
  type = 3,
  white.adjust = TRUE,
  detailed = TRUE,
  return_aov = TRUE,
  file = NULL,
  post_hoc_test = TRUE,
  base_size = 15
)
```

Arguments

df	dataframe
dv	names of dependent variables
wid	names of
within	names of within factors
within_full	names of within factors after data are collapsed to means per condition
between	names of between factors
within_covariates	names of within covariates
between_covariates	names of between covariates
observed	names in data that are already specified in either within or between that contain predictor variables that are observed variables (not manipulated)
diff	names of variables to collapse in a different score
reverse_diff	If TRUE, triggers reversal of the difference collapse requested by diff
type	sum of squares 1 2 3
white.adjust	if TRUE corrects for heteroscedasticity
detailed	if TRUE returns detailed information
return_aov	if TRUE returns aov object
file	output filename
post_hoc_test	if TRUE outputs post hoc in file
base_size	base font size

Examples

```

set.seed(12345)
df<-data.frame(id=rep(seq(1,80),each=81,1),
               IV1=rep(LETTERS[1:3],each=1,2160),
               IV2=rep(LETTERS[4:6],each=3,720),
               IV3=rep(LETTERS[7:9],each=9,240),
               IV4=rep(LETTERS[10:12],each=27,80),
               stringsAsFactors=FALSE)
cdf<-data.frame(matrix(.01,ncol=4,nrow=4))
correlation_martix<-as.matrix(cdf)
diag(correlation_martix)<-1
cdf<-generate_correlation_matrix(correlation_martix,nrows=nrow(df))+10
names(cdf)<-paste0("DV",1:4)
df<-data.frame(df,cdf)
df$DV2<-df$DV2+10
df$DV3<-df$DV3+20
df$DV4<-df$DV4+30
df[df$IV1=="A",]$DV1<-df[df$IV1=="A",]$DV1+1
df[df$IV1=="B",]$DV1<-df[df$IV1=="B",]$DV1+2
df[df$IV1=="C",]$DV1<-df[df$IV1=="C",]$DV1+3
cdf(df)
r1<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=c("IV1","IV2"),within_full=c("IV1","IV2"),
                           between=NULL,
                           within_covariates=NULL,between_covariates=NULL,
                           file="anova_within",
                           post_hoc=TRUE)
r2<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=NULL,within_full=NULL,
                           between=c("IV1","IV2"),
                           within_covariates=NULL,between_covariates=NULL,
                           file="anova_between",
                           post_hoc=TRUE)
r3<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=c("IV3","IV4"),within_full=c("IV3","IV4"),
                           between=c("IV1","IV2"),
                           within_covariates=NULL,between_covariates=NULL,
                           file="anova_mixed",
                           post_hoc=FALSE)
r4<-report_factorial_anova(df=df,wid="id",dv=c("DV1","DV2"),
                           within=c("IV1","IV2"),within_full=c("IV1","IV2"),
                           between=NULL,
                           within_covariates=c("DV3","DV4"),between_covariates=NULL,
                           file="anova_within_cov",
                           post_hoc=TRUE)

```

Description

Report HLR

Usage

```
report_hlr(  
  df,  
  corlist,  
  factorlist,  
  predictor,  
  random_effect,  
  file = NULL,  
  sheet = "report"  
)
```

Arguments

df	dataframe
corlist	Numeric outcome index
factorlist	Numeric predictor index
predictor	Character predictor name
random_effect	Character random effect name
file	Character file
sheet	Character sheet

Examples

```
# report_hlr(df=infert,corlist=8,factorlist=1,  
#           predictor="case",random_effect="case")
```

report_irt	<i>Output for irt model</i>
------------	-----------------------------

Description

Output for irt model

Usage

```
report_irt(model, m2 = TRUE, file = NULL)
```

Arguments

model	object mirt
m2	if TRUE report m2 statistics
file	output filename

Examples

```

set.seed(12345)
cormatrix<-psych::sim.rasch(nvar=5,n=50000,low=-4,high=4,d=NULL,a=1,mu=0,sd=1)$items
irt_onefactor<-mirt::mirt(cormatrix,1,empiricalhist=TRUE,calcNull=TRUE)
irt_twofactor<-mirt::mirt(cormatrix,2,empiricalhist=TRUE,calcNull=TRUE)
irt_threefactor<-mirt::mirt(cormatrix,3,empiricalhist=TRUE,calcNull=TRUE)
report_irt(model=irt_onefactor,file="one_factor")
report_irt(model=irt_twofactor,file="two_factors")
report_irt(model=irt_threefactor,file="three_factors")

```

report_lda	<i>Report for MASS::lda</i>
------------	-----------------------------

Description

Report for MASS::lda

Usage

```
report_lda(model, file = NULL, w = 10, h = 10, base_size = 10, title = "")
```

Arguments

model	object from MASS::lda
file	output filename
w	width of pdf file
h	height of pdf file
base_size	base font size
title	plot title

Examples

```

model<-MASS::lda(case~.,data=infert)
report_lda(model=model)
report_lda(model=model,file="lda")
model<-MASS::lda(Species~.,data=iris)
report_lda(model=model,file="lda")

```

report_logistic	<i>Report logistic regression</i>
-----------------	-----------------------------------

Description

Report logistic regression

Usage

```
report_logistic(
  model,
  validation_data = NULL,
  file = NULL,
  title = "",
  w = 10,
  h = 10,
  base_size = 10,
  fast = FALSE
)
```

Arguments

model	object glm
validation_data	validation data
file	output filename
title	plot title
w	width of pdf file. Relevant only when file string is not empty
h	height of pdf file. Relevant only when file string is not empty
base_size	base font size
fast	if TRUE it will not output individual scores and residuals

Note

(1) Problematic values for standardized residuals $> \pm 1.96$

Standardized residuals are residuals divided by an estimated standard deviation and they can be interpreted as z scores in that:

95 99 99.99 (2) Problematic values for dfBeta ≥ 1

dfBeta estimates coefficients if the respective case is removed from the dataset

(3) Problematic values for Hat values (leverage) 2 or 3 times the average $(k+1)/n$

Hat values (leverage), gauge the influence of the observed value of the outcome variable over the predicted values

The average leverage value is defined as $(k+1)/n$, k =number of predictors, n =number of participants.

Leverage values lie between 0 (no influence) and 1 (complete influence over prediction)

If no cases exert undue influence over the model then all leverage values should be close to $(k+1)/n$

Hoaglin and Welsch (1978) recommends investigating cases with values greater than twice the average $(2(k+1)/n)$

Stevens (2002) recommends investigating cases with values greater than three times the average $(3(k+1)/n)$

(4) Problematic values for VIFs > 10

ASSUMPTIONS

(1) Linearity between continuous predictors and the logit (test whether the interaction term between the predictor and its log transformation is significant)

(2) Independence of errors

(3) No multicollinearity

Examples

```
modelcategoricalpredictor0<-glm(case~education,data=infert,family=binomial)
modelcategoricalpredictor1<-glm(case~education,data=infert,family=gaussian)
#modelcategoricalpredictor2<-glm(case~education,data=infert,family=Gamma)
#modelcategoricalpredictor3<-glm(case~education,data=infert,family=inverse.gaussian)
modelcategoricalpredictor4<-glm(case~education,data=infert,family=poisson)
modelcategoricalpredictor5<-glm(case~education,data=infert,family=quasi)
modelcategoricalpredictor6<-glm(case~education,data=infert,family=quasibinomial)
modelcategoricalpredictor7<-glm(case~education,data=infert,family=quasipoisson)
modelcontinuouspredictor0<-glm(case~stratum,data=infert,family=binomial)
modeltwopredictors0<-glm(case~education+stratum,data=infert,family=binomial)
modeltwopredictors1<-glm(case~education+stratum,data=infert,family=gaussian)
#modeltwopredictors2<-glm(case~education+stratum,data=infert,family=Gamma)
#modeltwopredictors3<-glm(case~education+stratum,data=infert,family=inverse.gaussian)
modeltwopredictors4<-glm(case~education+stratum,data=infert,family=poisson)
modeltwopredictors5<-glm(case~education+stratum,data=infert,family=quasi)
modeltwopredictors6<-glm(case~education+stratum,data=infert,family=quasibinomial)
modeltwopredictors7<-glm(case~education+stratum,data=infert,family=quasipoisson)
report_logistic(model=modelcategoricalpredictor0)
report_logistic(model=modelcategoricalpredictor1)
#report_logistic(model=modelcategoricalpredictor2)
#report_logistic(model=modelcategoricalpredictor3)
report_logistic(model=modelcategoricalpredictor4)
report_logistic(model=modelcategoricalpredictor5)
report_logistic(model=modelcategoricalpredictor6)
report_logistic(model=modelcategoricalpredictor7)
report_logistic(model=modelcontinuouspredictor0)
report_logistic(model=modeltwopredictors0)
report_logistic(model=modelcategoricalpredictor0,
                 file="logistic_categorical_predictor",
                 validation_data=infert)
report_logistic(model=modelcontinuouspredictor0,
                 file="logistic_continuous_predictor",
                 validation_data=infert)
report_logistic(model=modeltwopredictors0,
                 file="logistic_two_predictors",
                 validation_data=infert[1:10,])
```

report_manova	<i>Manova result</i>
---------------	----------------------

Description

Manova result

Usage

```
report_manova(model, file = NULL)
```

Arguments

model	object of manova model
file	output filename

Note

Pillai-Bartlett trace (V): Represents the sum of the proportion of explained variance on the discriminant functions. As such, it is similar to the ratio of SS_M / SS_T , which is known as R^2 .

Hotelling's T^2 : Represents the sum of the eigenvalues for each variate it compares directly to the F-ratio in ANOVA

Wilks's lambda (L): Represents the ratio of error variance to total variance (SS_R / SS_T) for each variate.

Roy's largest root: Represents the proportion of explained variance to unexplained variance (SS_M / SS_R) for the first discriminant function.

ASSUMPTIONS

Independence: Observations should be statistically independent.

Random sampling: Data should be randomly sampled from the population of interest and measured at an interval level.

Multivariate normality: In ANOVA, we assume that our dependent variable is normally distributed within each group. In the case of MANOVA, we assume that the dependent variables (collectively) have multivariate normality within groups.

Homogeneity of covariance matrices: In ANOVA, it is assumed that the variances in each group are roughly equal (homogeneity of variance). In MANOVA we must assume that this is true for each dependent variable, but also that the correlation between any two dependent variables is the same in all groups. This assumption is examined by testing whether the population variance-covariance matrices of the different groups in the analysis are equal.

Examples

```
## Set orthogonal contrasts.
op<-options(contrasts=c("contr.helmert", "contr.poly"))
model_mixed<-manova(cbind(yield, foo)~N*P*K, within(npk, foo<-rnorm(24)))
model_between<-manova(cbind(rnorm(24), rnorm(24))~round(rnorm(24), 0)*round(rnorm(24), 0))
report_manova(model=model_mixed)
report_manova(model=model_between)
```

report_normality_tests	<i>Normality tests</i>
------------------------	------------------------

Description

Shapiro-Wilk Anderson-Darling Cramer-von-Mises Shapiro-Francia Jarque-Bera Kolmogorov-Smirnov
Lilliefors Pearson X2

Usage

```
report_normality_tests(df, file = NULL)
```

Arguments

df	dataframe with continous or ordinal data
file	output filename

Details

returns xlsx file

Examples

```
vector<-generate_missing(rnorm(1000))  
df<-generate_missing(mtcars[,1:2])  
report_normality_tests(df=df)  
report_normality_tests(df=vector,file="normality_tests")
```

report_oneway	<i>One way</i>
---------------	----------------

Description

One way

Usage

```
report_oneway(  
  df,  
  dv,  
  iv,  
  file = NULL,  
  w = 10,  
  h = 10,  
  base_size = 10,
```

```

    note = "",
    title = "",
    type = "ci",
    plot_means = FALSE,
    plot_diagnostics = FALSE
)

```

Arguments

df	dataframe
dv	index of continous variables
iv	index of factors
file	output filename
w	width of pdf file
h	height of pdf file
base_size	base font size
note	text for footnote
title	plot title
type	type of bar to display "se" "ci" "sd" ""
plot_means	if TRUE it will output mean plots and descriptives for plots
plot_diagnostics	if TRUE it will output ANOVA diagnostics plots

Note

- (1) The Fisher procedure assumes heteroscedasticity
- (2) The Welch procedure does not assume heteroscedasticity
- (3) The Kruskal Wallis procedure does not assume normality but it is not an alternative for violations of heteroscedasticity
- (4) Posthoc Tuckey: not good for unequal sample sizes or heteroscedasticity
- (5) Posthoc Games Howell: good for unequal sample sizes and heteroscedasticity

Examples

```

report_oneway(df=mtcars,dv=2:4,iv=9:10,file="anova",
              plot_diagnostics=FALSE,plot_means=FALSE)
report_oneway(df=mtcars,dv=2:4,iv=9:10,file="anova_oneway_two_factor")
report_oneway(df=mtcars,dv=2:4,iv=9,file="anova_oneway_one_factor")
report_oneway(df=mtcars,dv=2:4,iv=9,file="anova_oneway_one_factor",
              plot_means=TRUE,plot_diagnostics=TRUE)

```

report_pdf

*Report pdf***Description**

Report pdf

Usage

```
report_pdf(
  ...,
  plotlist = NULL,
  file = NULL,
  title = NULL,
  w = 10,
  h = 10,
  print_plot = TRUE
)
```

Arguments

...	plot objects
plotlist	list of plot objects
file	output filename
title	output filename
w	width of pdf file
h	height of pdf file
print_plot	if TRUE it prints plot on graphics device

Examples

```
p1<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet,group=Chick))+
  geom_line()+
  ggtitle("Growth curve for individual chicks")+
  theme_bw()
p2<-ggplot(ChickWeight,aes(x=Time,y=weight,colour=Diet))+
  geom_point(alpha=.3)+
  geom_smooth(alpha=.2,size=1,method="loess",formula="y~x")+
  ggtitle("Fitted growth curve per diet")+theme_bw()
cars_plot_multiplot<-plot_multiplot(plotlist=plot_histogram(mtcars[,1:4]),cols=2)
cars_plot_base<-plot_normality_diagnostics(mtcars)
report_pdf(p1,p2,print_plot=TRUE)
report_pdf(p1,p2,file="report",print_plot=FALSE)
report_pdf(plotlist=cars_plot_multiplot,print_plot=TRUE)
report_pdf(plotlist=cars_plot_multiplot,file="report",print_plot=FALSE)
report_pdf(plotlist=cars_plot_base,print_plot=TRUE)
report_pdf(plotlist=cars_plot_base,file="report",print_plot=FALSE)
```

report_regression	<i>Regression</i>
-------------------	-------------------

Description

Regression

Usage

```
report_regression(
  model,
  base_size = 10,
  title = "",
  file = NULL,
  w = 10,
  h = 10,
  plot_diagnostics = TRUE
)
```

Arguments

model	object ml
base_size	base font size
title	plot title
file	output filename
w	width of pdf file. Relevant only when file string is not empty
h	height of pdf file. Relevant only when file string is not empty
plot_diagnostics	if TRUE it will output linear model diagnostics plots

Note

(1) Problematic values for standardized residuals $> \pm 1.96$
 Standardized residuals are residuals divided by an estimated standard deviation and they can be interpreted as z scores in that:
 95.00 99.00 99.99 (2) Studentized residuals indicate the the ability of the model to predict that case. They follow a t distribution
 (3) DFFits indicate the difference between the adjusted predicted value and the original predicted value. Adjusted predicted value for a case refers to the predicted value of that case, when that case is excluded from model fit.
 (4) Cook's distance indicates leverage. Problematic values for cook's distance > 1 Cook and Weisberg (1982).
 (5) Hat values indicate leverage. Problematic values for Hat values 2 or 3 times the average $(k+1/n)$
 The average leverage value is defined as $(k+1)/n$, k =number of predictors, n =number of participants. Leverage values lie between 0 (no influence) and 1 (complete influence over prediction)

Hoaglin and Welsch (1978) recommends investigating cases with values greater than twice the average $2(k+1)/n$

Stevens (2002) recommends investigating cases with values greater than three times the average $3(k+1)/n$

T-tests test the hypothesis that b's are different from 0

Multiple R^2 : Variance Explained

Adjusted R^2 : Indicates how much variance in Y would be accounted for if the model is derived from the population from which the sample was taken. Ideally, $R^2 = \text{Adjusted } R^2$

F-Statistic tests the null hypothesis is that the overall model has no effect

Covariance ratios critical values $CVR > 1 + [3(k+1)/n]$ $CRV < 1 - [3(k+1)/n]$. In general we should obtain small values or we may have to remove cases ASSUMPTIONS

(1) variable types: All predictors must be quantitative or categorical (with two levels), and the outcome variable must be quantitative (interval data), continuous and unbounded (no constraints on the variability of the outcome) (2) Non-zero variance

(3) No perfect multicollinearity

(4) Predictors are uncorrelated with -external variables-

(5) Homoscedasticity: At each level of the predictor variable(s), the variance of the residual terms should be constant. Residuals at each level of the predictor(s) should have similar variance (homoscedasticity)

(6) Independent errors: For any two observations the residual terms should be uncorrelated (or independent) This eventuality is sometimes described as a lack of autocorrelation. This assumption can be tested with the Durbin-Watson test, which tests for serial correlations between errors. Specifically, it tests whether adjacent residuals are correlated The size of the Durbin-Watson statistic depends upon the number of predictors in the model and the number of observations As a very conservative rule of thumb, values less than 1 or greater than 3 are definitely cause for concern; however, values closer to 2 may still be problematic depending on your sample and model R also provides a p-value of the autocorrelation. Be very careful with the Durbin-Watson test, though, as it depends on the order of the data: if you reorder your data, you'll get a different value

(7) Normally distributed errors: It is assumed that the residuals in the model are random, normally distributed variables with a mean of 0

(8) Independence: It is assumed that all of the values of the outcome variable are independent (in other words, each value of the outcome variable comes from a separate entity)

(9) Linearity: The mean values of the outcome variable for each increment of the predictor(s) lie along a straight line

Examples

```
form<-formula(mpg~qsec)
regressionmodel<-lm(form,data=mtcars)
multipleregressionmodel<-lm(mpg~qsec*hp*wt*drat,data=mtcars)
res<-report_regression(model=regressionmodel,plot_diagnostics=TRUE)
res<-report_regression(model=multipleregressionmodel)
res<-report_regression(model=regressionmodel,file="regression")
res<-report_regression(model=multipleregressionmodel,file="regression",plot_diagnostics=TRUE)
```

Description

T test

Usage

```
report_ttests(df, dv, iv, file = NULL, ...)
```

Arguments

- df dataframe
- dv index of continous variables
- iv index of factors
- file output filename
- ... Arguments passed on to `stats::t.test`
 x a (non-empty) numeric vector of data values.

Examples

```
report_ttests(df=mtcars,dv=2,iv=9)
report_ttests(df=mtcars,dv=2,iv=9:10)
report_ttests(df=mtcars,dv=2:3,iv=9)
report_ttests(df=mtcars,dv=2:3,iv=9:10,alternative="two.sided")
report_ttests(df=mtcars,dv=2:7,iv=9:10,alternative="less")
report_ttests(df=mtcars,dv=2:7,iv=9:10,alternative="greater")
report_ttests(df=mtcars,dv=2:7,iv=9:10,paired=FALSE)
#report_ttests(df=mtcars,dv=2:7,iv=9:10,paired=TRUE)
report_ttests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE)
report_ttests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE,file="ttest")
```

report_wtests	<i>Wilcoxon test</i>
---------------	----------------------

Description

Wilcoxon test

Usage

```
report_wtests(df, dv, iv, file = NULL, ...)
```


Arguments

df	dataframe
dv	index of continuous variables
iv	index of factors
file	output filename
...	Arguments passed on to <code>stats::wilcox.test</code>
	x numeric vector of data values. Non-finite (e.g., infinite or missing) values will be omitted.

Examples

```

report_wtests(df=mtcars,dv=2,iv=9)
report_wtests(df=mtcars,dv=2,iv=9:10)
report_wtests(df=mtcars,dv=2:3,iv=9)
report_wtests(df=mtcars,dv=2:3,iv=9:10,alternative="two.sided")
report_wtests(df=mtcars,dv=2:7,iv=9:10,alternative="less")
report_wtests(df=mtcars,dv=2:7,iv=9:10,alternative="greater")
report_wtests(df=mtcars,dv=2:7,iv=9:10,paired=FALSE)
#report_wtests(df=mtcars,dv=2:7,iv=9:10,paired=TRUE)
report_wtests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE)
report_wtests(df=mtcars,dv=1:7,iv=8:10,var.equal=TRUE,file="wilcoxontest")

```

report_xgboost	<i>Report for xgboost::xgb.train</i>
----------------	--------------------------------------

Description

Report for xgboost::xgb.train

Usage

```

report_xgboost(
  model,
  validation_data = NULL,
  label = NULL,
  file = "xgboost",
  w = 10,
  h = 10,
  base_size = 10,
  title = "",
  fast = FALSE
)

```

Arguments

model	object from xgboost::xgb.train
validation_data	validation data
label	outcome variable name
file	output filename
w	width of pdf file
h	height of pdf file
base_size	base font size
title	plot title
fast	if TRUE error values are not saved in output

Examples

```
infert_formula<-as.formula(factor(case)~age+parity+education+spontaneous+induced)
boston_formula<-as.formula(c("medv~",paste(names(MASS::Boston)[1:13],collapse="+")))
train_test_classification<-psycholaterfunctions::k_fold(df=infert,model_formula=infert_formula)
train_test_regression<-psycholaterfunctions::k_fold(df=MASS::Boston,model_formula=boston_formula)
xgb_classification<-xgboost::xgb.train(data=train_test_classification$xgb$f1$train,
                                     watchlist=train_test_classification$xgb$f1$watchlist,
                                     eta=.1,
                                     nthread=8,
                                     nround=20,
                                     objective="binary:logistic")
xgb_regression<-xgboost::xgb.train(data=train_test_regression$xgb$f1$train,
                                   watchlist=train_test_regression$xgb$f1$watchlist,
                                   eta=.3,
                                   nthread=8,
                                   nround=20)
report_xgboost(model=xgb_classification,
               validation_data=train_test_classification$f$test$f1,
               label=train_test_classification$outcome,
               file="Classification")
report_xgboost(model=xgb_regression,
               validation_data=train_test_regression$f$test$f1,
               label=train_test_regression$outcome,
               file="Regression")
```

response_dimension	<i>index parameter and items relative to their dimensions</i>
--------------------	---

Description

index parameter and items relative to their dimensions

Usage

```
response_dimension(response, dimensions, items)
```

Arguments

response	vector one to number of items
dimensions	number of dimensions
items	item comparisons

Examples

```
response_dimension(c(1:18),3,c(1,2))
response_dimension(c(1:18),3,c(1,3))
response_dimension(c(1:18),3,c(2,3))
```

response_frequency	<i>Response frequencies</i>
--------------------	-----------------------------

Description

returns count proportion percent

Usage

```
response_frequency(
  df,
  max = 10,
  uniqueitems = NULL,
  type = "percent",
  file = NULL
)
```

Arguments

df	dataframe
max	maximum score
uniqueitems	number of unique items
type	"frequency" "proportion" "percent" "all"
file	output filename

Details

returns dataframe

Examples

```

response_frequency(mtcars[,c("gear","carb")],uniqueitems=1:8,type="frequency")
response_frequency(mtcars[,c("gear")],uniqueitems=1:8,type="proportion")
response_frequency(mtcars[,c("gear","carb")],uniqueitems=1:8,type="percent")
response_frequency(mtcars[,c("gear","carb")],uniqueitems=1:8,type="all")
response_frequency(mtcars[,c("gear","carb")],uniqueitems=1:8,type="all",
                  file="descriptives")

```

result_confusion_performance

Plot performance of confusion matrix for different cut off points

Description

Plot performance of confusion matrix for different cut off points

Usage

```

result_confusion_performance(
  observed,
  predicted,
  step = 0.1,
  base_size = 10,
  title = ""
)

```

Arguments

observed	vector of observed outcomes
predicted	vector of predicted outcome probability
step	stepping for tested cut values
base_size	base font size
title	plot title

Examples

```

df<-data.frame(matrix(.999,ncol=2,nrow=2))
correlation_martix<-as.matrix(df)
diag(correlation_martix)<-1
df<-generate_correlation_matrix(correlation_martix,nrows=1000)
df$X1<-ifelse(abs(df$X1)<1,0,1)
df$X2<-abs(df$X2)
df$X2<-(df$X2-min(df$X2))/(max(df$X2)-min(df$X2))
result_confusion_performance(observed=round(abs(df$X1),0),
                             predicted=abs(df$X2),
                             step=.01)
result_confusion_performance(observed=c(1,2,3,1,2,3),predicted=abs(rnorm(6,0,sd=.1)))

```

round_dataframe	<i>Round dataframe</i>
-----------------	------------------------

Description

It only processes numeric values in a dataframe

Usage

```
round_dataframe(df, digits = 0, type = "round")
```

Arguments

df	dataframe
digits	decimal points to return. It works only with "round" type
type	"round" "ceiling" "floor" "tenth"

Examples

```
round_dataframe(df=change_data_type(df=mtcars,type="factor"),digits=0)
round_dataframe(df=change_data_type(df=mtcars,type="character"),digits=0)
round_dataframe(df=mtcars,digits=0)
round_dataframe(df=mtcars,digits=0,type="ceiling")
round_dataframe(df=mtcars,digits=0,type="floor")
round_dataframe(df=mtcars*100,digits=2,type="tenth")
```

shrout	<i>Shrout reliability</i>
--------	---------------------------

Description

Shrout reliability

Usage

```
shrout(sperson, spersonitem, stime, spersontime, serror, m, k)
```

Arguments

sperson	variance component of participant
spersonitem	variance component of participant by item interaction
stime	variance component of time
spersontime	variance component of participant by time interaction
serror	variance component of error
m	m item reports
k	k time points

Examples

```
design<-expand.grid(time=1:3,item=1:2,person=1:10)
design<-change_data_type(design,type="factor")
design$response<-rnorm(30,0,0.1)
model<-mixlm::lm(response~r(time)*r(person)+r(item)*r(person),data=design)
result<-extract_components(model)
vc<-result$components
shrout(spersion=vc[2,3],spersionitem=vc[5,3],stime=vc[1,3],
      spersiontime=vc[4,3],serror=vc[6,3],3,3)
```

simulate_cfa_fit	<i>Simulate CFA from coefficients</i>
------------------	---------------------------------------

Description

Simulates cfa from coefficients Simulates cfa from correlations of obeserved data Returns fit indices for predefined set of sample sizes

Usage

```
simulate_cfa_fit(
  model_sim = NULL,
  model = NULL,
  df = NULL,
  minnobs = 50,
  maxnobs = 1000,
  stepping = 10,
  file = NULL,
  w = 10,
  h = 10
)
```

Arguments

model_sim	lavaan model spesification with defined coefficients
model	lavaan model spesification with free coefficients
df	dataframe
minnobs	start sample size
maxnobs	end sample size
stepping	stepping
file	output filename
w	width of pdf file
h	height of pdf file

Examples

```

model_sim='LATENT=~1*X1+0.5*X2+1.5*X3+1.5*X4+X5'
model='LATENT=~X1+X2+X3+X4+X5'
df<-lavaan::simulateData(model=model_sim,model.type="cfa",
                          return.type="data.frame",sample.nobs=1000)
# simulate_cfa_fit(model_sim=model_sim,model=model,
#                  minnobs=50,maxnobs=1000,stepping=100,file="report")
# simulate_cfa_fit(model=model,df=df,
#                  minnobs=50,maxnobs=1000,stepping=100,file="report")

```

```
simulate_correlation_from_sample
```

Generate a dataframe that produces the same correlation matrix as the input dataframe

Description

Generate a dataframe that produces the same correlation matrix as the input dataframe

Usage

```
simulate_correlation_from_sample(cordata, nrows = 10)
```

Arguments

cordata	dataframe
nrows	number of rows to generate

Examples

```

correlation_matrix<-generate_correlation_matrix()
stats::cor(correlation_matrix)
simulate_correlation_from_sample(correlation_matrix,nrows=1000)
stats::cor(simulate_correlation_from_sample(correlation_matrix,nrows=1000))

```

split_str	<i>Split string to dataframe</i>
-----------	----------------------------------

Description

Split string to dataframe

Usage

```
split_str(vector, split = "/", include_original = FALSE)
```

Arguments

vector String
split Separation character
include_original if TRUE it will return the input on a separate column

Examples

```
string<-paste0(1:10,"/",  
               generate_string(nchar=2,vector_length=10),"/",  
               generate_string(nchar=2,vector_length=10),"/",  
               generate_string(nchar=2,vector_length=10))  
split_str(string,split="/")
```

split_str_df	<i>Split string in dataframe</i>
--------------	----------------------------------

Description

Split string in dataframe

Usage

```
split_str_df(df, split = "/", type = "row", index, ...)
```

Arguments

df dataframe
split Separation character
type "row" "column" if "row" it will split the string of row names and it will display it on seperate columns if "column" it will split the string of a spesified column and it will display it on separate collumns
index Numeric index of column to split. This is only relevant if type="column"
... arguments passed to split_str

Examples

```
df<-generate_correlation_matrix()  
string<-paste0(1:nrow(df),"/",  
               generate_string(nchar=2,vector_length=nrow(df)),"/",  
               generate_string(nchar=2,vector_length=nrow(df)),"/",  
               generate_string(nchar=2,vector_length=nrow(df)))  
row.names(df)<-string  
split_str_df(df,split="/",type="row")  
df[,1]<-string  
split_str_df(df,split="/",type="column",index=1)
```

string_aes	<i>Adjust string aesthetics</i>
------------	---------------------------------

Description

Treats spesific characters such as ".", as separating characters and separates strings with space.
Trims leading and trailing spaces and capitalizes the first letter of the string and lowers the rest.

Usage

```
string_aes(  
  vector,  
  characterlist = c(".", "_", "-", " ", "$", "<p>", "</p>", "<br>", "<br/>", "<B>",  
    "</B>", "<BR/>", "|", "/", "&nbsp;"),  
  proper = TRUE  
)
```

Arguments

- vector Vector
- characterlist List the list of characters to treat as separating characters
- proper Logical TRUE capitalizes the first letter in sentence format

Examples

```
vector<-c("TES.T", "TES<p>T", "TES&nbsp;T")  
string_aes(vector=vector)  
string_aes(vector=vector,proper=FALSE)  
string_aes(vector=vector,proper=TRUE)
```

sub_str	<i>Return n characters from left or right</i>
---------	---

Description

Return n characters from left or right

Usage

```
sub_str(x, n = 2, type)
```

Arguments

- x Character
- n Number of characters to return
- type "right" "left"

Examples

```
sub_str("12345",n=2,type="right")
sub_str("12345",n=2,type="left")
```

swap	<i>Reverse a numeric vector</i>
------	---------------------------------

Description

Reverse a numeric vector

Usage

```
swap(vector)
```

Arguments

vector numeric

Examples

```
swap(c(1:10,1,2,3))
```

symmetric_matrix	<i>Symmetric Matrix</i>
------------------	-------------------------

Description

Symmetric Matrix

Usage

```
symmetric_matrix(matrix, duplicate = "lower", diagonal = NULL)
```

Arguments

matrix matrix
duplicate "upper" duplicates upper triangle "lower" duplicates lower triangle
diagonal diagonal values

Examples

```
m_lower<-matrix_triangle(matrix(1:9,nrow=3,ncol=3),type="lower",diagonal=NA)
m_upper<-matrix_triangle(matrix(11:19,nrow=3,ncol=3),type="upper",diagonal=NA)
symmetric_matrix(matrix=m_lower,duplicate="lower",diagonal=NA)
symmetric_matrix(matrix=m_upper,duplicate="upper",diagonal=NA)
```

tag_pos	<i>Part of speech tagging</i>
---------	-------------------------------

Description

Part of speech tagging

Usage

```
tag_pos(text)
```

Arguments

text	character vector
------	------------------

Examples

```
text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
tag_pos(text)
```

text_similarity	<i>Text similarity measures</i>
-----------------	---------------------------------

Description

Text similarity measures

Usage

```
text_similarity(text1, text2)
```

Arguments

text1	character vector
text2	character vector

Examples

```

text1<-"word_one word_two word_three"
text2<-"word_three word_four word_six"
text3<-"All the Lorem Ipsum generators on the Internet tend to repeat predefined
chunks as necessary, making this the first true generator on the Internet."
text4<-"It uses a dictionary of over 200 Latin words, combined with a handful of
model sentence structures, to generate Lorem Ipsum which looks reasonable."
text5<-"The generated Lorem Ipsum is therefore always free from repetition,
injected humour, or non-characteristic words etc."
text<-c(text1,text2,text3,text4,text5)
text<-unlist(strsplit(text,split=" "))
text1<-unlist(strsplit(text1,split=" "))
text2<-unlist(strsplit(text2,split=" "))
text3<-unlist(strsplit(text3,split=" "))
text4<-unlist(strsplit(text4,split=" "))
text5<-unlist(strsplit(text5,split=" "))
text_similarity(text1,text1)
text_similarity(text1,text2)
text_similarity(text1,text3)
text_similarity(text1,text4)

```

trim_df

Trim whitespace in dataframe

Description

Trim whitespace in dataframe

Usage

```
trim_df(df)
```

Arguments

df dataframe

Examples

```

string<-data.frame(str1=rep(paste0(sample(c(LETTERS,rep(" ",10))),collapse=""),10),
                    str2=rep(paste0(sample(c(LETTERS,rep(" ",10))),collapse=""),10),
                    num1=rnorm(10),
                    stringsAsFactors=FALSE)
trim_df(string)

```

ts_smoothing	<i>Smoothing</i>
--------------	------------------

Description

smoothing for timeseries. uses base plot

Usage

```
ts_smoothing(  
  df,  
  start = 0.01,  
  stop = 2,  
  step = 0.001,  
  title = "",  
  type = "kernel"  
)
```

Arguments

df	ts object
start	start value
stop	stop value
step	step
title	plot title
type	"default" "kernel" "lowess" "friedman" "splines" "polynomial" "linear"

Details

returns plot

Examples

```
ts_data<-ts(UKDriverDeaths,start=1969,end=1984,frequency=12)  
par(mfrow=c(2,2))  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="default")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="polynomial")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="linear")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="kernel")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="lowess")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="friedman")  
ts_smoothing(ts_data,start=.01,stop=2,step=.01,title="Driver Deaths in UK",type="splines")
```

wrapper	<i>Wrap string</i>
---------	--------------------

Description

Wrap string

Usage

```
wrapper(x, ...)
```

Arguments

x	title
...	arguments passed to strwrap

Examples

```
wrapper(rep("sting",50),30)
```

write_txt	<i>Log console in file</i>
-----------	----------------------------

Description

Logs console in file and then displays log in console

Usage

```
write_txt(input, file = NULL)
```

Arguments

input	Script to log in log file
file	Filename of log

Examples

```
write_txt(mtcars)
write_txt(mtcars,file="mtcars")
```

Index

* ANOVA

- compute_aov_es, [12](#)
- compute_kruskal_wallis_test, [19](#)
- compute_one_way_test, [21](#)
- compute_posthoc, [22](#)
- plot_interaction, [69](#)
- plot_oneway, [76](#)
- plot_oneway_diagnostics, [77](#)
- report_factorial_anova, [99](#)
- report_manova, [106](#)
- report_oneway, [107](#)
- report_ttests, [111](#)
- report_wtests, [112](#)

* EFA

- compute_residual_stats, [24](#)
- model_loadings, [60](#)
- plot_loadings, [71](#)
- plot_scree, [82](#)
- report_efa, [98](#)

* IRT

- cfa_icc_index, [7](#)
- compute_ability, [10](#)
- compute_dummy_comparisons, [15](#)
- compute_icc_thurstonian, [16](#)
- compute_info_1pl, [17](#)
- compute_info_2pl, [17](#)
- compute_info_3pl, [18](#)
- compute_map, [20](#)
- compute_scores, [24](#)
- compute_scores_parallel, [25](#)
- compute_se_theta, [26](#)
- compute_unidimensional_ability, [28](#)
- compute_unidimensional_theta, [29](#)
- generate_comparisons_matrix, [45](#)
- generate_matrix_A, [47](#)
- generate_matrix_lambda_hat, [48](#)
- generate_unique_comparisons_index, [50](#)
- get_mplus_thu_3t, [51](#)

- icc_cfa, [53](#)
- increase_index, [54](#)
- plot_icc_thurstonian, [68](#)
- plot_irt_onefactor, [70](#)
- rank3_to_triplets, [87](#)
- rank_df_to_binary, [87](#)
- rank_to_binary, [88](#)
- report_irt, [102](#)
- response_dimension, [114](#)

* ML

- report_lda, [103](#)
- report_xgboost, [113](#)

* NLP

- clear_stopwords, [8](#)
- clear_text, [9](#)
- tag_pos, [123](#)
- text_similarity, [123](#)

* SEM

- plot_cfa, [65](#)
- report_cfa, [95](#)
- simulate_cfa_fit, [118](#)

* Thurstonian

- cfa_icc_index, [7](#)
- compute_ability, [10](#)
- compute_dummy_comparisons, [15](#)
- compute_icc_thurstonian, [16](#)
- compute_map, [20](#)
- compute_scores, [24](#)
- compute_scores_parallel, [25](#)
- generate_comparisons_matrix, [45](#)
- generate_matrix_A, [47](#)
- generate_matrix_lambda_hat, [48](#)
- generate_unique_comparisons_index, [50](#)
- get_mplus_thu_3t, [51](#)
- icc_cfa, [53](#)
- increase_index, [54](#)
- plot_icc_thurstonian, [68](#)
- rank3_to_triplets, [87](#)

- rank_df_to_binary, [87](#)
- rank_to_binary, [88](#)
- response_dimension, [114](#)
- * assumptions**
 - outlier_summary, [61](#)
 - plot_boxplot, [64](#)
 - plot_histogram, [67](#)
 - plot_normality_diagnostics, [75](#)
 - plot_outlier, [78](#)
 - plot_qq, [78](#)
 - remove_outliers, [92](#)
 - report_normality_tests, [107](#)
- * check**
 - cdf, [6](#)
- * correlation**
 - compute_power_r, [22](#)
 - compute_power_r_matrix, [23](#)
 - deg2rad, [35](#)
 - plot_corrplot, [66](#)
 - rad2deg, [86](#)
 - report_choric_serial, [95](#)
 - report_correlation, [96](#)
- * dataframe**
 - cdf, [6](#)
- * data**
 - generate_correlation_matrix, [45](#)
 - generate_data, [46](#)
 - generate_factor, [47](#)
 - generate_missing, [48](#)
 - generate_multiple_responce_vector, [49](#)
 - generate_string, [49](#)
 - simulate_correlation_from_sample, [119](#)
- * descriptives**
 - compute_aggregate, [11](#)
 - compute_crosstable, [13](#)
 - compute_descriptives, [14](#)
 - compute_frequencies, [15](#)
 - plot_crosstable, [67](#)
 - plot_mosaic, [73](#)
 - plot_response_frequencies, [79](#)
 - response_frequency, [115](#)
- * dimensions**
 - get_questionnaire_reference, [52](#)
 - get_reversals, [52](#)
 - questions_by_keys, [85](#)
 - questions_dimensions_dataframe, [85](#)
- * functions**
 - c_bind, [33](#)
 - call_to_string, [6](#)
 - cdf, [6](#)
 - change_data_type, [7](#)
 - comparison_combinations, [9](#)
 - compute_adjustment, [11](#)
 - compute_confidence_interval, [13](#)
 - compute_dissatenuation, [14](#)
 - compute_kurtosis, [19](#)
 - compute_skewness, [26](#)
 - compute_standard, [27](#)
 - compute_standard_error, [28](#)
 - confusion, [31](#)
 - confusion_matrix_percent, [31](#)
 - data_frame_index, [33](#)
 - detach_package, [35](#)
 - display_upper_lower_triangle, [36](#)
 - dotnames, [36](#)
 - drop_levels, [37](#)
 - dummy_arrange, [37](#)
 - duplicate_y_axis, [38](#)
 - environment_options, [39](#)
 - excel_confusion_matrix, [39](#)
 - excel_critical_value, [40](#)
 - excel_generic_format, [41](#)
 - excel_matrix, [42](#)
 - flatten_list, [44](#)
 - generate_correlation_matrix, [45](#)
 - generate_data, [46](#)
 - generate_factor, [47](#)
 - generate_missing, [48](#)
 - generate_multiple_responce_vector, [49](#)
 - generate_string, [49](#)
 - get_coding, [51](#)
 - get_questionnaire_reference, [52](#)
 - get_reversals, [52](#)
 - getfwp, [50](#)
 - hinvert_title_grob, [53](#)
 - install_all_packages, [54](#)
 - install_load, [55](#)
 - json2dataframe, [55](#)
 - k_fold, [56](#)
 - k_sample, [57](#)
 - matrix_triangle, [57](#)
 - mgsb, [59](#)
 - min_max_index, [59](#)

- off_diagonal_index, 61
- output_separator, 62
- padNA, 63
- plot_confusion, 66
- plot_multiplot, 74
- plot_roc, 80
- plot_separability, 82
- proper, 84
- proportion_accurate, 84
- questions_by_keys, 85
- questions_dimensions_dataframe, 85
- rbind_all, 89
- recode_dataframe, 90
- recode_scale_dummy, 90
- remove_nc, 91
- remove_user_packages, 93
- replace_na_with_previous, 93
- report_dataframe, 97
- report_pdf, 109
- result_confusion_performance, 116
- round_dataframe, 117
- simulate_correlation_from_sample, 119
- split_str, 119
- split_str_df, 120
- string_aes, 121
- sub_str, 121
- swap, 122
- symmetric_matrix, 122
- trim_df, 124
- wrapper, 126
- write_txt, 126
- * generate**
 - generate_correlation_matrix, 45
 - generate_data, 46
 - generate_factor, 47
 - generate_missing, 48
 - generate_multiple_responce_vector, 49
 - generate_string, 49
 - simulate_correlation_from_sample, 119
- * keys**
 - get_questionnaire_reference, 52
 - get_reversals, 52
 - questions_by_keys, 85
 - questions_dimensions_dataframe, 85
- * logistic**
 - compute_y_logistic, 30
 - output_compare_model_logistic, 62
 - plot_logistic_model, 72
 - report_logistic, 104
- * matrix**
 - display_upper_lower_triangle, 36
 - matrix_triangle, 57
 - off_diagonal_index, 61
 - symmetric_matrix, 122
- * plot**
 - duplicate_y_axis, 38
 - hinvert_title_grob, 53
 - plot_multiplot, 74
 - report_pdf, 109
 - wrapper, 126
- * regression**
 - compute_y_logistic, 30
 - output_compare_model_logistic, 62
 - plot_logistic_model, 72
 - plot_scatterplot, 80
 - report_logistic, 104
 - report_regression, 110
- * reliability**
 - alpha_diagnostics, 5
 - extract_components, 44
 - key_to_cfa_model, 56
 - mean_sd_alpha, 58
 - plot_mtm, 73
 - raw_alpha, 89
 - report_alpha, 94
 - shrout, 117
- * series**
 - compute_moving_average, 21
 - plot_acf, 64
 - plot_ts, 83
 - ts_smoothing, 125
- * statistical**
 - compute_adjustment, 11
 - compute_confidence_interval, 13
 - compute_dissatenuation, 14
 - compute_kurtosis, 19
 - compute_skewness, 26
 - compute_standard, 27
 - compute_standard_error, 28
- * strings**
 - call_to_string, 6
 - mgsb, 59
 - output_separator, 62

- proper, 84
- split_str, 119
- split_str_df, 120
- string_aes, 121
- sub_str, 121
- trim_df, 124
- * **timestamp**
 - convert_excel_unix_timestamp, 32
 - decompose_datetime, 34
- * **time**
 - compute_moving_average, 21
 - plot_acf, 64
 - plot_ts, 83
 - ts_smoothing, 125
- * **unidimensional**
 - compute_info_1pl, 17
 - compute_info_2pl, 17
 - compute_info_3pl, 18
 - compute_se_theta, 26
 - compute_unidimensional_ability, 28
 - compute_unidimensional_theta, 29
- alpha_diagnostics, 5
- c_bind, 33
- call_to_string, 6
- cdf, 6
- cfa_icc_index, 7
- change_data_type, 7
- clear_stopwords, 8
- clear_text, 9
- comparison_combinations, 9
- compute_ability, 10
- compute_adjustment, 11
- compute_aggregate, 11
- compute_aov_es, 12
- compute_confidence_inteval, 13
- compute_crosstable, 13
- compute_descriptives, 14
- compute_dissatenuation, 14
- compute_dummy_comparisons, 15
- compute_frequencies, 15
- compute_icc_thurstonian, 16
- compute_info_1pl, 17
- compute_info_2pl, 17
- compute_info_3pl, 18
- compute_kruskal_wallis_test, 19
- compute_kurtosis, 19
- compute_map, 20
- compute_moving_average, 21
- compute_one_way_test, 21
- compute_posthoc, 22
- compute_power_r, 22
- compute_power_r_matrix, 23
- compute_residual_stats, 24
- compute_scores, 24
- compute_scores_parallel, 25
- compute_se_theta, 26
- compute_skewness, 26
- compute_standard, 27
- compute_standard_error, 28
- compute_unidimensional_ability, 28
- compute_unidimensional_theta, 29
- compute_y_logistic, 30
- confusion, 31
- confusion_matrix_percent, 31
- convert_excel_unix_timestamp, 32
- data_frame_index, 33
- decompose_datetime, 34
- deg2rad, 35
- detach_package, 35
- display_upper_lower_triangle, 36
- dotnames, 36
- drop_levels, 37
- dummy_arrange, 37
- duplicate_y_axis, 38
- environment_options, 39
- excel_confusion_matrix, 39
- excel_critical_value, 40
- excel_generic_format, 41
- excel_matrix, 42
- extract_components, 44
- flatten_list, 44
- generate_comparisons_matrix, 45
- generate_correlation_matrix, 45
- generate_data, 46
- generate_factor, 47
- generate_matrix_A, 47
- generate_matrix_lambda_hat, 48
- generate_missing, 48
- generate_multiple_responce_vector, 49
- generate_string, 49
- generate_unique_comparisons_index, 50
- get_coding, 51

get_mplus_thu_3t, 51
get_questionnaire_reference, 52
get_reversals, 52
getfw, 50

hinvert_title_grob, 53

icc_cfa, 53
increase_index, 54
install_all_packages, 54
install_load, 55

json2dataframe, 55

k_fold, 56
k_sample, 57
key_to_cfa_model, 56

matrix_triangle, 57
mean_sd_alpha, 58
mgsub, 59
min_max_index, 59
model_loadings, 60

off_diagonal_index, 61
outlier_summary, 61
output_compare_model_logistic, 62
output_separator, 62

padNA, 63
plot_acf, 64
plot_boxplot, 64
plot_cfa, 65
plot_confusion, 66
plot_corrplot, 66
plot_crosstable, 67
plot_histogram, 67
plot_icc_thurstonian, 68
plot_interaction, 69
plot_irt_onefactor, 70
plot_loadings, 71
plot_logistic_model, 72
plot_mosaic, 73
plot_mtm, 73
plot_multiplot, 74
plot_normality_diagnostics, 75
plot_oneway, 76
plot_oneway_diagnostics, 77
plot_outlier, 78
plot_qq, 78

plot_response_frequencies, 79
plot_roc, 80
plot_scatterplot, 80
plot_scee, 82
plot_separability, 82
plot_ts, 83
proper, 84
proportion_accurate, 84

questions_by_keys, 85
questions_dimensions_dataframe, 85

rad2deg, 86
rank3_to_triplets, 87
rank_df_to_binary, 87
rank_to_binary, 88
raw_alpha, 89
rbind_all, 89
recode_dataframe, 90
recode_scale_dummy, 90
remove_nc, 91
remove_outliers, 92
remove_user_packages, 93
replace_na_with_previous, 93
report_alpha, 94
report_cfa, 95
report_choric_serial, 95
report_correlation, 96
report_dataframe, 97
report_efa, 98
report_factorial_anova, 99
report_hlr, 101
report_irt, 102
report_lda, 103
report_logistic, 104
report_manova, 106
report_normality_tests, 107
report_oneway, 107
report_pdf, 109
report_regression, 110
report_ttests, 111
report_wtests, 112
report_xgboost, 113
response_dimension, 114
response_frequency, 115
result_confusion_performance, 116
round_dataframe, 117

shrout, 117

`simulate_cfa_fit`, [118](#)
`simulate_correlation_from_sample`, [119](#)
`split_str`, [119](#)
`split_str_df`, [120](#)
`stats::t.test`, [112](#)
`stats::wilcox.test`, [113](#)
`string_aes`, [121](#)
`sub_str`, [121](#)
`swap`, [122](#)
`symmetric_matrix`, [122](#)

`tag_pos`, [123](#)
`text_similarity`, [123](#)
`trim_df`, [124](#)
`ts_smoothing`, [125](#)

`wrapper`, [126](#)
`write_txt`, [126](#)