

<sup>1</sup> Binocular combination in the autonomic nervous system

<sup>2</sup> Federico G. Segala<sup>1\*</sup>, Aurelio Bruno<sup>1,2</sup>, Joel T. Martin<sup>1,3</sup>,  
Anisa Y. Morsi<sup>1</sup>, Alex R. Wade<sup>1,4</sup> & Daniel H. Baker<sup>1,4</sup>

<sup>3</sup> <sup>1</sup>Department of Psychology, University of York, Heslington, York, YO10 5DD, United Kingdom.

<sup>4</sup> <sup>2</sup>School of Psychology and Vision Sciences, University of Leicester, Leicester, LE1 7RH, United Kingdom.

<sup>5</sup> <sup>3</sup>School of Philosophy, Psychology and Language Sciences, University of Edinburgh, Edinburgh, EH8 9AD,  
<sup>6</sup> United Kingdom.

<sup>7</sup> <sup>4</sup>York Biomedical Research Institute, University of York, Heslington, York, YO10 5NG, United Kingdom.

<sup>8</sup> \*Corresponding author

<sup>9</sup> E-mail:[federico.segala@york.ac.uk](mailto:federico.segala@york.ac.uk) (FGS)

## <sup>10</sup> 1 Abstract

<sup>11</sup> Pupil diameters are regulated by the autonomic nervous system, which combines light signals across the eyes  
<sup>12</sup> independently of the visual cortex. Distinct classes of retinal photoreceptor are involved in this process, with  
<sup>13</sup> cones and rods driving the initial constriction and intrinsically photosensitive retinal ganglion cells main-  
<sup>14</sup> taining diameter over prolonged time periods. We investigated binocular combination by targeting different  
<sup>15</sup> photoreceptor pathways using a novel binocular multiprimary system to modulate the input spectra via silent  
<sup>16</sup> substitution. At the first harmonic of the modulation frequency, luminance and S-cone responses showed  
<sup>17</sup> strong binocular facilitation, and weak interocular suppression. Melanopsin responses were invariant to the  
<sup>18</sup> number of eyes stimulated. Notably, the L-M pathway involved binocular inhibition, whereby responses to  
<sup>19</sup> binocular stimulation were weaker than for monocular stimulation. The second harmonic involved strong  
<sup>20</sup> interocular suppression in all pathways, but with some evidence of binocular facilitation. Our results are  
<sup>21</sup> consistent with a computational model of binocular signal combination (implemented in a Bayesian hier-  
<sup>22</sup> archical framework), in which the weight of interocular suppression differs across pathways. We also find  
<sup>23</sup> pathway differences in response phase, consistent with different lag times for phototransduction. This work  
<sup>24</sup> demonstrates for the first time the algorithm governing binocular combination in the autonomic nervous  
<sup>25</sup> system.

## <sup>26</sup> 2 Introduction

<sup>27</sup> The autonomic nervous system regulates many involuntary bodily processes, including the constriction and  
<sup>28</sup> dilation of the pupils in response to light [1]. The anatomical pathway from the retina to the subcortical  
<sup>29</sup> nuclei controlling the pupillary light response (PLR) is well established: it includes the Pretectal Olivary  
<sup>30</sup> nucleus (PON), the Superior Cervical ganglion and the Edinger-Westphal nucleus, which project to the iris  
<sup>31</sup> sphincter muscles that directly control the pupil size [1]. Evidence of a binocular component to the PLR is  
<sup>32</sup> shown by the consensual response of the pupil (stimulation of one eye will cause constriction of the other  
<sup>33</sup> eye) [2]. The anatomical segregation of the subcortical pathway from the rest of the brain means that this  
<sup>34</sup> binocular combination of signals must occur independently of the cortical processes of binocular integration  
<sup>35</sup> required for visual perception. Our recent work [3] has shown that the algorithm underlying binocular  
<sup>36</sup> combination of light in the pupil pathway differs from that in the cortex. Here we extend this paradigm to  
<sup>37</sup> compare binocular combination in different photoreceptor pathways that feed into the autonomic nervous  
<sup>38</sup> system.

<sup>39</sup> Different classes of retinal photoreceptors, including cones, rods and melanopsin-containing intrinsically

40 photosensitive retinal ganglion cells (ipRGCs), are directly involved in controlling and maintaining the size  
41 of the pupils [e.g. 4,5–8]. Cones drive the initial rapid constriction of the pupils [9], while the slower and  
42 longer activation of the ipRGCs maintains constriction over a prolonged period of time and regulates the  
43 post-illumination pupillary response [4,10]. The ipRGCs are a recently discovered photoreceptor class [11]  
44 that express the photopigment melanopsin, and are involved in the regulation of the circadian rhythm [12,13],  
45 forming a major input to the PON [14]. The first direct evidence of the involvement of the ipRGCs in the  
46 PLR was shown in melanopsin knockout rats [15], resulting in the loss of the intrinsic photosensitivity of  
47 the cells and a reduced pupil constriction. Similar behaviour was later observed in primates and humans  
48 [16] using silent substitution (see Methods), where it was demonstrated that the PLR continues during light  
49 presentation even when cone and rod signalling is blocked, indicating the primary role of the ipRGCs in  
50 maintaining pupil constriction over a prolonged time.

51 Binocular combination has been extensively studied in visual perception, where it is mediated by neurons in  
52 primary visual cortex [17]. For pattern vision, binocular summation occurs at threshold, such that a lower  
53 contrast is required to detect a target shown to both eyes than a target shown to one eye [18,19]. At higher  
54 contrasts, the phenomenon of ‘ocularity invariance’ is observed, in which the response to monocularly- and  
55 binocularly-presented patterns is equal [20,21]. This is explained by a process of interocular suppression  
56 that cancels out the additional excitatory drive caused by stimulating two eyes. Our recent work [3] showed  
57 that cortical signal combination for luminance flicker is substantially more linear than for spatial patterns,  
58 whereas there is evidence of interocular suppression in the pupil pathway. Here we measure the amplitude  
59 of pupil modulations in response to flickering stimuli presented as light flux, or directed towards the L-M  
60 cone, S-cone, and melanopsin pathways (see Figure 1e-l). Our key comparisons are between monocular and  
61 binocular stimulation, and in a dichoptic masking condition where the two eyes are stimulated at different  
62 frequencies. The results are interpreted using a contemporary model of binocular vision [22] implemented  
63 within a hierarchical Bayesian framework.

### 64 **3 Materials and methods**

#### 65 **3.1 Participants**

66 Twenty-four participants were recruited for each of the four experiments for a total of ninety-six adult  
67 participants (28 male, 68 female), whose ages ranged from 18 to 41. All participants had normal or corrected  
68 to normal vision, no known abnormalities of binocular or colour vision, and gave written informed consent.  
69 Our procedures were approved by the Ethics Committee of the Department of Psychology at the University

70 of York (identification number 184). Participant recruitment began on 3 March 2023 and concluded on 8  
71 Decemeber 2023.

## 72 3.2 Apparatus & stimuli

73 To present synchronised stimulus modulations independently to each eye, two light engines (Spec-  
74 traTuneLAB, which are thermally stable across time with active cooling according to the manufacturers:  
75 LEDMOTIVE Technologies, LLC, Barcelona, Spain), each with 10 independently addressable LED colour  
76 channels, were integrated into a customised binocular viewing system. The light engines were operated via  
77 a Python interface to their REST API [23], which supports synchronous launch and playback of spectral  
78 sequences prepared in advance and stored in JSON format. A special command was commissioned from  
79 LEDMOTIVE to allow two different stimulus files to be launched simultaneously from the two devices. The  
80 synchronisation of the spectra from the two devices was tested and showed that they were synchronised to  
81 within approximately 3ms.

82 When preparing the spectral sequences, the age of participants was used to account for the yellowing of  
83 the lenses. We used the silent substitution technique [24] to selectively stimulate specific photoreceptor  
84 classes. Silent substitution exploits the fact that each photoreceptor class has a distinct spectral tuning that  
85 overlaps with the others. Using a multiprimary system, in which the primaries (i.e. LEDs) have different  
86 spectra, it is possible to target one class of photoreceptors while maintaining the others at a constant  
87 activity level, effectively silencing them [25,26, this paper also offers a clear explanation of how to implement  
88 silent substitution]. We calculated silent substitution solutions using the *PySilSub* toolbox [27], using linear  
89 algebra. The outputs of the two light engines (see Figure 1b,c) were calibrated using an Ocean Optics  
90 Jaz spectroradiometer, which was wavelength-calibrated to an Argon lamp and intensity calibrated using  
91 a NIST-traceable light source. Each primary was also linearised using a polynomial fit (see Figure S1 for  
92 details). We used the 10-degree cone fundamentals [28], and estimates of melanopsin absorbance spectra  
93 from CIE S 026 (discussed in a previous paper [27]) to calculate  $\alpha$ -opic irradiance.

94 The output from the light engines was directed through liquid light guides (LLG3-8H: Thorlabs Ltd, Cam-  
95 bridgeshire, UK) and diffused onto semi-opaque and highly diffusive white glass discs with a diameter of 50  
96 mm for even illumination (34-473: Edmund Optics, York, UK). The light guide gaskets were butt-coupled  
97 to the light engine diffusers with threaded adapters (SM1A9, AD3LLG: Thorlabs Ltd, Cambridgeshire,  
98 UK) and the exiting ends of the light guides were mated with 51 mm depth optical cylinders (SM2L20:  
99 Thorlabs Ltd, Cambridgeshire, UK) via appropriately threaded adapters (AD3LLG, SM2A6: Thorlabs Ltd,  
100 Cambridgeshire, UK). The stimulus diffuser discs were retained at the front end of the optical cylinders

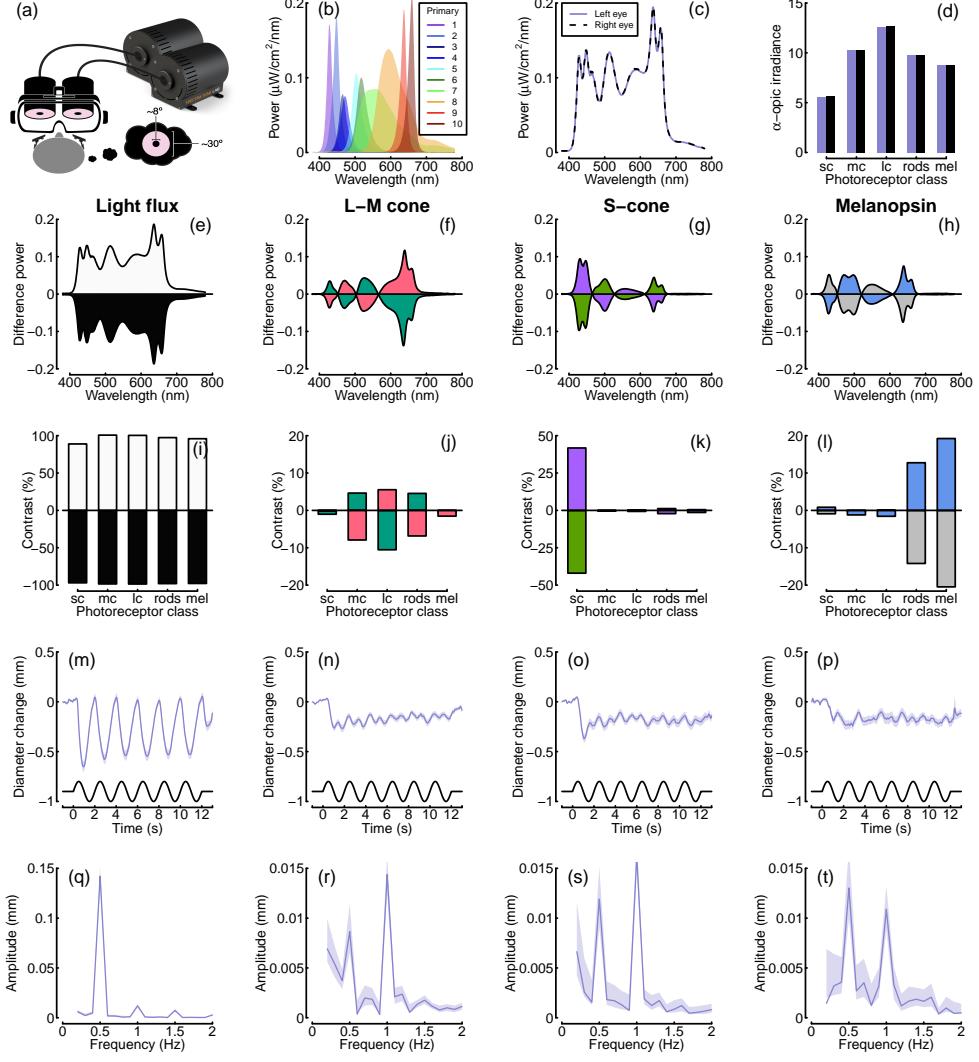


Figure 1: Summary of the spectral power distributions and alpha-optic irradiances for the background and each condition, as well as averaged pupil diameters and Fourier spectra. Panel (a) shows a schematic of the binocular stimulation system for presenting spectrally tuned modulations independently to each eye. The VR headset was attached to a clamp stand that the experimenters could use to adjust the height and align the headset with the eyes of the participant. The participant's head was supported by a chin rest to keep it in position throughout the experiment. Panel (b) shows the outputs of each LED primary at maximum intensity, and panels (c) and (d) show the overall spectral power distributions and the alpha-optic irradiances of the background spectra used for both eyes. The subsequent rows show the power differences (e-h), and photoreceptor contrasts (i-l) relative to the background, averaged pupil diameter waveforms (m-p) and Fourier spectra (q-t) for binocular stimulation. Column headings indicate the pathway stimulated, and shaded regions in panels m-t indicate bootstrapped 95% confidence intervals.

101 approximately 51 mm from the light source, at which distance the output beam was sufficiently dispersed to  
102 afford even illumination of the diffuser when viewed from the front. To guarantee safe illumination levels, a  
103 circular neutral-density filter with the same diameter of the white glass discs (50 mm) and an optical density  
104 of 0.6 log units was placed in the optical path between the light source and the diffusers. A small circular  
105 piece of blackout material with a diameter of approximately 8 degrees (10 mm) was positioned centrally on  
106 the front of each diffuser disc to aid as a fusion lock, as a fixation point, and to occlude the fovea.

107 The diffuser discs were positioned in the objective planes of the lenses of a modified VR headset (SHINECON  
108 SC-G01, Dongguan Shinecon Industrial Co. Ltd., Guangdong, China), which was used by the participants to  
109 view the stimuli. The stimuli were two discs of flickering light with a diameter of approximately 30 degrees,  
110 which were fused together into a cyclopean percept resembling a donut-shaped ring of light, similar to that  
111 used in other studies [e.g., 5,6,7,29,30]. The VR headset modifications allowed for small adjustments to  
112 account for individual differences in interpupillary distance and focal length. The use of this set up allowed  
113 us to modulate the stimuli in three different ocular configurations, similar to the ones we used in our previous  
114 study [3]: monocular, binocular and dichoptic. In the monocular configuration, the unstimulated eye still  
115 saw a non-flickering disc of mean light flux. A schematic of the stimulation system is shown in Figure  
116 1a. Pupillometry data were collected using a binocular Pupil Core eye-tracker headset (Pupil Labs GmbH,  
117 Berlin, Germany [31]) running at 120 Hz, and the signals were recorded with the Pupil Capture software.

118 Our previous study [3] used a temporal frequency of 2Hz for foveal luminance flicker, and recorded EEG  
119 data simultaneously with pupillometry. Initial pilot experiments indicated that this frequency was too high  
120 to elicit measurable responses when stimulating individual photoreceptor pathways. For all experiments, we  
121 therefore used a primary flicker frequency of 0.5Hz, as previous literature showed that this was slow enough  
122 to elicit a pupil response from all photoreceptor classes [7]. We also focussed on only recording pupillometry  
123 data as this frequency would be too slow to elicit steady-state EEG responses [32].

124 For all experiments, sinusoidal temporal modulations were presented against the same background spectrum  
125 (matched between the eyes), which was used to achieve silent substitution in the three photoreceptor mod-  
126 ulation experiments. The background spectra were defined by setting all channels to half maximum output  
127 for the brighter of the two devices (STLab 1, left eye) and then using the STLab 1/STLab 2 calibration  
128 ratio to find the equivalent settings for the companion device (STLab 2, right eye). The background spec-  
129 trum illuminance was approximately 74 lux, or 68.5 cd/m<sup>2</sup>. The spectral power distributions and  $\alpha$ -opic  
130 irradiances of the background spectra for both eyes are shown in Figure 1c-d.

131 Silent substitution stimuli were prepared and calibrated for each participant with custom Python software [27]

132 and Python scripts. Estimates of photoreceptor spectral sensitivities for each participant were constructed  
133 from the known photopigment absorbance spectra [28], taking account of the peak axial density of the  
134 respective photopigments, as well as lens [28,33,34] and macular pigment density [34,35], in accordance with  
135 the field size and age-dependant CIEPO06 observer model [36]. The melanopic and rhodopic action spectra  
136 of the 32-year-old standard observer were taken from CIE S 026 [37] and then adjusted for age-related lens  
137 transmittance with a spectral correction function, in line with the standard. Macular pigment correction  
138 was not applied to the rhodopic and melanopic action spectra because rods are not present at the fovea and  
139 ipRGCs sit above the retinal pigment layer [38].

140 In the light flux experiment, the stimulus intensity was increased and decreased relative to the background,  
141 which we expected to modulate all photoreceptor classes (see Figure 1e,i). In the L-M cone modulation  
142 experiment, we used silent substitution to increase the L-cone activity, and simultaneously decrease the  
143 M-cone activity, during the first half-cycle of the sine wave. In the second half-cycle the polarity of the  
144 modulation reversed (see Figure 1f,j). The maximum available L-M contrast was approximately 10%. In the  
145 S-cone modulation experiment, we increased and decreased S-cone-directed signals, whilst keeping activity  
146 in the other photoreceptors constant (see Figure 1g,k). Our system allowed a maximum contrast of 45%.  
147 Finally, in the melanopsin experiment, we modulated the activity of the melanopsin-containing intrinsically  
148 photoreceptive retinal ganglion cells, whilst keeping cone activity constant (see Figure 1h,l). The maximum  
149 available melanopsin contrast was 22%. We assume that the activity of rods was constant at the high  
150 background luminance intensity used here, and so did not attempt to silence rod activity in any condition, as  
151 this would have greatly reduced the available dynamic range. Splatter on nominally silenced photoreceptors  
152 was very small (see Figure 1j-l), well below the levels that would be expected to generate measurable pupil  
153 modulations, although we can observe that the rods may not be saturated in the L-M and melanopsin-  
154 directed stimuli (Figure 1j and 1l) and could intrude in these two experiments. We also estimated activation  
155 of penumbral L and M cones [5,29,39], which was minimal ( $\leq 1.5$  contrast; less than splatter on the open-field  
156 cones) for melanopsin-directed stimuli. We note that the temporal frequency of our modulation (0.5Hz) is  
157 well below the range where penumbral cone activation can elicit visible percepts, and that such percepts  
158 fade after around 1 second [29], and do not typically affect pupil responses [7].

### 159 3.3 Procedure

160 Before the start of each experiment, participants adjusted the objective planes of the lenses with the help  
161 of the experimenter until the stimulus was in focus and they perceived the two pieces of blackout material  
162 as one fused disc. Pupil responses to binocular temporal contrast modulations were examined in a factorial

163 design that combined six ocular conditions and five temporal contrast levels: 6, 12, 24, 48 and 96% of the  
164 available dynamic range. This design, similar to that used in our previous studies [3,40,41], was applied  
165 in four separate experiments, each with a different mode of photoreceptor stimulation. In the first three  
166 conditions, the discs flickered at 0.5 Hz, in either a monocular, binocular or dichoptic arrangement. In the  
167 dichoptic condition the non-target eye saw a flickering fixed contrast of 48% of the available dynamic range.  
168 In the remaining three conditions (the cross-frequency conditions) one eye's disc flickered at 0.4 Hz, and  
169 the other eye's disc flickered at 0.5 Hz. This included monocular responses at 0.4 Hz, as well as binocular  
170 (one eye sees each frequency at the target contrast) and dichoptic (target stimulus flickering at 0.5 Hz, mask  
171 contrast of 48% at 0.4 Hz in the other eye) arrangements. We counterbalanced presentation of the target  
172 stimulus across the left and right eyes.

173 The experiments were conducted in a windowless room, in which the only source of light was the modified  
174 VR headset. The participants sat as close as possible to the VR headset, leaving enough space for the  
175 eye-tracker to record the eyes. Each experiment was carried out in a single session of around 45-60 minutes,  
176 divided into three blocks of 15-17 minutes each. In each block, there were a total of 60 trials lasting 15  
177 seconds each (12s of stimulus presentation, followed by 3s of interstimulus interval). The participants were  
178 given no task other than look at the black fixation dot while trying to minimise their blinking during the  
179 presentation period. For all experiments other than the light flux condition, participants adapted to the  
180 unmodulated background luminance for two minutes before stimulation began.

181 Before the start of the L-M experiment, participants completed a luminance nulling perceptual calibration  
182 procedure in L-M cone space on an Iiyama VisionMaster<sup>TM</sup> Pro 510 display (800 x 600 pixels, 60 Hz refresh  
183 rate). During the task, participants were presented with a disc flickering within the L-M cone space (between  
184 magenta and cyan). Using a trackball, participants adjusted the angle in cone space to find their subjective  
185 isoluminant point, which resulted in changing the flickering intensity of the stimulus until the amplitude of  
186 the flicker appeared to be minimised. The result was used to modify the requested contrasts during stimulus  
187 preparation so as to account for individual differences affecting perceived illuminance, principally the L:M  
188 cone ratio [42,43].

### 189 3.4 Data analysis

190 The pupillometry data were analysed using the same method we used in our previous study [3]. The data  
191 were converted from mp4 videos to a csv text file using the Pupil Player software [31], which estimated  
192 pupil diameter for each eye on each frame using a 3D model of the eyeball. The individual data were then  
193 loaded into R for analysis, where a ten-second waveform for each trial in each eye was extracted (excluding

194 the first two seconds after stimulus onset). We interpolated across any dropped or missing frames to ensure  
 195 regular and continuous sampling over time. The Fourier transform was calculated for each waveform, and all  
 196 repetitions of each condition were pooled across eye and then averaged. Finally, data were averaged across  
 197 all participants to obtain the group results. We used coherent averaging and at each stage we excluded data  
 198 points with a Mahalanobis distance exceeding  $D = 3$  from the complex-valued mean [44]. For monocular  
 199 stimulation, we confirmed that the consensual response was equivalent to the response in the stimulated eye.  
 200 For all experiments, we used a bootstrapping procedure with  $10^4$  iterations to estimate standard errors across  
 201 participants. All analysis and figure construction was conducted using a single R-script, available online,  
 202 making this study fully computationally reproducible: <https://osf.io/gdvt4/>.

### 203 3.5 Computational model and parameter estimation

204 To quantitatively summarise our data, we used the same model described in our previous study [3]. The  
 205 model has the same general form as the first stage of the contrast gain control model proposed by Meese  
 206 and colleagues [22], but omits the second stage. For the previous model that we used [3], the exponent of  
 207 the numerator and denominator had fixed values of 2 and (implicitly) 1. Here, we allow these parameters  
 208 (called  $p$  and  $q$ ) to be free, in order to permit different shapes of contrast response function, e.g. accelerating  
 209 or saturating. The responses of the left eye and right eye channels are as follows:

$$resp_L = \frac{L^p}{Z + L^q + \omega R^q}, \quad (1)$$

$$resp_R = \frac{R^p}{Z + R^q + \omega L^q}, \quad (2)$$

210 where  $L$  and  $R$  are the contrast signals from the left and right eyes,  $p$  and  $q$  are exponents,  $Z$  is a saturation  
 211 constant that shifts the contrast-response function laterally, and  $\omega$  is the weight of suppression from the  
 212 other eye.

213 The responses from the two eyes are then summed binocularly:

$$resp_B = R_{max}(resp_L + resp_R) + k, \quad (3)$$

214 where  $k$  is a noise parameter, and  $R_{max}$  scales the overall response amplitude.

215 The models were fit using a hierarchical Bayesian framework implemented in Stan [45]. The data for each  
216 photoreceptor type and response frequency was fit separately, for a total of 8 model fits. The prior for the  $\omega$   
217 parameter was Gaussian, with a mean of 1 and standard deviation of 0.5. Priors for the other free parameters  
218 were also Gaussian, with mean values based on previous work [3]. We calculated over  $10^6$  posterior samples,  
219 and retained 10% for plotting.

## 220 4 Results

221 Figure 1m-p shows averaged waveforms of pupil diameter in response to binocular stimulation. For light  
222 flux stimuli (Figure 1m) a strong modulation is apparent at the stimulation frequency (0.5Hz), which is also  
223 clear in the Fourier amplitude spectrum (Figure 1q). For binocular stimulation of the L-M pathway, S-cone  
224 pathway and melanopsin pathway, pupil modulations were less than 10% of the amplitude of the light flux  
225 modulation (note the change in y-axis scale for the Fourier spectra), but still apparent at 0.5Hz in the Fourier  
226 spectra (Figure 1n-p). We also observed a second harmonic response (at 1Hz) for all conditions, which was  
227 weaker than the first harmonic for light flux and melanopsin stimulation, but stronger for L-M and S-cone  
228 stimulation. The second harmonic is also apparent in the pupil waveforms shown in Figure 1n-p. Our main  
229 analysis therefore focuses on the amplitude of the pupil modulations at both the first and second harmonic  
230 frequencies across different stimulus conditions.

231 Figure 2a-d shows contrast response functions across stimulation conditions for responses at the first harmonic  
232 of the main stimulation frequency (0.5Hz). In each plot, the response to monocular stimulation is given by  
233 the red circles and typically increases monotonically as a function of stimulus (temporal) contrast. Relative  
234 to monocular stimulation, binocular stimulation led to higher response amplitudes, indicating a binocular  
235 facilitation effect, for the light flux and S-cone conditions (Figure 2a,c), and to some extent for the melanopsin  
236 condition (Figure 2d). However, the L-M cone condition (Figure 2b) produced a binocular suppression effect,  
237 where the response to binocular stimulation was weaker than the response to monocular stimulation (blue  
238 squares below red circles). These results indicate that the magnitude of binocular facilitation differs across  
239 photoreceptor pathway, suggesting heterogeneity in the underlying neural computation.

240 In contemporary models of binocular signal combination, the amount of binocular facilitation is determined  
241 by the magnitude of interocular suppression, with strong suppression reducing facilitation [46]. We can  
242 estimate the strength of interocular suppression by measuring how much monocular responses are reduced  
243 when a dichoptic ‘mask’ is shown to the other eye. In our paradigm, the two components flickered at different  
244 frequencies (0.5 and 0.4Hz) so that their responses remained distinct in the Fourier spectrum [e.g. 47]. The

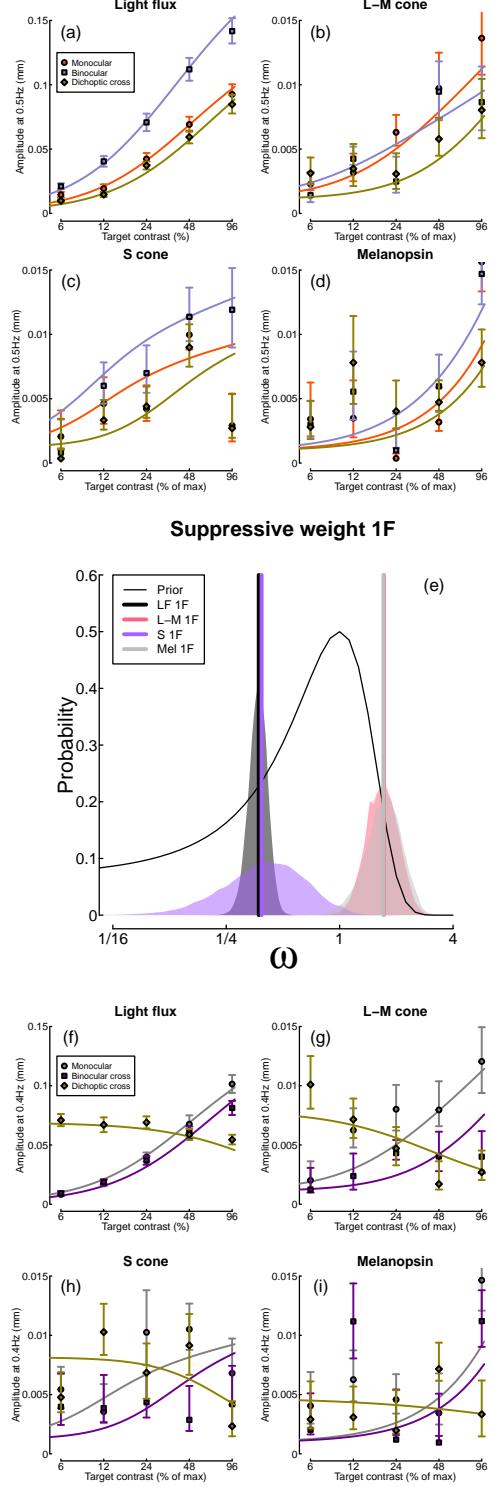


Figure 2: Contrast response functions for pupil modulations in response to flicker at 0.5Hz (panels a-d), and 0.4Hz (panels f-i), and posterior parameter distributions for the weight of interocular suppression (panel e). Within each panel (except panel e), data points are the coherently averaged amplitudes for each condition, and error bars indicate bootstrapped 95% confidence intervals. Curves show model fits using the maximum a posteriori (MAP) parameter values (see Table 1). In panel (e), vertical lines show the MAP estimates, and the thin black curve indicates the prior (note the logarithmic x-axis).

245 yellow diamond symbols in Figure 2a-d show the target responses in this condition, and in most cases were  
 246 weaker than the monocular responses (red circles). The strongest dichoptic masking is found in the L-M  
 247 condition, where we also observed the binocular suppression effect. Suppression can also be estimated from  
 248 the responses at 0.4Hz (Figure 2f-i). The reduced amplitude in the binocular cross condition (where the  
 249 two eyes received different temporal frequencies; purple squares) relative to the 0.4Hz monocular condition  
 250 (grey circles), and the progressive decline in amplitude of the dichoptic cross response (yellow diamonds)  
 251 also differ across photoreceptor conditions, showing similar differences to those observed at 0.5Hz.

252 To estimate the extent of interocular suppression for each photoreceptor pathway, we fitted each data set  
 253 using a Bayesian hierarchical implementation of a simple binocular combination model [3,22]. Our primary  
 254 objective was to compare posterior distributions of the weight of interocular suppression, which are shown in  
 255 Figure 2e. Consistent with our earlier observations, the strongest suppressive weight corresponds to the L-M  
 256 and Melanopsin conditions, and the weakest suppression corresponds to the light flux and S-cone conditions,  
 257 with virtually no overlap between the posterior distributions for weak and strong suppression. The model  
 258 fits were of good quality, as shown by the curves in Figure 2a-d,f-i. The fitted model parameters are given  
 259 in Table 1.

Table 1: Summary of maximum a posteriori (MAP) parameter estimates for each data set.

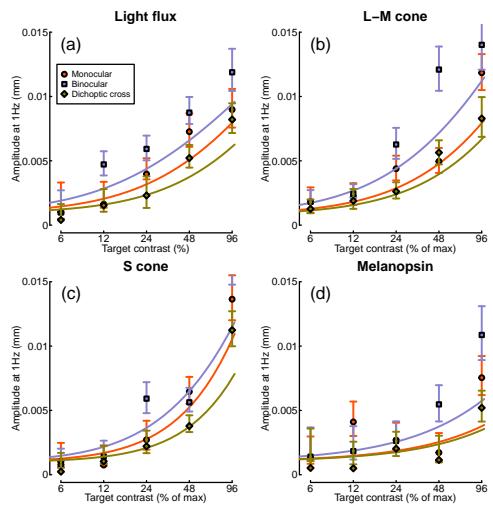
Experiment	Z	k	w	p	q	Rmax
Light flux 1F	95.77	0.00083	0.37	1.32	1.24	0.0905
L-M cone 1F	49.82	0.00109	1.71	1.50	1.21	0.0032
S cone 1F	63.00	0.00121	0.39	1.94	1.79	0.0042
Melanopsin 1F	79.61	0.00094	1.71	1.29	0.76	0.0025
Light flux 2F	48.81	0.00089	1.24	1.15	0.88	0.0039
L-M cone 2F	66.00	0.00075	0.95	1.10	0.82	0.0050
S cone 2F	75.94	0.00097	2.22	1.41	0.83	0.0018
Melanopsin 2F	67.94	0.00101	1.37	0.90	0.49	0.0035

260 At the second harmonic frequencies, the levels of suppression were more uniform across different photore-  
 261 ceptor pathways (see Figure 3). In general, suppression estimates were near or above 1 (see lower rows of  
 262 Table 1), with substantial overlap between the posterior distributions (Figure 3e). The contrast response  
 263 functions also looked more uniform, and generally involved less binocular facilitation and more interocular  
 264 suppression than were seen at the first harmonics. The L-M condition now featured the weakest suppressive

265 weight, and the strongest binocular facilitation, which was the opposite pattern seen at the first harmonic.  
266 Finally, we inspected the response phase of our four stimulation conditions, given previous reports that  
267 these differ across pathways [7], and may be in antiphase for melanopsin and S-cone signals. Figure 4 shows  
268 the phase angles for the first (a) and second (b) harmonic frequencies for binocular stimulation (monocular  
269 stimulation produced very similar results). At the first harmonic, melanopsin and S-cone signals differed in  
270 phase by more than 90 degrees, though they were not fully in antiphase. The light flux and L-M responses  
271 were approximately in antiphase to each other, however this is likely due to our choice to modulate L+M- in  
272 the first half-cycle of the sine wave (corresponding to a luminance increase in the light flux condition), and  
273 L-M+ in the second half-cycle (corresponding to a luminance decrease in the light flux condition). Had we  
274 reversed this phase arrangement, we would likely have seen a close phase correspondence between the L-M  
275 and light flux conditions (another way to think about this is that L-cone decreases and M-cone increases  
276 are processed like luminance increases). Phase differences between the light flux, S-cone and melanopsin  
277 conditions are likely attributable to different lags in phototransduction at the earliest stage (i.e. in the  
278 retina). At the second harmonic frequency (Figure 4b), the light flux condition was again out of phase with  
279 the other three, and was approximately in quadrature phase with the L-M condition, and in antiphase with  
280 both the S-cone and melanopsin conditions (which were in phase with each other). We note that the marked  
281 phase differences between conditions make the possibility that our results are dominated by rod activity  
282 relatively unlikely (e.g. in the L-M and melanopsin conditions, which have very different phase profiles).

## 283 5 Discussion

284 We used binocular pupillometry and silent substitution to measure monocular and binocular responses of  
285 the pupils to flickering stimuli when stimulating specific photoreceptor pathways. In all four experiments, we  
286 were able to record contrast response functions at both the first and the second harmonic frequencies. All  
287 experiments showed that binocular combination in the autonomic nervous system happens in a non-linear  
288 manner, with evidence of different magnitudes of interocular suppression depending on the photoreceptor  
289 pathway. This pattern of results was confirmed by a computational model, which allowed us to compare the  
290 weight of interocular suppression for each pathway. We found that at the first harmonic frequency the L-M  
291 and melanopsin pathways involved strong suppression, whereas the light flux and S-cone pathways involved  
292 weaker suppression. Suppression was strong in all four pathways at the second harmonic frequency. Finally,  
293 the phase of the pupil response revealed different lag times for the different pathways at the first and second  
294 harmonic frequencies.



**Suppressive weight 2F**

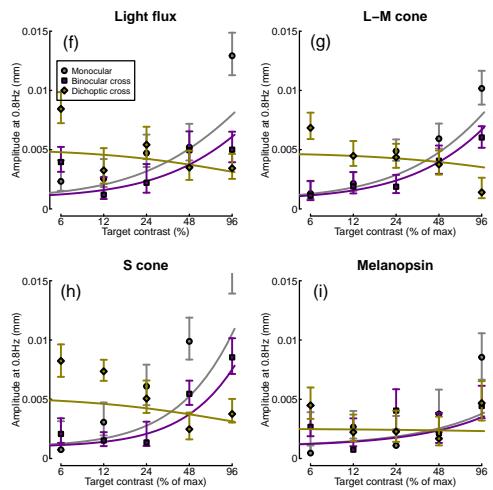
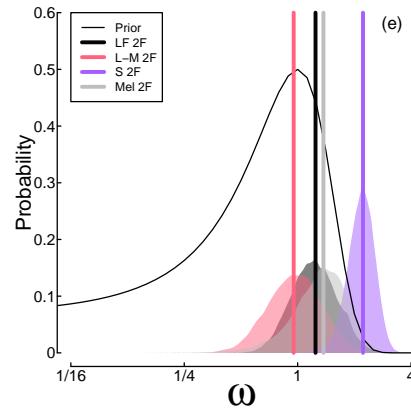


Figure 3: Responses at the second harmonic frequencies (1Hz and 0.8Hz), in the same format as Figure 2.

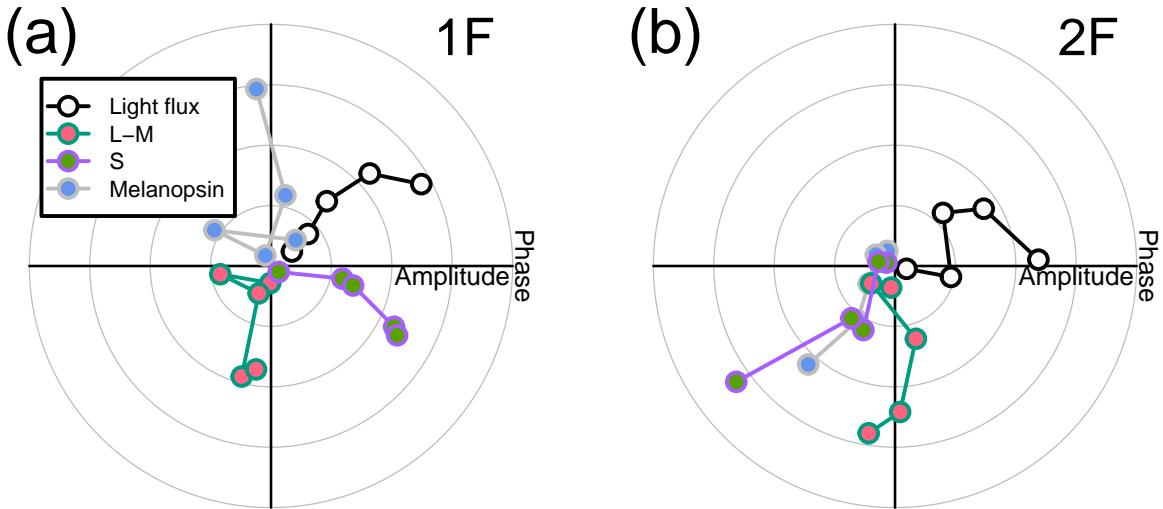


Figure 4: Pupil phase plots at the first and second harmonic frequencies for the light flux, melanopsin, L-M pathway and the S-cone pathway conditions. Panel (a) shows the pupil response at the first harmonic frequency during binocular stimulation. Panel (b) shows the pupil response at the second harmonic frequency during binocular stimulation. The five data points for each pathway correspond to the five contrast levels displayed. In panel (a) the light flux amplitudes have been scaled down by a factor of 10 to enable comparison with the other conditions.

295 This is the first study to investigate binocular interactions in the melanopsin pathway directly. A previous  
 296 meta-analysis [48] indicated that there may be a substantial binocular facilitation effect in the circadian  
 297 pathway, as indexed by melatonin suppression (melatonin is a hormone released by the pineal gland; its  
 298 production is suppressed by exposure to bright light, particularly when the melanopsin-containing ipRGCs  
 299 are stimulated). In brief, monocular stimulation of the ipRGCs requires up to ten times the signal strength  
 300 to produce an equivalent effect to binocular stimulation. This superadditive effect implies an absence of  
 301 interocular suppression, and perhaps the presence of either a highly compressive nonlinearity, or an AND-style  
 302 neural operation. Our findings here are very different - we do not see substantial binocular facilitation effects  
 303 in response to melanopsin modulation, and our data indicate that interocular suppression is very strong.  
 304 Moreover, by measuring the full contrast response function, we can rule out compressive nonlinearities,  
 305 because the functions accelerate at both the first and second harmonic frequencies. The explanation for this  
 306 difference could be due to different anatomical pathways. Binocular combination in the circadian system  
 307 likely takes place in the suprachiasmatic nucleus, whereas in the pupil constriction circuit the Edinger-  
 308 Westphal nucleus is the most likely site of binocular integration [9]. Presumably these anatomical differences,  
 309 and the practical constraints of the two systems, lead to differences in response.

310 Our recent psychophysical work [49] has looked at binocular interactions in the L-M and S-cone pathways, and  
311 compared these to the light flux pathway. Using a contrast discrimination paradigm with spatial modulations  
312 of light flux and colour, we found equally strong interocular suppression in all three pathways [49]. This  
313 is rather different from our pupillometry results here, which demonstrate weaker suppression in the light  
314 flux and S-cone pathways than in the L-M pathway (see Figure 2). However, the current experiments  
315 involve temporal modulations, which are quite distinct from the spatial modulations used in our previous  
316 work [49]. Our other recent work [3] has shown that temporal luminance modulations involve much weaker  
317 interocular suppression in the cortical response than do spatial luminance modulations [40]. These different  
318 normalization processes might reflect different priorities for spatial and temporal vision. Spatial vision aims  
319 to fuse images to provide binocular single vision, and benefits from ‘ocularity invariance’ [20], in which  
320 visual appearance is constant when viewed with one eye or two. Temporal vision is critical for motion  
321 perception, which can involve alternative binocular computations, such as calculating velocity differences  
322 between the eyes [50]. Of course the present measurements were of pupil size, which may be subject to  
323 different anatomical and functional constraints from those in the cortex. Future research should aim to  
324 extend our current findings to perception using psychophysical approaches.

## 325 6 Conclusions

326 We have demonstrated that binocular combination of temporal flickering light in the autonomic nervous  
327 system depends on the photoreceptor pathway stimulated. We were able to elicit pupil responses by stimu-  
328 lating the periphery of the retina and we were able to record contrast response functions for all photoreceptor  
329 pathways. While all pathways showed non-linear combination, they varied in how the signals are combined,  
330 particularly in the weight of interocular suppression. This was strong ( $\omega \geq 1$ ) for L-M and melanopsin  
331 signals at the first harmonic, and all pathways at the second harmonic. Suppression was weaker ( $\omega < 1$ ) in  
332 the light flux pathway (consistent with previous work), and also for S-cone directed modulations.

## 333 7 References

- 334 1. McDougal DH, Gamlin PD. Autonomic control of the eye. *Comprehensive Physiology*. 2015;5: 439–  
335 473. doi:[10.1002/cphy.c140014](https://doi.org/10.1002/cphy.c140014)
- 336 2. Wyatt HJ, Musselman JF. Pupillary light reflex in humans: Evidence for an unbalanced pathway from  
nasal retina, and for signal cancellation in brainstem. *Vision Res.* 1981;21: 513–25. doi:[10.1016/0042-6989\(81\)90097-3](https://doi.org/10.1016/0042-6989(81)90097-3)

- 338 3. Segala FG, Bruno A, Martin JT, Aung MT, Wade AR, Baker DH. Different rules for binoc-  
339 ular combination of luminance flicker in cortical and subcortical pathways. *eLife*. 2023;12.  
doi:[10.7554/elife.87048](https://doi.org/10.7554/elife.87048)
- 340 4. McDougal DH, Gamlin PD. The influence of intrinsically-photosensitive retinal ganglion cells on the  
341 spectral sensitivity and response dynamics of the human pupillary light reflex. *Vision Research*.  
2010;50: 72–87. doi:[10.1016/j.visres.2009.10.012](https://doi.org/10.1016/j.visres.2009.10.012)
- 342 5. Barrionuevo PA, Cao D. Luminance and chromatic signals interact differently with melanopsin acti-  
343 vation to control the pupil light response. *Journal of Vision*. 2016;16(11): 29. doi:[10.1167/16.11.29](https://doi.org/10.1167/16.11.29)
- 344 6. Murray IJ, Kremers J, McKeeffry D, Parry NRA. Paradoxical pupil responses to isolated  
345 M-cone increments. *Journal of the Optical Society of America A*. 2018;35: B66–B71.  
doi:[10.1364/JOSAA.35.000B66](https://doi.org/10.1364/JOSAA.35.000B66)
- 346 7. Spitschan M, Jain S, Brainard DH, Aguirre GK. Opponent melanopsin and s-cone signals in the human  
347 pupillary light response. *Proceedings of the National Academy of Sciences*. 2014;111: 15568–15572.  
doi:[10.1073/pnas.1400942111](https://doi.org/10.1073/pnas.1400942111)
- 348 8. Woelders T, Leenheers T, Gordijn MCM, Hut RA, Beersma DGM, Wams EJ. Melanopsin- and L-  
349 cone-induced pupil constriction is inhibited by S- and M-cones in humans. *Proceedings of the National  
Academy of Sciences*. 2018;115: 792–797. doi:[10.1073/pnas.1716281115](https://doi.org/10.1073/pnas.1716281115)
- 350 9. Mathôt S. Pupilometry: Psychology, physiology, and function. *J Cogn.* 2018;1: 16.  
doi:[10.5334/joc.18](https://doi.org/10.5334/joc.18)
- 351 10. Markwell EL, Feigl B, Zele AJ. Intrinsically photosensitive melanopsin retinal ganglion cell contribu-  
352 tions to the pupillary light reflex and circadian rhythm. *Clinical and Experimental Optometry*.  
2010;93: 137–149. doi:[10.1111/j.1444-0938.2010.00479.x](https://doi.org/10.1111/j.1444-0938.2010.00479.x)
- 353 11. Provencio I, Rodriguez IR, Jiang G, Hayes WP, Moreira EF, Rollag MD. A Novel Human Opsin  
354 in the Inner Retina. *Journal of Neuroscience*. 2000;20: 600–605. doi:[10.1523/JNEUROSCI.20-02-00600.2000](https://doi.org/10.1523/JNEUROSCI.20-02-00600.2000)
- 355 12. Panda S, Sato TK, Castrucci AM, Rollag MD, DeGrip WJ, Hogenesch JB, et al. Melanopsin  
356 (Opn4) requirement for normal light-induced circadian phase shifting. *Science*. 2002;298: 2213–2216.  
doi:[10.1126/science.1076848](https://doi.org/10.1126/science.1076848)
- 357 13. Ruby NF, Brennan TJ, Xie X, Cao V, Franken P, Heller HC, et al. Role of melanopsin in circadian  
358 responses to light. *Science*. 2002;298: 2211–2213. doi:[10.1126/science.1076701](https://doi.org/10.1126/science.1076701)

- 360 14. Dacey DM, Peterson BB, Robinson FR, Gamlin PD. Fireworks in the primate retina: In vitro  
photodynamics reveals diverse LGN-projecting ganglion cell types. *Neuron*. 2003;37: 15–27.  
doi:[https://doi.org/10.1016/S0896-6273\(02\)01143-1](https://doi.org/10.1016/S0896-6273(02)01143-1)
- 361
- 362 15. Lucas RJ, Hattar S, Takao M, Berson DM, Foster RG, Yau K-W. Diminished pupillary  
light reflex at high irradiances in melanopsin-knockout mice. *Science*. 2003;299: 245–247.  
doi:[10.1126/science.1077293](https://doi.org/10.1126/science.1077293)
- 363
- 364 16. Gamlin PDR, McDougal DH, Pokorny J, Smith VC, Yau K-W, Dacey DM. Human and macaque  
pupil responses driven by melanopsin-containing retinal ganglion cells. *Vision Research*. 2007;47:  
946–954. doi:[10.1016/j.visres.2006.12.015](https://doi.org/10.1016/j.visres.2006.12.015)
- 365
- 366 17. Hubel DH, Wiesel TN. Receptive fields, binocular interaction and functional architecture in the cat's  
visual cortex. *J Physiol*. 1962;160: 106–54. doi:[10.1113/jphysiol.1962.sp006837](https://doi.org/10.1113/jphysiol.1962.sp006837)
- 367
- 368 18. Baker DH, Lygo FA, Meese TS, Georgeson MA. Binocular summation revisited: Beyond  $\sqrt{2}$ . *Psychol  
Bull.* 2018;144: 1186–1199. doi:[10.1037/bul0000163](https://doi.org/10.1037/bul0000163)
- 369
- 370 19. Campbell FW, Green DG. Monocular versus binocular visual acuity. *Nature*. 1965;208: 191–2.  
doi:[10.1038/208191a0](https://doi.org/10.1038/208191a0)
- 371
- 372 20. Baker DH, Meese TS, Georgeson MA. Binocular interaction: Contrast matching and con-  
trast discrimination are predicted by the same model. *Spat Vis*. 2007;20: 397–413.  
doi:[10.1163/156856807781503622](https://doi.org/10.1163/156856807781503622)
- 373
- 374 21. Moradi F, Heeger DJ. Inter-ocular contrast normalization in human visual cortex. *J Vis*. 2009;9:  
13.1–22. doi:[10.1167/9.3.13](https://doi.org/10.1167/9.3.13)
- 375
- 376 22. Meese TS, Georgeson MA, Baker DH. Binocular contrast vision at and above threshold. *Journal of  
Vision*. 2006;6: 1224–43. doi:[10.1167/6.11.7](https://doi.org/10.1167/6.11.7)
- 377
- 378 23. Martin JT, Pinto J, Bulte D, Spitschan M. PyPlr: A versatile, integrated system of hardware and  
software for researching the human pupillary light reflex. *Behavior Research Methods*. 2022;54:  
2720–2739. doi:[10.3758/s13428-021-01759-3](https://doi.org/10.3758/s13428-021-01759-3)
- 379
- 380 24. Estévez O, Spekreijse H. The "silent substitution" method in visual research. *Vision Research*.  
1982;22: 681–691. doi:[10.1016/0042-6989\(82\)90104-3](https://doi.org/10.1016/0042-6989(82)90104-3)
- 381
- 382 25. Shapiro AG, Pokorny J, Smith VC. Cone–rod receptor spaces with illustrations that use CRT phos-  
phor and light-emitting-diode spectra. *Journal of the Optical Society of America A*. 1996;13: 2319.  
doi:[10.1364/josaa.13.002319](https://doi.org/10.1364/josaa.13.002319)
- 383

- 384 26. Spitschan M, Woelders T. The Method of Silent Substitution for Examining Melanopsin Contributions  
385 to Pupil Control. *Frontiers in Neurology*. 2018;9. doi:[10.3389/fneur.2018.00941](https://doi.org/10.3389/fneur.2018.00941)
- 386 27. Martin JT, Boynton GM, Baker DH, Wade AR, Spitschan M. PySilSub: An open-source python  
387 toolbox for implementing the method of silent substitution in vision and nonvisual photoreception  
research. *Journal of Vision*. 2023;23: 10. doi:[10.1167/jov.23.7.10](https://doi.org/10.1167/jov.23.7.10)
- 388 28. Stockman A, Sharpe LT. The spectral sensitivities of the middle- and long-wavelength-sensitive  
389 cones derived from measurements in observers of known genotype. *Vision Res.* 2000;40: 1711–37.  
doi:[10.1016/s0042-6989\(00\)00021-3](https://doi.org/10.1016/s0042-6989(00)00021-3)
- 390 29. Spitschan M, Aguirre GK, Brainard DH. Selective stimulation of penumbral cones reveals perception in  
391 the shadow of retinal blood vessels. *PLoS One*. 2015;10: e0124328. doi:[10.1371/journal.pone.0124328](https://doi.org/10.1371/journal.pone.0124328)
- 392 30. Zele AJ, Feigl B, Adhikari P, Maynard ML, Cao D. Melanopsin photoreception contributes to human  
393 visual detection, temporal and colour processing. *Scientific Reports*. 2018;8. doi:[10.1038/s41598-018-22197-w](https://doi.org/10.1038/s41598-018-22197-w)
- 394 31. Kassner M, Patera W, Bulling A. Pupil: An open source platform for pervasive eye tracking and mobile  
395 gaze-based interaction. *Proceedings of the 2014 ACM international joint conference on pervasive and  
ubiquitous computing: Adjunct publication*. ACM; 2014. doi:[10.1145/2638728.2641695](https://doi.org/10.1145/2638728.2641695)
- 396 32. Norcia AM, Appelbaum LG, Ales JM, Cottreau BR, Rossion B. The steady-state visual evoked  
397 potential in vision research: A review. *Journal of Vision*. 2015;15(6): 4. doi:[10.1167/15.6.4](https://doi.org/10.1167/15.6.4)
- 398 33. Pokorny J, Smith VC, Lutze M. Aging of the human lens. *Applied Optics*. 1987;26: 1437.  
399 doi:[10.1364/ao.26.001437](https://doi.org/10.1364/ao.26.001437)
- 400 34. Stockman A, Sharpe LT, Fach C. The spectral sensitivity of the human short-wavelength sensitive  
401 cones derived from thresholds and color matches. *Vision Research*. 1999;39: 2901–2927. doi:[https://doi.org/10.1016/S0042-6989\(98\)00225-9](https://doi.org/10.1016/S0042-6989(98)00225-9)
- 402 35. Bone RA, Landrum JT, Fernandez L, Tarsis SL. Analysis of the macular pigment by HPLC: Retinal  
403 distribution and age study. *Investigative Ophthalmology & Visual Science*. 1988;29: 843–849.
- 404 36. CIE. Fundamental chromaticity diagram with physiological axes. CIE Central Bureau; 2006. Available:  
405 <https://cie.co.at/publications/fundamental-chromaticity-diagram-physiological-axes-part-1>
- 406 37. CIE. CIE system for metrology of optical radiation for ipRGC-influenced responses to light. CIE  
407 Central Bureau; 2018. doi:[10.25039/S026.2018](https://doi.org/10.25039/S026.2018)

- 408 38. Trieschmann M, Kuijk FJGM van, Alexander R, Hermans P, Luthert P, Bird AC, et al. Macular  
pigment in the human retina: Histological evaluation of localization and distribution. *Eye*. 2007;22:  
132–137. doi:[10.1038/sj.eye.6702780](https://doi.org/10.1038/sj.eye.6702780)
- 409
- 410 39. Zele AJ, Adhikari P, Cao D, Feigl B. Melanopsin driven enhancement of cone-mediated visual pro-  
411 cessing. *Vision Res.* 2019;160: 72–81. doi:[10.1016/j.visres.2019.04.009](https://doi.org/10.1016/j.visres.2019.04.009)
- 412 40. Baker DH, Wade AR. Evidence for an optimal algorithm underlying signal combination in human  
413 visual cortex. *Cereb Cortex*. 2017;27: 254–264. doi:[10.1093/cercor/bhw395](https://doi.org/10.1093/cercor/bhw395)
- 414 41. Baker DH, Vilidaite G, McClarnon E, Valkova E, Bruno A, Millman RE. Binaural summation of am-  
415 plitude modulation involves weak interaural suppression. *Sci Rep.* 2020;10: 3560. doi:[10.1038/s41598-020-60602-5](https://doi.org/10.1038/s41598-020-60602-5)
- 416 42. Carroll J, Neitz J, Neitz M. Estimates of L:M cone ratio from ERG flicker photometry and genetics.  
417 *Journal of Vision*. 2002;2: 1. doi:[10.1167/2.8.1](https://doi.org/10.1167/2.8.1)
- 418 43. Hofer H, Carroll J, Neitz J, Neitz M, Williams DR. Organization of the human trichromatic cone  
419 mosaic. *The Journal of Neuroscience*. 2005;25: 9669–9679. doi:[10.1523/jneurosci.2414-05.2005](https://doi.org/10.1523/jneurosci.2414-05.2005)
- 420 44. Baker DH. Statistical analysis of periodic data in neuroscience. *Neurons, Behavior, Data analysis,  
421 and Theory*. 2021;5. doi:[10.51628/001c.27680](https://doi.org/10.51628/001c.27680)
- 422 45. Carpenter B, Gelman A, Hoffman MD, Lee D, Goodrich B, Betancourt M, et al. Stan: A probabilistic  
423 programming language. *J Stat Softw.* 2017;76. doi:[10.18637/jss.v076.i01](https://doi.org/10.18637/jss.v076.i01)
- 424 46. Kingdom FAA, Libenson L. Dichoptic color saturation mixture: Binocular luminance contrast pro-  
425 motes perceptual averaging. *J Vis.* 2015;15: 2. doi:[10.1167/15.5.2](https://doi.org/10.1167/15.5.2)
- 426 47. Busse L, Wade AR, Carandini M. Representation of concurrent stimuli by population activity in  
427 visual cortex. *Neuron*. 2009;64: 931–42. doi:[10.1016/j.neuron.2009.11.004](https://doi.org/10.1016/j.neuron.2009.11.004)
- 428 48. Spitschan M, Cajochen C. Binocular facilitation in light-mediated melatonin suppression? *J Pineal  
429 Res.* 2019;67: e12602. doi:[10.1111/jpi.12602](https://doi.org/10.1111/jpi.12602)
- 430 49. Baker DH, Hansford KJ, Segala FG, Morsi AY, Huxley RJ, Martin JT, et al. Binocular integration  
431 of chromatic and luminance signals. *J Vis.* 2024;24(12): 7. doi:[10.1167/jov.24.12.7](https://doi.org/10.1167/jov.24.12.7)
- 432 50. Kaestner M, Maloney RT, Wailes-Newson KH, Bloj M, Harris JM, Morland AB, et al. Asymmetries  
433 between achromatic and chromatic extraction of 3D motion signals. *Proc Natl Acad Sci U S A.*  
2019;116: 13631–13640. doi:[10.1073/pnas.1817202116](https://doi.org/10.1073/pnas.1817202116)

## 8 Supplementary material

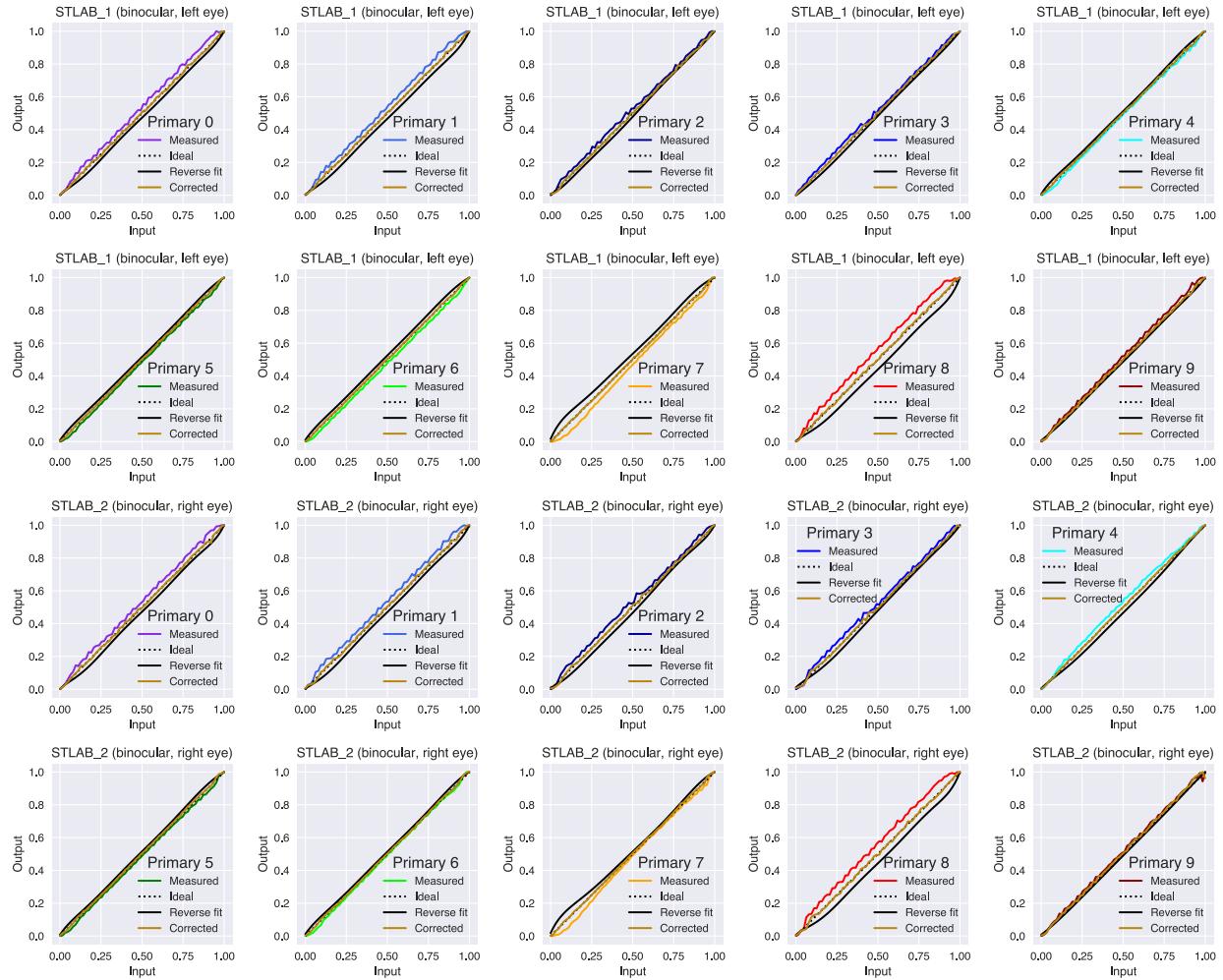


Figure S1: Linearity of primaries for each device. The calibration measures were summed (i.e., total unweighted irradiance), and the input-output relationship summarised by a 7th order polynomial reverse curve fit. By applying the coefficients of the regression, it is possible to achieve a linear output in the primaries.