

## Supplementary Figures

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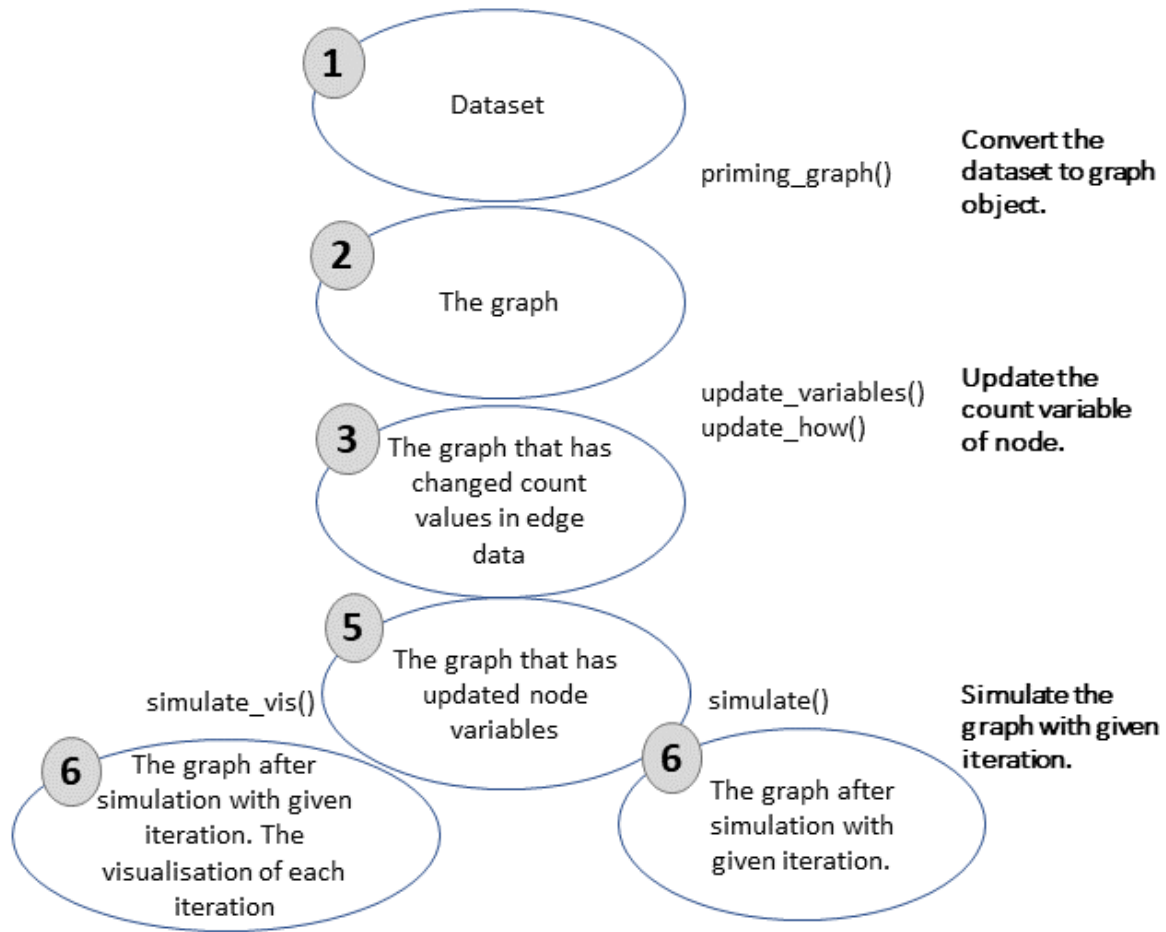


Figure S1: Workflow for simulation of competing endogenous RNA regulations. Graph object in steps 2-6 is saved and updated continuously.

```
#install.packages("devtools")
#devtools::install_github("selcenari/ceRNAetsim")
library(ceRNAetsim)
```

*minsamp* dataset analysis in absence of interaction factors.

```
minsamp<-readRDS("data/minsamp.RDS")
```

```
priming_graph(minsamp,  
  competing_count = Competing_expression,  
  miRNA_count = miRNA_expression)%>%  
vis_graph(Competing_color = "navajowhite3",  
  mirna_color = "ivory4",  
  title = "Minimal dataset in\nsteady-state conditions")
```

## Minimal dataset in steady-state conditions

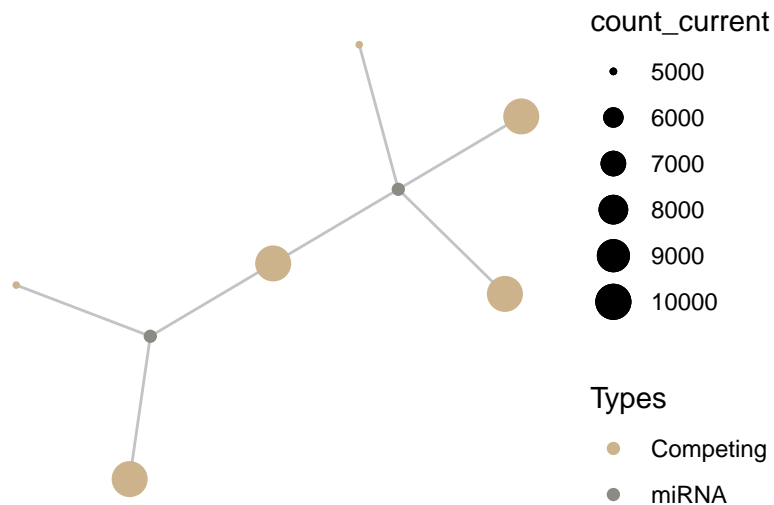


Figure S2: Minimal Dataset in Steady-state

```
priming_graph(minsamp,  
  competing_count = Competing_expression,  
  miRNA_count = miRNA_expression)%>%  
update_how("Gene2", 2)%>%  
vis_graph(Competing_color = "navajowhite3",  
  mirna_color = "ivory4",  
  Upregulation = "red",  
  title = "Gene2 Upregulation\nwithout interaction factors")
```

## Gene2 Upregulation without interaction factors

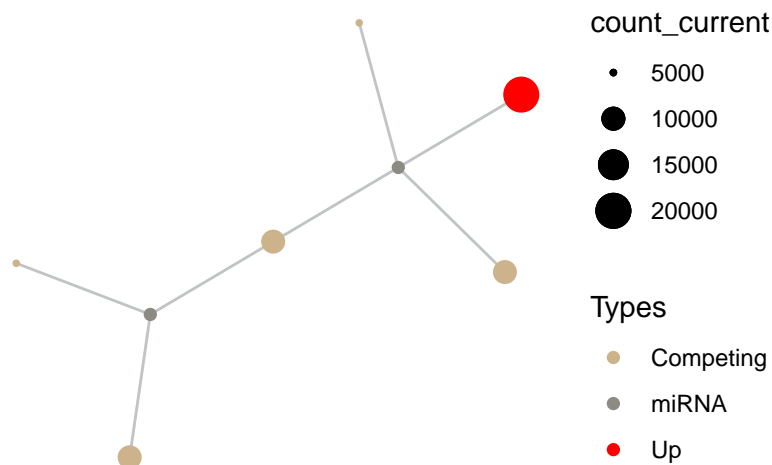


Figure S3: Gene2 Upregulation on Minimal Dataset

```
priming_graph(minsamp,
               competing_count = Competing_expression,
               miRNA_count = miRNA_expression) %>%
update_how("Gene2", 2) %>%
simulate(cycle=1) %>%
vis_graph(Competing_color = "navajowhite3",
           mirna_color = "ivory4",
           Upregulation = "red",
           Downregulation = "blue",
           title = "A")

priming_graph(minsamp,
               competing_count = Competing_expression,
               miRNA_count = miRNA_expression) %>%
update_how("Gene2", 2) %>%
simulate(2) %>%
vis_graph(Competing_color = "navajowhite3",
           mirna_color = "ivory4",
           Upregulation = "red",
           Downregulation = "blue",
           title = "B")
```

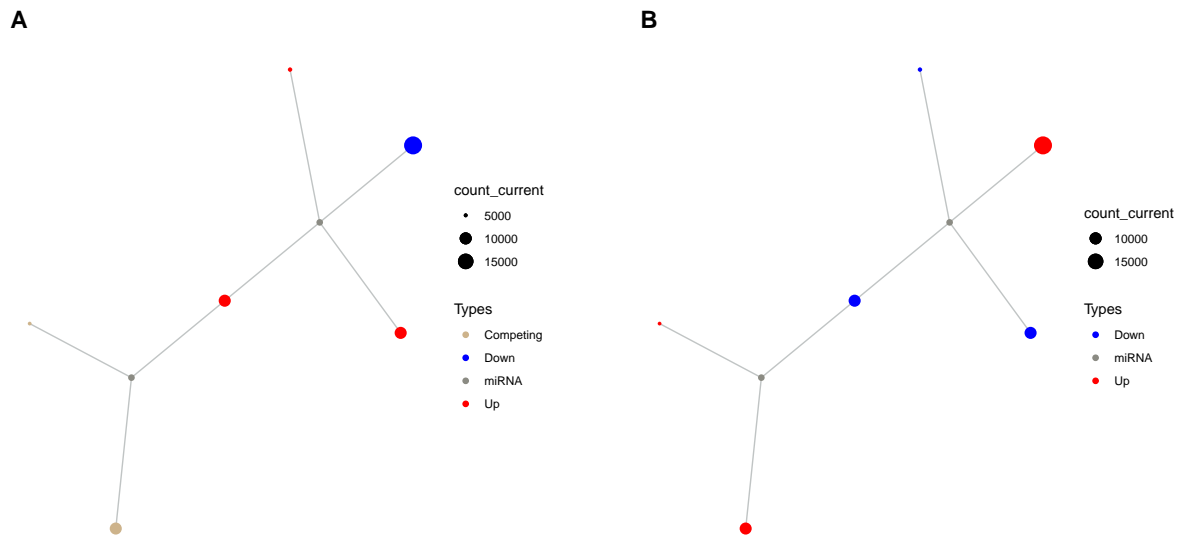


Figure S4: Sequential iteration of minsamp data. A) First response of system to Gene2 upregulation (2nd iteration). B) Spreading of perturbation on system (3th iteration)

### *minsamp* dataset analysis with interaction factors.

```
priming_graph(minsamp,
  competing_count = Competing_expression,
  miRNA_count = miRNA_expression,
  aff_factor = c(energy, seed_type),
  deg_factor = region)%>%
vis_graph(Competing_color = "navajowhite3",
  mirna_color = "ivory4",
  title = "Minimal dataset with interaction\nfactors in steady-state conditions")
```

## Minimal dataset with interaction factors in steady-state conditions

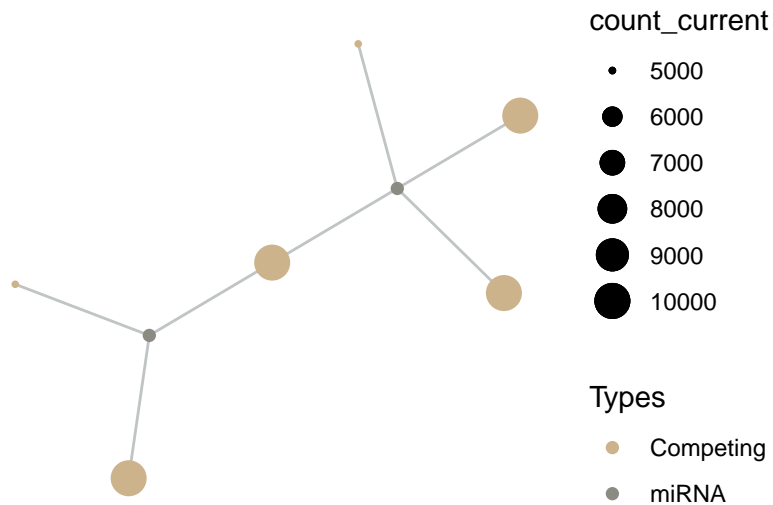


Figure S5: Minimal Dataset with interaction factors in Steady-state

```
priming_graph(minsamp,
               competing_count = Competing_expression,
               miRNA_count = miRNA_expression,
               aff_factor = c(energy, seed_type),
               deg_factor = region)%>%
update_how("Gene2", 2)%>%
vis_graph(Competing_color = "navajowhite3",
          mirna_color = "ivory4",
          Upregulation = "red",
          title = "Gene2 Upregulation\nwith interaction factors")
```

## Gene2 Upregulation with interaction factors

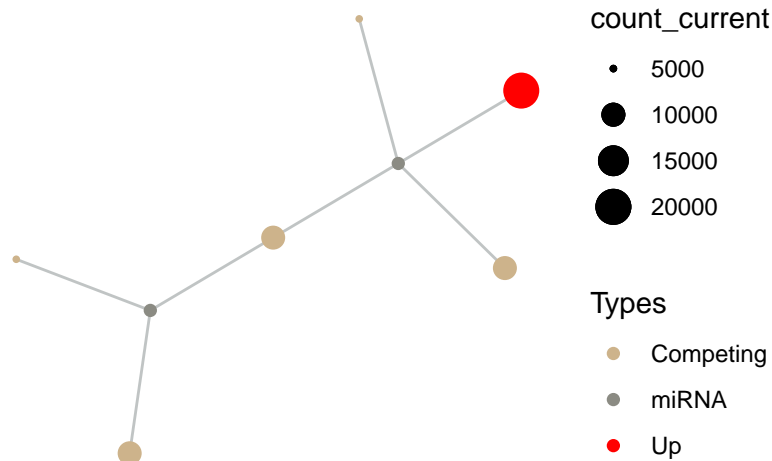


Figure S6: When Gene2 is upregulated on Minimal Dataset with interaction factors

```
priming_graph(minsamp,
               competing_count = Competing_expression,
               miRNA_count = miRNA_expression,
               aff_factor = c(energy, seed_type),
               deg_factor = region)%>%
update_how("Gene2", 2)%>%
simulate(cycle = 1)%>%
vis_graph(Competing_color = "navajowhite3",
           mirna_color = "ivory4",
           Upregulation = "red",
           Downregulation = "blue",
           title = "A")

priming_graph(minsamp,
               competing_count = Competing_expression,
               miRNA_count = miRNA_expression,
               aff_factor = c(energy, seed_type),
               deg_factor = region)%>%
update_how("Gene2", 2)%>%
simulate(cycle = 2)%>%
vis_graph(Competing_color = "navajowhite3",
           mirna_color = "ivory4",
           Upregulation = "red",
           Downregulation = "blue",
           title = "B")
```

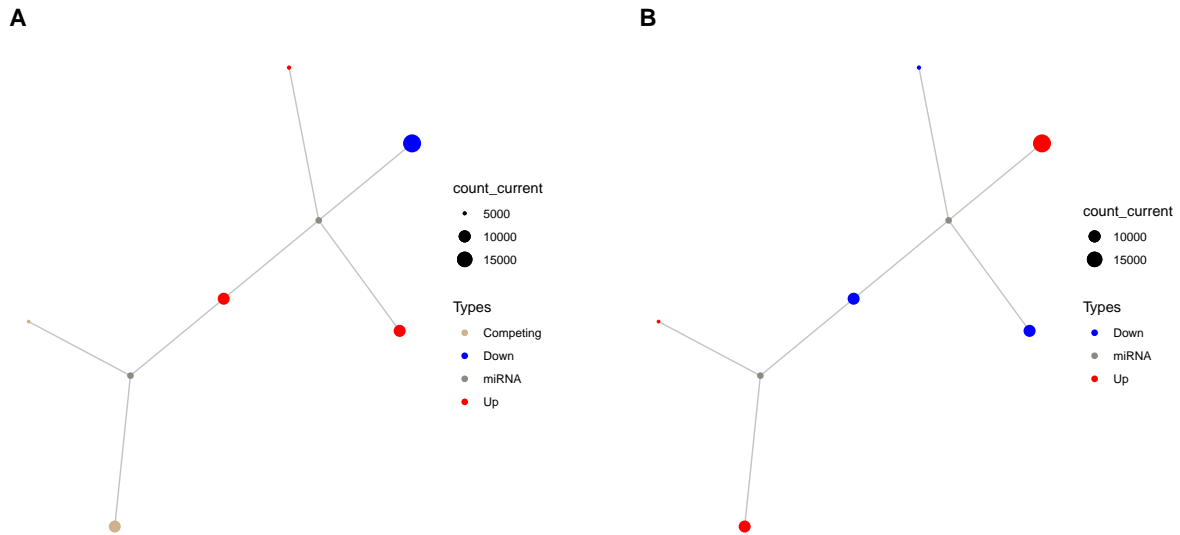


Figure S7: Sequential iteration of minsamp data with interaction factors A)First response of system to Gene2 upregulation (2nd iteration). B)Spreading of perturbation on system (3th iteration)

### Common target perturbation in *minsamp* dataset.

```
priming_graph(minsamp,
  competing_count = Competing_expression,
  miRNA_count = miRNA_expression,
  aff_factor = c(energy, seed_type),
  deg_factor = region)%>%
vis_graph(Competing_color = "navajowhite3",
  mirna_color = "ivory4",
  Upregulation = "red",
  title = "A")

priming_graph(minsamp,
  competing_count = Competing_expression,
  miRNA_count = miRNA_expression,
  aff_factor = c(energy, seed_type),
  deg_factor = region)%>%
update_how("Gene4", 2)%>%
vis_graph(Competing_color = "navajowhite3",
  mirna_color = "ivory4",
  Upregulation = "red",
  title = "B")

priming_graph(minsamp,
  competing_count = Competing_expression,
  miRNA_count = miRNA_expression,
  aff_factor = c(energy, seed_type),
  deg_factor = region)%>%
update_how("Gene4", 2)%>%
```

```

simulate(cycle=1)%>%
vis_graph(Competing_color = "navajowhite3",
  mirna_color = "ivory4",
  Upregulation = "red",
  title = "C")

priming_graph(minsamp,
  competing_count = Competing_expression,
  miRNA_count = miRNA_expression,
  aff_factor = c(energy, seed_type),
  deg_factor = region)%>%
update_how("Gene4", 2)%>%
simulate(cycle=2)%>%
vis_graph(Competing_color = "navajowhite3",
  mirna_color = "ivory4",
  Upregulation = "red",
  title = "D")

```



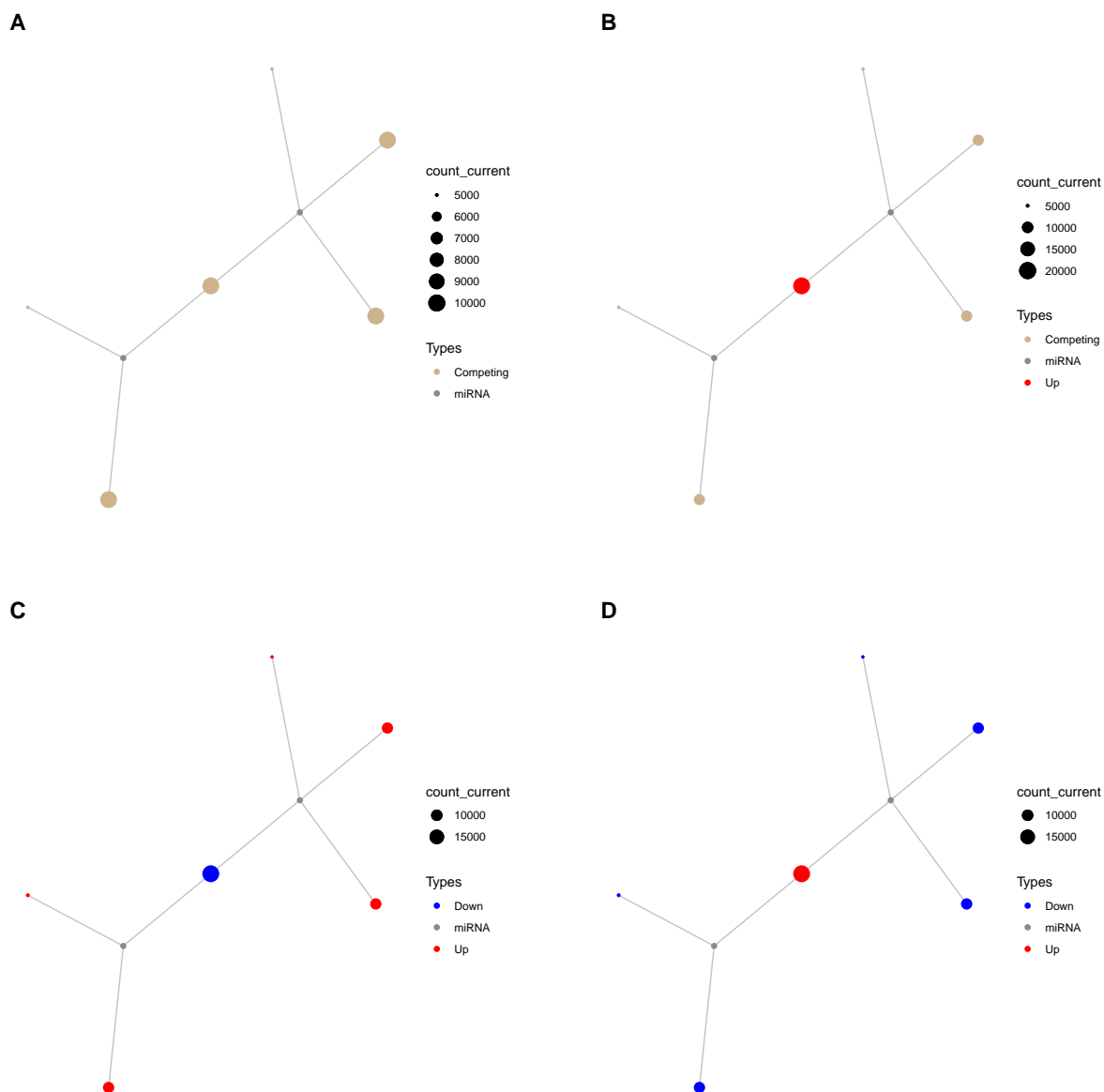


Figure S8: Perturbation of Gene4 on minsamp data with interaction factors. A) Network at steady-state. B) Upregulation of Gene4. C) Primary response of network to upregulation of Gene4. D) Re-regulation of whole nodes on system (3th iteration)

## Determination of optimum iteration for Upregulation of SERPINE2 gene from Breast cancer patient dataset

```
E9GE_mirnagenenormal<-readRDS("data/E9GE_mirnagenenormal.RDS")
head(E9GE_mirnagenenormal)
```

```
## # A tibble: 6 x 7
```

```
## hgnc_symbol miRNA_name mirna_RPM GE_normal Energy seed_type_effect
## <chr> <chr> <dbl> <dbl> <dbl> <dbl>
## 1 CCNG1 hsa-let-7... 111204. 5245 -25.1 0.05
## 2 DICER1 hsa-let-7... 111204. 3285 -24.4 0.43
## 3 SESN1 hsa-let-7... 111204. 1179 -22.2 0.05
## 4 NIPBL hsa-let-7... 111204. 4503 -22.1 0.05
## 5 INTS12 hsa-let-7... 111204. 600 -21.9 0.05
## 6 FNIP1 hsa-let-7... 111204. 1248 -21.8 0.43
## # ... with 1 more variable: region_effect <dbl>
```

```
as.data.frame(E9GE_mirnagenenormal)%>%
  priming_graph(competing_count = GE_normal,
                miRNA_count = mirna_RPM,
                aff_factor = c(Energy, seed_type_effect),
                deg_factor = region_effect)%>%
  update_how("SERPINE2",2.75) %>%
  simulate(150) %>%
  find_iteration(limit=1, plot= TRUE)
```

```
## Warning in priming_graph(., competing_count = GE_normal, miRNA_count = mirna_RPM, : First column is p
```

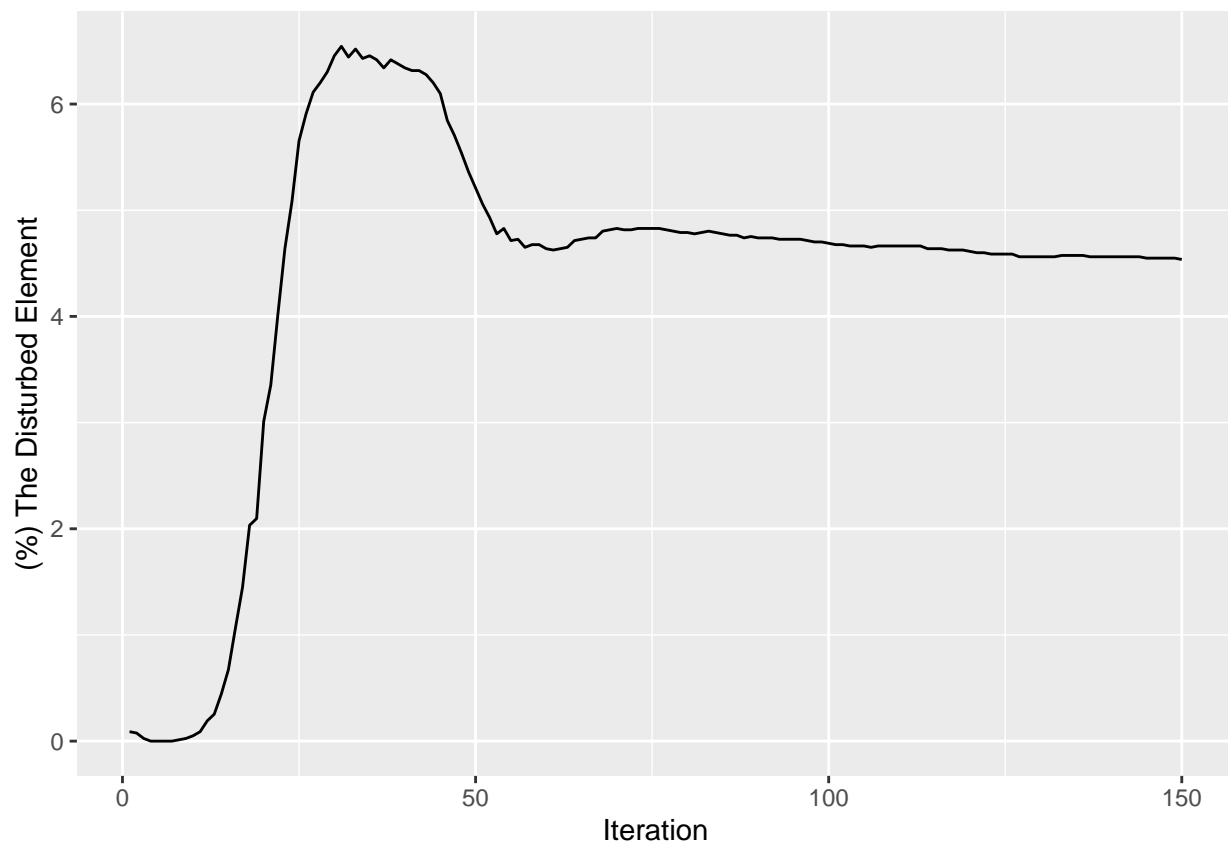


Figure S9: Percentage of affected nodes of each iteration for SERPINE2 Gene

*#limit=0 describes the change that is not taken into account.*

## Highly effective perturbing elements in network

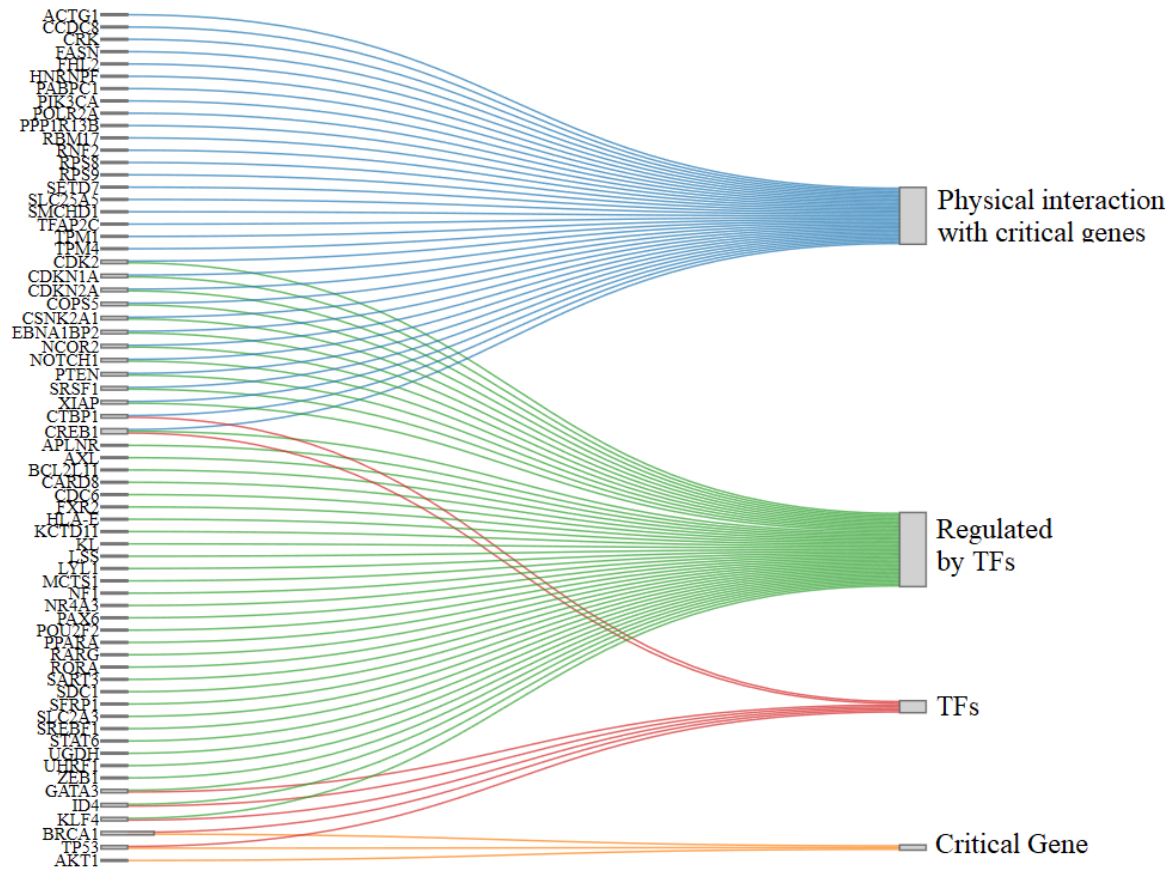


Figure S10: Interactions of genes that have high perturbation efficiency. Critical gene, represents crucial genes for breast cancer, provided by eDGAR; TFs, transcription factors; Regulated by TFs, regulated genes with these transcription factors; Physical interaction with critical genes; shared genes in physical interactions between critical genes