Main Figures

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Figures

This file consists analysis steps and codes of figures in manuscript.

```
mirtarbase_node_perturbations <- readRDS("mirtarbase_node_perturbations.RDS") # data includes results
```

Venn diagrams Figure 4A

```
gene_dist <- mirtarbase_node_perturbations %>% group_by(name) %>%
    summarise(perturbed = sum(perturbed_count > 0,
        na.rm = TRUE)) %>% ungroup()

## 'summarise()' ungrouping output (override with '.groups' argument)

effective_nodes <- mirtarbase_node_perturbations %>%
    left_join(gene_dist, by = "name") %>% filter(perturbed >
    10, perturbed_count > 78) %>% distinct(name)
```

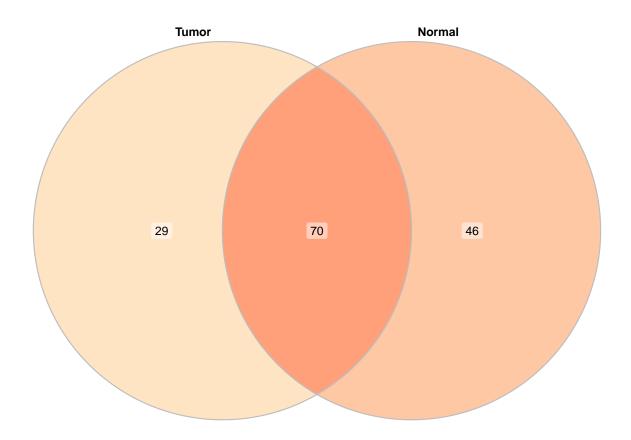
• Detecting highly perturbing gene nodes in tumor tissues

```
lim_1_gene_tumor <- mirtarbase_node_perturbations %>%
    left_join(gene_dist, by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
    "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
    effective_nodes$name, "effective", "non_effective")) %>%
    filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type !=
    "Normal", startsWith(name, "ENS")) %>% distinct(name) %>%
    pull()
```

• Detecting highly perturbing gene nodes in normal tissues

```
lim_1_gene_normal <- mirtarbase_node_perturbations %>%
   left_join(gene_dist, by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
   "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
   effective_nodes$name, "effective", "non_effective")) %>%
   filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type ==
   "Normal", startsWith(name, "ENS")) %>% distinct(name) %>%
   pull()
```

Venn diagram of genes

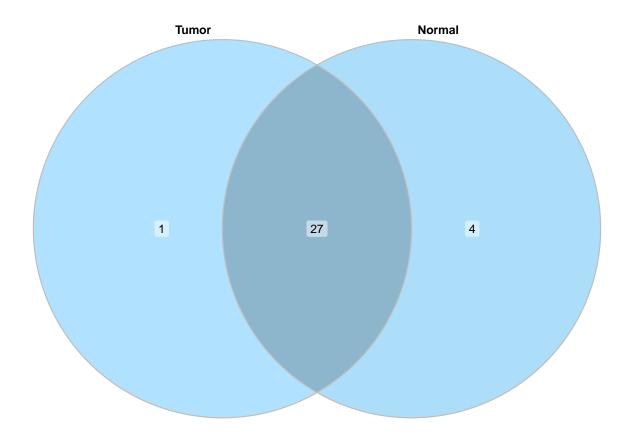


• Detecting highly perturbing mirna nodes in tumor tissues

```
lim_1_mirna_tumor <- mirtarbase_node_perturbations %>%
   left_join(gene_dist, by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
   "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
   effective_nodes$name, "effective", "non_effective")) %>%
   filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type !=
   "Normal", startsWith(name, "hsa")) %>% distinct(name) %>%
   pull()
```

• Detecting highly perturbing mirna nodes in normal tissues

```
lim_1_mirna_normal <- mirtarbase_node_perturbations %>%
   left_join(gene_dist, by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
   "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
   effective_nodes$name, "effective", "non_effective")) %>%
   filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type ==
   "Normal", startsWith(name, "hsa")) %>% distinct(name) %>%
   pull()
```



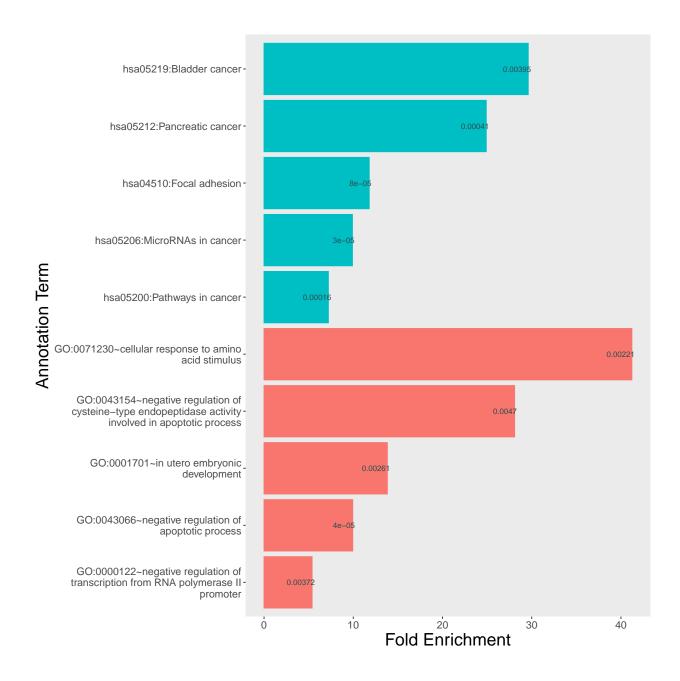
Functional Annotation Analysis

Anno_tumor_specific <- readRDS("Anno_tumor_specific.RDS") # includes functional annotation of 29 genes

#Fuctional annotation of Tumor specific 29 genes. Figure 4B

Tumor only:

```
tumor_only_annotation <- Anno_tumor_specific %>% filter(startsWith(Category,
    c("KEGG", "GO")), !str_detect(Category, "CC_DIRECT")) %>%
    filter(!str_detect(Category, "MF_DIRECT")) %>%
    clean_names() %>% group_by(category) %>% filter(row_number() <</pre>
    6)
position <- rev(c("hsa05219:Bladder cancer", "hsa05212:Pancreatic cancer",
    "hsa04510:Focal adhesion", "hsa05206:MicroRNAs in cancer",
    "hsa05200:Pathways in cancer", "GO:0071230~cellular response to amino acid stimulus",
    "G0:0043154~negative regulation of cysteine-type endopeptidase activity involved in apoptotic proce
    "GO:0001701~in utero embryonic development", "GO:0043066~negative regulation of apoptotic process",
    "GO:0000122~negative regulation of transcription from RNA polymerase II promoter"))
tumor_only_annotation %>% group_by(category) %>% arrange(-desc(fold_enrichment))
## # A tibble: 10 x 13
## # Groups: category [2]
##
      category term count percent p_value genes list_total pop_hits pop_total
              <chr> <dbl> <dbl>
                                    <dbl> <chr>
                                                      <dbl>
                                                               <dbl>
##
      <chr>
                                                                         <dbl>
                              20.7 3.72e-3 ENSG~
                                                                         16792
## 1 GOTERM_~ GO:O~
                                                         26
                                                                 720
## 2 KEGG_PA~ hsa0~
                         7
                             24.1 1.64e-4 ENSG~
                                                         17
                                                                 393
                                                                          6879
## 3 KEGG_PA~ hsa0~
                         7
                             24.1 2.76e-5 ENSG~
                                                                 286
                                                         17
                                                                          6879
## 4 GOTERM_~ GO:O~
                        7
                             24.1 4.38e-5 ENSG~
                                                         26
                                                                 455
                                                                         16792
## 5 KEGG_PA~ hsa0~
                             20.7 7.65e-5 ENSG~
                         6
                                                         17
                                                                 206
                                                                          6879
## 6 GOTERM_~ GO:0~
                             13.8 2.61e-3 ENSG~
                                                                 187
                                                                         16792
                        4
                                                         26
## 7 KEGG PA~ hsa0~
                             13.8 4.13e-4 ENSG~
                        4
                                                         17
                                                                 65
                                                                          6879
## 8 GOTERM_~ GO:0~
                        3
                             10.3 4.70e-3 ENSG~
                                                         26
                                                                  69
                                                                         16792
## 9 KEGG PA~ hsa0~
                        3
                             10.3 3.95e-3 ENSG~
                                                         17
                                                                  41
                                                                          6879
## 10 GOTERM_~ GO:0~
                        3
                             10.3 2.21e-3 ENSG~
                                                         26
                                                                  47
                                                                         16792
## # ... with 4 more variables: fold_enrichment <dbl>, bonferroni <dbl>,
## # benjamini <dbl>, fdr <dbl>
tumor_only_annotation %>% ggplot(aes(x = fold_enrichment,
   y = term)) + geom_col(aes(color = category, fill = category)) +
    geom_text(aes(label = round(p_value, 5)), hjust = 0.9,
        color = "darkslategrey", size = 3) + xlab("Fold Enrichment") +
   ylab("Annotation Term") + theme(panel.grid = element_blank(),
    axis.title = element_text(size = 20), axis.text = element_text(size = 12),
   legend.position = "none") + scale_y_discrete(labels = scales::wrap_format(40),
   position = "left", limits = position)
```



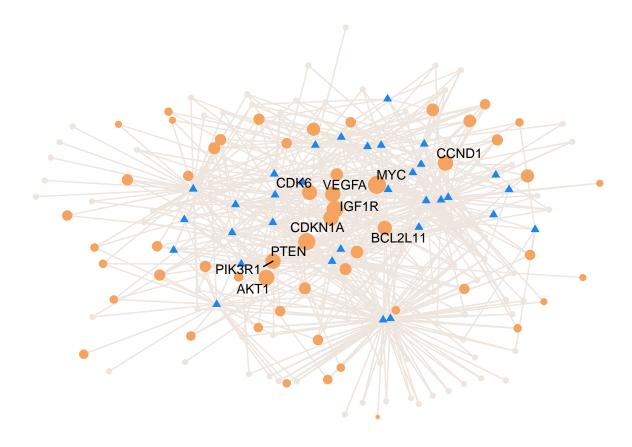
Network construction with overall functional annotation: Figure 3

significant_node_graph <- readRDS("significant_node_graph.RDS") #consists network of highly perturbing
hg19 <- readRDS("hg19.RDS") #For id matching (obtained via bioMaRt package)</pre>

Annotation network:

 $\texttt{Annotation_overall <- readRDS("Annotation_overall.RDS")} \quad \textit{\# consists functional annotation of 145 high particles}$

```
gene_top_annotation_graph <- Annotation_overall %>%
    filter(startsWith(Category, c("KEGG", "GO")), !str_detect(Category,
        "CC_DIRECT")) %>% filter(!str_detect(Category,
    "MF DIRECT")) %>% clean names() %>% group by(category) %>%
   filter(row_number() < 6) %>% dplyr::select(term,
    Gene_Count = count, p_value, genes) %>% mutate(genes2 = str_split(genes,
    ", ")) %>% unnest() %>% ungroup %>% mutate(genes2 = str_trim(genes2,
   side = "both"), genes2 = ifelse(endsWith(genes2,
    ","), substr(genes2, 0, (nchar(genes2) - 1)), genes2),
    interaction_type = term, Ensembl_Gene_Id = genes2,
    competing_name = genes2) %>% dplyr::select(interaction = term,
    Ensembl_Gene_Id, interaction_type, competing_name)
## Adding missing grouping variables: 'category'
## Warning: 'cols' is now required when using unnest().
## Please use 'cols = c(genes2)'
top_important <- significant_node_graph %>% bind_rows(gene_top_annotation_graph) %>%
    as_tbl_graph() %>% mutate(type = ifelse(startsWith(name,
    "hsa"), "KEGG", "GO"), type = ifelse(startsWith(name,
    "hsa-"), "miRNA", type), type = ifelse(startsWith(name,
    "ENSG"), "Gene", type)) %>% mutate(centrality = centrality degree(mode = "all"),
   annotation = ifelse(name %in% gene_top_annotation_graph$Ensembl_Gene_Id,
        "top_annotation", "other")) %>% as_tibble() %>%
   filter(annotation == "top_annotation", centrality >
        10) %>% dplyr::select(name) %>% pull()
significant_node_graph %>% bind_rows(gene_top_annotation_graph) %>%
    as_tbl_graph() %>% mutate(type = ifelse(startsWith(name,
    "hsa"), "KEGG", "GO"), type = ifelse(startsWith(name,
    "hsa-"), "miRNA", type), type = ifelse(startsWith(name,
    "ENSG"), "Gene", type)) %>% mutate(centrality = centrality_degree(mode = "all"),
    annotation = ifelse(name %in% gene top annotation graph$Ensembl Gene Id,
        "Enriched Genes", "other")) %>% left_join(hg19,
    by = c(name = "ensembl_gene_id")) %>% filter(type %in%
    c("miRNA", "Gene")) %>% ggraph(layout = "kk") +
    geom_edge_link(colour = "seashell2") + geom_node_point(color = "seashell2") +
    geom_node_point(aes(filter = annotation == "Enriched Genes",
        size = centrality, color = "Enriched Genes"),
        shape = 16) + geom_node_point(aes(filter = type ==
    "miRNA", color = "miRNA"), shape = 17, size = 2) +
    geom_node_point(aes(filter = type == "GO", size = centrality,
        color = "GO"), shape = 18) + geom_node_point(aes(filter = type ==
    "KEGG", size = centrality, color = "KEGG"), shape = 15) +
   theme_graph(base_family = "sans") + geom_node_text(aes(filter = name %in%
   top_important, label = hgnc_symbol), size = 3.5,
   repel = TRUE) + theme(plot.margin = margin(0, 0,
    0, 0, "cm"), legend.position = "none") + guides(size = FALSE,
    shape = FALSE) + scale_color_manual(name = "Node types",
    values = c('Enriched Genes' = "sandybrown", miRNA = "dodgerblue2",
       GO = "green", KEGG = "red"))
```



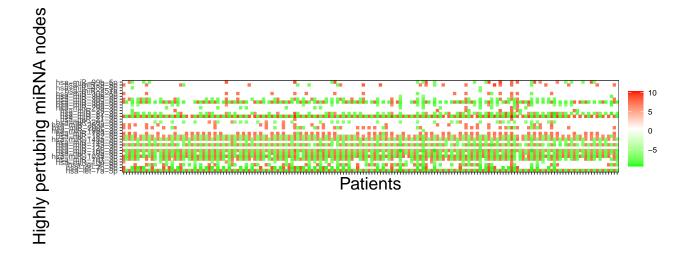
Heat-maps for miRNAs

 $+ Detecting\ perturbing\ miRNAs$

```
mirna_tumor <- mirtarbase_node_perturbations %>% left_join(gene_dist,
    by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
    "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
    effective_nodes$name, "effective", "non_effective")) %>%
    filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type !=
    "Normal", startsWith(name, "hsa")) %>% dplyr::select(file_name,
    name, tissue_type, perturbed_count)
mirna normal <- mirtarbase node perturbations %>% left join(gene dist,
    by = "name") %>% mutate(tissue_type = ifelse(endsWith(file_name,
    "01A"), "Tumor", "Normal"), node_type = ifelse(name %in%
    effective_nodes$name, "effective", "non_effective")) %>%
    filter(node_type == "effective", perturbed_count >
        78, perturbed > 1) %>% filter(tissue_type ==
    "Normal", startsWith(name, "hsa")) %>% dplyr::select(file_name,
    name, tissue_type, perturbed_count)
```

Heat map Figure 4C

```
mirna_tumor%>%
  full_join(mirna_normal, by =c("name", "file_name"), suffix= c("_tumor", "_normal"), fill.na = 0)%>%
  dplyr::select(-3, -5)\%>\%
  mutate(perturbed_count_tumor = ifelse(is.na(perturbed_count_tumor), 1, perturbed_count_tumor),
         perturbed_count_normal = ifelse(is.na(perturbed_count_normal), 1, perturbed_count_normal),
         log_FC= log2(perturbed_count_tumor/perturbed_count_normal))%>%
  ggplot(aes(x = file_name, y = name, fill = log_FC))+
  geom_tile(aes(colour = log_FC, fill = log_FC))+coord_fixed(ratio = 1)+
  theme test()+
  theme(axis.text.x = element_blank(), #element_text(angle = 90, vjust = 0, hjust=0)
       plot.title = element_text(hjust = 0.5, size =20),
       axis.title = element_text(size =20))+
  scale_colour_gradientn(colours = c("green", "white", "red"), aesthetics = c("colour", "fill"), na.valu
  ylab("Highly pertubing miRNA nodes")+
  xlab("Patients")+
  theme( legend.title = element_blank(), panel.background = element_rect(fill = "white"), plot.margin =
```



Heat map Figure 4D

```
axis.title = element_text(size =20))+
scale_colour_gradientn(colours = c("green","red", "black"), aesthetics = c("colour", "fill"), na.valu
ylab("Highly pertubing miRNA nodes")+
xlab("Patients")+
facet_grid(rows = "situ")+
theme( legend.title = element_blank(), panel.background = element_rect(fill = "gray93"), plot.margin
```

