

Supplementary Figures

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17 09 2019

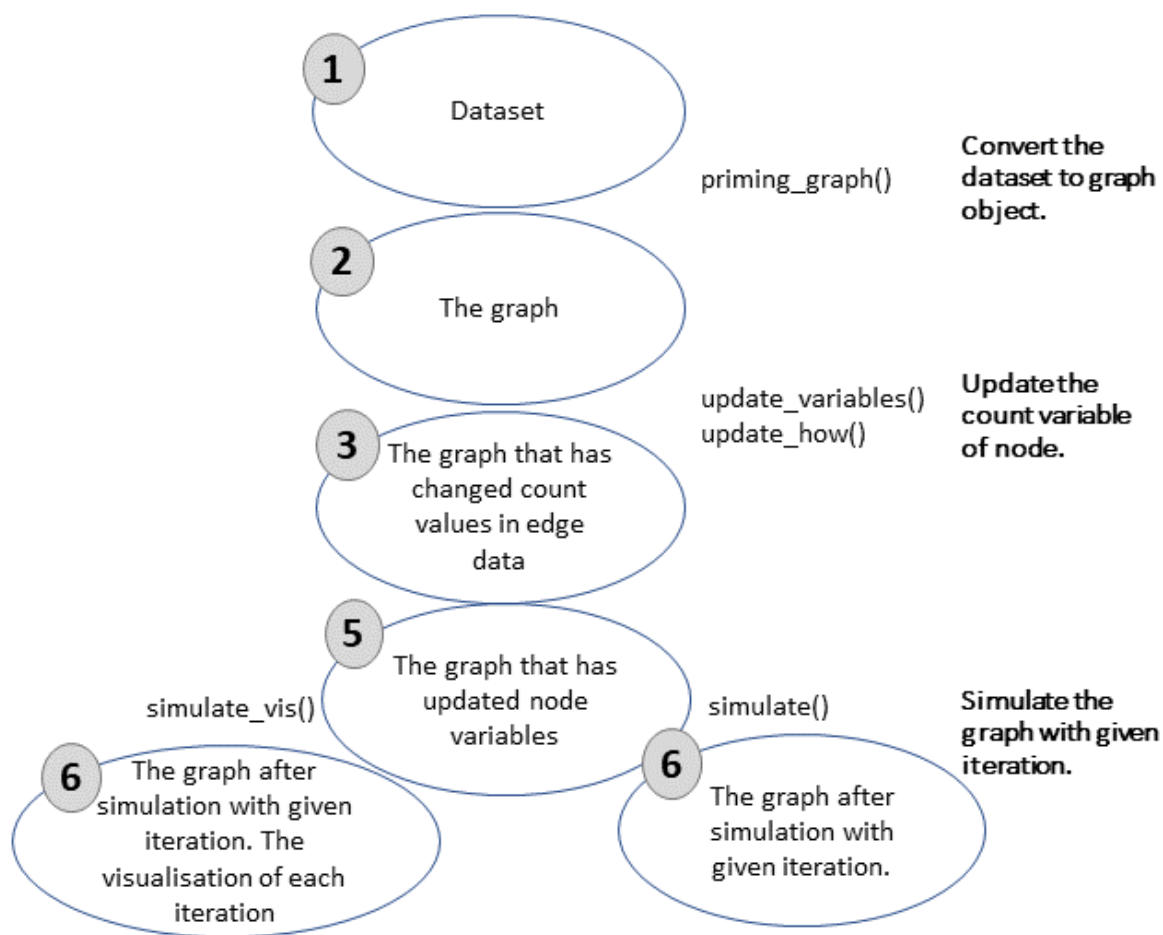


Figure S1: Workflow for simulation of competing endogenous RNA regulations. Graph object in steps 2-6 is saved and updated continuously.

```
#install.packages("devtools")
#devtools::install_github("selcenari/ceRNAetsim")
library(ceRNAetsim)
```

minsamp dataset analysis at lack of interaction factors.

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression)%>%
  vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", title = "Minimal dataset in steady state")
```

Minimal dataset in steady-state condit

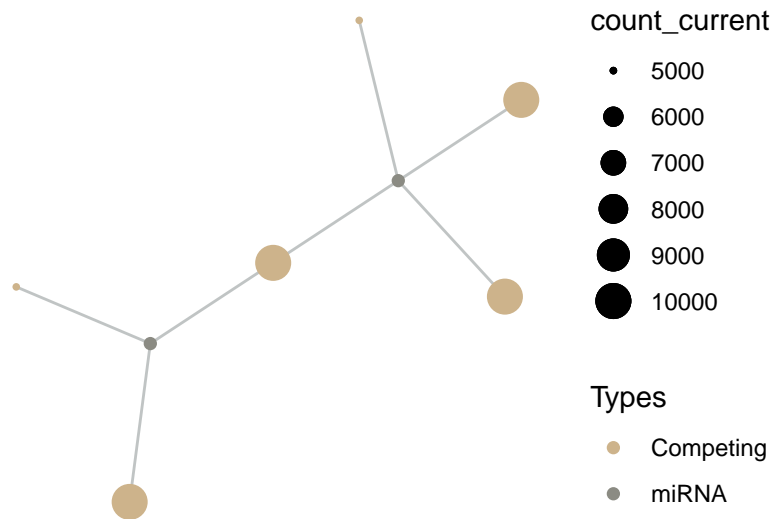


Figure S2: Minimal Dataset in Steady-state

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression)%>%
  update_how("Gene2", 2)%>%
  vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "Gene2")
```

Gene2 Upregulation without interaction

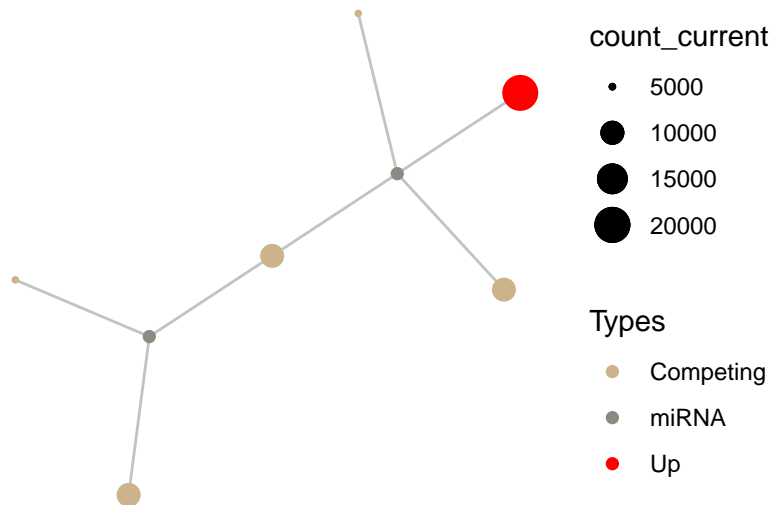


Figure S3: Gene2 Upregulation on Minimal Dataset

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression)%>%
  update_how("Gene2", 2)%>%
  simulate(cycle=1)%>%
  vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", Downregulation = "blue")

priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression)%>%
  update_how("Gene2", 2)%>%
  simulate(2)%>%
  vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", Downregulation = "blue")
```


Minimal dataset with interaction factor

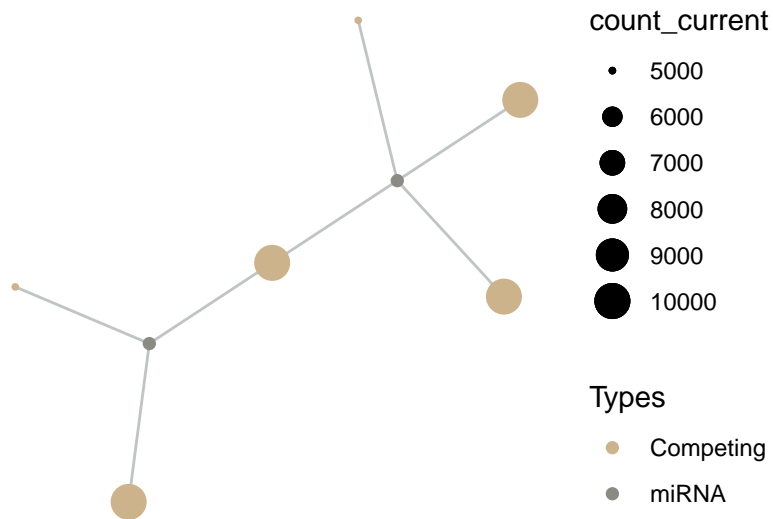


Figure S5: Minimal Dataset with interaction factors in Steady-state

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_factor = Aff_factor)
update_how("Gene2", 2)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "Gene2")
```

Gene2 Upregulation with interaction fa

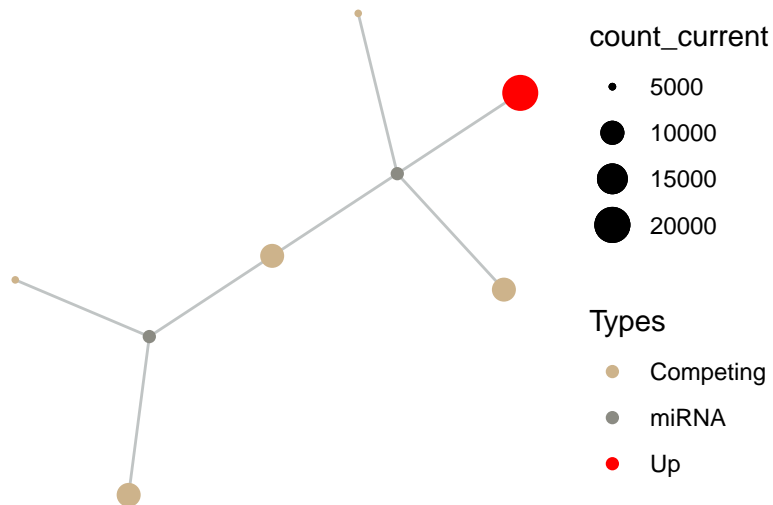


Figure S6: When Gene2 is upregulated on Minimal Dataset with interaction factors

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
update_how("Gene2", 2)%>%
simulate(cycle = 1)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", Downregulat

priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
update_how("Gene2", 2)%>%
simulate(cycle = 2)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", Downregulat
```

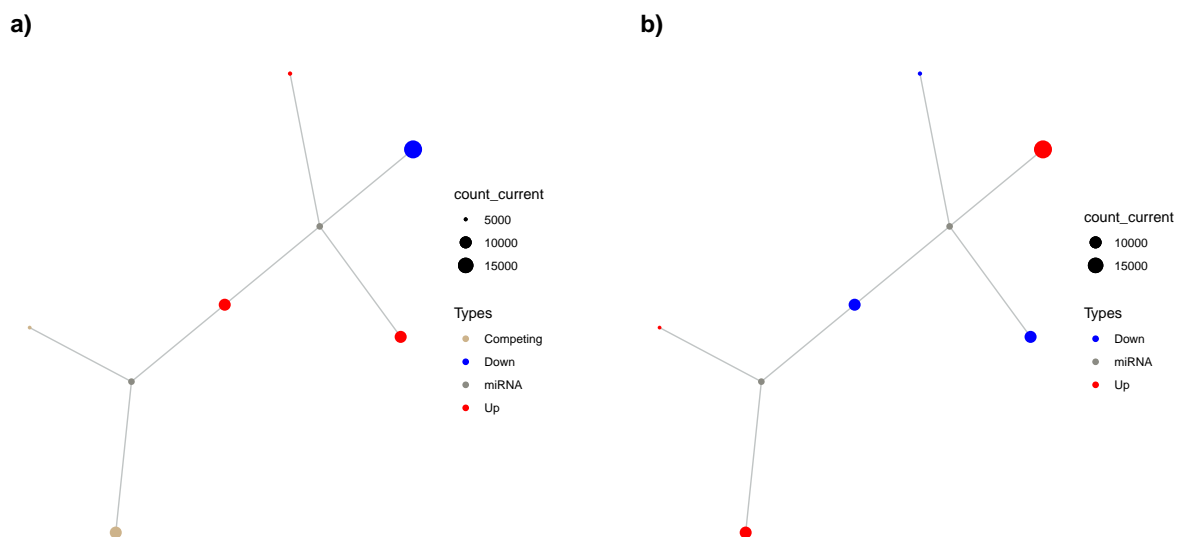


Figure S7: Sequential iteration of minsamp data with interaction factors a)First response of system to Gene2 upregulation (2nd iteration). b)Spreading of perturbation on system (3th iteration)

Common target perturbation in *minsamp* dataset.

```
priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "a)

priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
update_how("Gene4", 2)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "b)

priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
update_how("Gene4", 2)%>%
simulate(cycle=1)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "c)

priming_graph(minsamp, competing_count = Competing_expression, miRNA_count = miRNA_expression, aff_facto
update_how("Gene4", 2)%>%
simulate(cycle=2)%>%
vis_graph(Competing_color = "navajowhite3", mirna_color = "ivory4", Upregulation = "red", title = "d)
```

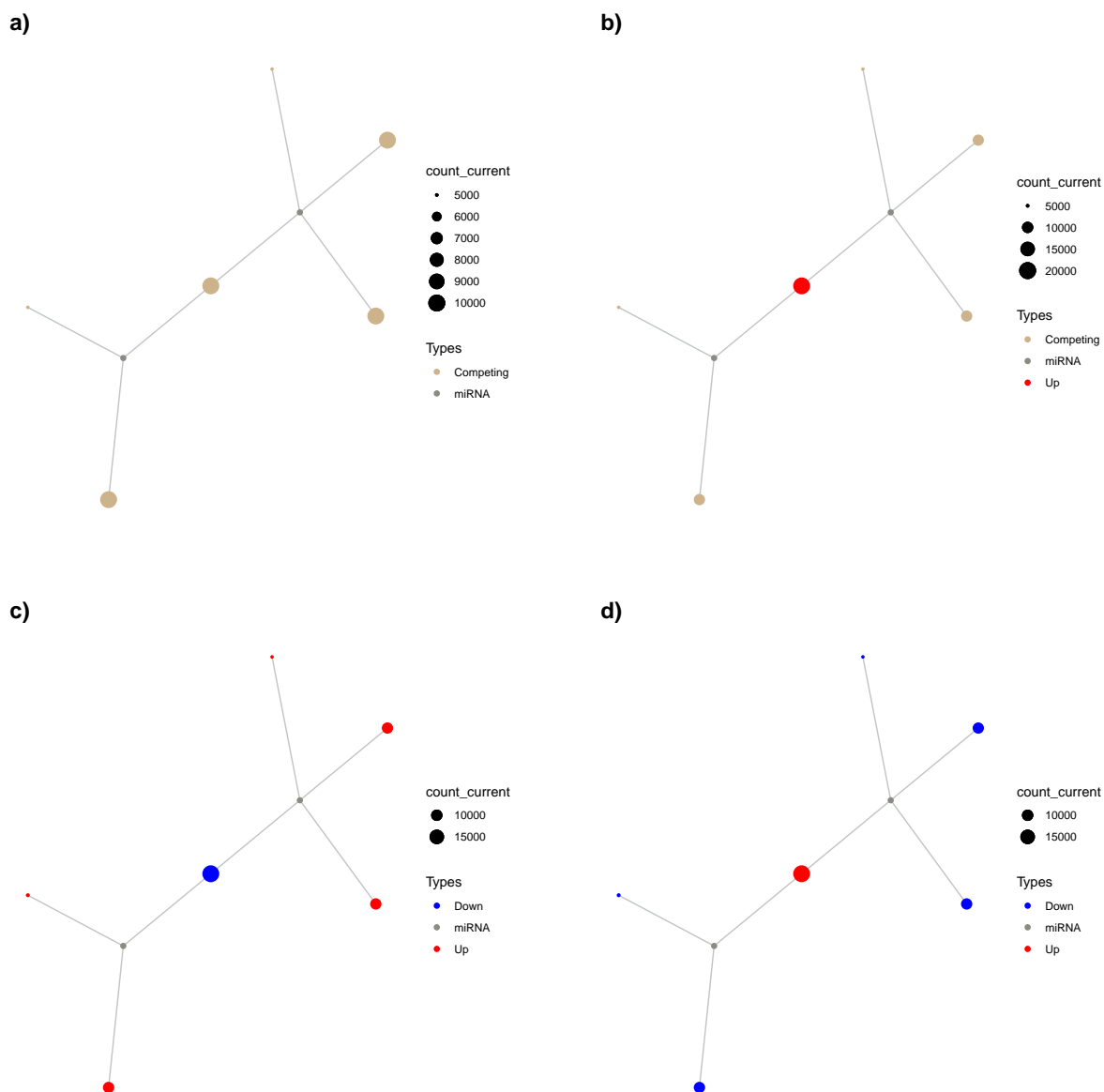


Figure S8: Perturbation of Gene4 on minsamp data with interaction factors. a) Network at steady-state. b) Upregulation of Gene4. c) Primary response of network to upregulation of Gene4. d) Re-regulation of whole nodes on system (3th iteration)

Determination of iteration for Upregulation of ANO6 gene from Breast cancer patient dataset

```
E9GE_mirnagenenormal<-readRDS("data/E9GE_mirnagenenormal.rda")
head(E9GE_mirnagenenormal)
```

```
## # A tibble: 6 x 7
```



```
## Hugo_Symbol miRNA_name mirna_RPM GE_normal energy seed_type_effect
## <chr> <chr> <dbl> <dbl> <dbl> <dbl>
## 1 ENAH hsa-let-7~ 111204. 7540 -22.7 0.01
## 2 GALNT2 hsa-let-7~ 111204. 2824 -20.5 0.14
## 3 RLF hsa-let-7~ 111204. 1144 -18 0.028
## 4 MAST2 hsa-let-7~ 111204. 2640 -22.6 0.24
## 5 DOCK1 hsa-let-7~ 111204. 4826 -20.5 0.01
## 6 ZBTB16 hsa-let-7~ 111204. 315 -24.3 0.14
## # ... with 1 more variable: region_effect <dbl>
```

```
as.data.frame(E9GE_mirnagenenormal)%>%
  priming_graph(competing_count = GE_normal, miRNA_count = mirna_RPM, aff_factor = c(energy, seed_type_e
  update_how("ANO6",3) %>%
  simulate(150) %>%
  find_iteration(limit=1, plot= TRUE) #limit=1 describes the change that is not taken into account.
```

```
## Warning in priming_graph(., competing_count = GE_normal, miRNA_count = mirna_RPM, : First column is p
```

```
## Warning: Removed 150 rows containing missing values (geom_path).
```

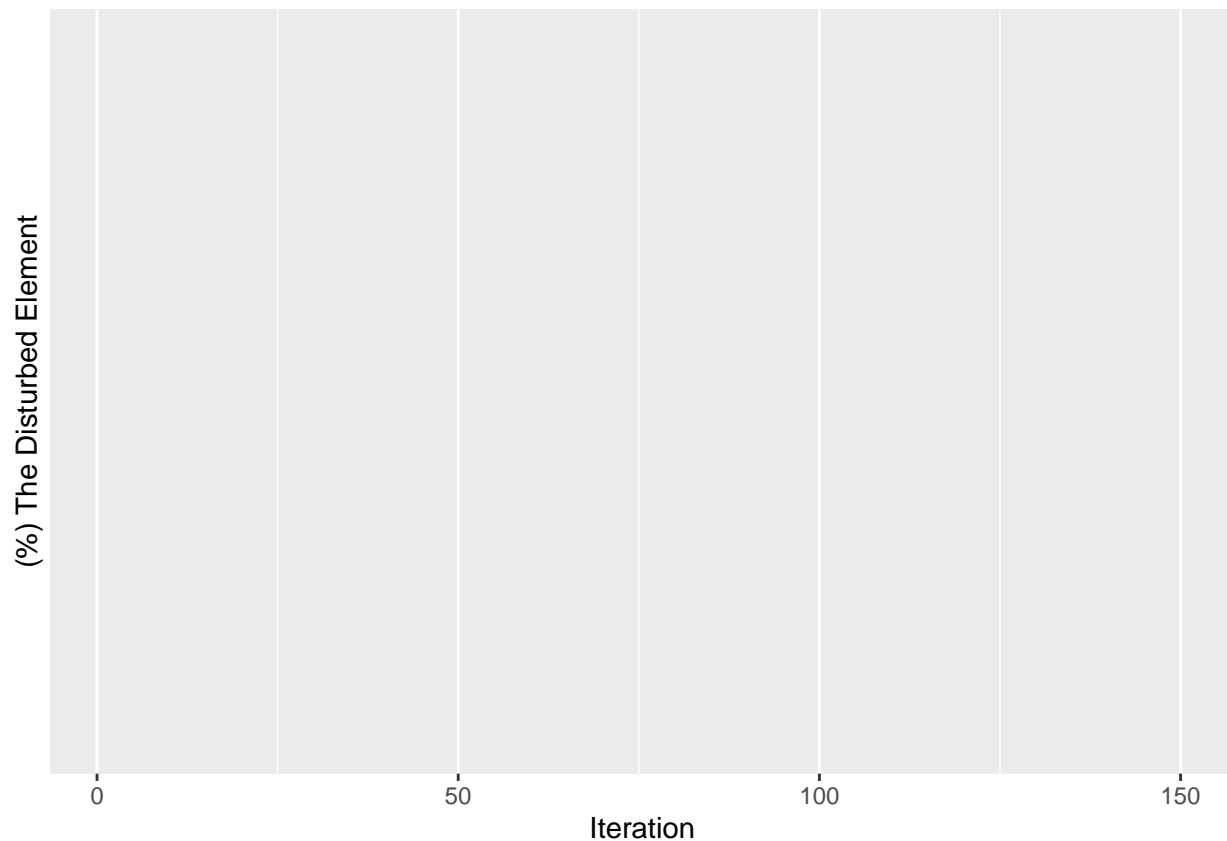


Figure S9: Percentage of affected nodes of each iteration for ANO6 Gene