

Follicular Growth Pattern Disease on Thyroid Fine-needle Aspiration Biopsy

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Thyroid nodules are a common worldwide health problem and a diagnostic challenge for clinicians and cytopathologists. Follicular growth pattern constitutes the majority of thyroid lesions. Thyroid nodules can be neoplastic or non-neoplastic, and neoplastic nodules can be classified as benign, malignant, or gray zone. Gray zone lesions include different benign and malignant entities that might be resulted in unnecessary thyroidectomies with risk of morbidity and higher

health care costs. Depending on the cellularity, most cases might fall into the follicular neoplasia (FN)/ suspicious for FN (SFN) category or follicular lesion of undetermined significance (FLUS) in The Bethesda System for Reporting Thyroid Cytopathology (TBSRTC). Pathologists must be aware of the relationship between this diagnostic category and follow-up patient management and avoid over-diagnosing by mastering the diagnostic criteria.

Thyroid lesions are global health problem, taking into account both non-neoplastic and neoplastic nodules, whereby a correct diagnosis represents an importance for patient management and follow up. Over diagnosis might lead to unnecessary surgeries, causing both morbidity and high health expenditure. On the other hand, underdiagnosis may cause to the exclusion of malignant lesion from follow-up or delayed intervention.¹⁻³

Fine-needle aspiration biopsy (FNAB) is a noninvasive and primary diagnostic method used in the management of thyroid nodules. Numerous studies have reported that more than 70% of all thyroid nodules are reported as benign, 5-10% are malignant, and the remaining 20% represent the so-called gray zone.^{4,5} Most gray zone lesions are follicular-patterned thyroid lesions which malignancy cannot be definitively detected via cytology due to the requirement of evaluating the capsular and vascular status of the pathology specimen. Thus, the FN/SFN category remains a limited technique in screening lesions. Gray zone lesions are segregated in the FN/SFN category of TBSRTC. Identification of nonneoplastic lesions, which can be managed conservatively, and neoplastic lesions,

which will be treated with surgical resection, is possible with the use of FNAB.^{6,7}

HISTORY OF VARIOUS CLASSIFICATION SYSTEMS OF FOLLICULAR PROLIFERATIONS

Several classification systems for reporting thyroid cytopathology have been introduced in recent decades, such as the UK, Japanese, Australian, Danish, and Italian classifications⁸⁻¹⁹. TBSRTC, which is a widely accepted reporting system, was proposed in 2007. It aimed to bring uniformity in the categorization of the wide spectrum of lesions, particularly gray zone lesions.²⁰ Over the years, the need for the regulation of TBSRTC arose, and the renewed system guidelines were published in 2017.^{7,21,22} A notable flaw of this system was an implied risk of malignancy for each diagnostic category combined with the lack of correct recommendations concerning the management of patients.

TBSRTC proposed three categories for indeterminate cytology: atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS), FN/SFN, and suspicious



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for malignancy (SM)²³⁻²⁶. The literature revealed the difficulties in AUS/FLUS and FN/SFN diagnoses that might lead to overdiagnosis and overtreatment, such as lobectomy or bilateral total thyroidectomy.²⁷⁻³³ These diagnostic problems in TBSRTC led to new pursuits and the application of ancillary techniques, such as immunocytochemistry (ICC) and molecular analysis, on cytology samples. Currently, these techniques can be performed in all cytology preparations. However, the use of liquid-based cytology (LBC) techniques might facilitate their application in exceeding the difficulties faced with conventional cytology.³⁴⁻⁴⁰ The cytopathologic features, as well as the differential diagnoses of the follicular lesions, will be discussed in this article using TBSRTC.

FOLLICULAR PATTERN LESIONS IN THE BETHESDA SYSTEM FOR REPORTING THYROID CYTOPATHOLOGY

Follicular neoplasms: follicular adenoma (FA) and follicular carcinoma (FC) are generally classified into the FN/SFN category of TBSRTC. In addition, the FN/SFN category includes a few cases of follicular variants of papillary carcinoma and noninvasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP), which cannot be diagnosed as malignant cytologically because they do not definitively show the nuclear features of papillary thyroid cancer (PTC). In the differential diagnosis of this category, the follicular pattern of medullary carcinoma, dyshormonogenetic goiter, parathyroid lesions, and poorly differentiated carcinoma should be considered^{41,42} (Table 1).

CYTOMORPHOLOGIC FEATURES OF THYROID FOLLICULAR LESIONS AND DIFFERENTIAL DIAGNOSIS

Thyroid aspiration slides should be evaluated with all elements together on the microscopic area. When the slides are predominantly composed of micro follicles, the background of the slides and the accompanying components should be carefully analyzed. In the presence of the thyroid follicular epithelial cell sheets in a honeycomb arrangement, the presence of abundant colloid, numerous histocytes, lymphocytes, stromal fragments, and cystic changes mostly indicate benign entities, whereas the uniform proliferation of micro follicles with scant colloid suggests neoplastic lesions.⁴¹⁻⁴⁶

TABLE 1. Differential Diagnosis of Follicular Pattern Thyroid Lesions in Fine Needle Aspiration Biopsy

Follicular pattern thyroid lesions in fine-needle aspiration biopsy
■ Adenomatous/hyperplastic nodules
■ Follicular adenoma
■ Follicular carcinoma
■ Follicular variant of papillary thyroid carcinoma
■ Noninvasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP)
■ Follicular pattern of medullary carcinoma
■ Dyshormonogenetic goiter
■ Parathyroid lesions
■ Poorly differentiated carcinoma

Cytological evaluation of the FN/SFN category is usually accompanied by thyroid follicular lesions. Although the diagnostic criteria of follicular lesions on FNA are not clearly defined, cellular aspirates with scanty amount of colloid, which are composed of follicular epithelial cell groups with microfollicular patterns >50-70%, are diagnosed as FN/SFN.¹ This category likewise has two alternative names. FN and SFN are synonymous terms and should not be used like two different diagnostic terms. Generally, SFN is preferred by most laboratories because histopathologic studies supported that a significant number of cases (up to 35%) prove not to be neoplasms but rather hyperplastic proliferative lesions, which are most commonly nodular goiter.⁴³⁻⁴⁶

Notably, the accurate definition of the microfollicular/macropolyfollicular terms is important. Microfollicles are composed of less than 15 overlapping thyroid follicular epithelial cells, which form a circle that is at least two-thirds complete. Macropolyfollicles are composed of small or large flat groups/sheets or even rows of follicular epithelial cells. Cell circles and overlapping are not characteristic findings for macropolyfollicles.⁴⁷ An important diagnostic clue is not to call the crowded small groups, which create a pseudo microfollicular appearance in a clot, such as microfollicles in bloody smears. In these slides, observing microfollicles out of the clot may be helpful for diagnosis. Sometimes, sampling from the microfollicular/cellular areas of benign nodules can lead to a cytological misdiagnosis of FN/SFN. Reactive/degenerative nuclear features that are not distinguishable but arouse suspicion may also lead to the overdiagnosis of PTC. The 2017 TBSRTC supported a modification to the definition and diagnostic criteria for the FN/SFN category in light of NIFTP. The new description reads as follows: "Follicular-patterned cases with mild nuclear changes (increased nuclear size, nuclear contour irregularity, and/or chromatin clearing) can be classified as FN/SFN so long as true papillae and intranuclear pseudoinclusions are absent, a note that some nuclear features raise-the possibility of an FVPTC or NIFTP, can be included".^{7,48}

In the following parts of this article, the cytological features and differential diagnoses of the major follicular growth pattern disease of the thyroid will be discussed.

Adenomatous, Hyperplastic Nodules

These lesions show single-layer groups, honeycomb-pattern follicles, and/or single thyroid follicular epithelial cells. The presence of abundant watery colloid is an important feature of smears. Follicular epithelial cells have small, round nuclei and scanty, abundant, mildly vacuolated, or oncocytic cytoplasm (Figure 1). Although they are simple diagnostic criteria, diagnostic difficulties are sometimes experienced in the FNA of hyperplastic nodules. In relation to this, sampling from the microfollicular/cellular areas of benign nodules can be a challenging condition for a cytopathologist, and such cases can be misdiagnosed as FN/SFN.^{43,49-53} It should also be noted that the nuclei of follicular lesions/neoplasms are expected to be round and with coarsely granular chromatin (rather hyperchromatic), which might be slightly larger than regular thyroid follicular epithelial cells.

The presence of abundant loose/watery colloids can help to the cytopathologist for away from over-diagnosis⁴⁸⁻⁵³.

Follicular Adenomas/Follicular Carcinomas

FAs/FCs are the most important and classical categories of follicular pattern lesions. Cytomorphological features do not allow the differentiation of FAs from FCs. The main differential diagnosis criteria of these lesions are based on capsular and vascular invasion, which can be diagnosed only with a thorough histologic evaluation of the nodule. With this knowledge, we are faced with the fact that FN is a screening test rather than a diagnostic test in this situation^{41,43,54}.

These lesions exhibit the main findings of the FN/FNS category. The cytological definition of FA/FC is “cellularity-rich aspirates consisting of follicular cells with crowding or microfollicle formation on colloid poor background.” The nuclei of follicular epithelial cells are round, slightly larger than normal thyroid follicular epithelial cells, and hyperchromatic with coarsely granular chromatin (Figure 2). Due to the inability to distinguish FAs from FCs with these findings, many studies have been explored clinicopathologic findings, as well as patient demographics, to increase diagnostic accuracy. Most of these studies have suggested that nodules larger than 3-4 cm predict malignancy. These findings

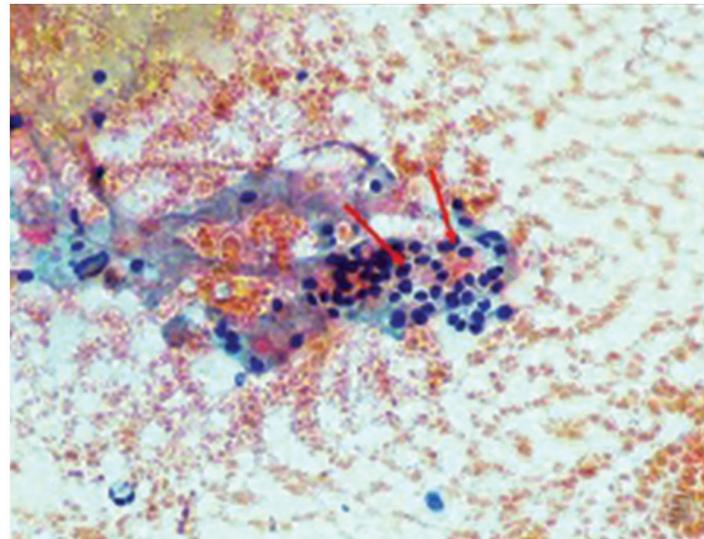
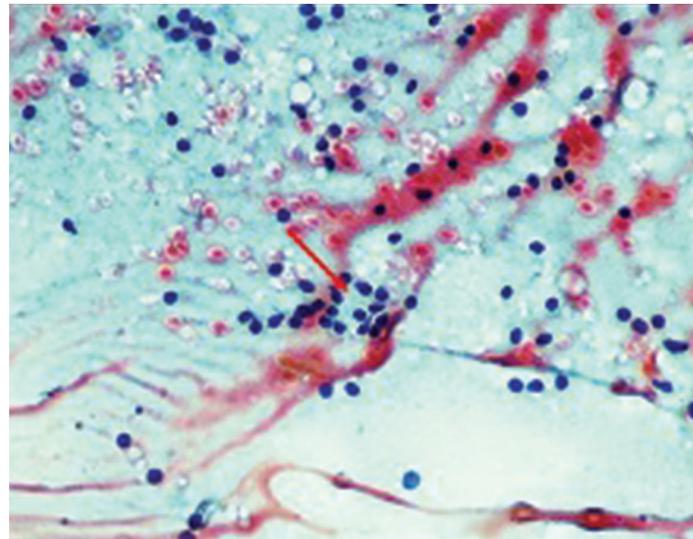


FIG. 1. Hyperplastic nodule on fine-needle aspiration biopsy. Benign follicle epithelial cells with few follicular structures (arrows) near colloidal material (PAP x200)

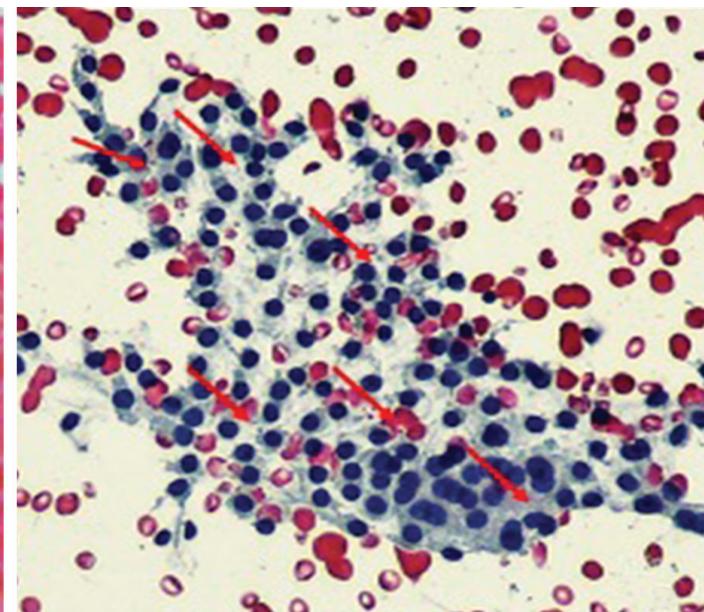
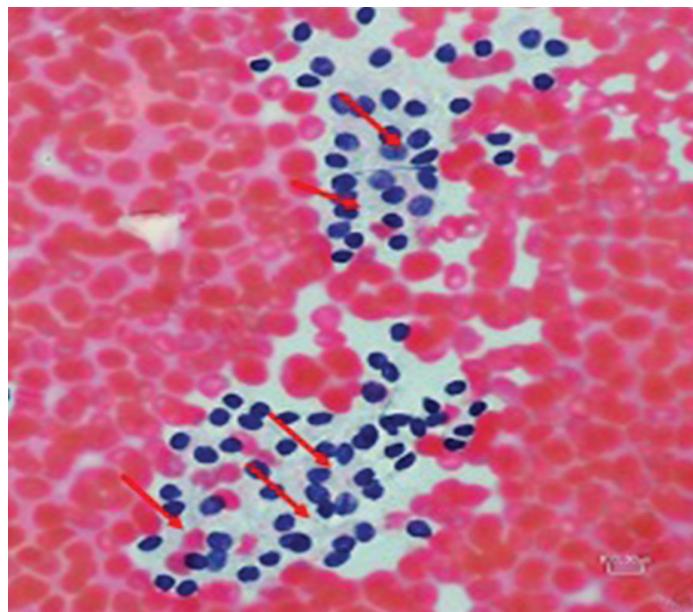


FIG. 2. Follicular neoplasms (FN)/suspicious for FN (SFN) with numerous microfollicles (arrows) on fine-needle aspiration biopsy (PAP x400)

prove once again the importance of evaluating cases with clinical findings in the pathology routine.^{7,51-54}

Although the definition of follicular lesions in FN remains unclear, cellular aspirates consisting of >50-70% microfollicles and insufficient colloid amount are considered FN/SFN.⁵⁵ Microfollicles show a uniform appearance; other features of these lesions include trabeculae, crowded cell groups, and isolated single cells in the background.^{56,57} Cases with scanty colloids with a macro/microfollicular pattern are not covered by this term even if the smear is hypercellular. These lesions should be considered benign nodules or AUS/FLUS depending on the amount of colloid, microfollicle ratio, and cellularity.^{7,43}

Follicular Variant of Papillary Thyroid Carcinoma/NIFT

The exclusion of FVPTC is the most important clue for the differential diagnosis of the FN/SFN category. In smears, these cases can have an abundance of microfollicles or monolayer fragments mimicking a follicular neoplasm. Undoubtedly, the most important criteria are the evaluation of specific nuclear features of PTC, such as nuclear enlargement, membrane irregularities, crowding, elongation, chromatin clearing, pseudo-inclusions, and grooves (Figure 3). However, unfortunately FVPTCs do not show always the characteristic nuclear features of PTC and present with subtle nuclear findings. In case of determination of nuclear features suspicious for PTC, should be reported as SM regardless of their prominent microfollicular pattern.^{13,24,45} But, the presence of subtle nuclear changes and microfollicles together can be misleading for cytological examination, and ignoring of this nuclear changes can result in an FN/SFN diagnosis. Thus, approximately 15-20% of cases diagnosed as FN/SFN later turn out to be FVPTC.^{51,55} The relationship between FVPTC and NIFT-P can be compared to FC and FA. Encapsulation as a differential diagnostic feature of these entities cannot be given by cytomorphology. In case of cytologic features suggestive of FVPTC/NIFT, optional notes in cytological diagnosis may be used to acknowledge NIFT: "Although the

architectural features suggest a follicular neoplasm, some nuclear features raise the possibility of an invasive FVPTC or its recently described indolent counterpart, NIFT; definitive distinction among these entities is not possible on cytological material".⁵⁶

Medullary Thyroid Carcinoma

Medullary thyroid carcinoma (MTC) is an aggressive neoplasm of the thyroid, which is typically characterized by many single cells and loose fragments; follicular formation is an unexpected finding. However, MTC may reveal nested/trabecular/loose microfollicular patterns and could be misdiagnosed as follicular neoplasm. Some tumors may show oncocytic cytoplasm mimicking oncocytic follicular neoplasm (Figure 4). For differential diagnosis, a predominantly dispersed cell pattern, presence of both plasmacytoid and spindle type cells, nuclear neuroendocrine type chromatin, binucleation, and eccentric nuclei can help identify MTC. If it can be identified, the presence of amyloid is a supportive diagnostic finding. ICC studies (calcitonin, CEA, TTF-1 etc.) may help provide the final diagnosis. Clinically requesting serum calcitonin analysis is important clue.^{1,56,57}

Dyshormonogenetic Goiter

These lesions are characterized by a microfollicular pattern and a lack of colloid. Therefore, they should be considered in the differential diagnosis of FN/FNS. The presence of congenital hypothyroidism is supported by dyshormonogenetic goiter and clinical history. Empty follicles (microfollicles without intrafollicular colloid) and anisokaryosis with bizarre cells are typical cytopathologic findings for dyshormonogenetic goiter.⁵⁸

Intrathyroidal Parathyroid Adenoma

Parathyroid adenomas (PAs) may show sheets/clusters or microfollicles and many bare nuclei. Due to these findings, patients are misdiagnosed as FN/FNS. Occasionally colloid-like parathyroid secretions can be seen in PA smears. Parathyroid cells typically reveal a neuroendocrine-type nuclear chromatin

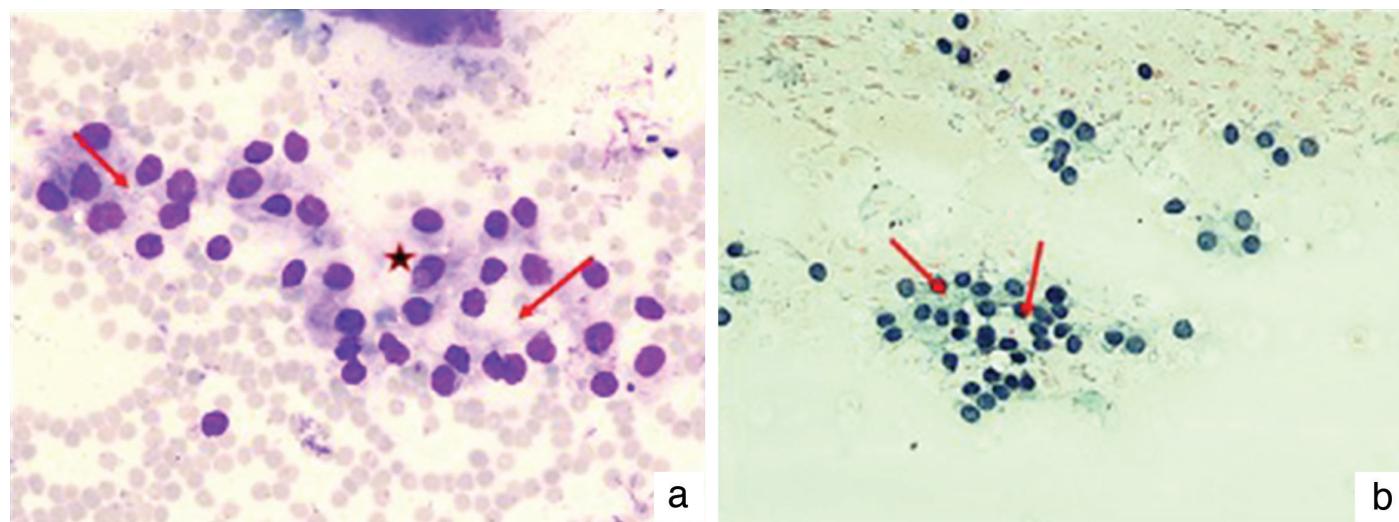


FIG. 3. Follicular variant papillary thyroid carcinoma with few follicular structures on fine-needle aspiration biopsy (arrows: microfollicles, asterisk: intranuclear pseudo inclusion), (a. Giemsa x400, b. PAP x200)

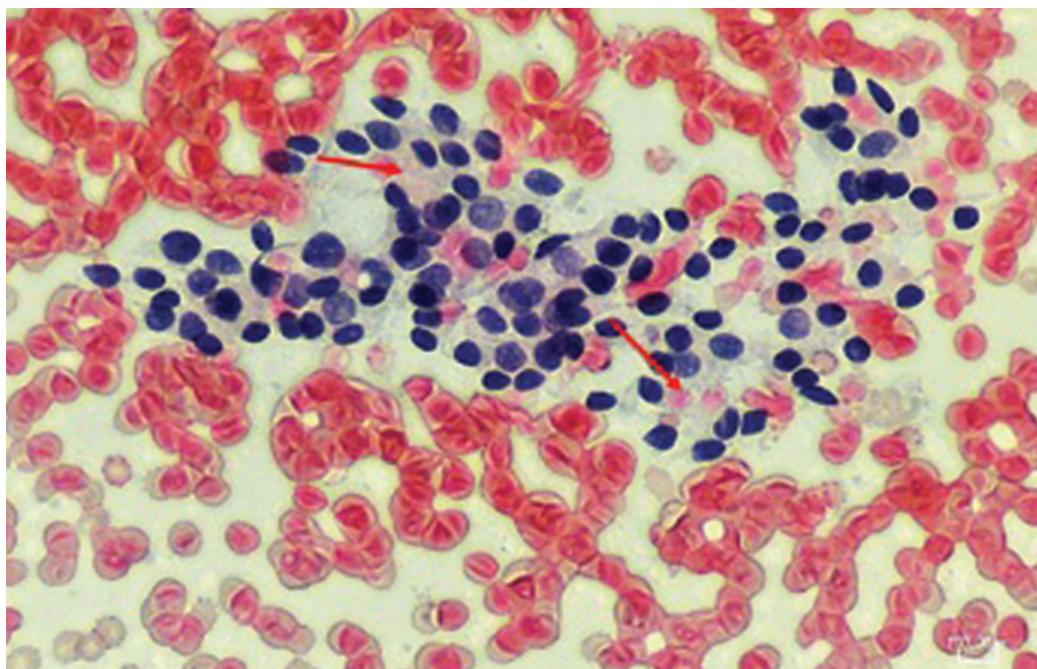


FIG. 4. Medullary carcinoma with few follicular structures (arrows) on fine-needle aspiration biopsy (PAP x400)

and prominent vascular network. Performing synchronous PTH assays on FNA and immunohistochemical studies on cell blocks are important diagnostic assistive techniques for PAs.⁵⁹

Poorly Differentiated Thyroid Carcinoma

Poorly differentiated thyroid carcinomas (PDTCs) are aggressive tumors derived from thyroid follicular epithelium. These thyroid malignancies commonly represent naked nuclei and solid/trabecular/insular patterns; however, microfollicles may sometimes be seen. If the microfollicular pattern is dominant, it frequently causes a misdiagnosis of FN/SFN, which can lead to inadequate patient management. Careful microscopic evaluation of nuclear details is important for differential diagnosis. The cells of PDTC are characterized by small, convoluted nuclei, speckled chromatin, and scant cytoplasm on smears. Nuclear pleomorphism, coarser chromatin, and anisokaryosis can be detected clearly in PDTCs at a high magnification.^{60,61}

ANCILLARY TECHNIQUES USED FOR DIFFERENTIAL DIAGNOSIS OF FOLLICULAR LESIONS

Immunocytochemistry

Diagnosing FNs by FNA is the main problem because one cannot evaluate the capsule and vascular status; thus, follicular-patterned dominant FNAs are the candidates for ancillary tests.⁶²⁻⁶⁷ The current version of guidelines for patients with thyroid nodules and differentiated thyroid cancer published by the American Thyroid Association suggests using these tests, and this approach has become popular among cytopathologists.^{10,68} However, the recommendation of TBSRTC or many other reports does not include any information about thyroid ICC and other molecular tests in the group of FN/FNS. The low specificity of ICC and possible false-positive and

false-negative results of ICC have been claimed as an important reason.^{69,70} It should be noted that LBC methods eliminate this problem to a great extent.^{71,72} There are many studies stating that ICC can be used in the differentiation of benign/malignant lesions as well as in the differentiation of lesions derived from thyroid follicle epithelial cells and C cells.⁷³⁻⁷⁷ Various studies that evaluated different antibodies supported the finding that HBME-1 and galectin-3 have the highest specificity and sensitivity, which is highly suggestive of thyroid malignancies. However, none of these antibodies presented enough sensitivity and specificity to be used in routine practice.⁷⁸⁻⁸³ In recent years, a specific monoclonal antibody (Ve1) against the mutated V600E BRAF protein began to be used for differential diagnosis and patient management. Few reports on the cytological application-assisted application of BRAF Ve1 antibody suggest that immunohistochemistry can be performed more reliably in cell block preparations, and false-positive results in direct smears may limit the benefit; thus, caution should be exercised in interpretation. VE1 expression may be an alternative method for detecting BRAF (V600E) when molecular detection is unavailable. Some studies suggest that the VE1 antibody can be used as a first-line approach for the evaluation of the BRAF mutation and case selection for molecular analysis; however, others do not support this approach.⁸⁴⁻⁹⁰

Molecular Tests

The application of molecular tests on cytology samples can contribute to the diagnosis and prediction of prognosis, and these findings may represent a new era for new targeted therapies aimed at individual molecular targets.^{10,62,63,68,84-93}

The application of molecular testing to indeterminate thyroid FNAs for detecting specific somatic mutations, gene rearrangements, or microRNA (miRNA) expression profiles is a new era for thyroid cytology since the last 20 years. These methods have a high predictive value for benign and malignant thyroid lesions.^{10,62,94}

Recent data from various studies have shown that molecular alterations of specific pathways play a pivotal role in thyroid carcinogenesis; thus, they may be used as markers of malignancy. The main mechanism for thyroid carcinogenesis works through the MAPK pathway, which is associated with cell proliferation, differentiation, and apoptosis. The BRAF gene results in the constitutive activation of MAPK and PI3K/AKT pathways and plays a major role in carcinogenesis. In particular, papillary carcinoma, the most common thyroid malignancy, may carry BRAF, RET/PTC, or NRAS mutations in 70% of cases.⁹⁵⁻¹⁰⁰

The presence of the V600E BRAF mutation is associated with higher aggressiveness and less favorable prognosis.¹⁰¹⁻¹⁰⁴ In our institutional experience, histopathologic features, presence of tumor capsular invasion, extrathyroidal extension, absence of pathologically detected lymphocytic thyroiditis, and radioiodine I-131 treatment were significantly higher in patients with the BRAF V600E point mutation (+) detected with DNA sequencing molecular tests.¹⁰⁵ Ancillary molecular tests can be done in-house using the commonly altered genes (BRAF, RAS, RET/PTC, and PPARc/PAX8), but there are three commercially available and established testing panels (Afirma gene expression classifier (GEC), ThyroSeq v.2, and ThyGenX/ThyraMIR). The Afirma GEC can be used to rule out malignancy for the AUS/FLUS and SFN/FN categories and has a high negative predictive value. In contrast, ThyroSeq v.2 and multipanel testing with ThyGenX/ThyraMIR can be used to rule in as well as rule out malignancy with high positive and negative predictive values.^{106,107}

In recent years, molecular-based studies have been focused on the effects of epigenetic changes on gene transcription and their relationship with oncogenic pathway activation in thyroid carcinomas. Data supported that TCs are strongly influenced by epigenetic alterations at the differentiation and proliferation mechanisms. The PTEN promoter gene, which presents with hypermethylation in ~50% of PTCs and nearly 100% of FTCs and FAs, is an important example. In contrast, activating mutations of BRAF in PTCs were linked to altered methylation of other genes (TIMP3, SLC5A8, DAPK, and RAR β 2) associated with aggressive behavior. The promoter methylation involving the RAS association family 1A (RASSF1A) tumor-suppressor gene was seen in approximately 30% of benign and malignant thyroid tumors, including anaplastic thyroid carcinoma; this finding has a role in regulating several key cell processes, suggesting that this change may occur early in tumorigenesis.¹⁰⁸⁻¹¹⁸

The FN/SFN category is the gray zone of thyroid cytology, and intra/interobserver variability is a diagnostic challenge.

When microfollicles dominate FNA, the background of aspiration should be evaluated carefully for the presence of sheets/macropollollicles, colloid, lymphocytes, and blood.

The presence of PTC nuclear features and other atypical nuclei should be considered and avoided while downgrading nuclear atypia.

Although FNA is a powerful diagnostic method in thyroid cytology, limitations in this diagnostic group should be considered.

Ancillary techniques, especially molecular tests, can be used in differential diagnosis and guiding treatment.

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REFERENCES

1. Cibas ES, Ali SZ. The 2017 Bethesda System for Reporting Thyroid Cytopathology. *Thyroid*. 2017;27:1341-1346. [\[CrossRef\]](#)
2. Trimboli P, Crescenzi A, Giovanella L. Performance of Italian Consensus for the Classification and Reporting of Thyroid Cytology (ICCRCT) in discriminating indeterminate lesions at low and high risk of malignancy. A systematic review and metaanalysis. *Endocrine*. 2018;60:31-35. [\[CrossRef\]](#)
3. Rossi ED, Morassi F, Santeusanio G, Zannoni GF, Fadda G. Thyroid fine needle aspiration cytology processed by ThinPrep: an additional slide decreased the number of inadequate results. *Cytopathology*. 2010;21:97-102. [\[CrossRef\]](#)
4. Duncan LD, Forrest L, Law WM Jr, Hubbard E, Stewart LE. Evaluation of thyroid fine-needle aspirations: can ThinPrep be used exclusively to appropriately triage patients having a thyroid nodule? *Diagn Cytopathol*. 2011;39:341-348. [\[CrossRef\]](#)
5. Ravetto C, Colombo L, Dottorini ME. Usefulness of fine-needle aspiration in the diagnosis of thyroid carcinoma: a retrospective study in 37,895 patients. *Cancer*. 2000;90:357-363. [\[CrossRef\]](#)
6. Poller DN, Ibrahim AK, Cummings MH, Mikel JJ, Boote D, Perry M. Fine-needle aspiration of the thyroid. *Cancer*. 2000;90:239-244. [\[CrossRef\]](#)
7. Siyed AZ, Cibas ES. The Bethesda System for Reporting Thyroid Cytopathology: Definitions, Criteria and Explanatory Notes, 2nd ed. Cham ham: Springer International Publishing, Switzerland; 2018. [\[CrossRef\]](#)
8. Guidelines of the Papanicolaou Society of Cytopathology for the examination of fine-needle aspiration specimens from thyroid nodules. The Papanicolaou Society of Cytopathology Task Force on Standards of Practice. *Mod Pathol*. 1996;9:710-715. [\[CrossRef\]](#)
9. American Thyroid Association (ATA) Guidelines Taskforce on Thyroid Nodules and Differentiated Thyroid Cancer, Cooper DS, Doherty GM, et al. Revised American Thyroid Association management guidelines for patients with thyroid nodules and differentiated thyroid cancer. *Thyroid*. 2009;19:1167-1214. [\[CrossRef\]](#)
10. Haugen BR, Alexander EK, Bible KC, et al. 2015 American Thyroid Association management guidelines for adult patients with thyroid nodules and differentiated thyroid cancer: The American Thyroid Association Guidelines Task Force on Thyroid Nodules and Differentiated Thyroid Cancer. *Thyroid*. 2016;26:1-133. [\[CrossRef\]](#)
11. Gharib H, Papini E, Valeavi R, et al. American Association of Clinical Endocrinologists and Associazione Medici Endocrinologi medical guidelines for clinical practice for the diagnosis and management of thyroid nodules. *Endocr Pract*. 2006;12:63-102. Erratum in: *Endocr Pract*. 2008;14:802-803. [\[CrossRef\]](#)
12. Cibas ED, Ali SZ; NCI Thyroid FNA State of the Science Conference. The Bethesda system for reporting thyroid cytopathology. *Am J Clin Pathol*. 2009;132:658-665. [\[CrossRef\]](#)
13. Baloch ZW, LiVolsi VA, Asa SL, et al. Diagnostic terminology and morphologic criteria for cytologic diagnosis of thyroid lesions: a synopsis of the National Cancer Institute fine-needle aspiration state-of-science conference. *Diagn Cytopathol*. 2008;36:425-437. [\[CrossRef\]](#)

14. Perros P, Clarke SE, Franklyn J. British Thyroid Association, Royal College of Physicians. Guidelines for the management of thyroid cancer, 2nd ed. Report of the Thyroid Cancer Guidelines Update Group. London: RCP, 2007. [\[CrossRef\]](#)
15. Lobo C, McQueen A, Beale T, Kocjan G. The UK royal college of pathologists thyroid fine-needle aspiration diagnostic classification is a robust tool for the clinical management of abnormal thyroid nodules. *Acta Cytol.* 2011;55:499-506. [\[CrossRef\]](#)
16. Perros P, Boelaert K, Colley S, et al. Guidelines for the management of thyroid cancer. *Clin Endocrinol (Oxf).* 2014;81:1-122. [\[CrossRef\]](#)
17. Fadda G, Basolo F, Bondi A, et al. Cytological classification of thyroid nodules. Proposal of the SIAPC-IAP Italian consensus working group. *Pathologica.* 2010;102:405-408. [\[CrossRef\]](#)
18. Nardi F, Basolo F, Crescenzi A, et al. Italian consensus for the classification and reporting of thyroid cytology. *J Endocrinol Invest.* 2014;37:593-599. [\[CrossRef\]](#)
19. Royal College of Pathologists of Australasia. Thyroid cytology structured reporting protocol. 2 nd ed. 2019. Available at: <https://www.rcpa.edu.au/getattachment/92c429b1-605f-4358-bbeb-26f66084ced9/Protocol-thyroid-FNA-cytology.aspx>. [\[CrossRef\]](#)
20. The Bethesda System for reporting thyroid cytopathology. Syed AZ, Cibas ES, editors. New York: Springer; 2010.
21. Rossi ED, Pusztaszeri M, Schmitt F, Bongiovanni M, Chandra A, Faquin WC. Thyroid FNA: international perspectives from the European congress of cytopathology: can we cross the bridge of classifications? *Cancer Cytopathol.* 2015;123:207-211. [\[CrossRef\]](#)
22. Poller DN, Baloch ZW, Fadda G, et al. Thyroid FNA: New classifications and new interpretations. *Cancer Cytopathol.* 2016;124:457-466. [\[CrossRef\]](#)
23. Bongiovanni M, Spitale A, Faquin WC, Mazzucchelli L, Baloch ZW. The Bethesda system for reporting thyroid cytopathology: a meta-analysis. *Acta Cytol.* 2012;56:333-339. [\[CrossRef\]](#)
24. Boonyaarnnate T, Olson MT, Ali SZ. 'Suspicious for a follicular neoplasm' before and after the Bethesda System for Reporting Thyroid Cytopathology: impact of standardized terminology. *Acta Cytol.* 2013;57:455-463. [\[CrossRef\]](#)
25. Olson MT, Boonyaarnnate T, Altinboga AA, Ali SZ. 'Suspicious for papillary thyroid carcinoma' before and after The Bethesda System for Reporting Thyroid Cytopathology: impact of standardized terminology. *Acta Cytol.* 2014;58:15-22. [\[CrossRef\]](#)
26. Baloch ZW, Mandel SJ, LiVolsi VA. Are we ready to modify the Bethesda thyroid fine-needle aspiration classification scheme? *Cancer Cytopathol.* 2013;121:171-174. [\[CrossRef\]](#)
27. Nagarkatti SS, Faquin WC, Lubitz CC, et al. Management of thyroid nodules with atypical cytology on fine-needle aspiration biopsy. *Ann Surg Oncol.* 2013;20:60-65. [\[CrossRef\]](#)
28. Olson MT, Clark DP, Erozan YS, Ali SZ. Spectrum of risk of malignancy in subcategories of 'atypia of undetermined significance'. *Acta Cytol.* 2011;55:518-525. [\[CrossRef\]](#)
29. Horne MJ, Chhieng DC, Theoharis C, et al. Thyroid follicular lesion of undetermined significance: Evaluation of the risk of malignancy using the two-tier sub-classification. *Diagn Cytopathol.* 2012;40:410-415. [\[CrossRef\]](#)
30. Hyeon J, Ahn S, Shin JH, Oh YL. The prediction of malignant risk in the category "atypia of undetermined significance/follicular lesion of undetermined significance" of the Bethesda System for Reporting Thyroid Cytopathology using subcategorization and BRAF mutation results. *Cancer Cytopathol.* 2014;122:368-376. [\[CrossRef\]](#)
31. Dincer N, Balci S, Yazgan A, et al. Follow-up of atypia and follicular lesions of undetermined significance in thyroid fine needle aspiration cytology. *Cytopathology.* 2013;24:385-390. [\[CrossRef\]](#)
32. Gocun PU, Karakus E, Bulutay P, Akturk M, Akin M, Poyraz A. What is the malignancy risk for atypia of undetermined significance? Three years' experience at a university hospital in Turkey. *Cancer Cytopathol.* 2014;122:604-610. [\[CrossRef\]](#)
33. Wu HH, Inman A, Cramer HM. Subclassification of "atypia of undetermined significance" in thyroid fine-needle aspirates. *Diagn Cytopathol.* 2014;42:23-29. [\[CrossRef\]](#)
34. Rodrigues HG, de Pontes AA, Adan LF. Use of molecular markers in samples obtained from preoperative aspiration of thyroid. *Endocr J.* 2012;59:417-424. [\[CrossRef\]](#)
35. Paunovic I, Isic T, Havelka M, Tatic S, Cvejic D, Savin S. Combined immunohistochemistry for thyroid peroxidase, galectin-3, CK19 and HBME-1 in differential diagnosis of thyroid tumors. *APMIS.* 2012;120:368-379. [\[CrossRef\]](#)
36. Chiu CG, Strugnell SS, Griffith OL, et al. Diagnostic utility of galectin-3 in thyroid cancer. *Am J Pathol.* 2010;176:2067-2081. [\[CrossRef\]](#)
37. Bartolazzi A, Orlandi F, Saggiorato E, et al. Galectin-3-expression analysis in the surgical selection of follicular thyroid nodules with indeterminate fine-needle aspiration cytology: a prospective multicentre study. *Lancet Oncol.* 2008;9:543-549. [\[CrossRef\]](#)
38. Prasad ML, Pellegata NS, Huang Y, Nagaraja HN, de la Chapelle A, Kloos RT. Galectin-3, fibronectin-1, CITED-1, HBME-1 and cytokeratin-19 immunohistochemistry is useful for the differential diagnosis of thyroid tumors. *Mod Pathol.* 2005;18:48-57. [\[CrossRef\]](#)
39. Rossi ED, Raffaelli M, Minimo C, et al. Immunocytochemical evaluation of thyroid neoplasms on thin-layer smears from fine-needle aspiration biopsies. *Cancer.* 2005;105:87-95. [\[CrossRef\]](#)
40. Herrmann ME, LiVolsi VA, Pasha TI, et al. Immunohistochemical expression of galectin-3 in benign and malignant thyroid lesions. *Arch Pathol Lab Med.* 2002;126:710-713. [\[CrossRef\]](#)
41. Canberk S, Firat P, Schmitt F. Pitfalls in the Cytological Assessment of Thyroid Nodules. *Turk Patoloji Derg.* 2015;31 Suppl 1:18-33. [\[CrossRef\]](#)
42. De May Richard. The Art and science of cytopathology. 2nd ed. Chicago: ASCP;2012. [\[CrossRef\]](#)
43. Syed AZ, Nayar R, Krane JF, Westra WH. Atlas of Thyroid cytopathology with histopathologic correlations. New York: Demos Medical Publishing; 2014. [\[CrossRef\]](#)
44. Yang J, Schnadig V, Logrono R, Wasserman PG. Fine-needle aspiration of thyroid nodules: a study of 4703 patients with histologic and clinical correlations. *Cancer.* 2007;111:306-315. [\[CrossRef\]](#)
45. Deveci MS, Deveci G, LiVolsi VA, Baloch ZW. Fine-needle aspiration of follicular lesions of the thyroid. Diagnosis and follow-Up. *Cytojournal.* 2006;3:9. [\[CrossRef\]](#)
46. Baloch ZW, Fleisher S, LiVolsi VA, Gupta PK. Diagnosis of "follicular neoplasm": a gray zone in thyroid fine-needle aspiration cytology. *Diagn Cytopathol.* 2002;26:41-44. [\[CrossRef\]](#)
47. Renshaw AA, Wang E, Wilbur D, et al. Interobserver agreement on microfollicles in thyroid fine-needle aspirates. *Arch Pathol Lab Med.* 2006;130:148-152. [\[CrossRef\]](#)
48. Faquin WC. Diagnosis and reporting of follicular-patterned thyroid lesions by fine needle aspiration. *Head Neck Pathol.* 2009;3:82-85. [\[CrossRef\]](#)
49. Damiani D, Suciu V, Vielh P. Cytopathology of follicular cell nodules. *Endocr Pathol.* 2015; 26:286-290. [\[CrossRef\]](#)
50. Baloch ZW, LiVolsi VA. Current role and value of fine-needle aspiration in nodular goitre. *Best Pract Res Clin Endocrinol Metab.* 2014;28:531-544. [\[CrossRef\]](#)
51. Clark DP, Faquin WC. Thyroid cytopathology. New York: Springer; 2005. [\[CrossRef\]](#)
52. Syed AZ, Cibas ES. The Bethesda System for reporting thyroid cytopathology. New York: Springer; 2010. [\[CrossRef\]](#)
53. Baloch ZW, Fleisher S, LiVolsi VA, Gupta PK. Diagnosis of "follicular neoplasm": a gray zone in thyroid fine-needle aspiration cytology. *Diagn Cytopathol.* 2002;26:41-44. [\[CrossRef\]](#)
54. Faquin WC, Baloch ZW. Fine-needle aspiration of follicular patterned lesions of the thyroid: Diagnosis, management, and follow-up according to National Cancer Institute (NCI) recommendations. *Diagn Cytopathol.* 2010;38:731-739. [\[CrossRef\]](#)
55. Schmitt F. Cytopathology. In: Schmitt, F, ed. Encyclopedia of Pathology. Cham, Switzerland: Springer; 2017:376-379. [\[CrossRef\]](#)
56. Canberk S, Firat P, Schmitt F. Pitfalls in the cytological assessment of thyroid nodules. *Turk Patoloji Derg.* 2015;31 Suppl 1:18-33. [\[CrossRef\]](#)
57. Liu CY, Chen CC, Bychkov A, et al. Constitutive Cytomorphologic Features of Medullary Thyroid Carcinoma Using Different Staining Methods. *Diagnostics (Basel).* 2021;11:1396. [\[CrossRef\]](#)
58. Deshpande AH, Bobhate SK. Cytological features of dyshormonogenetic goiter: case report and review of the literature. *Diagn Cytopathol.* 2005;33:252-254. [\[CrossRef\]](#)
59. Agarwal AM, Bentz JS, Hungerford R, Abraham D. Parathyroid fine-needle aspiration cytology in the evaluation of parathyroid adenoma: cytologic findings from 53 patients. *Diagn Cytopathol.* 2009;37:407-410. [\[CrossRef\]](#)
60. Lastra RR, LiVolsi VA, Baloch ZW. Aggressive variants of follicular cell-derived thyroid carcinomas: a cytopathologist's perspective. *Cancer Cytopathol.* 2014;122:484-503. [\[CrossRef\]](#)

61. Bongiovanni M, Bloom L, Krane JF, et al. Cytomorphologic features of poorly differentiated thyroid carcinoma: a multi-institutional analysis of 40 cases. *Cancer*. 2009;117:185-194. [\[CrossRef\]](#)
62. Schmitt FC, Barroca H. Role of ancillary studies in fine-needle aspiration from selected tumors. *Cancer Cytopathol*. 2012;120:145-160. [\[CrossRef\]](#)
63. Schmitt FC, Longatto-Filho A, Valent A, Vielh P. Molecular techniques in cytopathology practice. *J Clin Pathol*. 2008;61:258-267. [\[CrossRef\]](#)
64. Filho AL, Gonçalves AE, Martinho O, Schmitt FC, Reis RM. Liquid-based cytology in DNA-based molecular research. *Anal Quant Cytol Histol*. 2009;31:395-400. [\[CrossRef\]](#)
65. Nikiforova MN, Nikiforov YE. Molecular diagnostics and predictors in thyroid cancer. *Thyroid*. 2009;19:1351-1361. [\[CrossRef\]](#)
66. Nikiforov YE, Steward DL, Robinson-Smith TM, et al. Molecular testing for mutations in improving the fine-needle aspiration diagnosis of thyroid nodules. *J Clin Endocrinol Metab*. 2009;94:2092-2098. [\[CrossRef\]](#)
67. Ohori NP, Nikiforova MN, Schoedel KE, et al. Contribution of molecular testing to thyroid fine -needle aspiration cytology of "follicular lesion of undetermined significance/atypia of undetermined significance". *Cancer Cytopathol*. 2010;118:17-23. [\[CrossRef\]](#)
68. Gharib H, Papini E, Valcavi R, et al. American Association of Clinical Endocrinologists and Associazione Medici Endocrinologi medical guidelines for clinical practice for the diagnosis and management of thyroid nodules. *Endocr Pract*. 2006;12:63-102. [\[CrossRef\]](#)
69. Baloch ZW, LiVolsi VA, Asa SL, et al. Diagnostic terminology and morphologic criteria for cytologic diagnosis of thyroid lesions: a synopsis of the National Cancer Institute Thyroid Fine-Needle Aspiration State of Science Conference. *Diagn Cytopathol*. 2008;36:425-437. [\[CrossRef\]](#)
70. Cibas ES, Ali SZ; NCI Thyroid FNA State of the Science Conference. The Bethesda System For Reporting Thyroid Cytopathology. *Am J Clin Pathol*. 2001;132:658-665. [\[CrossRef\]](#)
71. Rossi ED, Morassi F, Santeusanio G, Zannoni GF, Fadda G. Thyroid fine needle aspiration cytology processed by ThinPrep: an additional slide decreased the number of inadequate results. *Cytopathology*. 2010;21:97-102. [\[CrossRef\]](#)
72. Cochand-Priollet B, Prat JJ, Polivka M, et al. Thyroid fine needle aspiration: the morphological features on ThinPrep slide preparations. Eighty cases with histological control. *Cytopathology*. 2003;14:343-349. [\[CrossRef\]](#)
73. Fadda G, Rossi ED, Raffaelli M, et al. Follicular thyroid neoplasms can be classified as low- and high-risk according to HBME-1 and galectin-3 expression on liquid-based fine-needle cytology. *Eur J Endocrinol*. 2011;165:447-453. [\[CrossRef\]](#)
74. de Matos LL, Del Giglio AB, Matsubayashi CO, de Lima Farah M, Del Giglio A, da Silva Pinhal MA. Expression of CK-19, galectin-3 and HBME-1 in the differentiation of thyroid lesions: systematic review and diagnostic meta-analysis. *Diagn Pathol*. 2012;7:97. [\[CrossRef\]](#)
75. Paunovic I, Isic T, Havelka M, Tatic S, Cvejic D, Savin S. Combined immunohistochemistry from thyroid peroxidase, galectin-3, CK19 and HBME-1 in differential diagnosis of thyroid tumors. *APMIS*. 2012;120:368-379. [\[CrossRef\]](#)
76. Papale F, Cafiero G, Grimaldi A, et al. Galectin-3 expression in thyroid fine needle cytology (t-FNAC) uncertain cases: validation of molecular markers and technology innovation. *J Cell Physiol*. 2013;228:968-974. [\[CrossRef\]](#)
77. Bartolazzi A, Orlandi F, Saggiorato E, et al. Italian Thyroid Cancer Study Group (ITCSG). Galectin-3-expression analysis in the surgical selection of follicular thyroid nodules with indeterminate fine-needle aspiration cytology: a prospective multicentre study. *Lancet Oncol*. 2008;9:543-549. [\[CrossRef\]](#)
78. Ramkumar S, Sivanandham S. The Combined Utility of HBME-1 and Galectin-3 Immunohistochemistry and BRAF V600E Mutations in the Diagnosis of Papillary Thyroid Carcinoma. *Cureus*. 2021;13:e20339. [\[CrossRef\]](#)
79. Abu-Sinna E, Hasan MY, El-Deftar MM, Amer SA, Abdelsalam LO, Nakhla JA. Galectin-3 and HBME-1 Expression on Agarose Cell Blocks from Fine-needle aspirates of Follicular Cell-derived Thyroid Tumors. *J Cytol*. 2018;35:27-32. [\[CrossRef\]](#)
80. Fadda G, Rossi ED, Raffaelli M, et al. Follicular thyroid neoplasms can be classified as low- and high-risk according to HBME-1 and Galectin-3 expression on liquid-based fine-needle cytology. *Eur J Endocrinol*. 2011;165:447-453. [\[CrossRef\]](#)
81. Margari N, Giovannopoulos I, Pouliakis A, et al. Application of Immunocytochemistry on Cell Block Sections for the Investigation of Thyroid Lesions. *Acta Cytol*. 2018;62:137-144. [\[CrossRef\]](#)
82. Chiu CG, Strugnell SS, Griffith OL, et al. Diagnostic utility of galectin-3 in thyroid cancer. *Am J Pathol*. 2010;176:2067-2081. [\[CrossRef\]](#)
83. Prasad ML, Pellegrata NS, Huang Y, Nagaraja HN, de la Chapelle A, Kloos RT. Galectin-3, fibronectin-1, CITED-1, HBME1 and cytokeratin-19 immunohistochemistry is useful for the differential diagnosis of thyroid tumors. *Mod Pathol*. 2005;18:48-57. [\[CrossRef\]](#)
84. Rodrigues HG, de Pontes AA, Adam LF. Use of molecular markers in samples obtained from preoperative aspiration of thyroid. *Endocr J*. 2012;59:417-424. [\[CrossRef\]](#)
85. Tastekin E, Keskin E, Can N, et al. CD56, CD57, HBME1, CK19, Galectin-3 and p63 immunohistochemical stains in differentiating diagnosis of thyroid benign/malignant lesions and NIFTP. *Pol J Pathol*. 2019;70:286-294. [\[CrossRef\]](#)
86. Straccia P, Brunelli C, Rossi ED, et al. The immunocytochemical expression of VE-1 (BRAF V600E-related) antibody identifies the aggressive variants of papillary thyroid carcinoma on liquid-based cytology. *Cytopathology*. 2019;30:460-467. [\[CrossRef\]](#)
87. Zhao H, Guo HQ, Zhang ZH, et al. [Value of the detection of BRAF(V600E) gene mutation and protein expression in auxiliary cytological diagnosis of papillary thyroid carcinoma]. *Zhonghua Er Bi Yan Hou Tou Jing Wai Ke Za Zhi*. 2019;54:18-22. [\[CrossRef\]](#)
88. Smith AL, Williams MD, Stewart J, et al. Utility of the BRAF p.V600E immunoperoxidase stain in FNA direct smears and cell block preparations from patients with thyroid carcinoma. *Cancer Cytopathol*. 2018;126:406-413. [\[CrossRef\]](#)
89. Zhang Y, Liu L, Liu Y, Cao N, Wang L, Xing C. Clinical significance of immunohistochemistry to detect BRAF V600E mutant protein in thyroid tissues. *Medicine (Baltimore)*. 2021; 100:e25566. [\[CrossRef\]](#)
90. Parker KG, White MG, Cipriani NA. Comparison of Molecular Methods and BRAF Immunohistochemistry (VE1 Clone) for the Detection of BRAF V600E Mutation in Papillary Thyroid Carcinoma: A Meta-Analysis. *Head Neck Pathol*. 2020;14:1067-1079. [\[CrossRef\]](#)
91. Filho AL, Gonçalves AE, Martinho O, Schmitt FC, Reis RM. Liquid-based cytology in DNA-based molecular research: viability and potential application. *Anal Quant Cytol Histol*. 2009;31:395-400. [\[CrossRef\]](#)
92. Rossi ED, Martini M, Capodimonti S, et al. BRAF (V600E) mutation analysis on liquid-based cytology-processed aspiration biopsies predicts bilaterality and lymph node involvement in papillary thyroid microcarcinoma. *Cancer Cytopathol*. 2013;121:291-297. [\[CrossRef\]](#)
93. Rossi ED, Martini M, Capodimonti S, et al. Diagnostic and prognostic value of immunocytochemistry and BRAF mutation analysis on liquid-based biopsies of thyroid neoplasms suspicious for carcinoma. *Eur J Endocrinol*. 2013;168:853-859. [\[CrossRef\]](#)
94. Nishino M. Molecular cytopathology for thyroid nodules: a review of methodology and test performance. *Cancer Cytopathol*. 2016;124:14-27. [\[CrossRef\]](#)
95. Nikiforov YE, Steward DL, Robinson-Smith TM, et al. Molecular testing for mutations in improving the fine-needle aspiration diagnosis of thyroid nodules. *J Clin Endocrinol Metab*. 2009;94:2092-2098. [\[CrossRef\]](#)
96. Soares P, Trovisco V, Rocha AS, et al. BRAF mutations and RET/PTC rearrangements are alternative events in the etiopathogenesis of PTC. *Oncogene*. 2003;22:4578-4580. [\[CrossRef\]](#)
97. Cheung CC, Carydis B, Ezzat S, Bedard YC, Asa SL. Analysis of ret/PTC gene rearrangements refines the fine-needle aspiration diagnosis of thyroid cancer. *J Clin Endocrinol Metab*. 2001; 86:2187-2190. [\[CrossRef\]](#)
98. Ohori NP, Nikiforova MN, Schoedel KE, et al. Contribution of molecular testing to thyroid fine-needle aspiration cytology of "follicular lesion of undetermined significance/atypia of undetermined significance". *Cancer Cytopathol*. 2010;118:17-23. [\[CrossRef\]](#)
99. Moses W, Weng J, Sansano I, et al. Molecular testing for somatic mutations improves the accuracy of thyroid fine-needle aspiration biopsy. *World J Surg*. 2010;34:2589-2594. [\[CrossRef\]](#)
100. Xing M. BRAF mutation in papillary thyroid cancer: pathogenic role, molecular bases, and clinical implications. *Endocrinol Rev*. 2007;28:742-762. [\[CrossRef\]](#)

101. Al-Masri M, Al-Shobaki T, Al-Najjar H, et al. BRAF V600E mutation in papillary thyroid carcinoma: its relation to clinical features and oncologic outcomes in a single cancer centre experience. *Endocr Connect.* 2021;10:1531-1537. [\[CrossRef\]](#)
102. Lin KL, Wang OC, Zhang XH, Dai XX, Hu XQ, Qu JM. The BRAF mutation is predictive of aggressive clinicopathological characteristics in papillary thyroid microcarcinoma. *Ann Surg Oncol.* 2010;17:3294-3300. [\[CrossRef\]](#)
103. Puxeddu E, Durante C, Avenia N, Filetti S, Russo D. Clinical implications of BRAF mutation in thyroid carcinoma. *Trends Endocrinol Metab.* 2008;19:138-145. [\[CrossRef\]](#)
104. Musholt TJ, Fottner C, Weber M, et al. Detection of papillary carcinoma by analysis of BRAF and RET/PTC1 mutations in fine-needle aspiration biopsies of thyroid nodules. *World J Surg.* 2010;34:2595-2603. [\[CrossRef\]](#)
105. Celik M, Bulbul BY, Ayturk S, et al. The relation between BRAFV600E mutation and clinicopathological characteristics of papillary thyroid cancer. *Med Glas (Zenica).* 2020;17:30-34. [\[CrossRef\]](#)
106. Sacks WL, Bose S, Zumsteg ZS, et al. Impact of Afirma gene expression classifier on cytopathology diagnosis and rate of thyroidectomy. *Cancer Cytopathol.* 2016;124:722-728. [\[CrossRef\]](#)
107. Nikiforov YE, Cartt SE, Chiosea SI, et al. Impact of the multi-gene ThyroSeq next-generation sequencing assay on cancer diagnosis in thyroid nodules with atypia of undetermined significance/follicular lesion of undetermined significance cytology. *Cytology Thyroid.* 2015; 25:1217-1223. [\[CrossRef\]](#)
108. Labourier E, Shifrin A, Busseniers AE, et al. Molecular testing for miRNA, mRNA, and DNA on Fine-Needle aspiration improves the preoperative diagnosis of thyroid nodules with indeterminate cytology. *J Clin Endocrinol Metab.* 2015;100:2743-2750. [\[CrossRef\]](#)
109. Nishino M. Molecular cytopathology for thyroid nodules: a review of methodology and test performance. *Cancer Cytopathol.* 2016;124:14-27. [\[CrossRef\]](#)
110. McIver B, Castro MR, Morris JC, et al. An independent study of a gene expression classifier (Afirma) in the evaluation of cytologically indeterminate thyroid nodules. *J Clin Endocrinol Metab.* 2014;99:4069-4077. [\[CrossRef\]](#)
111. Harrell RM, Bimston DN. Surgical utility of Afirma: effects of high cancer prevalence and oncocytic cell types in patients with indeterminate thyroid cytology. *Endocr Pract.* 2014; 20:364-369. [\[CrossRef\]](#)
112. Alvarez-Nuñez F, Bussaglia E, Mauricio D, et al. PTEN promoter methylation in sporadic thyroid carcinomas. *Thyroid.* 2006;16:17-23. [\[CrossRef\]](#)
113. Hu S, Liu D, Tufano RP, et al. Association of aberrant methylation of tumor suppressor genes with tumor aggressiveness and BRAF mutation in papillary thyroid cancer. *Int J Cancer.* 2006; 119:2322-2329. [\[CrossRef\]](#)
114. Xing M. Gene methylation in thyroid tumorigenesis. *Endocrinology.* 2007;148:948-953. [\[CrossRef\]](#)
115. Schagdarsurenjin U, Gimm O, Hoang-Vu C, Dralle H, Pfeifer GP, Dammann R. Frequent epigenetic silencing of the CpG island promoter of RASSF1A in thyroid carcinoma. *Cancer Res.* 2002;62:3698-3701. [\[CrossRef\]](#)
116. Nakamura N, Carney JA, Jin L, et al. RASSF1A and NORE1A methylation and BRAFV600E mutations in thyroid tumors. *Lab Invest.* 2005;85:1065-1075. [\[CrossRef\]](#)
117. Xing M, Cohen Y, Mambo E, et al. Early occurrence of RASSF1A hypermethylation and its mutual exclusion with BRAF mutation in thyroid tumorigenesis. *Cancer Res.* 2004;64:1664-1668. [\[CrossRef\]](#)
118. Canberk S, Lima AR, Pinto M, Máximo V. Translational Potential of Epigenetic-Based Markers on Fine-Needle Aspiration Thyroid Specimens. *Front Med (Lausanne).* 2021;8:640460. [\[CrossRef\]](#)