



OPEN ACCESS

Genetic Causes of Inner Ear Anomalies: a Review from the Turkish Study Group for Inner Ear Anomalies

Emre Ocak¹, Duygu Duman^{2,3}, Mustafa Tekin^{4,5,6}

¹Department of Otolaryngology, Ankara University School of Medicine, Ankara, Turkey

²Division of Genetics, Department of Pediatrics, Ankara University School of Medicine, Ankara, Turkey

³Department of Audiology, Ankara University Faculty of Health Sciences, Ankara, Turkey

⁴John P. Hussman Institute for Human Genomics, University of Miami Miller School of Medicine, Miami, USA

⁵Department of Otolaryngology, University of Miami Miller School of Medicine, Miami, USA

⁶Dr. John T. Macdonald Department of Human Genetics, University of Miami Miller School of Medicine, Miami, USA

Inner ear anomalies diagnosed using a radiological study are detected in almost 30% of cases with congenital or prelingual-onset sensorineural hearing loss. Inner ear anomalies can be isolated or occur along with a part of a syndrome involving other systems. Although astonishing progress has been made in research aimed at revealing the genetic causes of hearing loss in the past few decades, only a few genes have been linked to inner ear anomalies. The aim of this review is to discuss the known genetic causes of inner ear anomalies.

Identifying the genetic causes of inner ear anomalies is important for guiding clinical care that includes empowered reproductive decisions provided to the affected individuals. Furthermore, understanding the molecular underpinnings of the development of the inner ear in humans is important to develop novel treatment strategies for people with hearing loss.

Keywords: Anomaly, genetics, inner ear, syndrome

Hearing loss (HL) is the most commonly diagnosed sensory disorder affecting 1-2 of every 1000 newborns (1). Previous reports suggest that more than half of sensorineural HL (SNHL) is caused by genetic factors (2). Although the majority of cases with SNHL do not have additional findings, more than 400 genetic syndromes have been described as having SNHL as an accompanying feature. Genetic or environmental causes disrupting the development of the inner ear (IE) lead to IE anomalies (IEAs) associated with SNHL. Bamiou et al. (3) reported that IEAs can be found in up to one-third of children with SNHL through a computerized tomography or a magnetic resonance imaging study.

Early diagnosis and timely management of SNHL is important as early intervention will have a large impact on the treatment outcomes. Understanding the genetic basis of SNHL associated with IEAs is important. This will help in making appropriate treatment decisions (e.g., feasibility of a cochlear implant), diagnosing other family members early, and empowering the affected individuals with family planning. The diagnosis may also be a sign of a syndrome involving other organ systems, which would necessitate screening for additional abnormalities.

Although significant progress has recently been made in understanding the physiology of hearing and pathologies leading to

SNHL in humans, our understanding of the molecular mechanisms underlying the human IE development is astonishingly limited. Excellent studies have been performed in various animal models, particularly in zebrafish, chicken, and mouse, shedding light on the fundamental mechanisms involved in the vertebrate IE development (4-6). However, mutations in only a few of the genes identified in animal model systems have been shown to cause IEAs in humans, suggesting that the molecular mechanisms regulating IE development are only partially shared between different species. Identification of the genetic basis of IEAs will provide much needed information about the development of the human IE, which could form the basis for novel therapies in the future. This review aims to summarize the current understanding of the genetic etiology of IEAs, specifically its clinical presentation, the genes involved, and the roles they play in embryology.

MOLECULAR DEVELOPMENT of the INNER EAR

Most of the current knowledge about the molecular basis of IE development has been gained by studying model animals, specifically mouse, chicken, and zebrafish. There are general and organism-specific writing formats for genes and proteins. In general, symbols for genes are italicized, whereas symbols for

Address for Correspondence: Mustafa Tekin, John P. Hussman Institute for Human Genomics, University of Miami Miller School of Medicine, Miami, USA

Phone: 305-243-2381

e-mail: mtekin@med.miami.edu

ORCID: orcid.org/0000-0002-3525-7960

Received: 16 April 2019

Accepted: 24 May 2019

DOI: 10.4274/balkanmedj.galenos.2019.2019.4.66



Available at www.balkanmedicaljournal.org

Cite this article as:

Ocak E, Duman D, Tekin M. Genetic Causes of Inner Ear Anomalies: a Review from the Turkish Study Group for Inner Ear Anomalies. Balkan Med J 2019;36:206-11

©Copyright 2019 by Trakya University Faculty of Medicine / The Balkan Medical Journal published by Galenos Publishing House.

proteins are not italicized. Human genes and proteins are written entirely in upper case, the first letter is capitalized for mouse genes and proteins, and for zebrafish, they are written entirely in lower case.

Thickening of the ectoderm, which is known as the otic placode, constitutes the embryological basis of the IE. The development of the IE begins in the preplacodal region, which is a primitive embryological region that surrounds the neural plate to form the otic placode. Then, the otic vesicle forms from this thickened epithelium (Figure 1). This highly complicated process includes modeling the structure with morphogenetic movements, expression of cell-specific proteins, and differentiation of the highly specialized cell types. For later development of IE, planar polarization is necessary because stereociliary bundles are sensitive to vibrations only on a single plane, and it is also required for convergent extension, a polarized cellular movement that occurs during neural tube closure and cochlear extension (4). Several genes and numerous pathways work during these processes. Another issue on this subject is the influence of environmental factors on the molecular development of the IE structures. Epigenetic modifications and mtDNA play an important role in the molecular development of the IE (7).

Preplacodal region

Studies have demonstrated that the interaction of the proteins in the fibroblast growth factor pathway, bone morphogenic protein (Bmp), Wnt, and their antagonists play a major role in the development of the preplacodal region in the ectoderm of the developing embryo (8). Gbx2 and Otx2 mutually modulate the anterior-posterior differentiation of the preplacodal region. Among them, Gbx is especially important in the development of the otic placode (9). The transcriptional coactivator Eya1 and the homeobox gene Six1 are believed to participate in the early differentiation of hair cells and neurons (10), which are in turn regulated by Foxi1 (11). Eya1 and Six1 together with Dach form a regulatory network that leads to transcriptional activation, cell proliferation, and organogenesis of the IE (12).

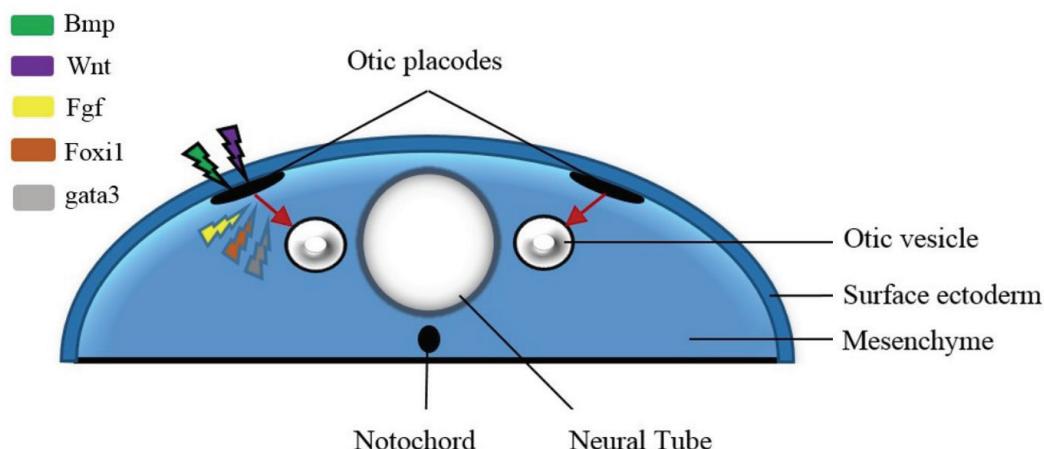


FIG. 1. Differentiation of the otic placode in the preplacodal region and formation of the otic vesicle. The various molecules affecting the different stages of the process are shown in the figure.

Otic vesicle

The otic vesicle is formed from the invagination of the otic placode. In a study investigating the conditions that influence the response to Fgf during otic placode induction (13), signaling of this factor has been shown to induce the expression of zebrafish otic genes such as *pax8*, *pax2a*, *fjf24*, and *sox3* throughout the preplacodal region, which is important in setting the pattern of the otic vesicle. This indicates that the fibroblast growth factor pathway continues to be important in the induction of the otic placode. The abovementioned transformation is believed to be regulated by the *Hoxa1* gene (14). This pathway is also influenced by foxi1 (inhibition effect) and gata3 (promotion effect) during otic development (15). Sai et al. (16) investigated the RhoA activity for apical constriction during phase 2 IE placode invagination and found that invagination of the otic placode to form the otic vesicle occurs via the activation of myosin-II through not only fibroblast growth factor signaling but also the RhoA-ROCK pathway. After the development of the otic vesicle, the otocyst gives rise to the mature IE structures, including the vestibular system in the dorsal plane and the auditory system in the ventral plane.

Molecules and factors involved in the neurogenesis of the inner ear

Several factors play important roles in the subsequent development of the neural IE structures after the otic vesicle stage. Inhibition of Sox2 transcription has been shown to be required for the progression of neurogenesis in the IE (17). In order for neuronal precursors to commit to a neuronal fate, Fgf10 signals the expression of Ngn1 and Neurod1, which act to inhibit the activity of Sox2 (17,18). Notch signaling also plays a role in specifying the sensory domains within the otic placode by inducing the proliferation of undifferentiated presensory cells, upregulating Sox2 activity, and inhibiting Ngn1 activity (19). Furthermore, tfap2a is believed to modulate the activity of fibroblast growth factor and notch by activating the inhibitor bmp7a, while playing a key role in neural development as well (20). Inner radial bundle formation is then mediated by Eph/Ephrin signaling, which is a target of Pou3f4 transcription factor activity (21,22).

Cochlea formation

In the topographic anatomy of the temporal bone, the cochlea is located in the ventral plane in the anteroposterior development axis after the otic vesicle stage. Several factors and molecules are involved in the development of the cochlea such as *Jag1*, *Sox2*, and *Lfng* (23). Deletion of p27^{kip1} can cause some changes in the cochlea, such as overproduction of hair cells, interestingly causing SNHL (24). *Atoh1* is the earliest discovered factor expressed in the prosensory domain associated with sensory hair cells. In the early period, it can be detected in the entire cochlea; however, over time, the expression of *Atoh1* is restricted to hair cell progenitors. Numerous factors that can upregulate or downregulate *Atoh1* have been described. Among them, *Sox2* is one of the most investigated molecules. Although being required for the expression, *Sox2* also downregulates *Atoh1*. Other regulators of this transcription factor are the Id (inhibitor of differentiation) genes (*Id1*, *Id2*, and *Id3*). These genes are known to negatively regulate *Atoh1* (25). Cochlear lumen formation begins at the base of the cochlea and proceeds toward the apex. This is partially controlled by fluid secretion in the vestibular labyrinth, which is then absorbed into the endolymphatic sac, a process mediated by *Slc26a4*-encoded channels (26).

Semicircular canal development

The vestibular system is located in the dorsal plane in the IE. The semicircular canals and their neural elements are derived from the two prominences of the otocyst, the horizontal and vertical canal pouches. The hindbrain is the source of the ventral–dorsal axial signaling for the IE. Wnts, from the dorsal hindbrain, are important signals for the development of the semicircular canals (27). *Dlx5* has been shown to be one of the downstream genes that respond to Wnt signaling. Previous studies have demonstrated that the lack of *Dlx5* can affect canal and crista formation (28). Another molecule that is also required for appropriate semicircular canal formation is *Hmx3*. On the other hand, *Chd7* encodes a chromodomain-containing helicase protein and has been proposed to act as a selector gene that encodes transcription factors essential for the formation of semicircular canals (29).

CLASSIFICATION OF INNER EAR ANOMALIES

IE malformations will be represented here according to the Sennaroglu classification system as it is one of the most widely used and accepted classification systems (30). From a systematic perspective, these malformations can be classified into eight distinct types as follows:

- Complete labyrinthine aplasia: This anomaly is also known as Michel deformity. There is an absence of the cochlea, vestibule, semicircular canals, and vestibular and cochlear aqueducts. There are three subgroups according to the imaging findings.
- Rudimentary otocysts: These are millimetric representations of the otic capsule without an internal acoustic canal. This type of anomaly represents between a complete labyrinthine aplasia and the common cavity.

- Cochlear aplasia: The cochlea is completely absent. The components of the vestibular system are in their normal anatomic locations.

- Common cavity: There is a cystic cavity representing the cochlea and the vestibule, but without showing any differentiation into the cochlea and the vestibule.

- Cochlear hypoplasia: Dimensions of the cochlea are smaller than normal. There is a differentiation between the cochlea and the vestibule with various internal architecture deformities. There are four distinct types of cochlear hypoplasia.

- Incomplete partition: This type of anomaly represents the most common IE malformation. There is a differentiation between the cochlea and the vestibule. The external dimensions are normal with various internal architecture defects. This group of malformations is subclassified into three subgroups as IP-I, -II, and -III.

- Enlarged vestibular aqueduct: The vestibular aqueduct is enlarged in the presence of a normal cochlea, vestibule, and semicircular canals.

- Cochlear aperture abnormalities: The cochlear aperture can be hypoplastic or aplastic, which can be detected in computerized tomography scans. This malformation may be accompanied by a narrow internal acoustic canal.

Syndromic causes of inner ear anomalies in humans

IEAs can be sporadic or occur along with a part of a syndrome involving other systems. Numerous syndromes can be associated with IEAs in humans. The most frequently diagnosed ones with clinical findings and causative genes are summarized in Table 1.

Nonsyndromic causes of inner ear anomalies

Nonsyndromic deafness is a type of hearing impairment in which HL is the only clinical finding in the patient. In these cases, the underlying etiology is generally a single gene mutation. Among all patients with nonsyndromic deafness, only a few genes have been discovered to cause IEAs when they have mutations.

SLC26A4

In 1997, the gene responsible for Pendred syndrome was identified as *SLC26A4* (31). *SLC26A4* encodes a transmembrane protein termed as pendrin. Subsequently, *SLC26A4* mutations were also discovered in individuals with autosomal recessive nonsyndromic deafness associated with enlargement of the vestibular aqueduct (32). More than 200 mutations have so far been reported, which can be related to sporadic and familial forms of Pendred syndrome and nonsyndromic SNHL with enlargement of the vestibular aqueduct.

Recent studies applying molecular testing for *SLC26A4* mutations and radiological imaging of temporal bones have demonstrated that enlargement of the vestibular aqueduct can be recognized as the most penetrant feature of Pendred syndrome (33). Enlargement of the vestibular aqueduct is the most common radiological anomaly of the IE that is primarily identified in either of two different contexts, nonsyndromic enlargement of the vestibular aqueduct or Pendred syndrome. Although the mutant alleles of *SLC26A4* have been shown to correlate with the auditory and thyroid phenotypes, no connections between the type of mutation and the thyroid

TABLE 1. Syndromic causes of inner ear anomalies

Syndrome	Clinical features	Gene involved
Waardenburg	Auditory-pigmentary syndrome characterized by pigmentary abnormalities of the hair, including a white forelock and premature graying; pigmentary changes of the iris, such as heterochromia iridis and brilliant blue eyes; and congenital sensorineural hearing loss. All types (I-IV) are inherited in an autosomal dominant manner, while a subgroup of type IV is autosomal recessive	<i>PAX3, MITF, SOX10, EDN3, SLUG</i>
Branchiootorenal	Autosomal dominant syndrome associated with malformations of the outer, middle, and inner ear associated with HL, branchial fistulae and cysts, and renal abnormalities. Inner ear presentations of patients include dysplastic vestibule, cochlear hypoplasia with defective modiolus, EVA, hypoplastic semicircular canals, and dilated internal auditory canals. Onset of hearing loss ranges from childhood to young adulthood	<i>EYAI, SIX1</i>
Pendred	Autosomal recessive condition with congenital severe-to-profound SNHL associated with EVA and either abnormal perchlorate discharge test or goiter	<i>SLC26A4</i>
Kabuki	Autosomal dominant disorder characterized by minor skeletal anomalies, unique facial features, “fetal” fingertip pads, mild-to-moderate intellectual disability, and growth deficiencies. Patients may have inner ear anomalies such as cochlear aplasia, vestibular dysplasia, and EVA	<i>KMT2D, KDM6A</i>
CHARGE	Autosomal dominant disorder standing for Coloboma, Heart defects, Choanal Atresia, Retarded growth and development, Genital abnormalities, and Ear anomalies	<i>CHD7, SEMA3E</i>
LAMM	Autosomal recessive condition standing for Labyrinthine Aplasia, Microtia, and Microdontia, which are its characteristic presentations along with profound congenital SNHL. The cerebellopontine angle may show the bilateral absence of cochleovestibular nerve with otherwise normal cerebral and cerebellar structures	<i>FGF3</i>
Branchiooculofacial	Autosomal dominant condition characterized by branchial skin defects, ocular anomalies such as cataract, facial anomalies, malformed pinnae, and HL. Patients may have EVA, cochlear dysplasia, and poorly differentiated cochlear turns	<i>TFAP2A</i>

EVA: enlarged vestibular aqueduct; HL: hearing loss; SNHL: sensorineural hearing loss

phenotype have been reported (34). Mutation in the *SLC26A4* gene is one of the leading causes of nonsyndromic SNHL and is inherited in an autosomal recessive manner.

Although variants in *FOXI1* and *KCNJ10* have been reported to cause SNHL with enlargement of the vestibular aqueduct when the same person is heterozygous for an *SLC26A4* mutation (i.e., digenic inheritance), this observation has not yet been confirmed by subsequent reports (35).

POU3F4

Variants in the *POU3F4* gene at the locus DFN3 are a major cause of X-linked deafness worldwide. The X-linked *POU3F4* gene is the first nuclear gene implicated in nonsyndromic deafness. The type of HL may be SNHL or mixed and associated with IP-III (incomplete partition type 3), cochlear hypoplasia, and/or stapes fixation (DFN3) (36-39). In addition to mutations located within the gene, copy number variants not involving the coding part of the gene have been reported; de Kok et al. (40) identified a hot spot for microdeletions in patients with X-linked deafness 900 kb proximal to the DFN3 gene.

From a clinical perspective, when *POU3F4* is identified as the HL-causing gene, the surgeon should be alert for a perilymphatic gusher during stapes surgery. Another important issue to consider is that if the HL is found to be X-linked and associated with an IE malformation, mutations in *POU3F4* gene should be evaluated.

COCH

The *COCH* gene is located at 14q11.2-q13 and encodes a secretory protein known as Cochlin (41). The pathogenetic mechanism of *COCH* gene-related HL is the accumulation of acidophilic deposits in the area of the spiral osseous lamina, spiral ligament, and vestibular nerve

channels (42). Several reports indicate a high probability of a possible link between the mutations in the *COCH* gene and the presentation of IEAs. Hildebrand et al. (43) described a patient who presented with semicircular canal dehiscence associated with a mutation in the *COCH* gene. Dodson et al. (44) described a patient heterozygous for a mutation in the *COCH* gene and who showed an enlargement of the vestibular aqueduct upon computerized tomography imaging. Finally, de Varebeke described nine patients with the same mutation in the *COCH* gene. On computerized tomography imaging, eight of them were found to have sclerotic lesions and/or narrowing of the semicircular canals, and in one patient, the posterior vestibule was also affected (45). Based on these findings, *COCH* mutations are a possible autosomal dominant inherited cause of IEAs and have been postulated to play a role, along with type II collagen bundles, in laying down the structure of the IE (46).

ROR1

ROR1 is an integral transmembrane protein consisting of extracellular and intracellular conserved domains. There is one reported family with two children having congenital autosomal recessive nonsyndromic SNHL and a common cavity anomaly who had an *ROR1* gene mutation as the cause (47).

FOXF2

Fox transcription factors regulate diverse biological processes throughout development and adult life. Mutations in some Fox genes are associated with human diseases. In a very recent paper, our study group published a homozygous mutation in the *FOXF2* gene required for cochlear development. In that study, the *FOXF2* gene mutation was found to be responsible for the IP-I anomaly (48).

Embryological development of the IE is a complex process in which various molecules are involved at different stages. Till date, only a few genes have been found to cause IEAs when their functions are disrupted by mutations. As the link between genetics and IEAs has been well established, future studies will help increase the knowledge regarding the molecular biology of IEAs.

Acknowledgements: This paper is submitted on behalf of The Turkish Study Group for Inner Ear Anomalies. We thank the members of the study group, Armağan İncesulu, Müzeyyen Yıldırım, Nazım Bozan, Suat Fitoz, Levent Olgun, Seyra Erbek, Tahir Atik, Mehmet Alıkaşifoğlu, Ferda Özkinsa, Güney Bademci, Demir Bajin, and Levent Sennaroğlu for their great support.

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: This work was supported by the National Institutes of Health Grants R01DC012836 and R01DC009645 to M.T.

REFERENCES

- Lantos PM, Maradiaga-Panayotti G, Barber X, Raynor E, Tucci D, Hoffman K, et al. Geographic and Racial Disparities in Infant Hearing Loss. *Otolaryngol Head Neck Surg* 2018;194:599-818803305.
- Nance WE. The genetics of deafness. *Ment Retard Dev Disabil Res Rev* 2003;9:109-19.
- Bamiou DE, Phelps P, Sirimanna T. Temporal bone computed tomography findings in bilateral sensorineural hearing loss. *Arch Dis Child* 2000;82:257-60.
- Chen J, Tambalo M, Barembaum M, Ranganathan R, Simões-Costa M, Bronner ME, et al. A systems-level approach reveals new gene regulatory modules in the developing ear. *Development* 2017;144:1531-43.
- Torii H, Yoshida A, Katsuno T, Nakagawa T, Ito J, Omori K, et al. Septin7 regulates inner ear formation at an early developmental stage. *Dev Biol* 2016;419:217-28.
- Whitfield TT, Riley BB, Chiang MY, Phillips B. Development of the zebrafish inner ear. *Dev Dyn* 2002;223:427-58.
- Montcouquiol M, Kelley MW. Development and Patterning of the Cochlea: From Convergent Extension to Planar Polarity. *Cold Spring Harb Perspect Med* 2019. pii: a033266.
- Litsiou A, Hanson S, Streit A. A balance of FGF, BMP and WNT signalling positions the future placode territory in the head. *Development* 2005;132:4051-62.
- Steventon B, Mayor R, Streit A. Mutual repression between Gbx2 and Otx2 in sensory placodes reveals a general mechanism for ectodermal patterning. *Dev Biol* 2012;367:55-65.
- Zou D, Silvius D, Fritzsch B, Xu PX. Eya1 and Six1 are essential for early steps of sensory neurogenesis in mammalian cranial placodes. *Development* 2004;131:5561-72.
- Ishihara T, Sato S, Ikeda K, Yajima H, Kawakami K. Multiple evolutionarily conserved enhancers control expression of Eya1. *Dev Dyn* 2008;237:3142-56.
- Li X, Oghi KA, Zhang J, Krones A, Bush KT, Glass CK, et al. Eya protein phosphatase activity regulates Six1-Dach-Eya transcriptional effects in mammalian organogenesis. *Nature* 2003;426:247-54.
- Padanad MS, Bhat N, Guo B, Riley BB. Conditions that influence the response to Fgf during otic placode induction. *Dev Biol* 2012;364:1-10.
- Makki N, Capecchi MR. Identification of novel Hoxa1 downstream targets regulating hindbrain, neural crest and inner ear development. *Dev Biol* 2011;357:295-304.
- Yao D, Zhao F, Wu Y, Wang J, Dong W, Zhao J, et al. Dissecting the differentiation process of the preplacodal ectoderm in zebrafish. *Dev Dyn* 2014;243:1338-51.
- Sai X, Yonemura S, Ladher RK. Junctionally restricted RhoA activity is necessary for apical constriction during phase 2 inner ear invagination. *Dev Biol* 2014;394:206-16.
- Evsen L, Sugahara S, Uchikawa M, Kondoh H, Wu DK. Progression of neurogenesis in the inner ear requires inhibition of Sox2 transcription by neurogenin1 and neurod1. *J Neurosci* 2013;33:3879-90.
- Alsina B, Abelló G, Ulloa E, Henrique D, Pujades C, Giraldez F. FGF signaling is required for determination of otic neuroblasts in the chick embryo. *Dev Biol* 2004;267:119-34.
- Jeon SJ, Fujioka M, Kim SC, Edge AS. Notch signaling alters sensory or neuronal cell fate specification of inner ear stem cells. *J Neurosci* 2011;31:8351-8.
- Kantarci H, Edlund RK, Groves AK, Riley BB. Tfap2a promotes specification and maturation of neurons in the inner ear through modulation of Bmp, Fgf and notch signaling. *PLoS Genet* 2015;11:e1005037.
- Coate TM, Raft S, Zhao X, Ryan AK, Crenshaw EB, Kelley MW. Otic mesenchyme cells regulate spiral ganglion axon fasciculation through a Pou3f4/EphA4 signaling pathway. *Neuron* 2012;73:49-63.
- Raft S, Coate TM, Kelley MW, Crenshaw EB, Wu DK. Pou3f4-mediated regulation of ephrin-b2 controls temporal bone development in the mouse. *PLoS One* 2014;9:e109043.
- Wu DK, Kelley MW. Molecular mechanisms of inner ear development. *Cold Spring Harb Perspect Biol* 2012;4:a008409.
- Chen P, Segil N. p27(Kip1) links cell proliferation to morphogenesis in the developing organ of Corti. *Development* 1999;126:1581-90.
- Jones JM, Montcouquiol M, Dabdoub A, Woods C, Kelley MW. Inhibitors of differentiation and DNA binding (Id)s regulate Math1 and hair cell formation during the development of the organ of Corti. *J Neurosci* 2006;26:550-8.
- Kim HM, Wangemann P. Failure of fluid absorption in the endolymphatic sac initiates cochlear enlargement that leads to deafness in mice lacking pendrin expression. *PLoS One* 2010;5:e14041.
- Riccomagno MM, Takada S, Epstein DJ. Wnt-dependent regulation of inner ear morphogenesis is balanced by the opposing and supporting roles of Shh. *Genes Dev* 2005;19:1612-23.
- Acampora D, Merlo GR, Paleari L, Zerega B, Postiglione MP, Mantero S, et al. Craniofacial, vestibular and bone defects in mice lacking the Distal-less-related gene Dlx5. *Development* 1999;126:3795-809.
- Hurd EA, Micucci JA, Reamer EN, Martin DM. Delayed fusion and altered gene expression contribute to semicircular canal defects in Chd7 deficient mice. *Mech Dev* 2012;129:308-23.
- Sennaroğlu L, Bajin MD. Classification and Current Management of Inner Ear Malformations. *Balkan Med J* 2017;34:397-411.
- Everett LA, Glaser B, Beck JC, Idol JR, Buchs A, Heyman M, et al. Pendred syndrome is caused by mutations in a putative sulphate transporter gene (PDS). *Nat Genet* 1997;17:411-22.
- Li XC, Everett LA, Lalwani AK, Desmukh D, Friedman TB, Green ED, et al. A mutation in PDS causes non-syndromic recessive deafness. *Nat Genet* 1998;18:215-7.
- Phelps PD, Coffey RA, Trembath RC, Luxon LM, Grossman AB, Britton KE, et al. Radiological malformations of the ear in Pendred syndrome. *Clin Radiol* 1998;53:268-73.
- Maciaszczyk K, Lewiński A. Phenotypes of SLC26A4 gene mutations: Pendred syndrome and hypoacusis with enlarged vestibular aqueduct. *Neuro Endocrinol Lett* 2008;29:29-36.
- Wu CC, Lu YC, Chen PJ, Yeh PL, Su YN, Hwu WL, et al. Phenotypic analyses and mutation screening of the SLC26A4 and FOXI1 genes in 101 Taiwanese families with bilateral nonsyndromic enlarged vestibular aqueduct (DFNB4) or Pendred syndrome. *Audiol Neurotol* 2010;15:57-66.
- Gong WX, Gong RZ, Zhao B. HRCT and MRI findings in X-linked non-syndromic deafness patients with a POU3F4 mutation. *Int J Pediatr Otorhinolaryngol* 2014;78:1756-62.
- Choi BY, An YH, Park JH, Jang JH, Chung HC, Kim AR, et al. Audiological and surgical evidence for the presence of a third window effect for the conductive hearing loss in DFNX2 deafness irrespective of types of mutations. *Eur Arch Otorhinolaryngol* 2013;270:3057-62.
- Anger GJ, Crocker S, McKenzie K, Brown KK, Morton CC, Harrison K, et al. X-linked deafness-2 (DFNX2) phenotype associated with a paracentric inversion upstream of POU3F4. *Am J Audiol* 2014;23:1-6.
- de Kok YJ, van der Maarel SM, Bitner-Glindzicz M, Huber I, Monaco AP, et al. Association between X-linked mixed deafness and mutations in the POU domain gene POU3F4. *Science* 1995;267:685-8.
- de Kok YJ, Vossenaar ER, Cremers CW, Dahl N, Laporte J, Hu LJ, et al. Identification of a hot spot for microdeletions in patients with X-linked deafness type 3 (DFN3) 900 kb proximal to the DFN3 gene POU3F4. *Hum Mol Genet* 1996;5:1229-35.
- Parzefall T, Frohne A, Koenighofer M, Kirchnawy A, Streubel B, Schoefer C, et al. Identification of a rare COCH mutation by whole-exome sequencing : Implications

- for personalized therapeutic rehabilitation in an Austrian family with non-syndromic autosomal dominant late-onset hearing loss. *Wien Klin Wochenschr* 2018;130:299-306.
42. Robertson NG, Lu L, Heller S, Merchant SN, Eavey RD, McKenna M, et al. Mutations in a novel cochlear gene cause DFNA9, a human nonsyndromic deafness with vestibular dysfunction. *Nat Genet* 1998;20:299-303.
 43. Hildebrand MS, Tack D, Deluca A, Hur IA, Van Rybroek JM, McMordie SJ, et al. Mutation in the COCH gene is associated with superior semicircular canal dehiscence. *Am J Med Genet A* 2009;149A:280-5.
 44. Dodson KM, Georgolios A, Barr N, Nguyen B, Sismanis A, Arnos KS, et al. Etiology of unilateral hearing loss in a national hereditary deafness repository. *Am J Otolaryngol* 2012;33:590-4.
 45. de Varebeke SP, Termote B, Van Camp G, Govaerts PJ, Schepers S, Cox T, et al. Focal sclerosis of semicircular canals with severe DFNA9 hearing impairment caused by a P51S COCH-mutation: is there a link? *Otol Neurotol* 2014;35:1077-86.
 46. Shindo S, Ikezono T, Ishizaki M, Sekiguchi S, Mizuta K, Li L, et al. Spatiotemporal expression of cochlin in the inner ear of rats during postnatal development. *Neurosci Lett* 2008;444:148-52.
 47. Diaz-Horta O, Abad C, Sennaroglu L, Foster J, DeSmidt A, Bademci G, et al. ROR1 is essential for proper innervation of auditory hair cells and hearing in humans and mice. *Proc Natl Acad Sci USA* 2016;113:5993-8.
 48. Bademci G, Abad C, Incesulu A, Elian F, Reyahi A, Diaz-Horta O, et al. FOXF2 is required for cochlear development in humans and mice. *Hum Mol Genet* 2019;28:1286-97.