

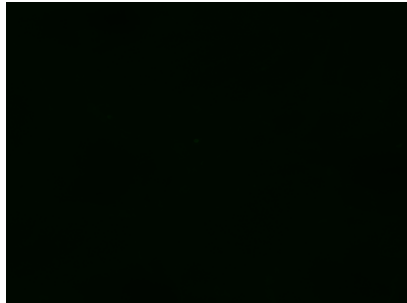
Coton trempé 10 min dans tu TEP concentré à 0.05% pendant 10min avec 10 μ L de PHK67, et rincé pendant 20s

Conditions expérimentales:

- Objectif x10
- Fluorescence en vert
- Temps d'exposition 10 – 800ms
- Gain 1
- Echantillon : coton imbibé de TEP 0.05% rincé 20s
- Ajout de DI water sur la lamelle
- Date de préparation et d'observation : 22 mai 2019

Images brutes

10ms



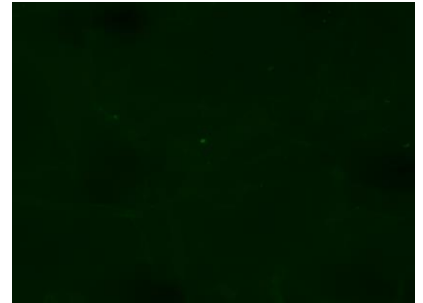
20ms



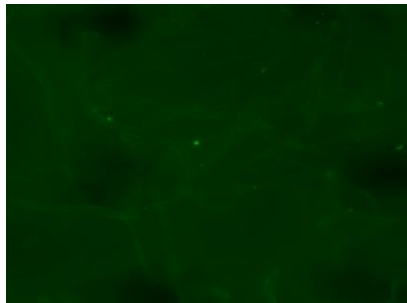
50ms



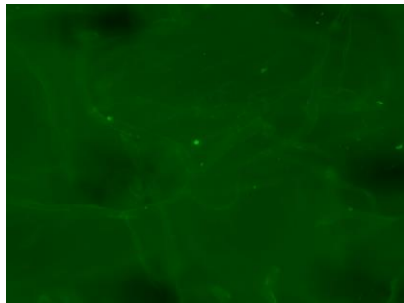
100ms



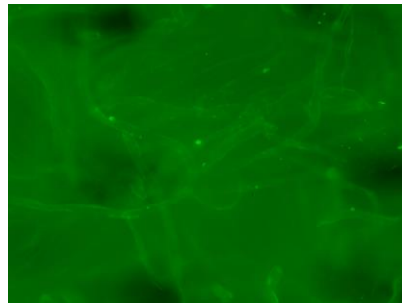
200ms



300ms



500ms



800ms

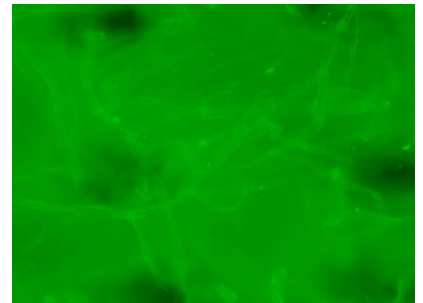
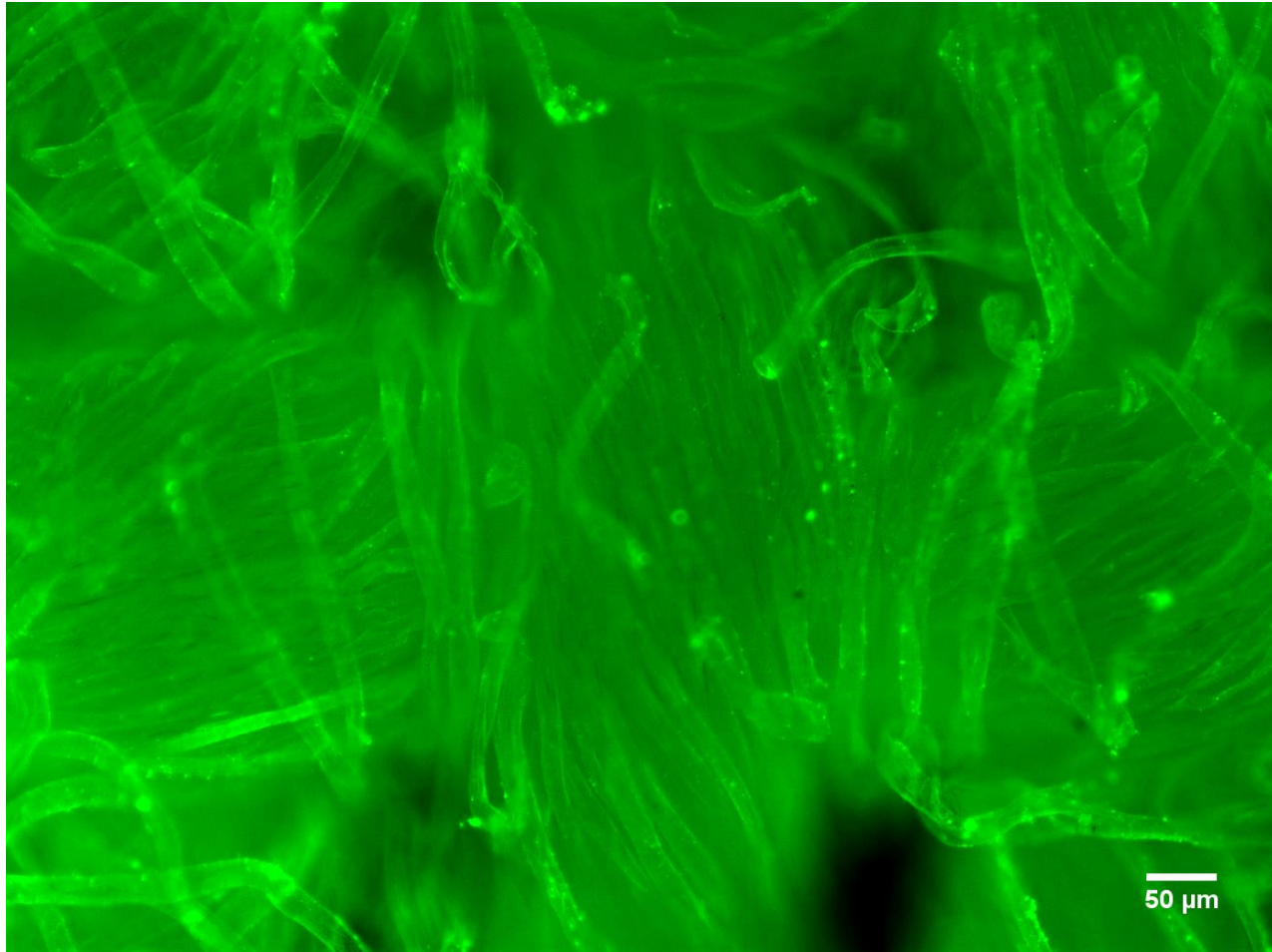
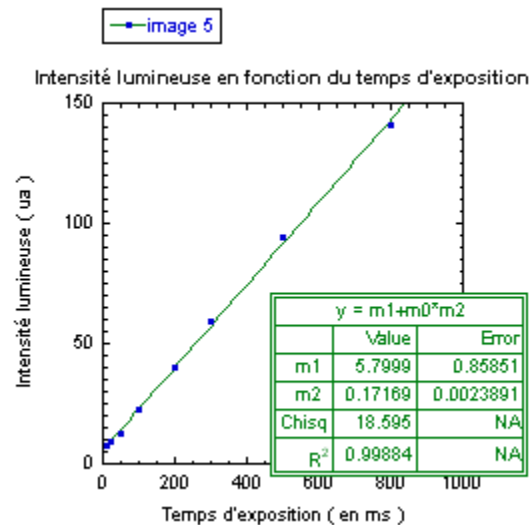
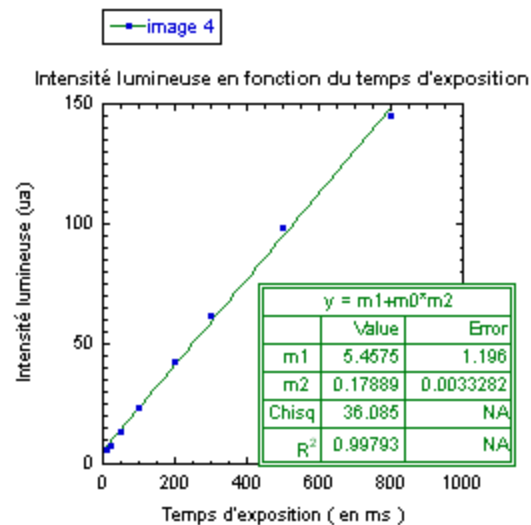
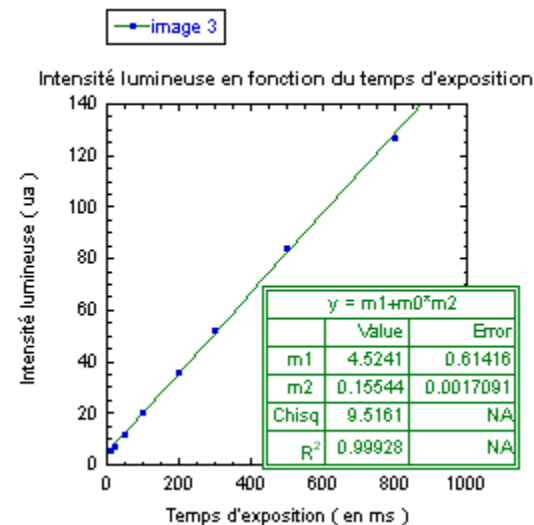
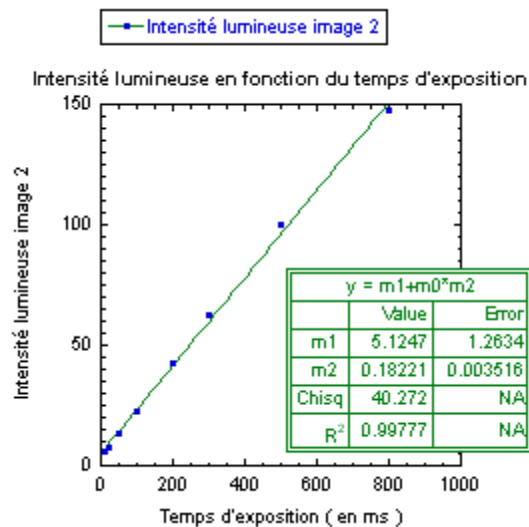
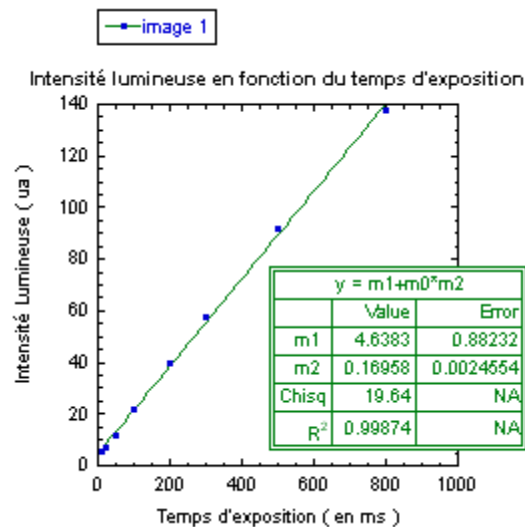
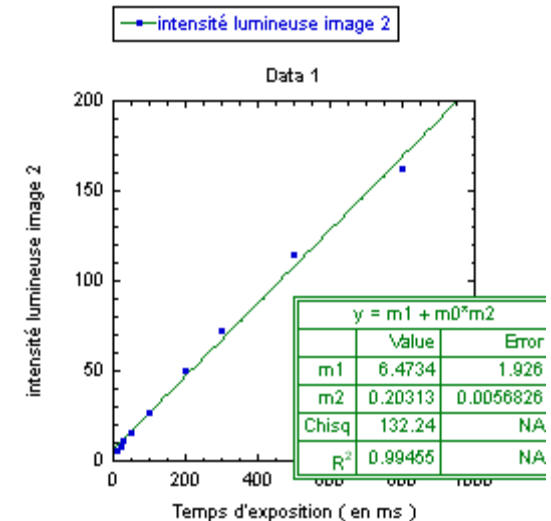
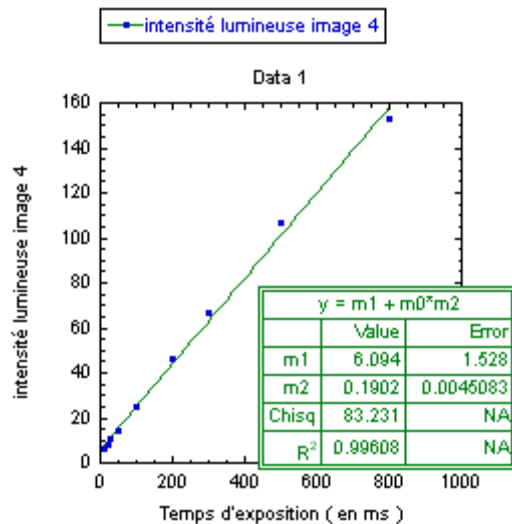
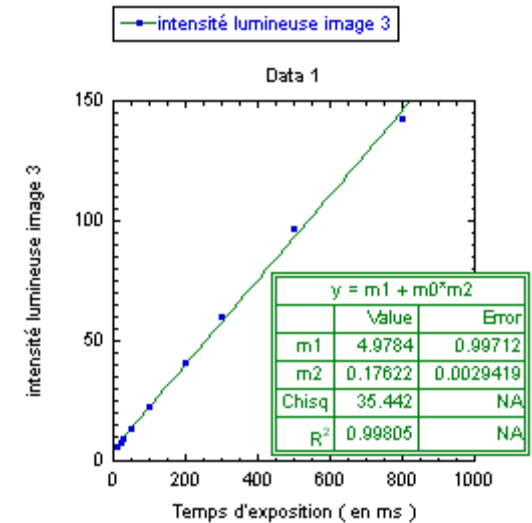
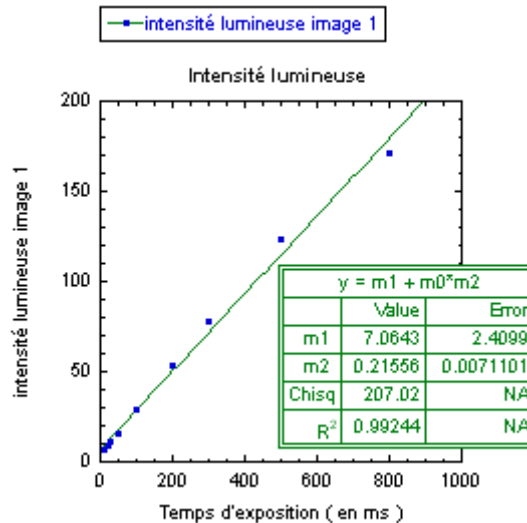


Image traitée

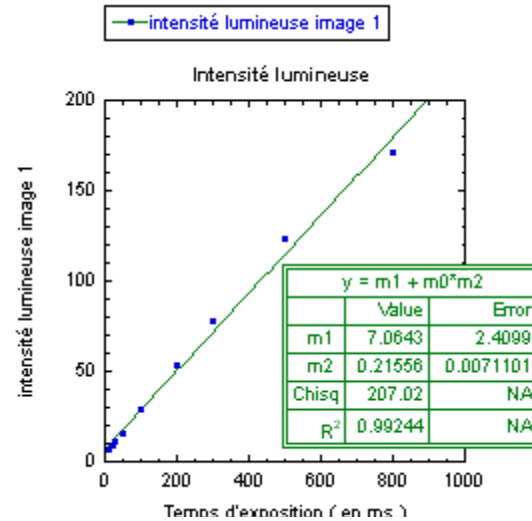
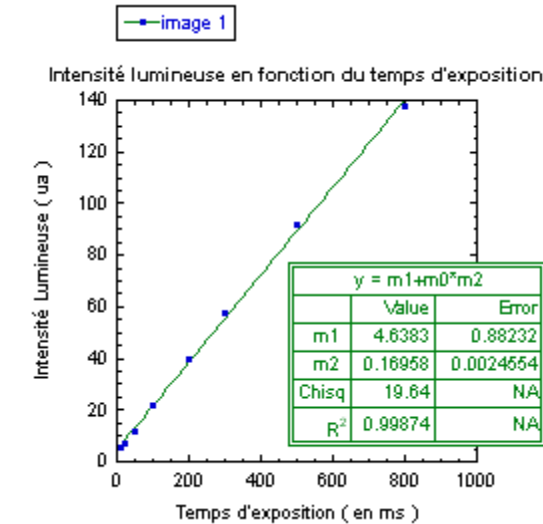




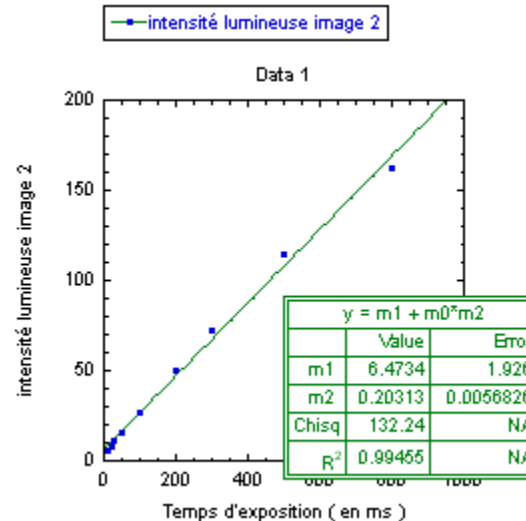
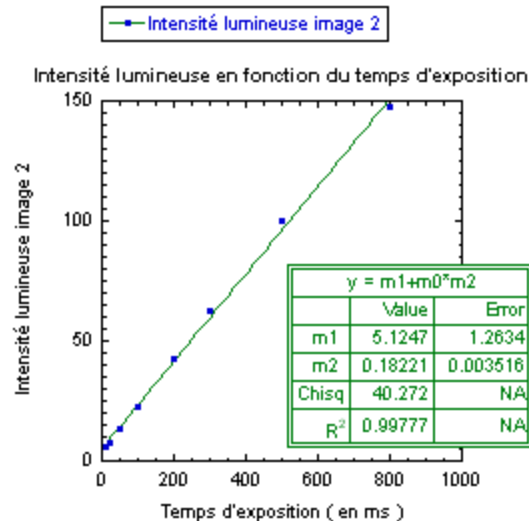
Coton ayant suivi le même protocole mais n'ayant pas été rincé



Comparaison entre le coton rincé 20s et celui observé sans rinçage



A gauche: le coton rincé 20s
A droite: celui non rincé



Conclusion

- Je retrouve plus ou moins la même ordonnée à 0 comme prévu c'est-à-dire 4 . Je n'ai pas enlevé les points qui saturent. Pour le coton rincé, il semble que la saturation ne soit pas atteinte. Pour celui non rincé, il faudrait enlever le point à 800ms.

Conditions expérimentales

- TEP 0.05%
- 10 μ L PKH67
- Trempé du 22 au 27 mai
- Rinçage 1min
- Microscope fluo x10 intensité 1 gain 1

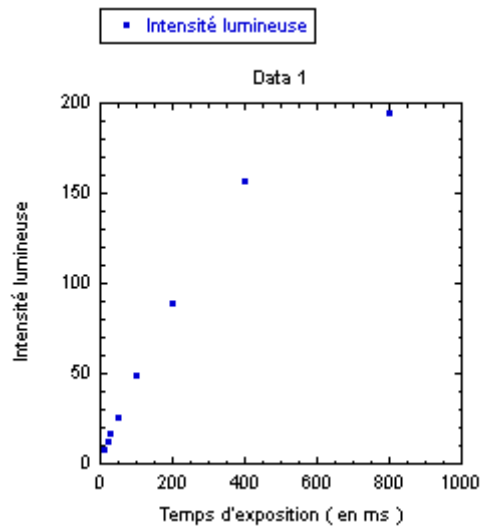


Image 5

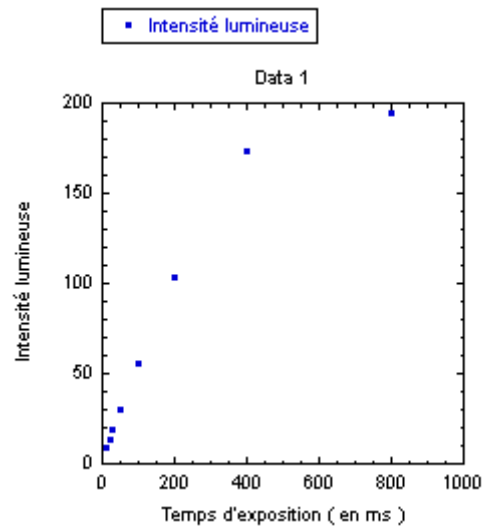


Image 4

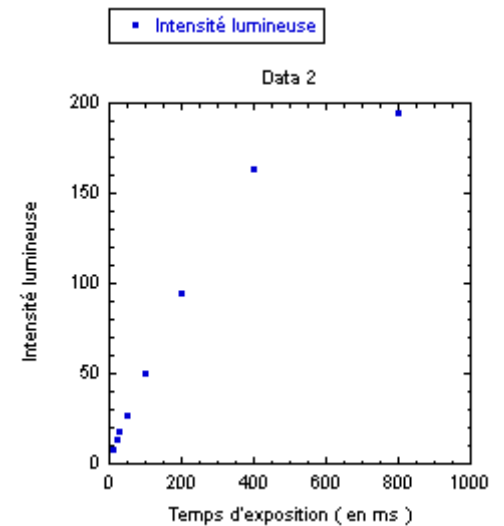


Image 3

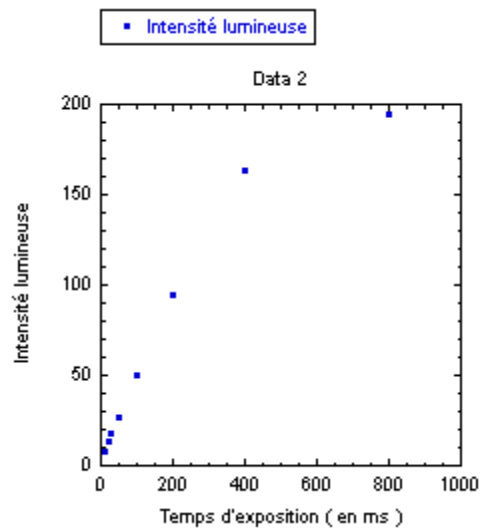


Image 2

On voit bien ici
qu'il y a une
saturation :
l'évolution doit
être linéaire

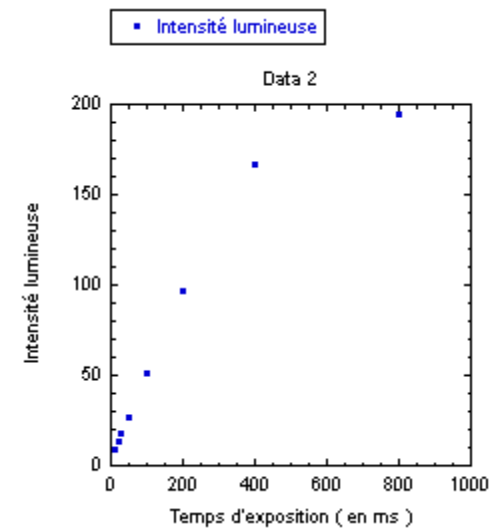


Image 1

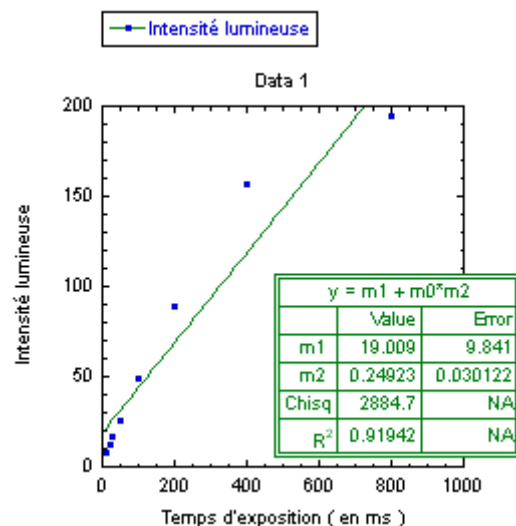


Image 5

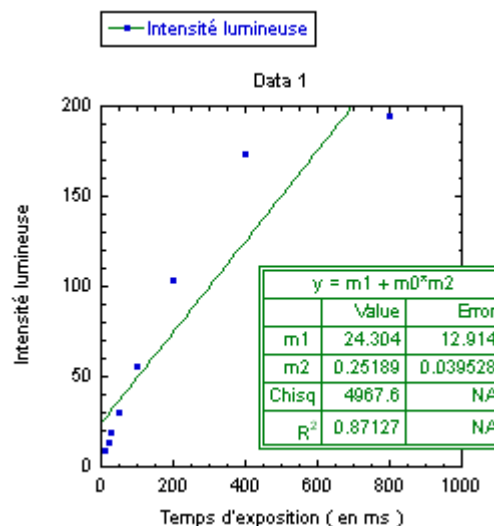


Image 4

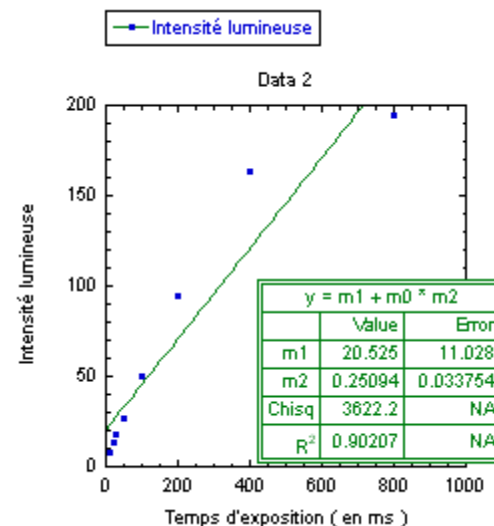


Image 3

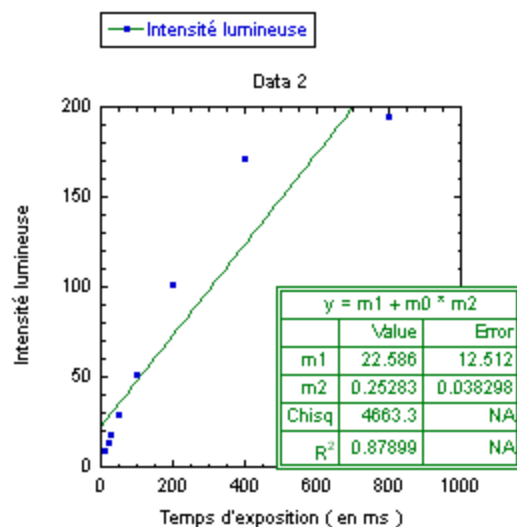


Image 2

Ici le R^2 n'est pas bon.
On va exclure les
points a 400 et 800
ms

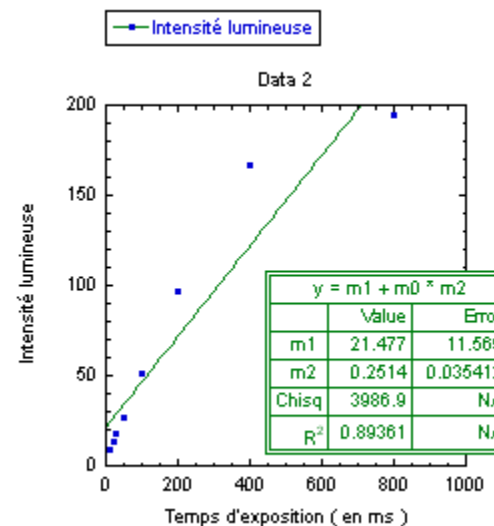


Image 1

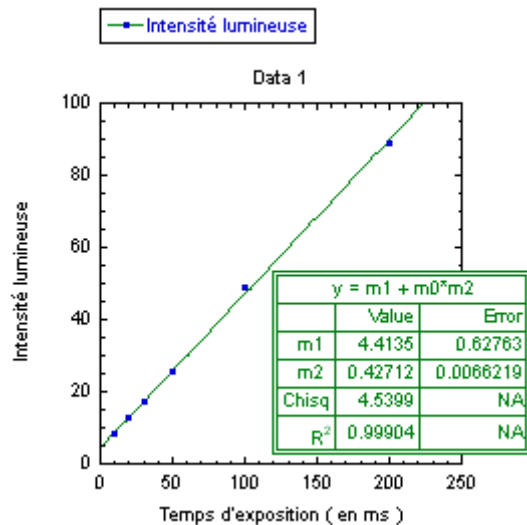


Image 5

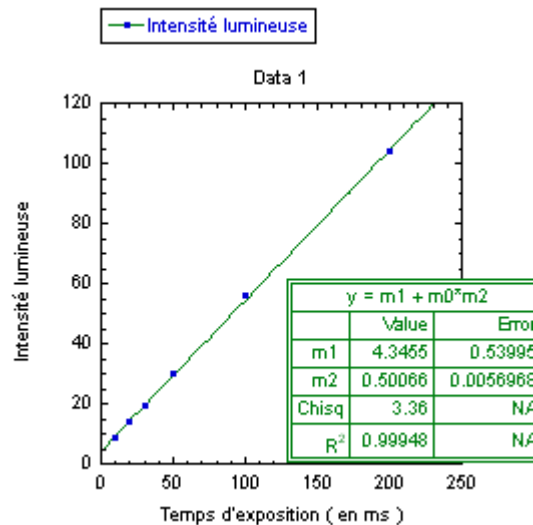


Image 4

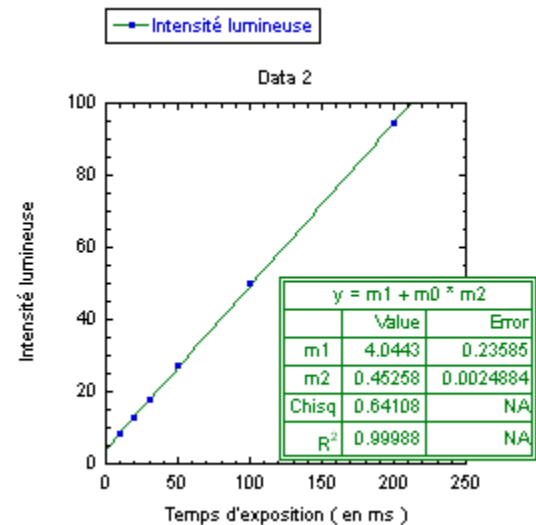


Image 3

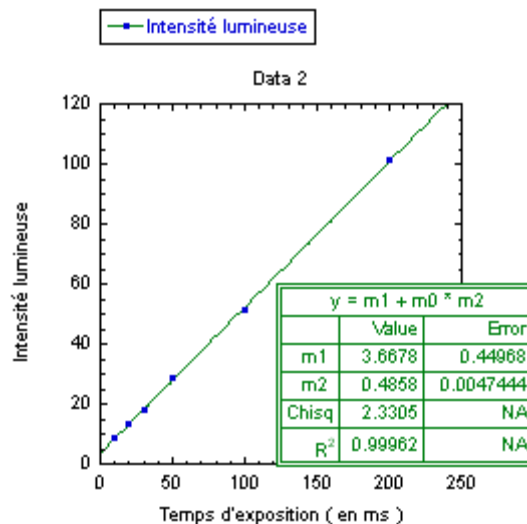


Image 2

Quand on enlève les points de la saturation on retrouve bien l'ordonnée à 0 de 4 environ.

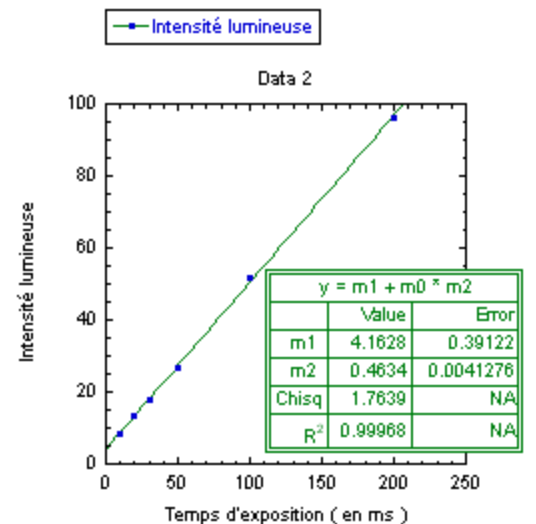
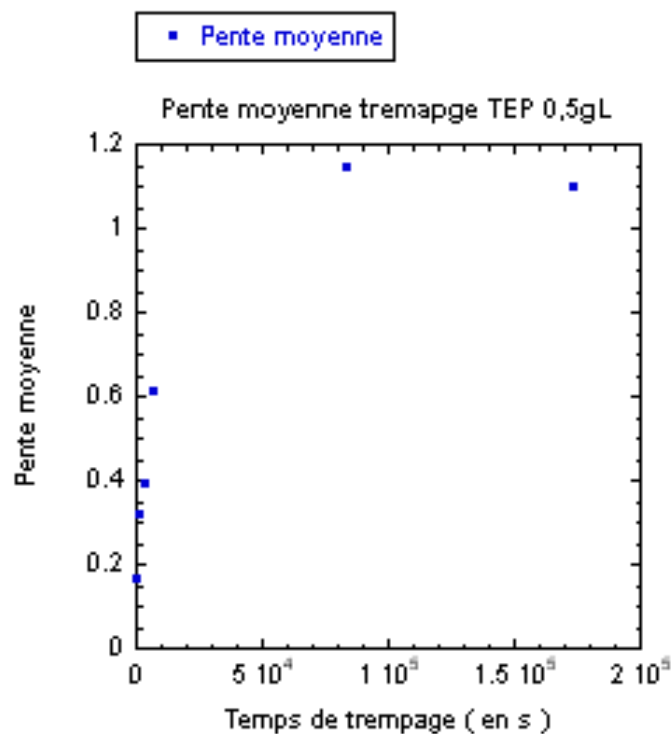


Image 1

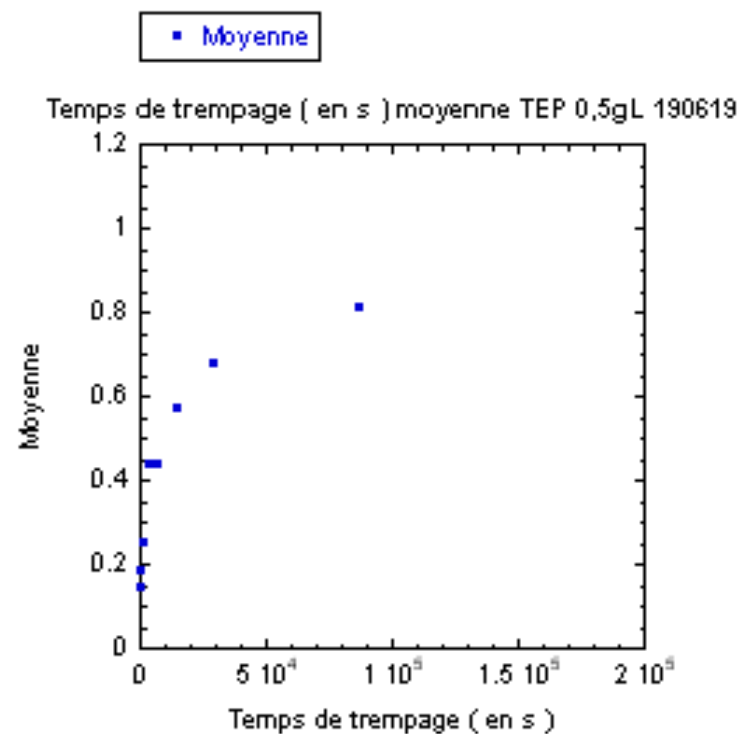
Conditions expérimentales

- On a 0,5g/L de TEP, manip faite deux fois.
- 0,03g de coton trempé dans 3mL de TEP + EAU + PKH67.
- Trempage entre 30s et 48h.
- Observé microscope x10 intensité 1 gain 1

TEP 0,5g/L 140619



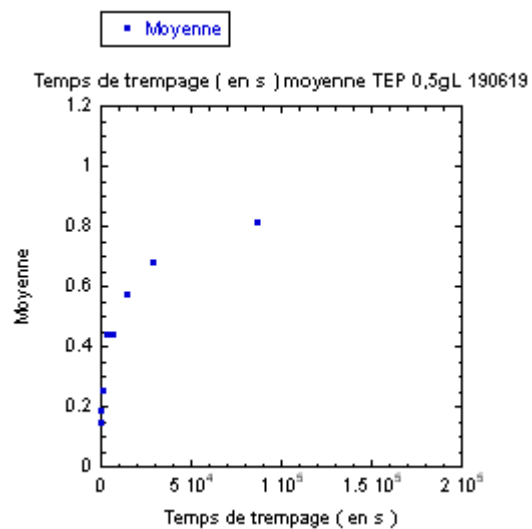
TEP 0,5g/L 190619



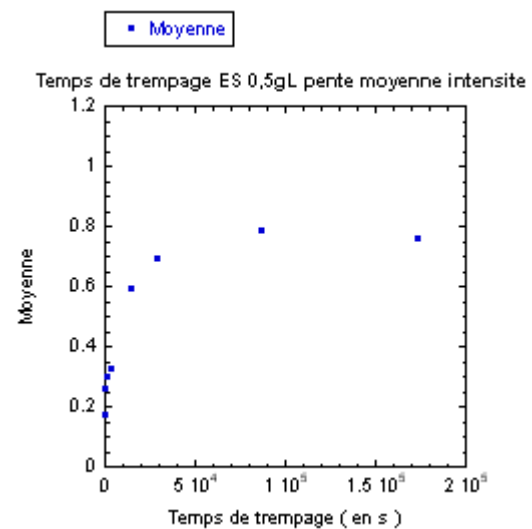
Conditions expérimentales

- ES 0,5g/L
- 0,03g de coton trempé dans ES.
- Mêmes conditions que pour TEP

TEP 0,5g/L 190619



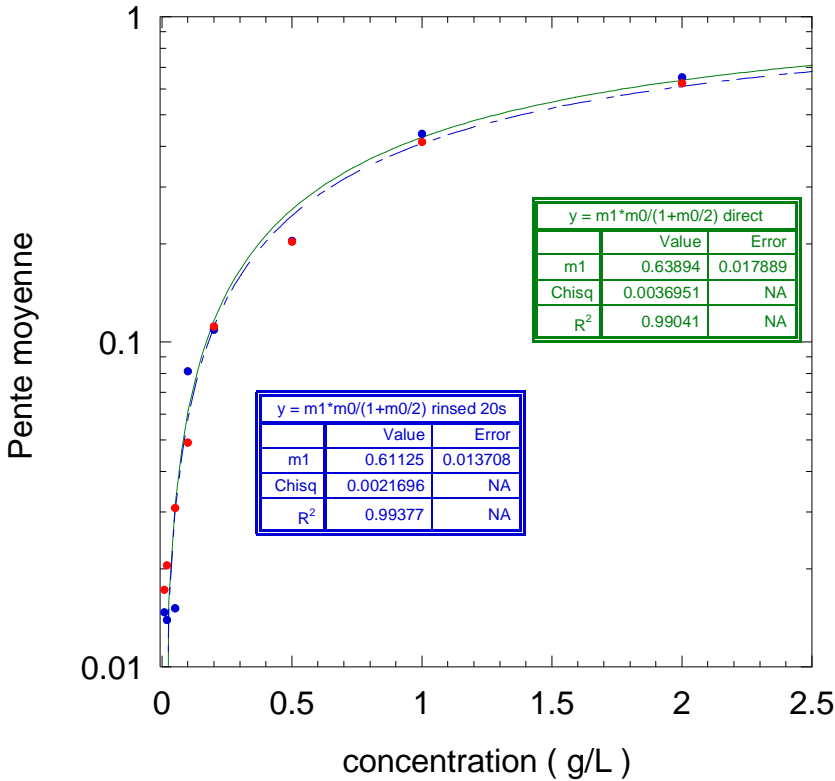
ES 0,5g/L 170619



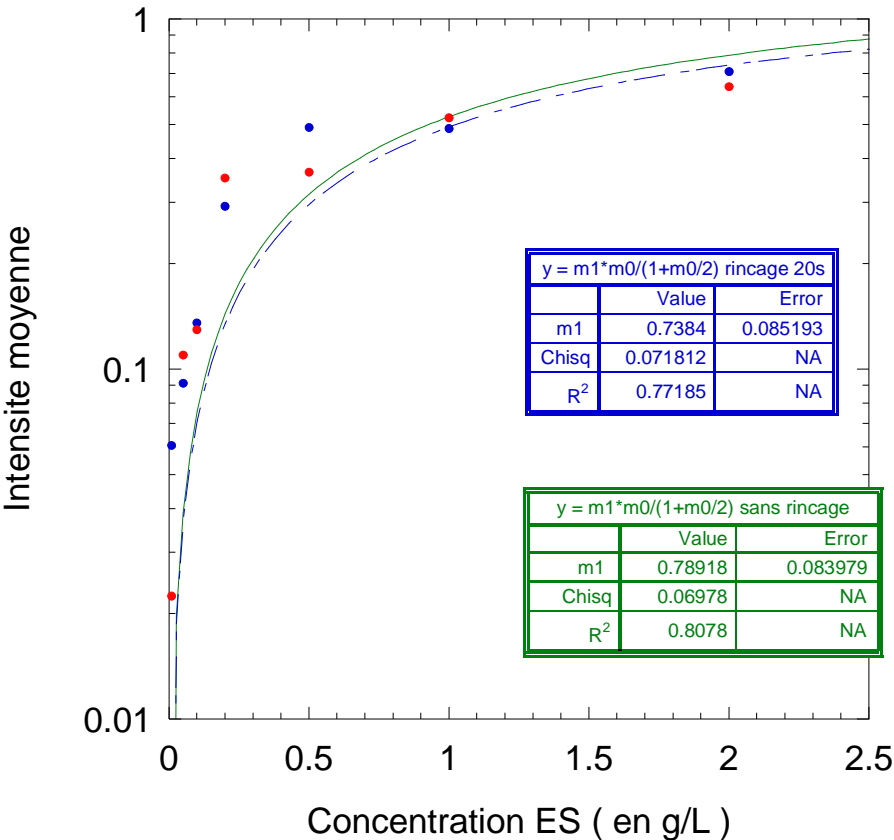
Concentration et intensité lumineuse

- TEP et ES concentré de 0.01g/L à 2g/L
- Rincé avec milliQ W pendant 20s ou directement.
- 0.03g de coton.
- Analysé sur lamelle avec 40µL de milliQ.
- Pas de prise en compte des points qui saturent.
- Microscopie fluo x10 intensité 1 gain 1

TEP

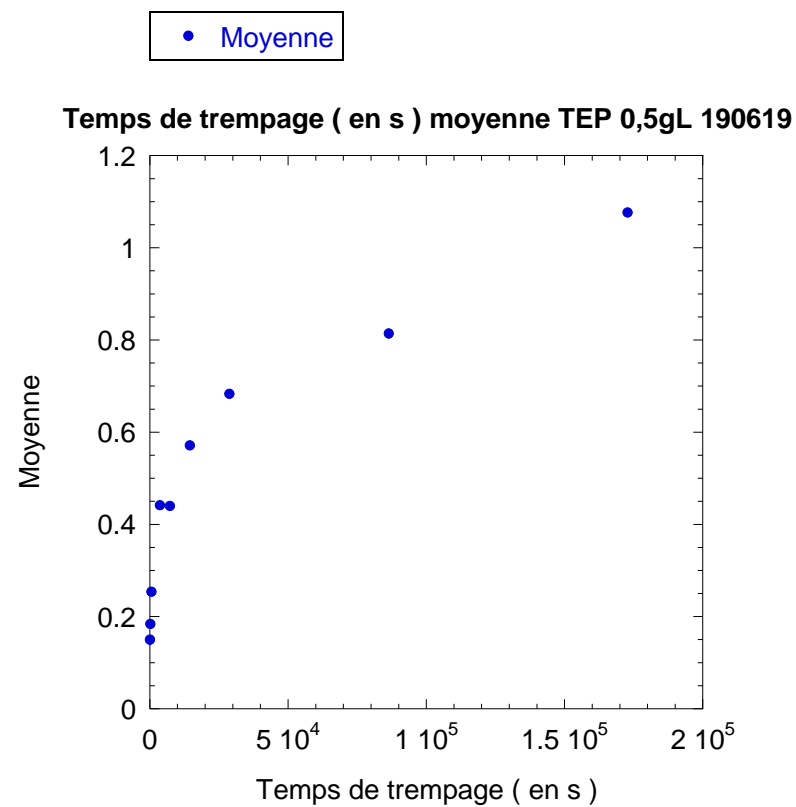
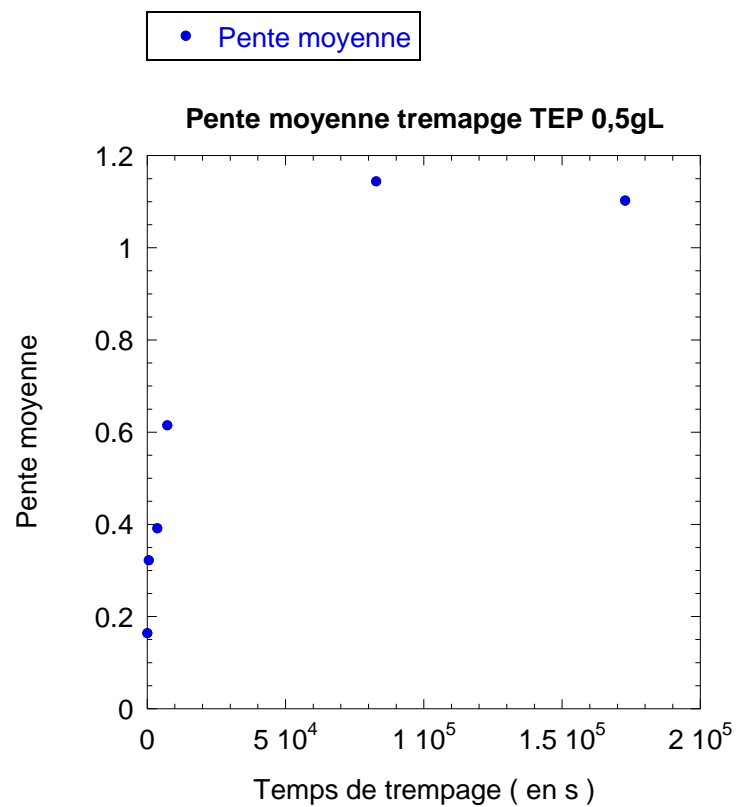


ES



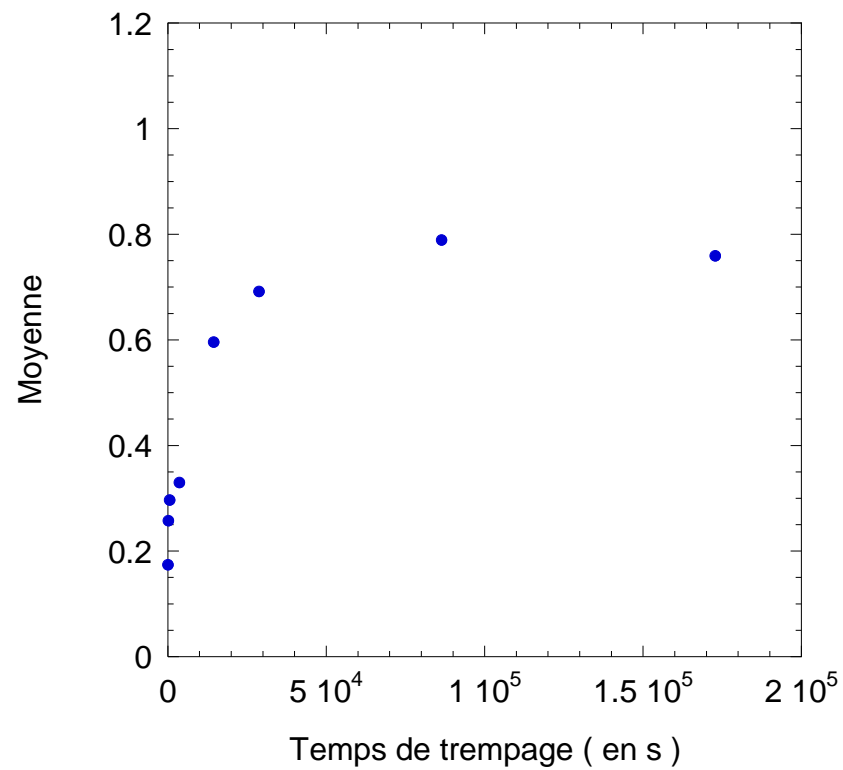
Influence temps de trempage

- ES et TEP 0,5g/L analysé avec 0.03g de coton
- Rincé pendant 20s dans milliQ water.
- Analysé sur lamelle avec 40 μ L de milliQ.
- Trempé pendant 30s, 3min ,10min, 1h, 2h, 4h, 8h, 24h et 48h.
- Microscopie fluo x10 intensité 1 gain 1





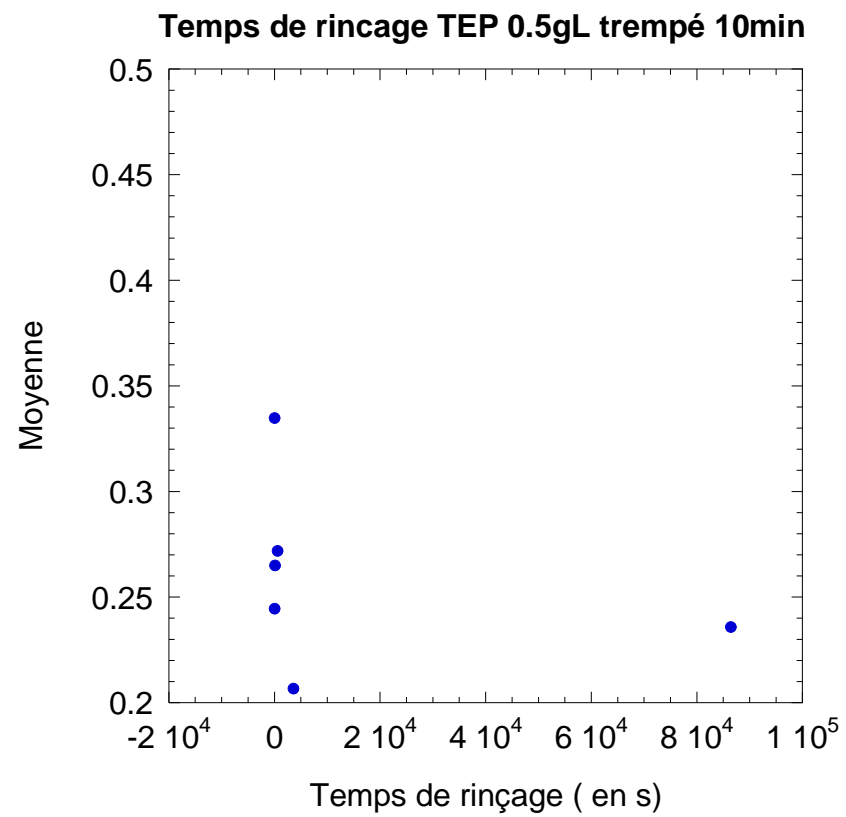
Temps de trempage ES 0,5gL pente moyenne intensite



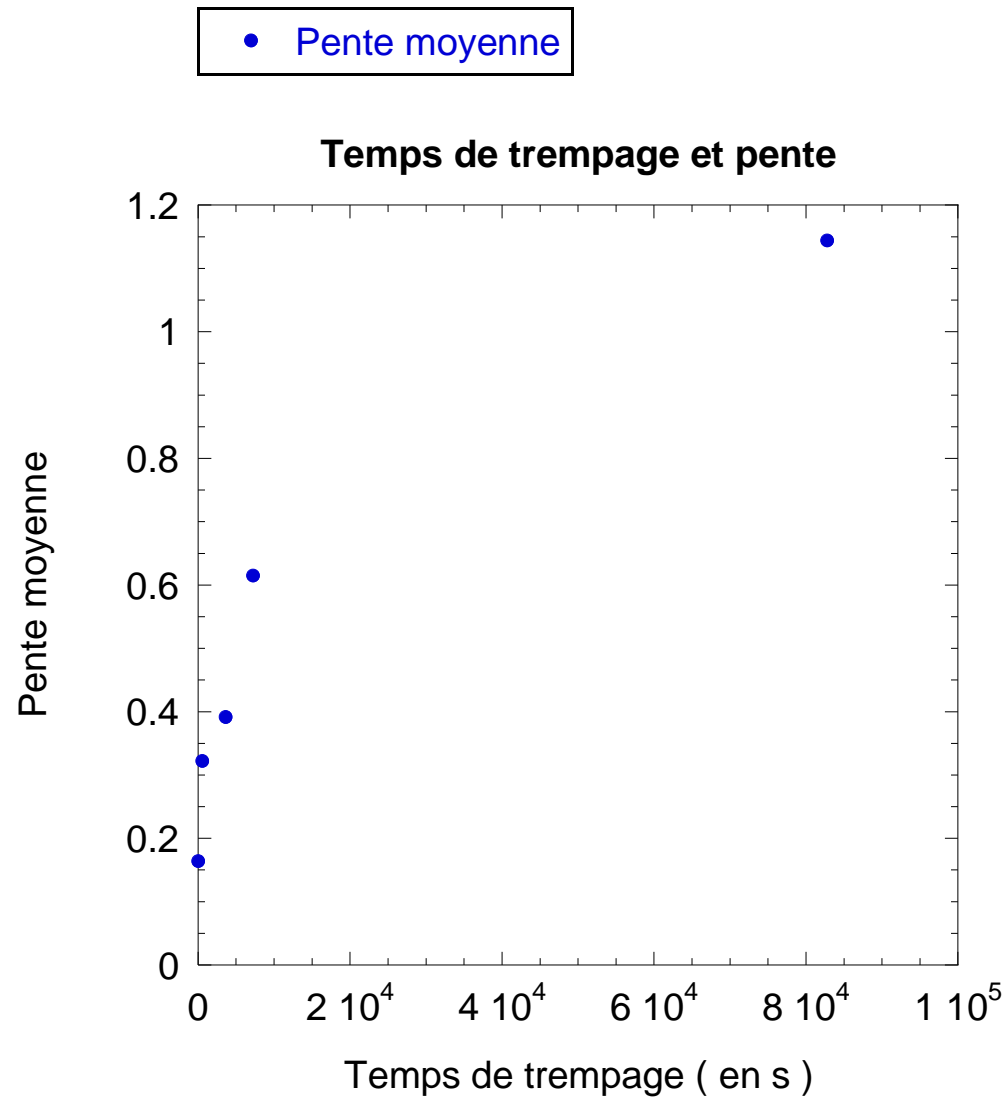
Influence du temps de trempage

- TEP à 0,5g/L trempé 10 min +PKH67 10μL
- Plusieurs temps de rinçage
- 0,03g de coton dans 3mL de solution rincé à la MilliQ et mis sur la lamelle avec 40μL de MilliQ

• Moyenne



- On trempe 0.03g de coton dans 3mL de TEP + PKH67 10 μ L concentré à 0.5g/L.
- Dans le premier cas, on regarde l'influence du temps de trempage dans la dispersion. On a laissé tremper 30s, 10min, 1h, 2h et 23h.
- Dans le second cas, on a laissé tremper 10 min et on a rincé dans de l'eau DI 0s (direct) 20s, 2 min, 10 min, 1h et 24h

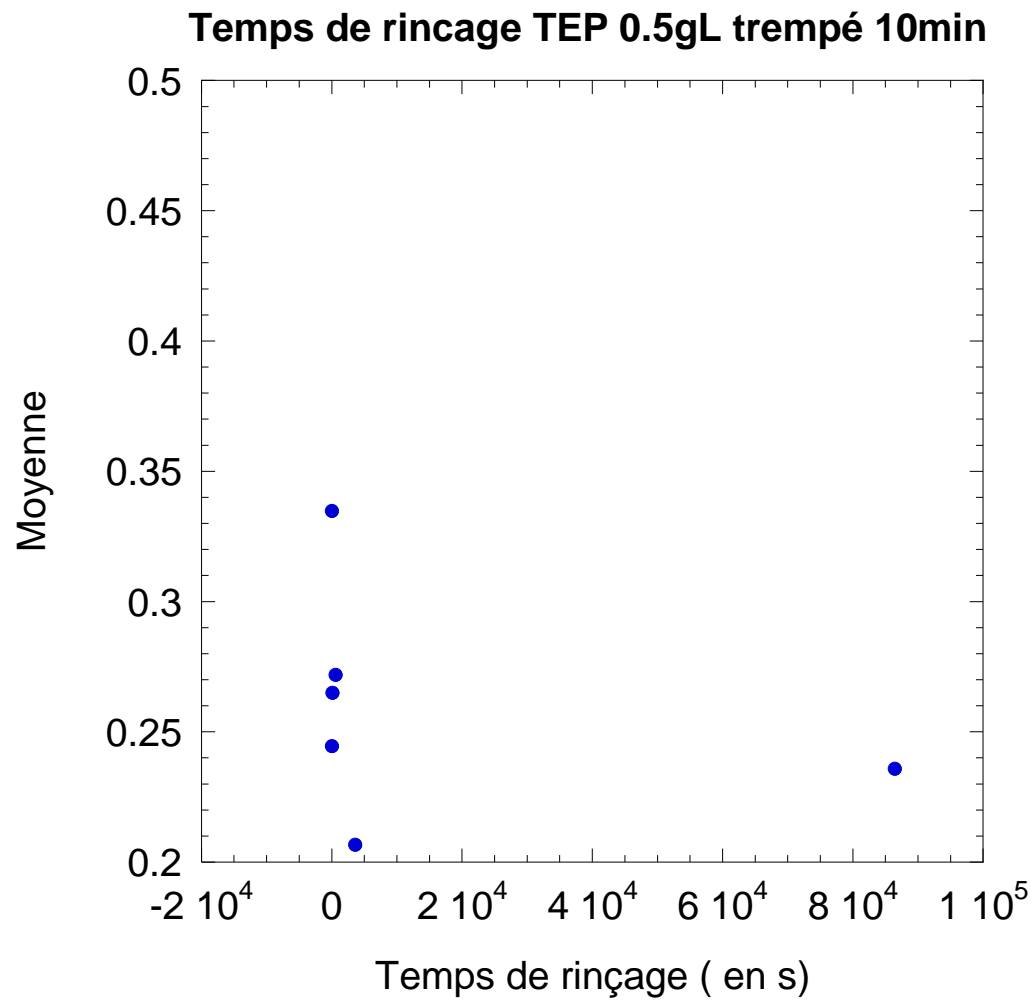


Conclusion

- Il semble que le temps de trempage dans la dispersion ait un impact sur la quantité de vésicules déposées sur les fibres de coton. En effet, plus on laisse longtemps, plus l'intensité lumineuse est grande donc plus il y a a priori de vésicules sur le coton. L'évolution a l'air d'atteindre un palier de saturation => à vérifier en ajoutant des points entre 2h et 23h et après 23h de trempage.

Etude influence temps de rinçage

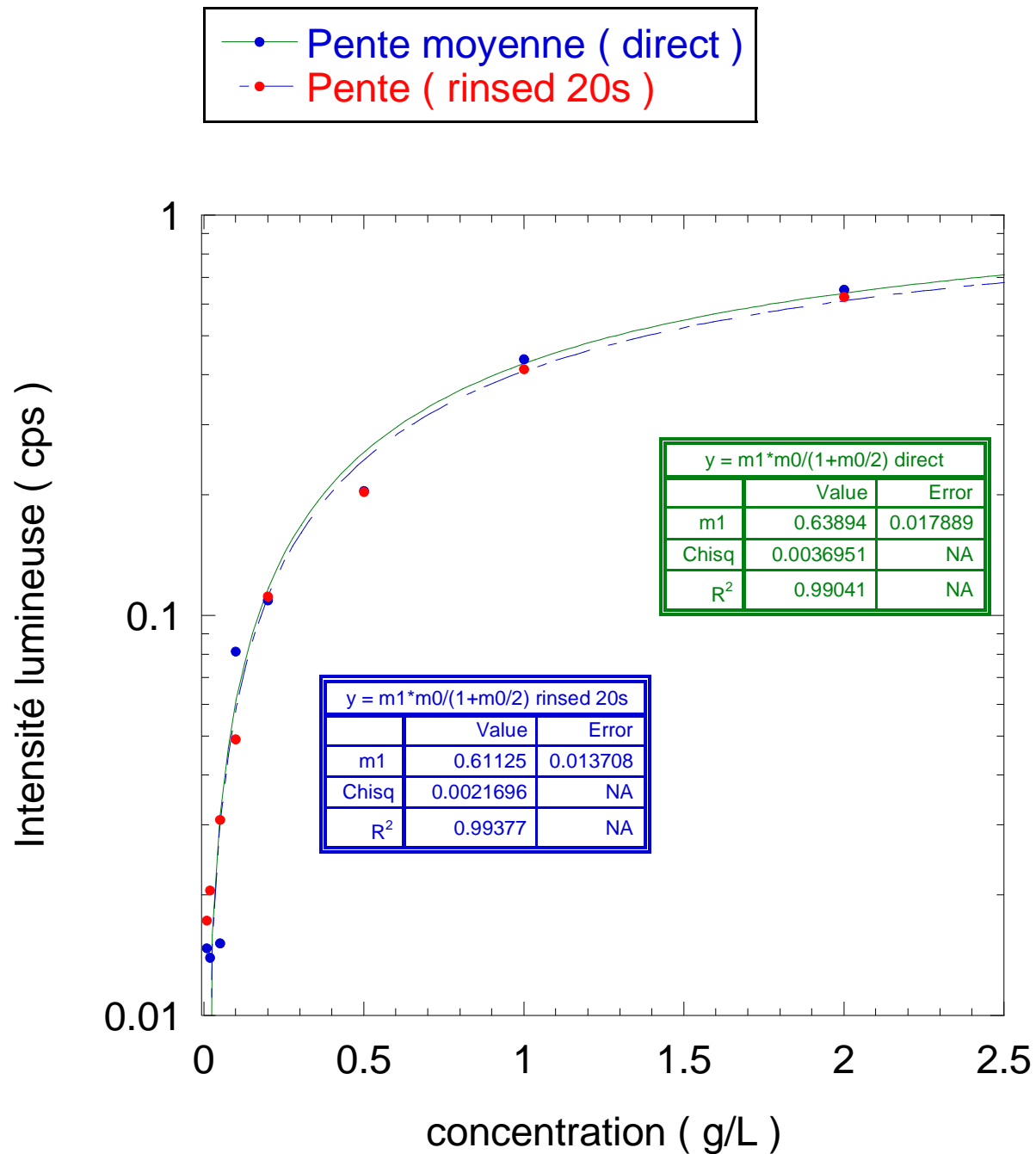
- Conditions expérimentales
 - 0,03g de coton trempé dans 3mL de TEP 0,5g/L avec 10 μ L de PKH67 pendant 10 min
 - Plusieurs temps de rinçage (0 s à 24h)
 - Microscopie fluo x10 intensité 1 gain 1



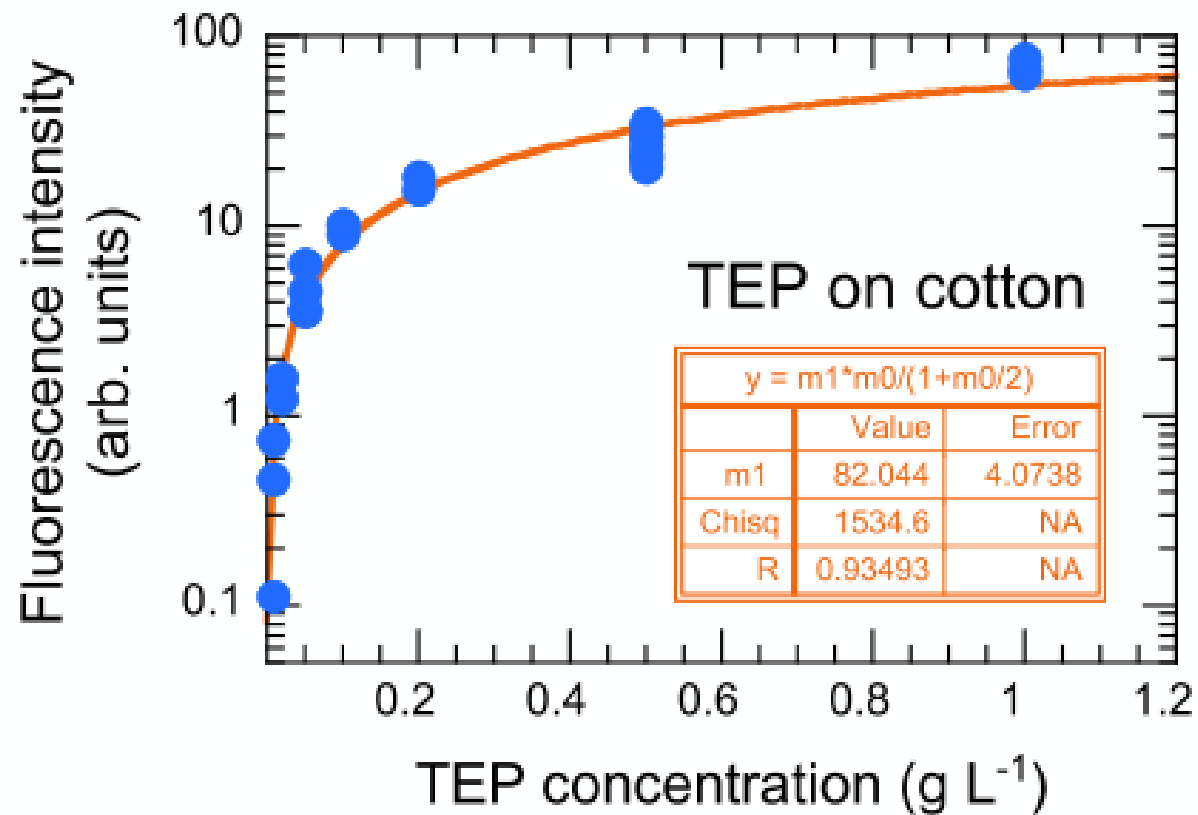
- Il semble que le rinçage et le temps de rinçage n'ont pas d'effet vraiment notable sur la quantité de vésicule sur le coton. Observé directement sans rinçage, le coton semble légèrement plus lumineux, mais peu importe combien de temps on le rince il reste plus ou moins toujours aussi lumineux.

Concentration TEP

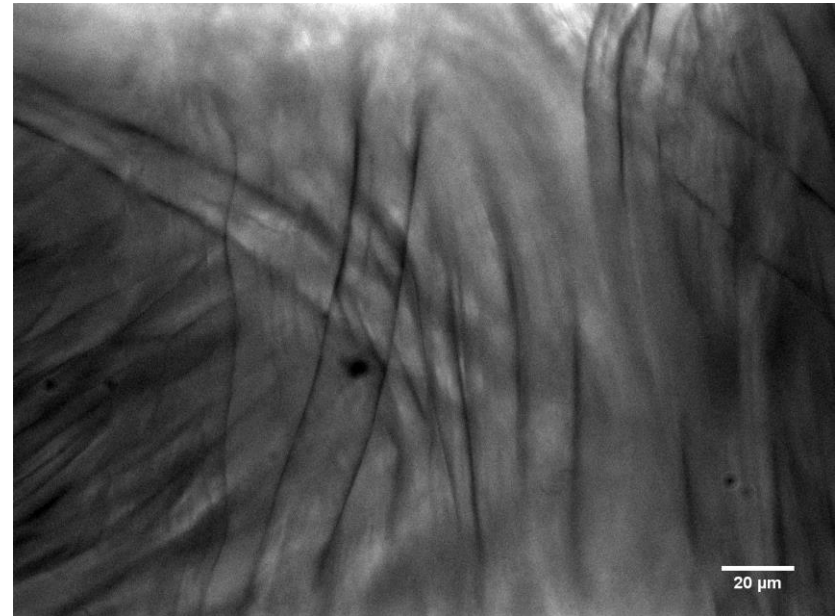
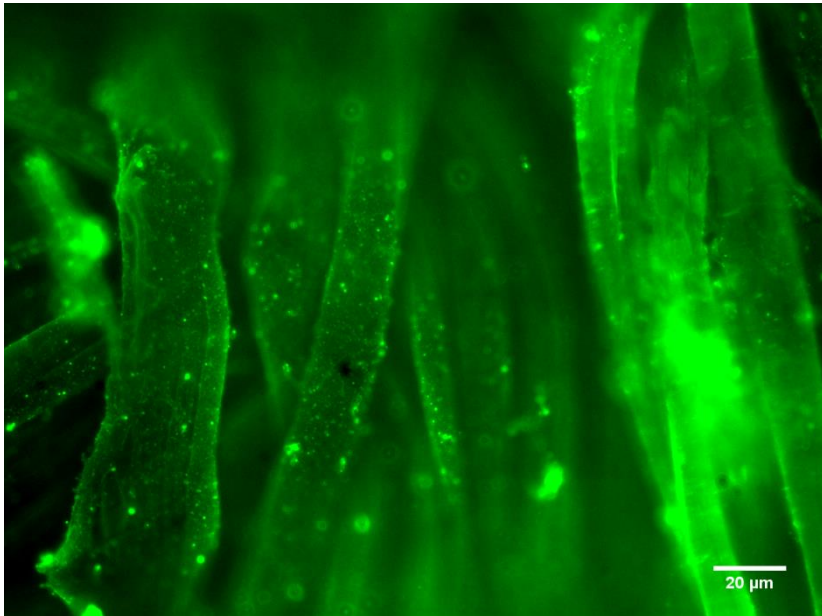
- 0.0,3 de coton TEP de 0,01 à 2g/L pendant 10min
- Rinçage soit 0s soit 20s
- Microscope fluo x10 intensité 1 gain 1 sur lamelle avec 40 μ L milliQ water



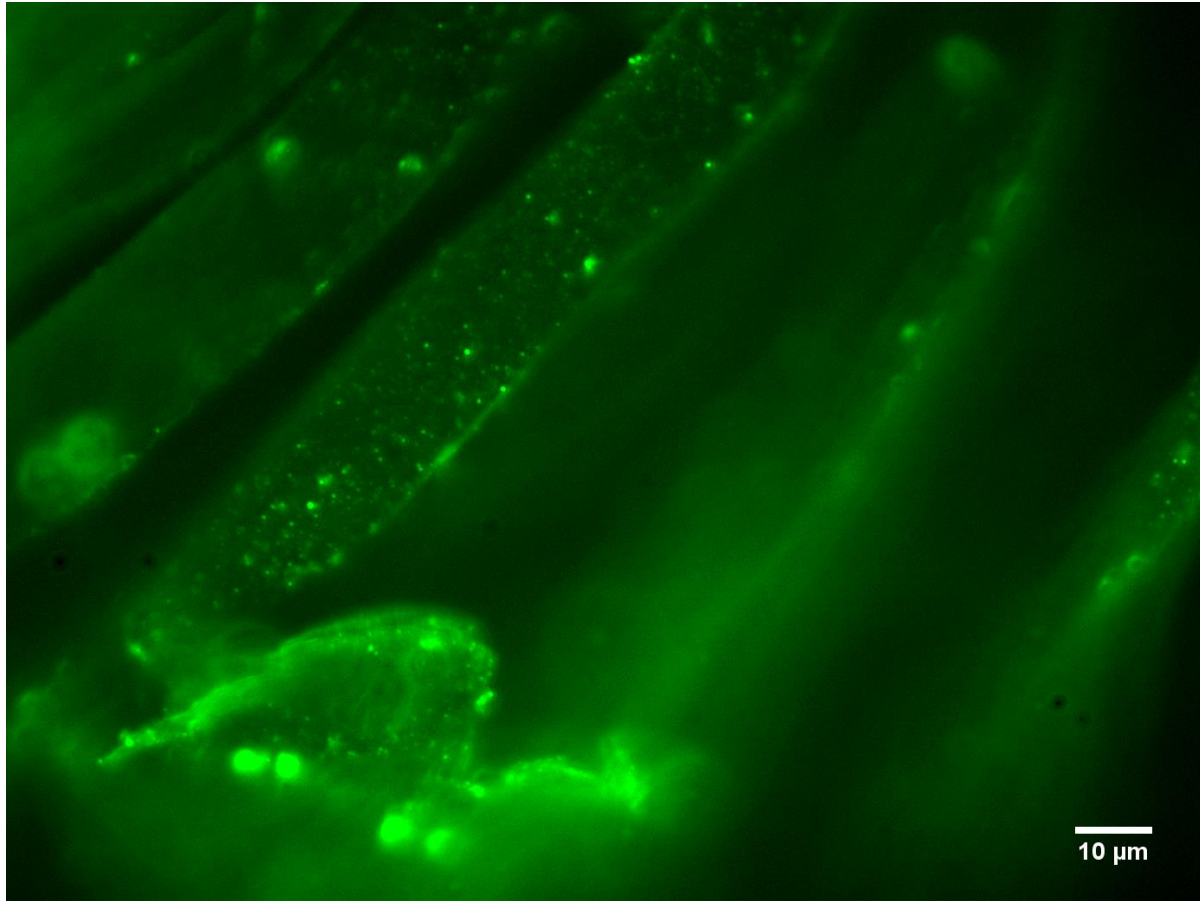
The fitting curve is a Langmuir isotherm



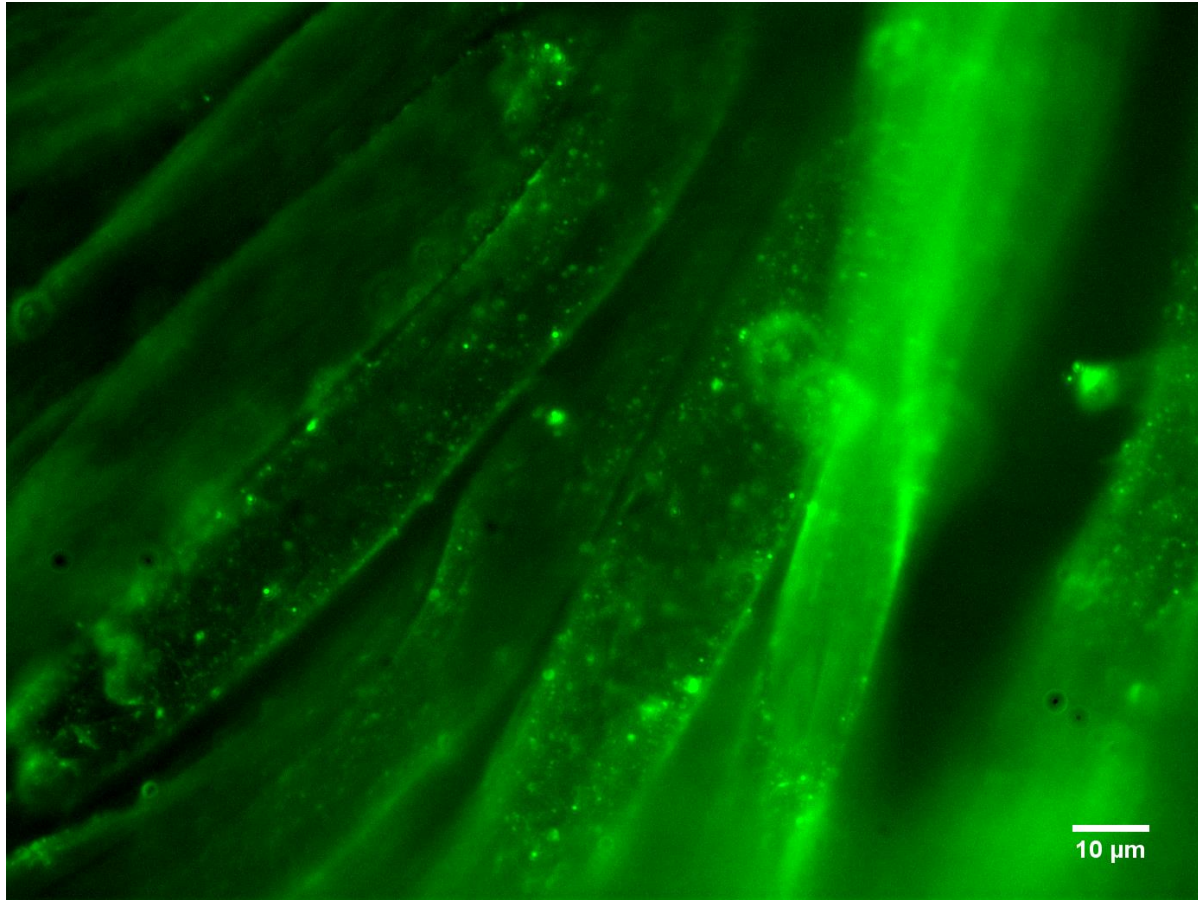
TEP 0.05% x40 opposition fluo / phase contrast



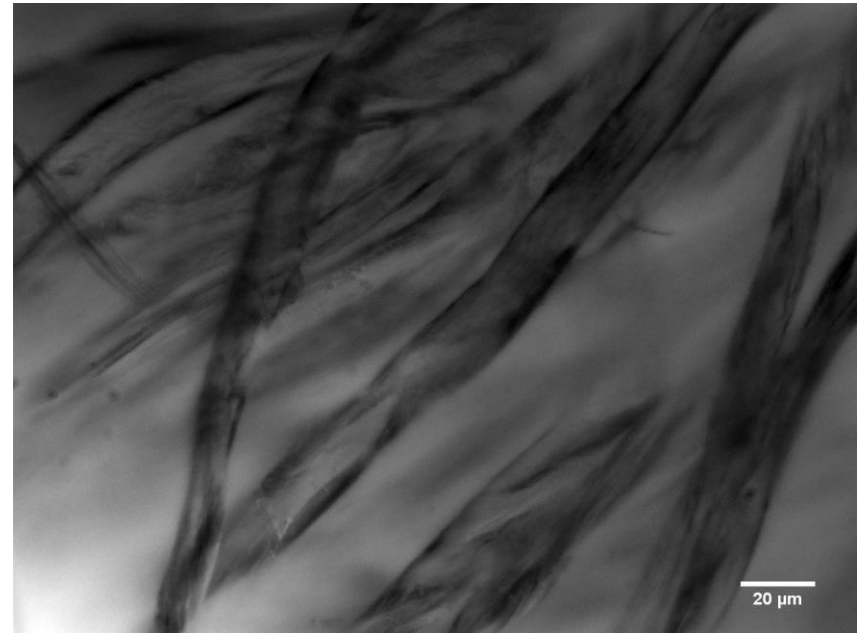
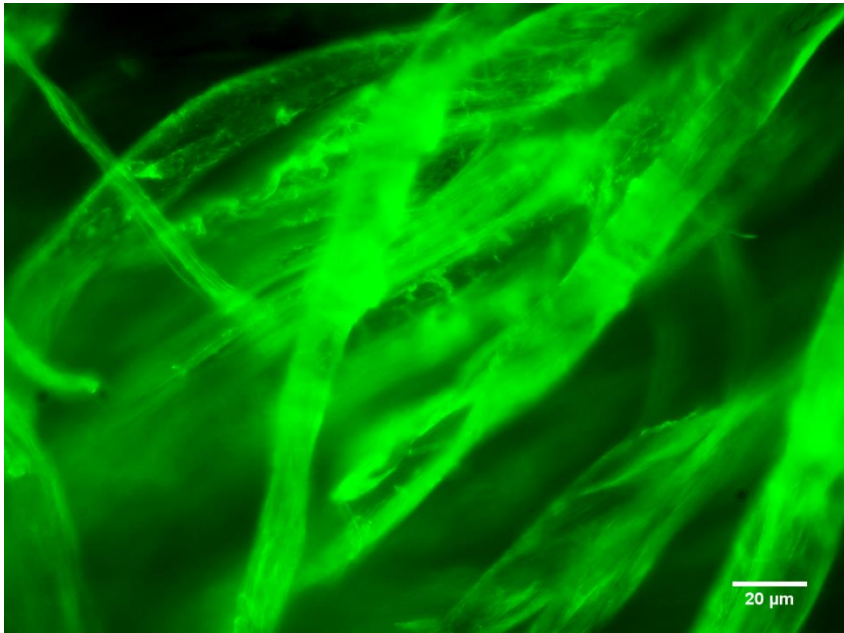
TEP 0.05% rincé 20s x60



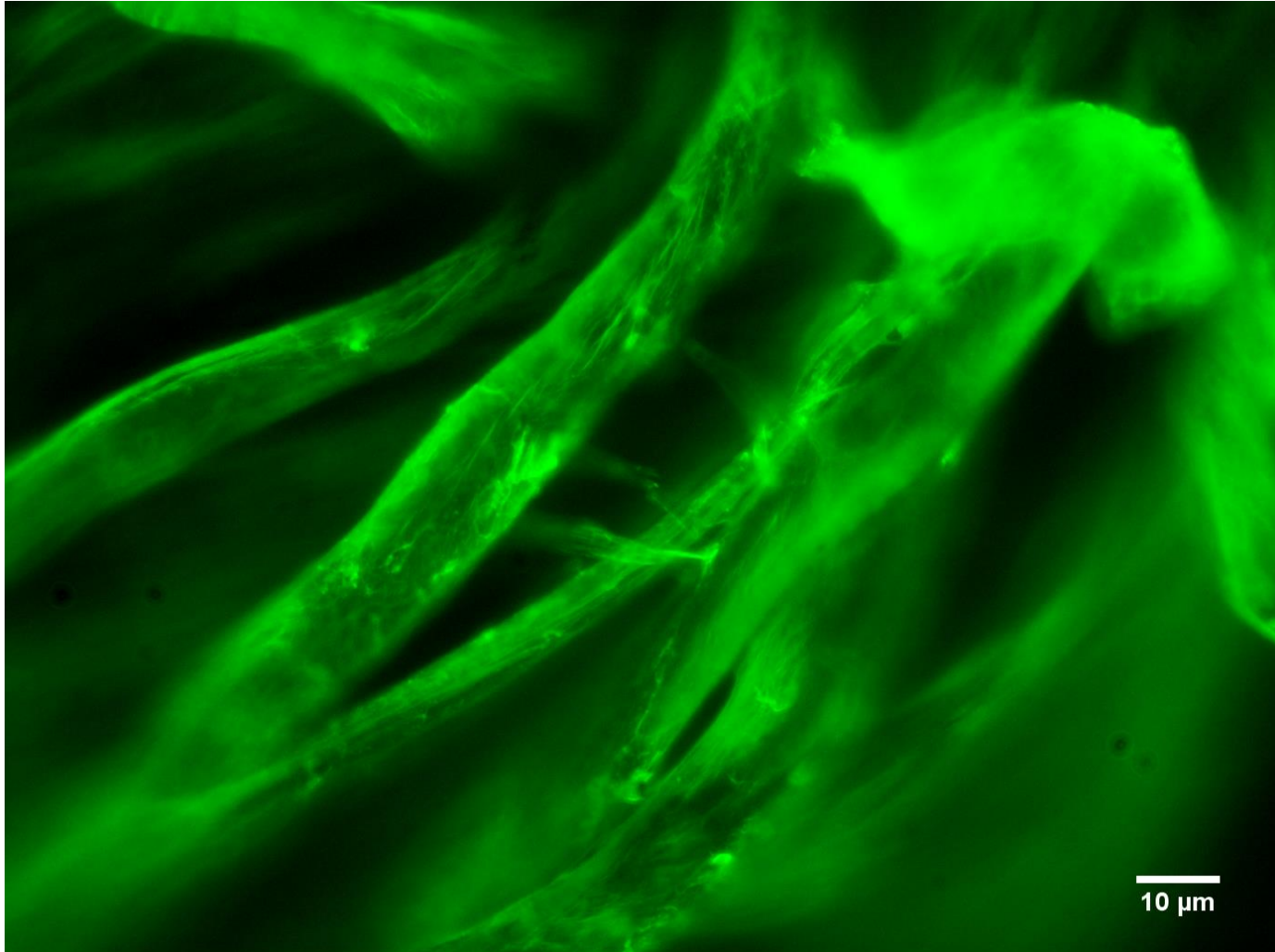
TEP 0.05% rincé 20s x60



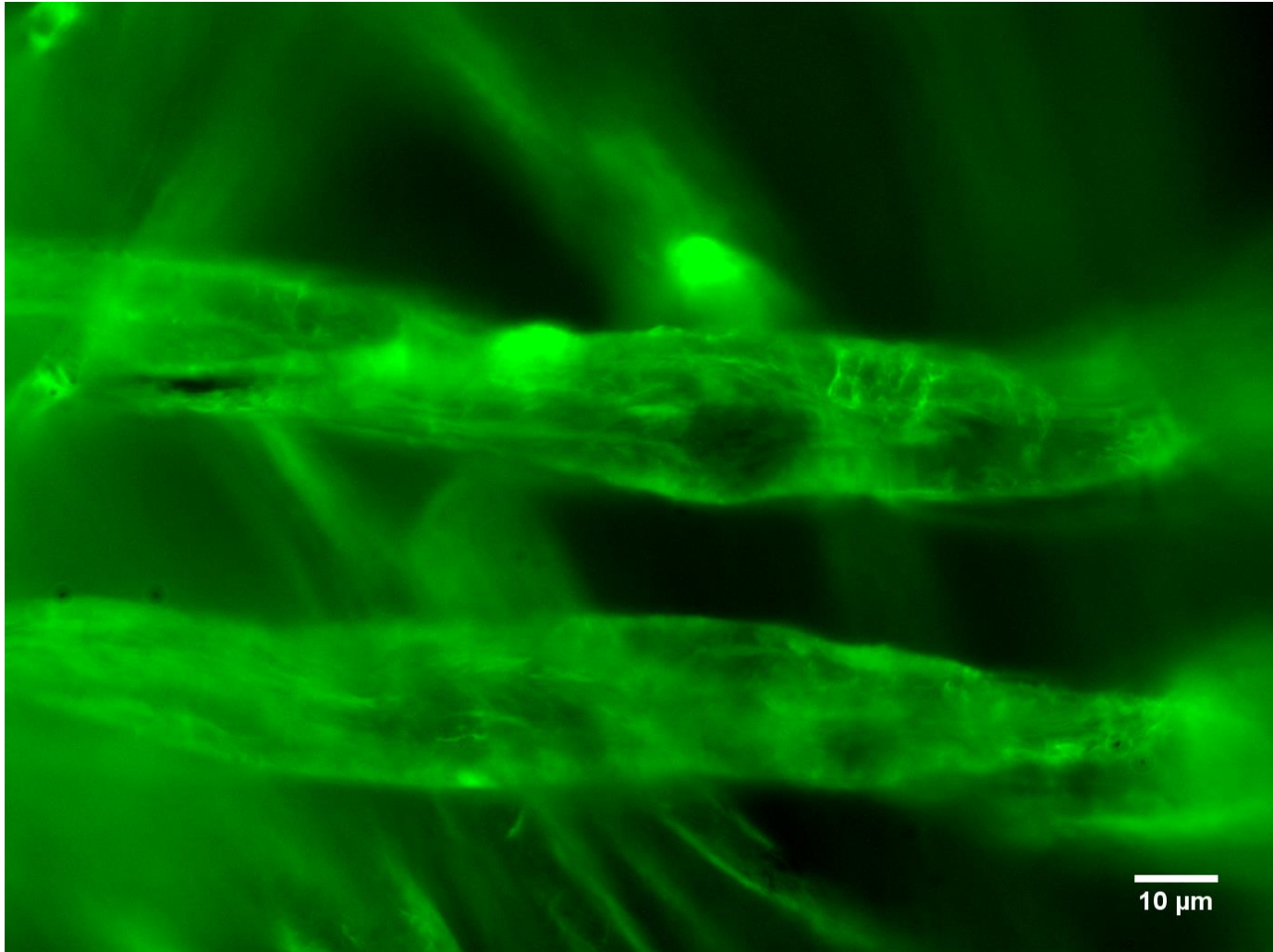
**TEP 0.05% rinsed 20s dried
x40 opposition fluo /
bright field**



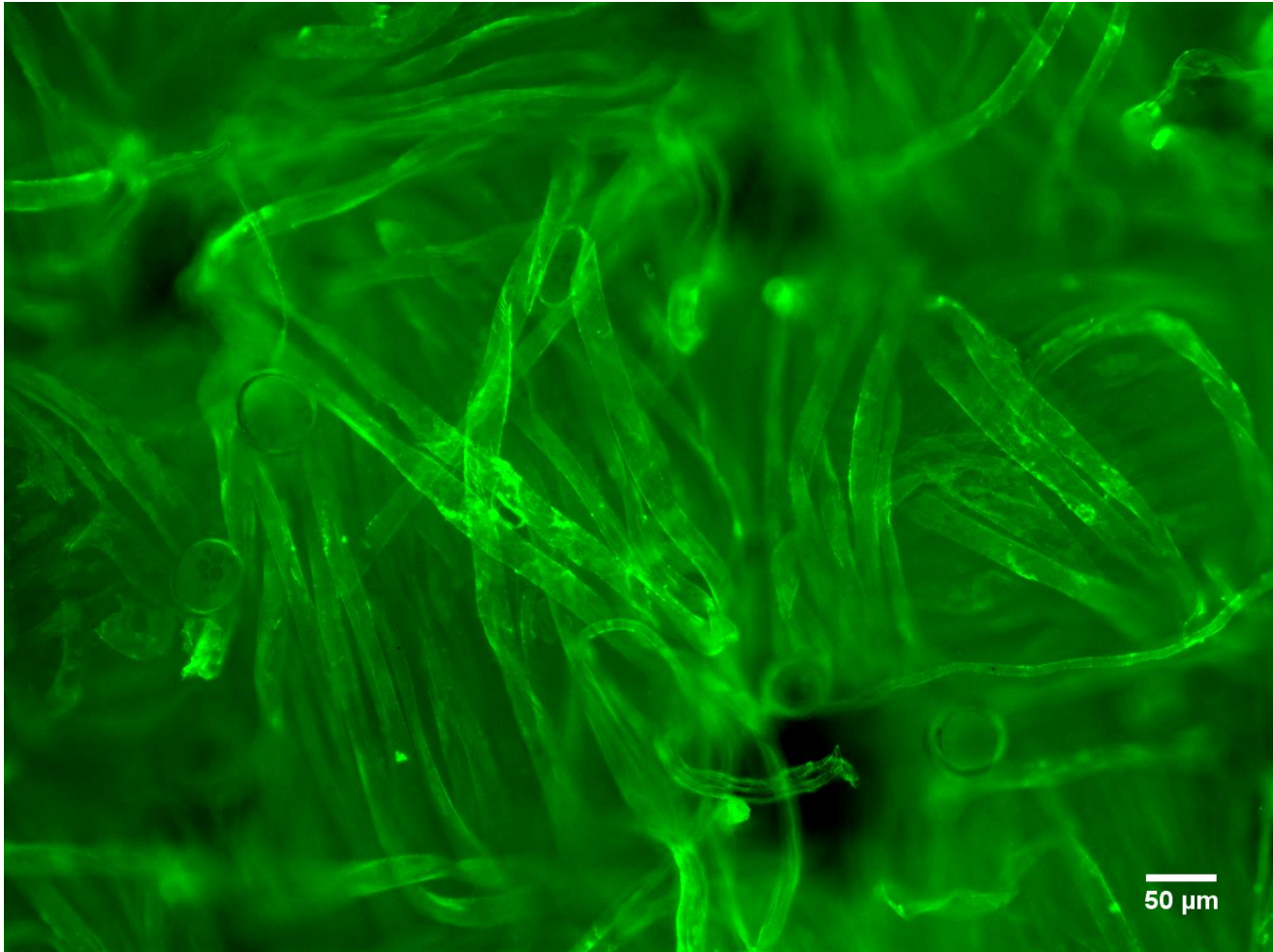
**TEP 0.05% rincé 20s
séché x60**



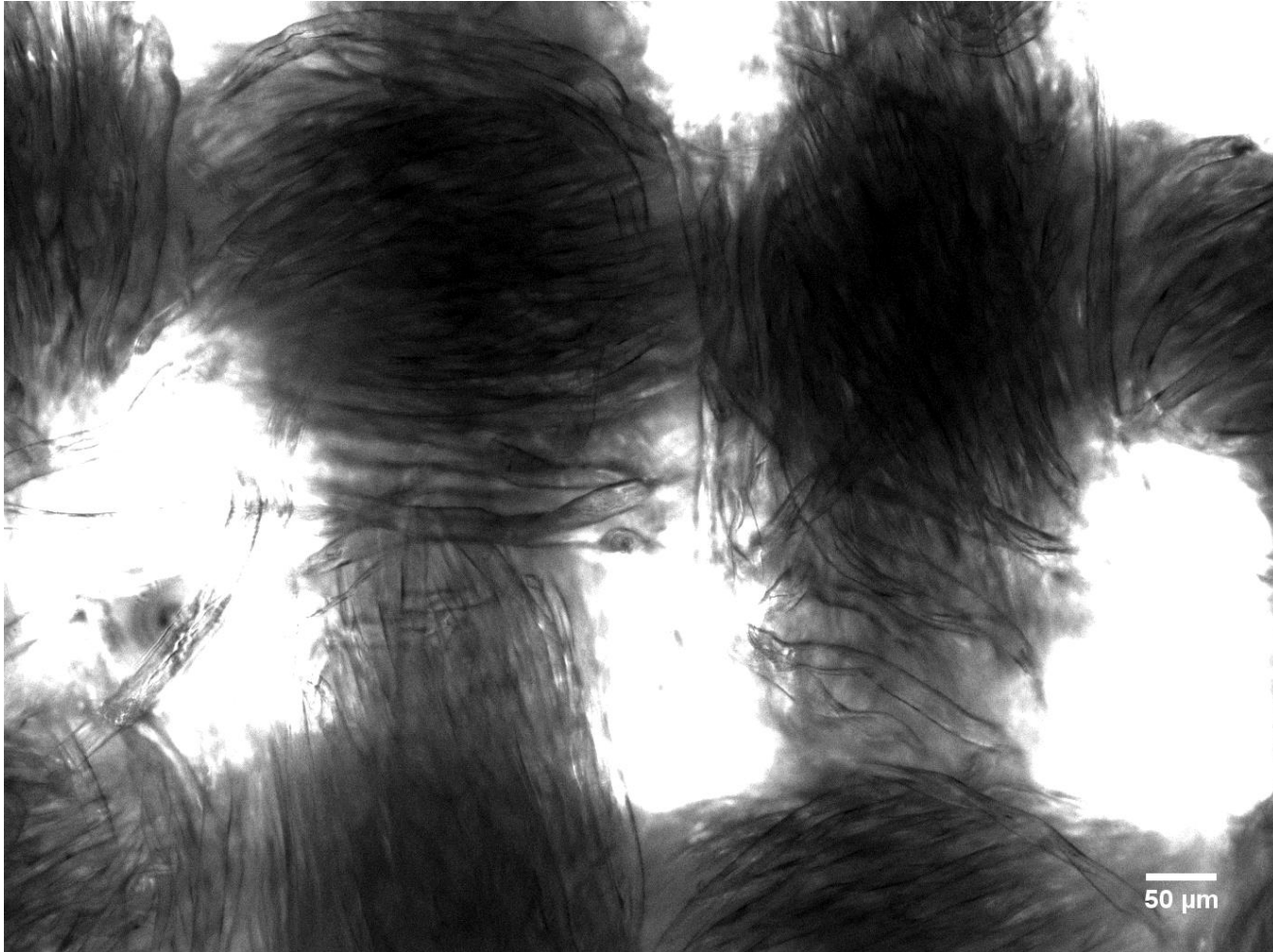
**TEP 0.05% rincé 20s
séché x60**



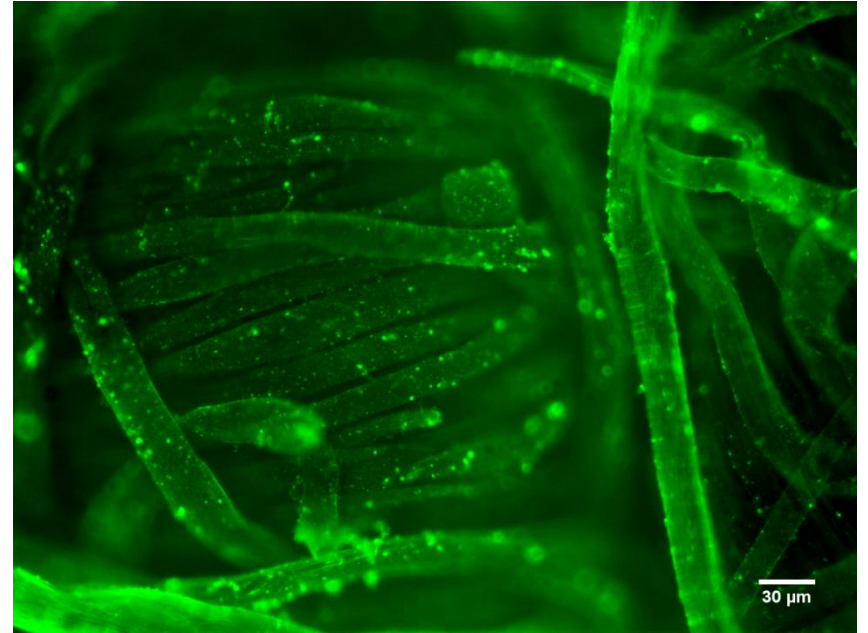
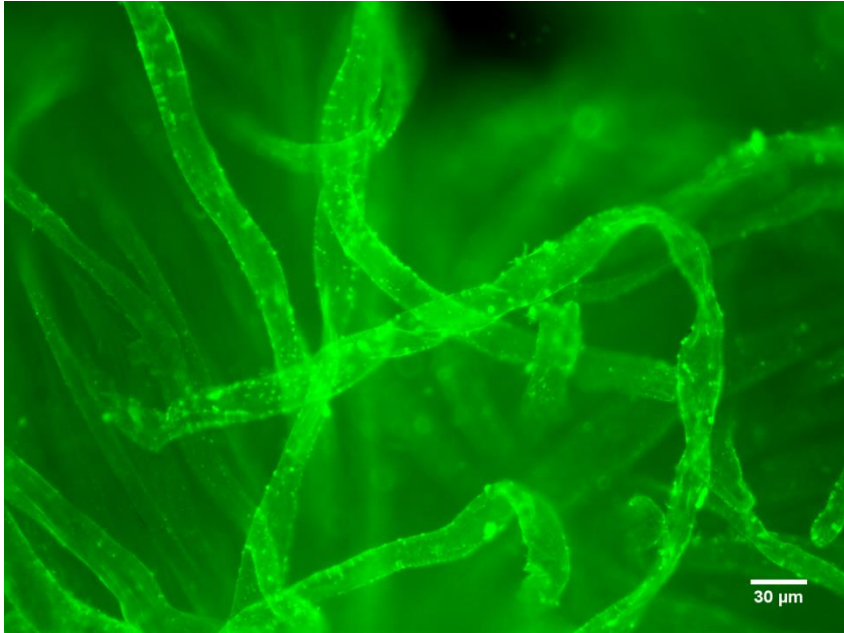
TEP 0.05% séché remouilléx10



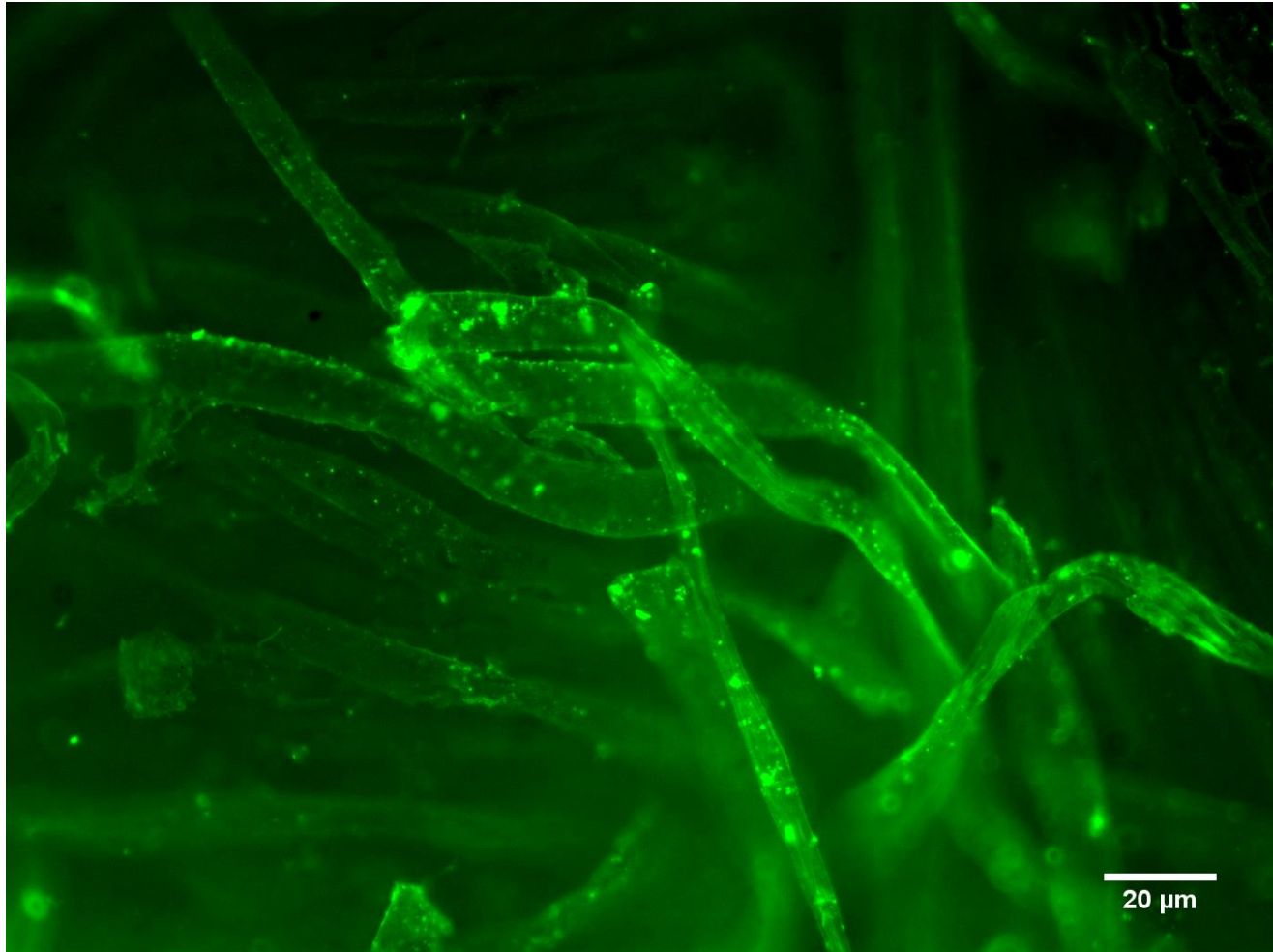
TEP 0.05% observé sans rinçage x10 bright field



TEP 0.05% observé sans rinçage x20



TEP 0.05% observé sans rinçage x40



04-05/06/2019

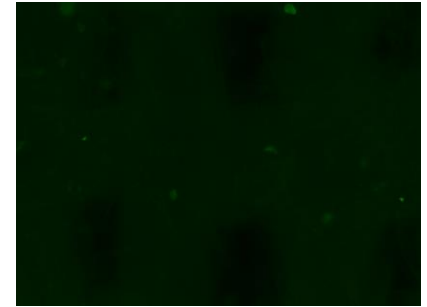
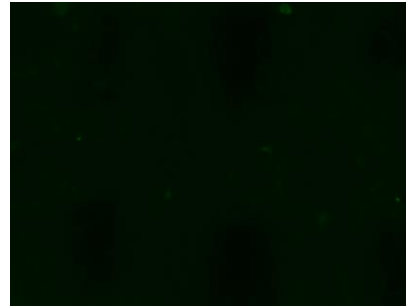
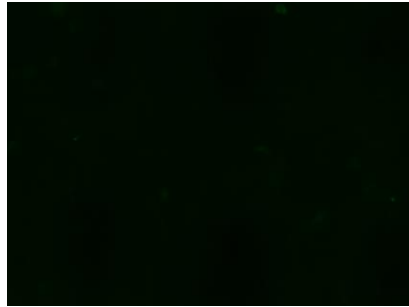
- TEP concentré à 0.02 – 0.05 – 0.1 – 0.2 – 0.5 – 1 g/L
- Préparé le 04 et le 05/06 2019
- Echantillon analysé au microscope fluo, x10, gain1 et lampe intensité 1 avec des temps d'exposition de 10 a 4000 ms
- Coton rincé 20s dans milliq water, ou non rincé. Posé sur la lamelle avec 40µL de milliq water.

10ms

20 ms

30 ms

50 ms

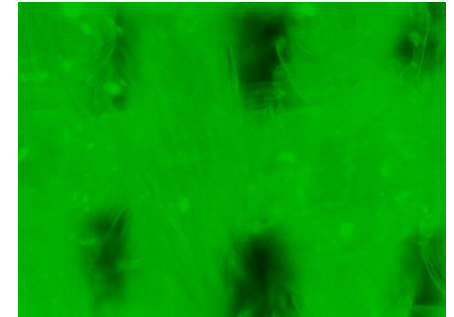
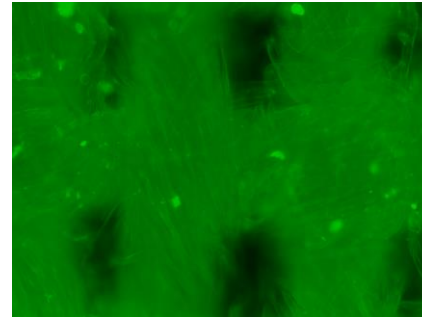
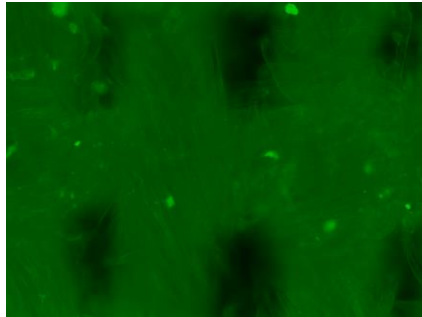
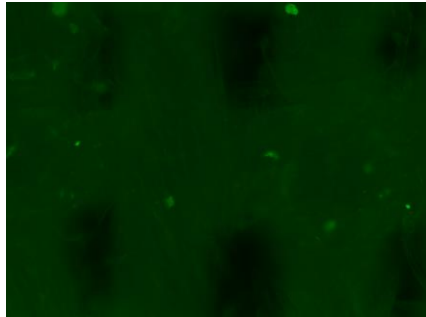


100 ms

200 ms

300 ms

500 ms

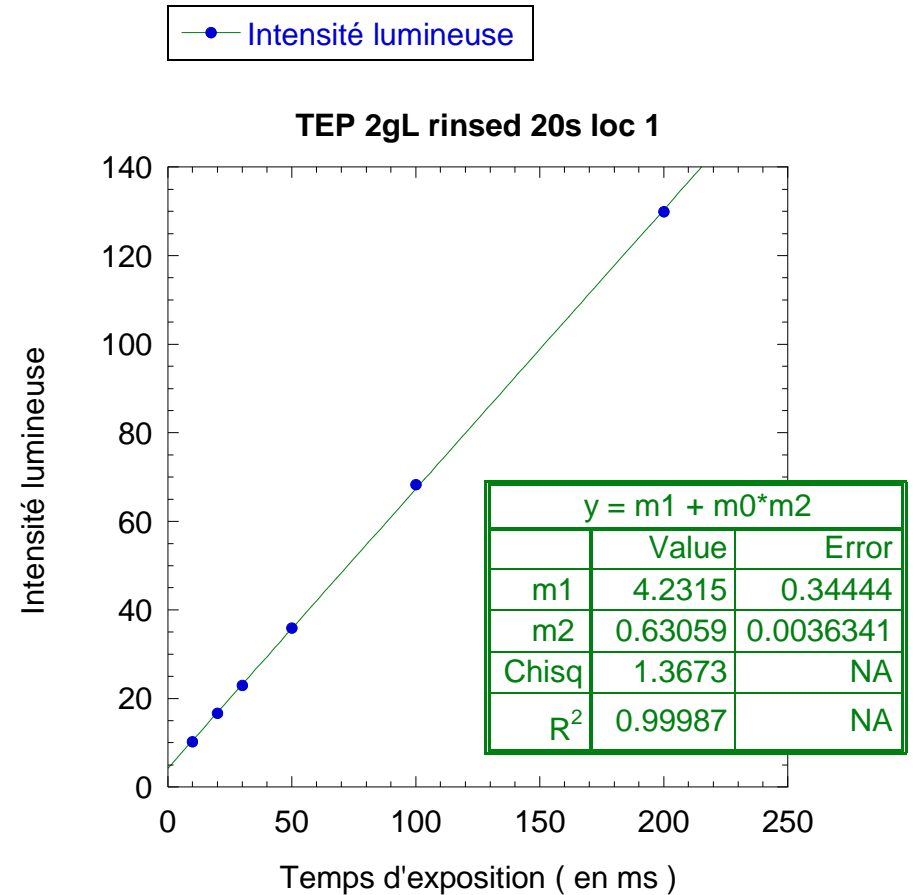
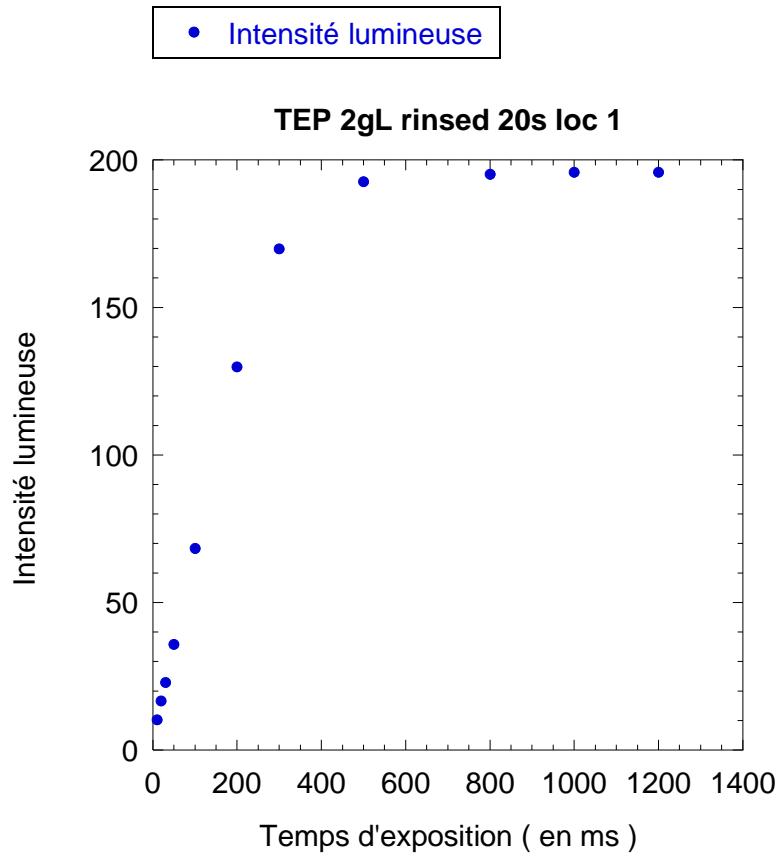


800 ms

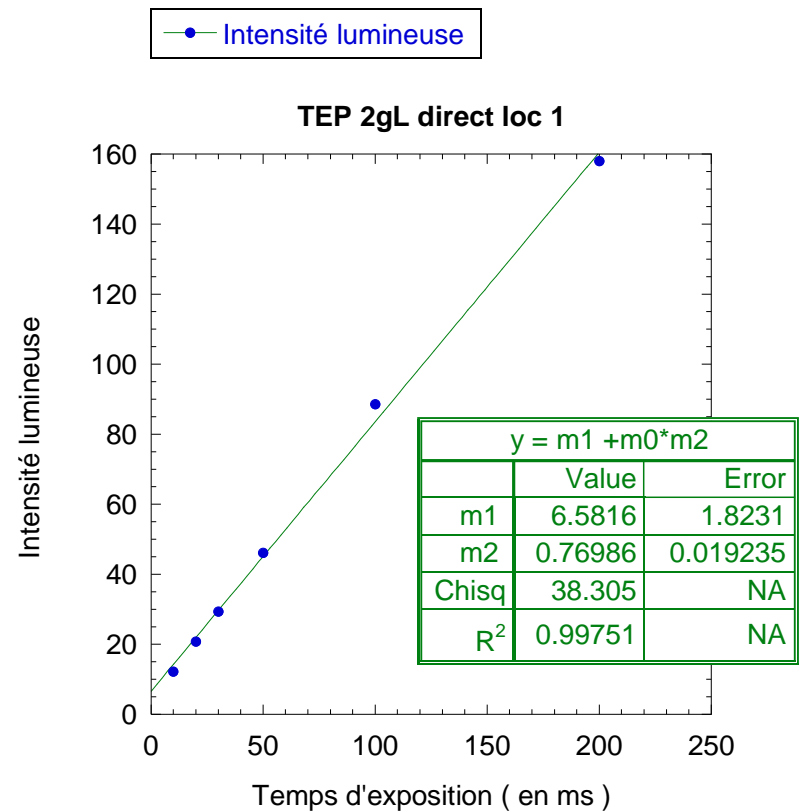
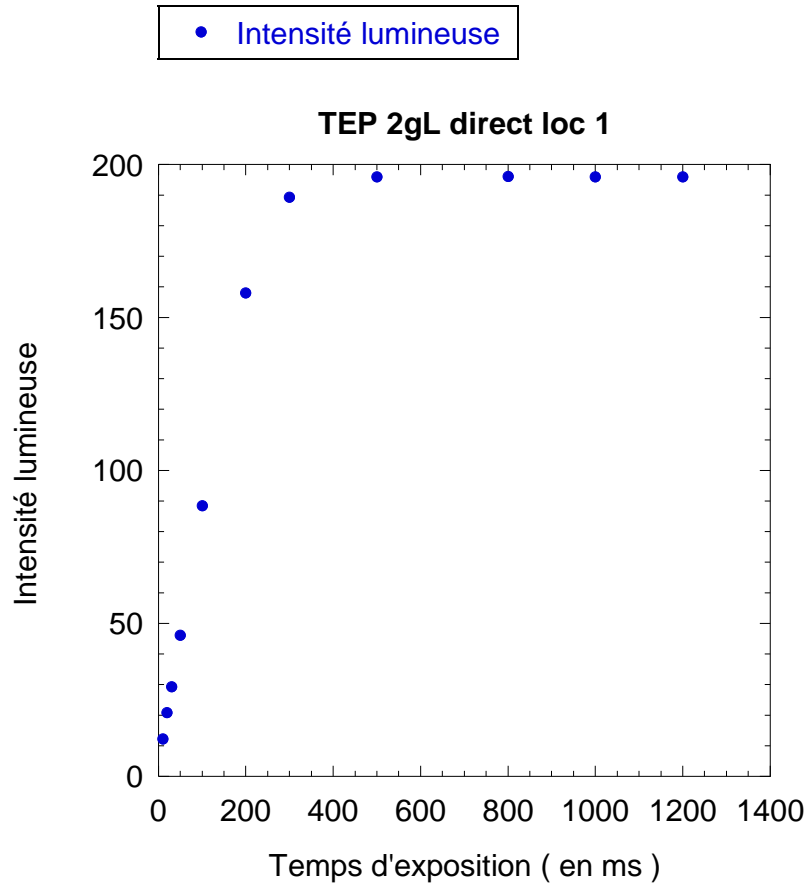
1000 ms



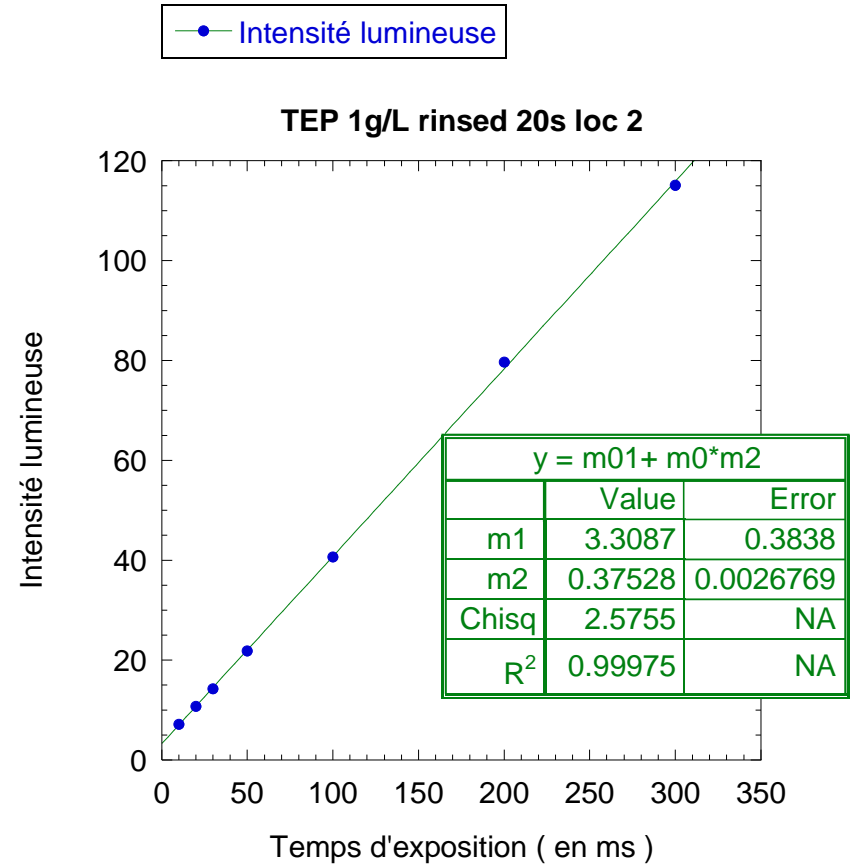
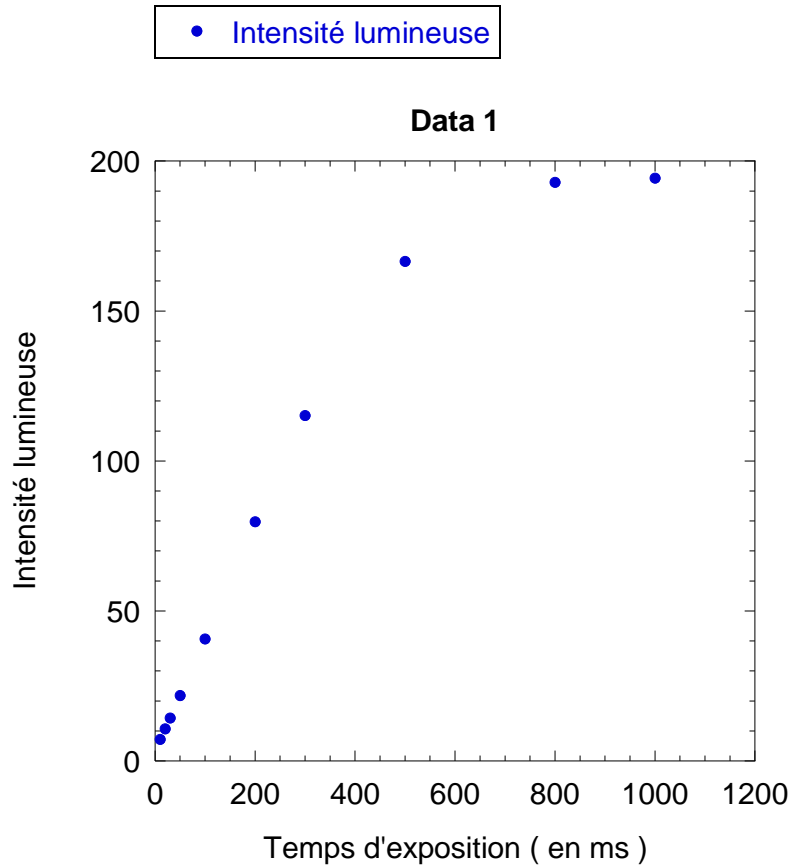
TEP 2g/L rincé 20s



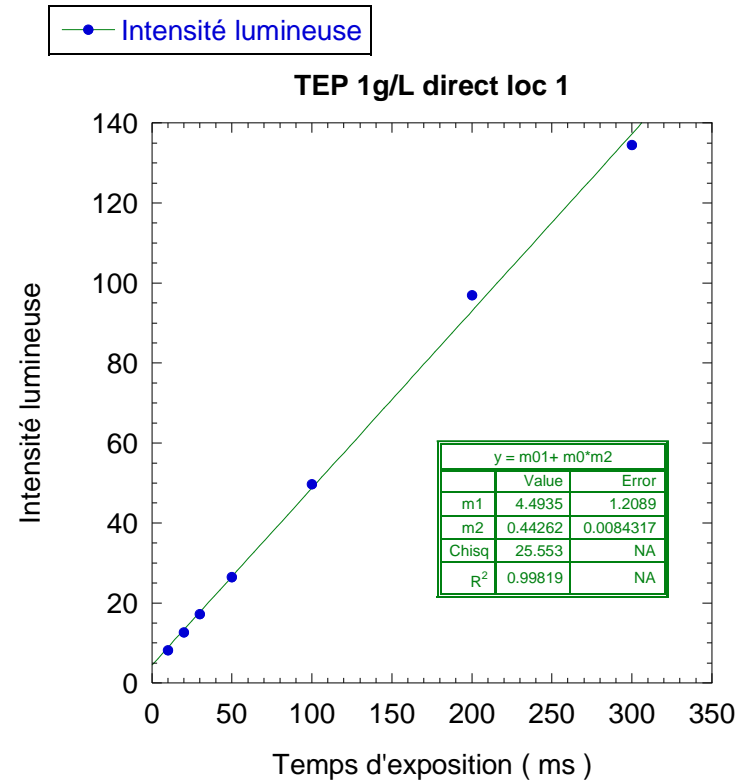
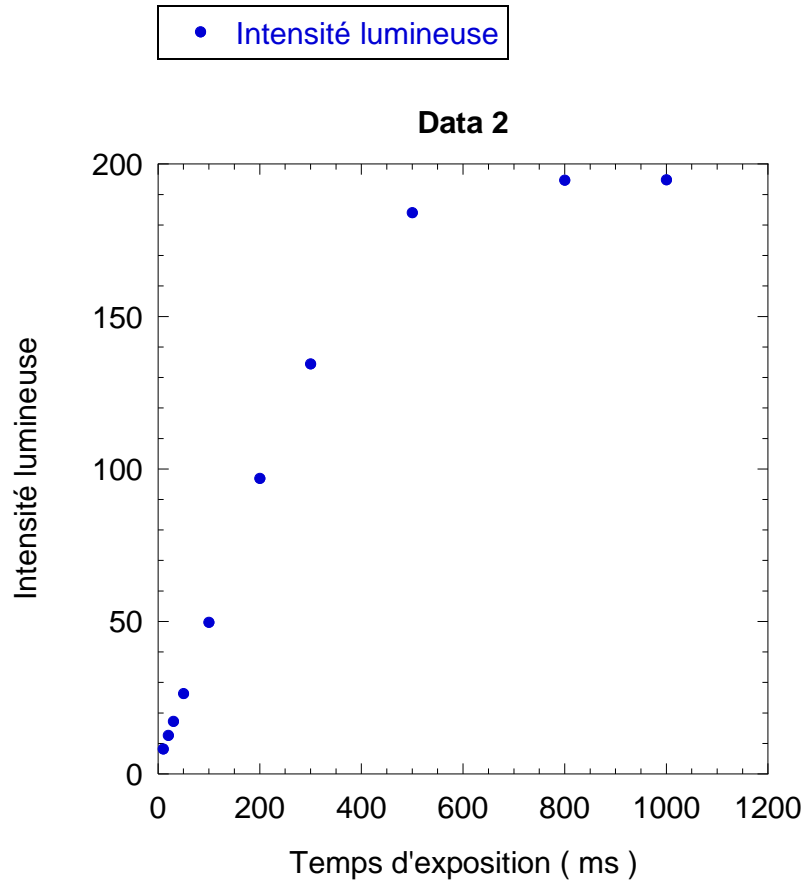
TEP 2g/L direct



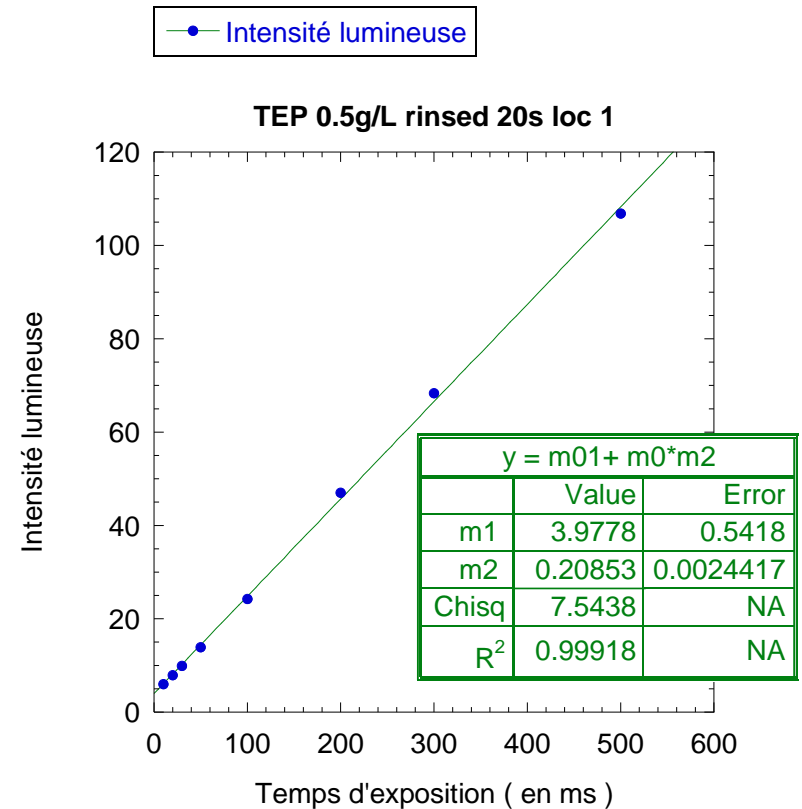
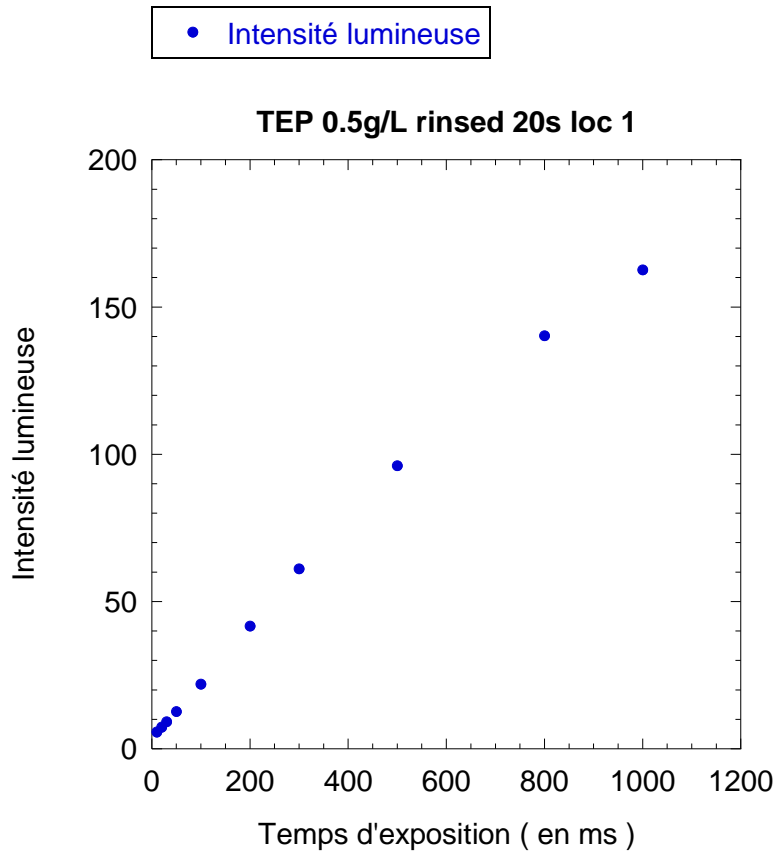
TEP 1g/L rincé 20s



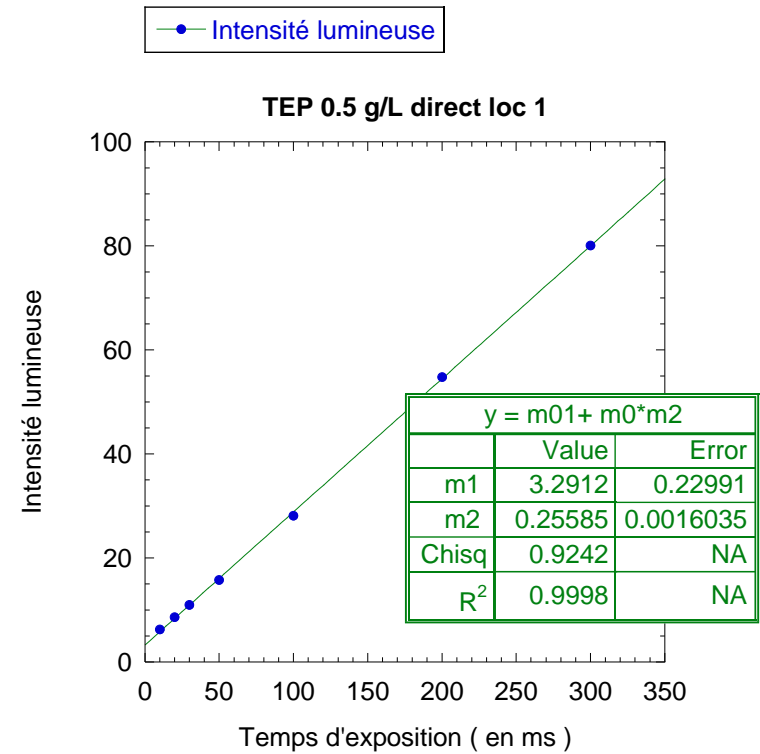
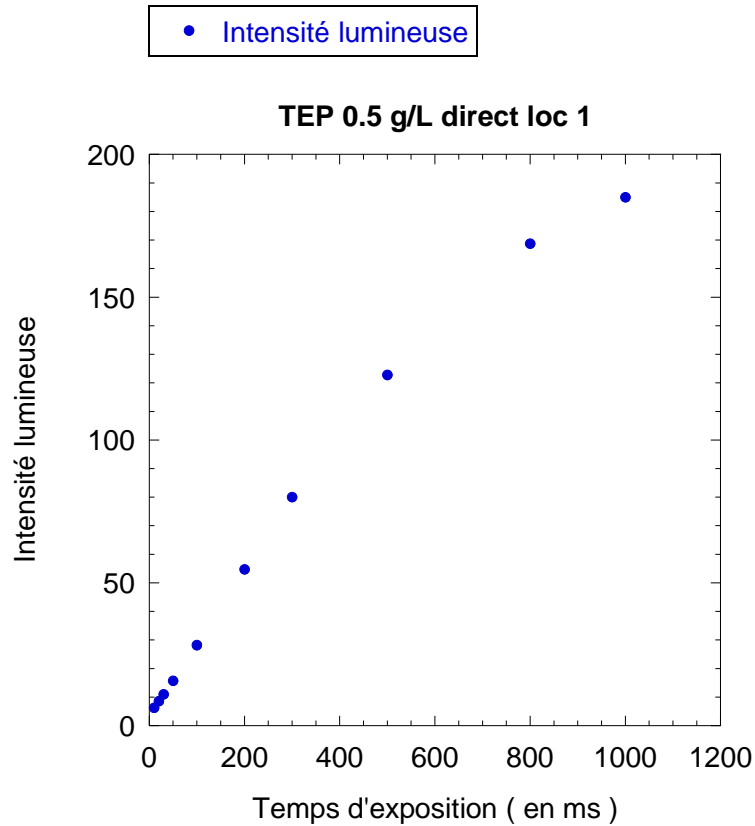
TEP 1g/L direct



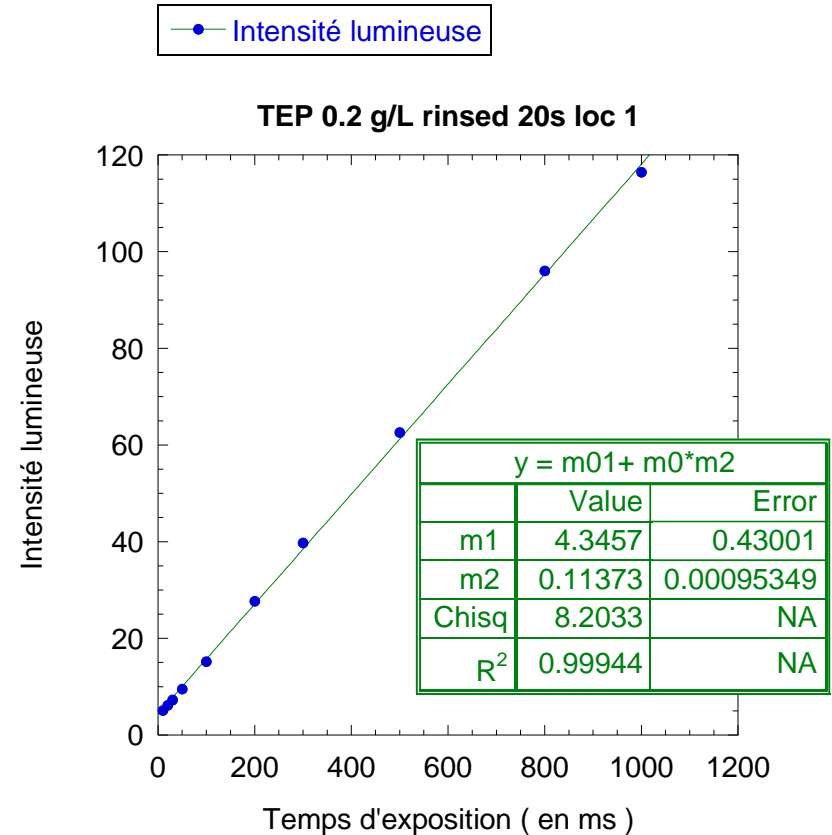
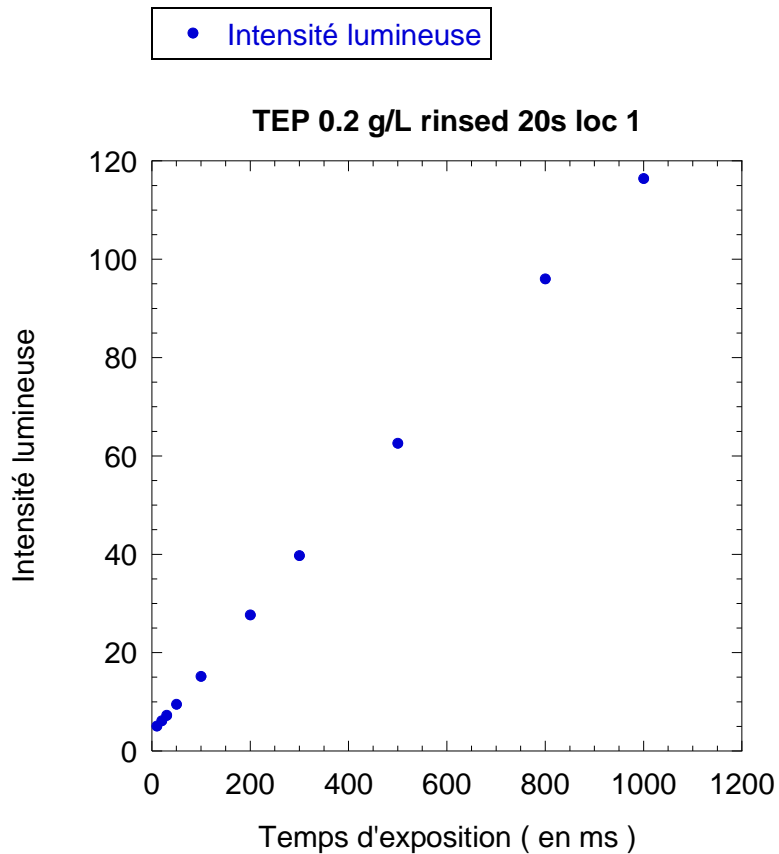
TEP 0.5 g/L rincé 20s



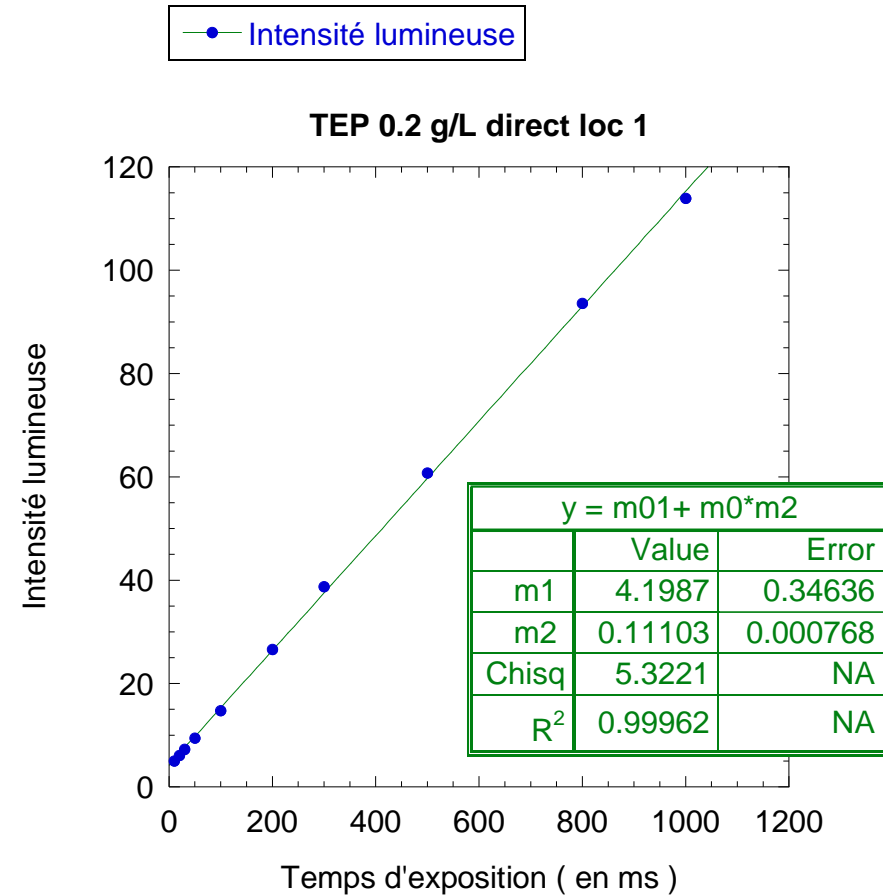
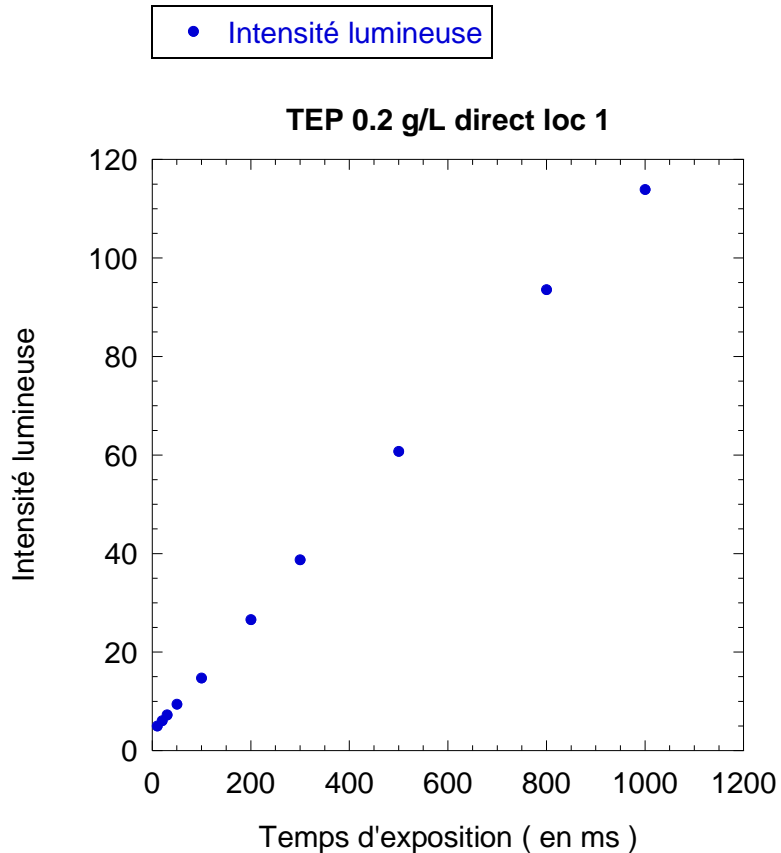
TEP 0.5 g/L direct



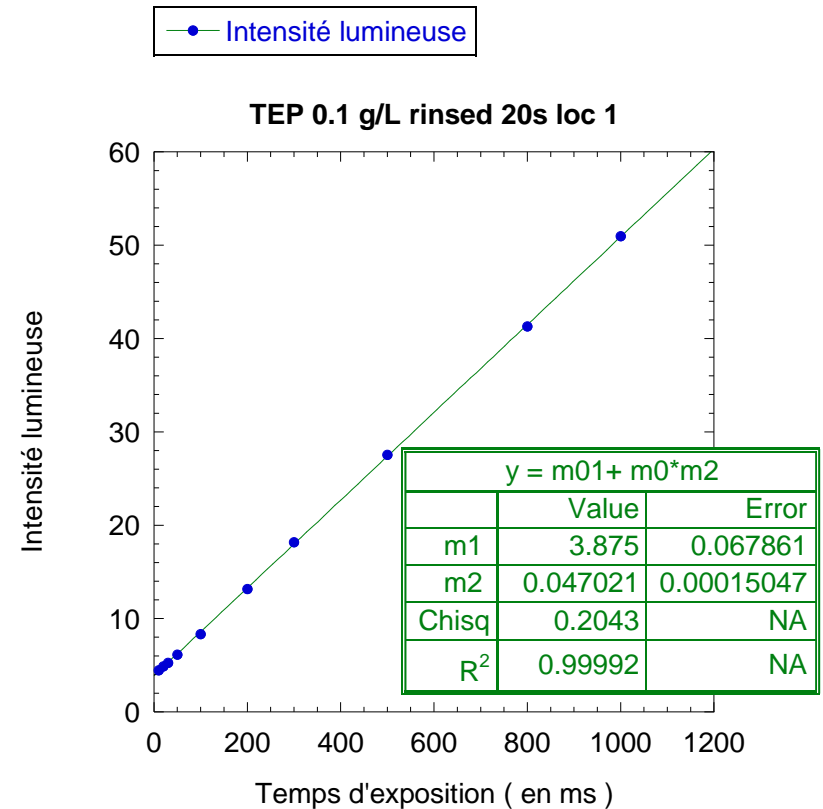
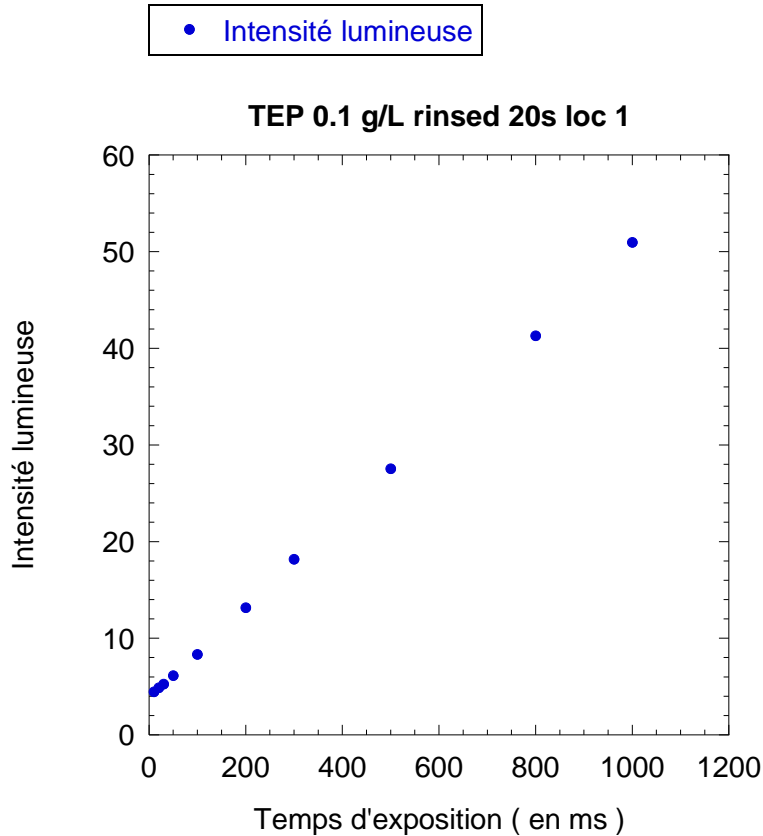
TEP 0.2 g/L rincé 20s



TEP 0.2 g/L direct



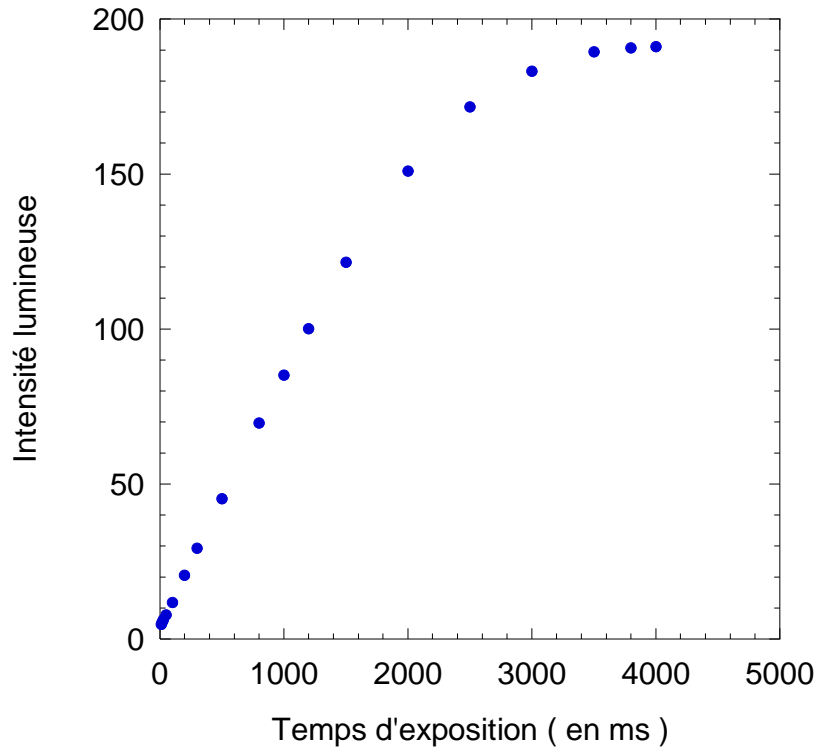
TEP 0.1 g/L rincé 20s



TEP 0.1g/L direct

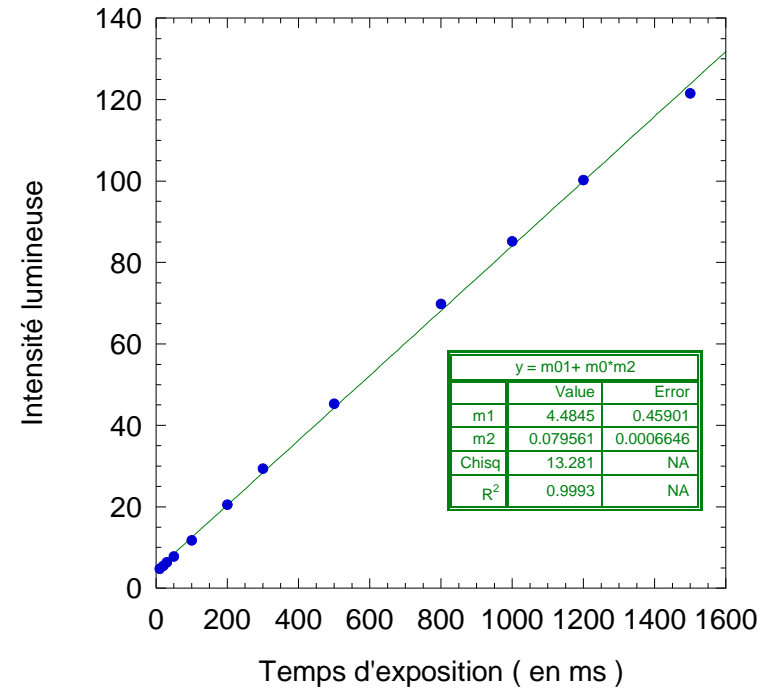
• Intensité lumineuse

TEP 0.1 g/L direct loc 1

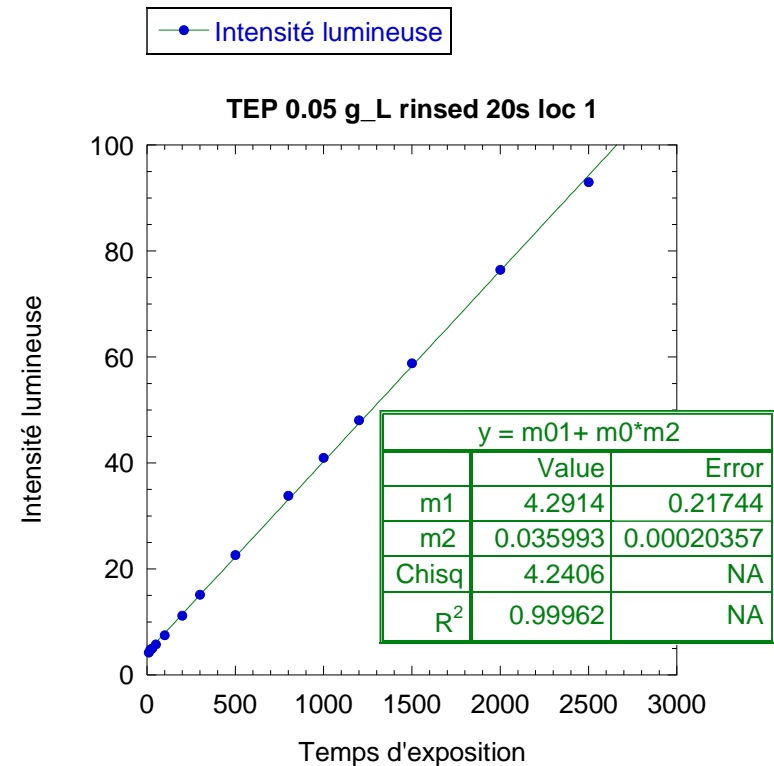
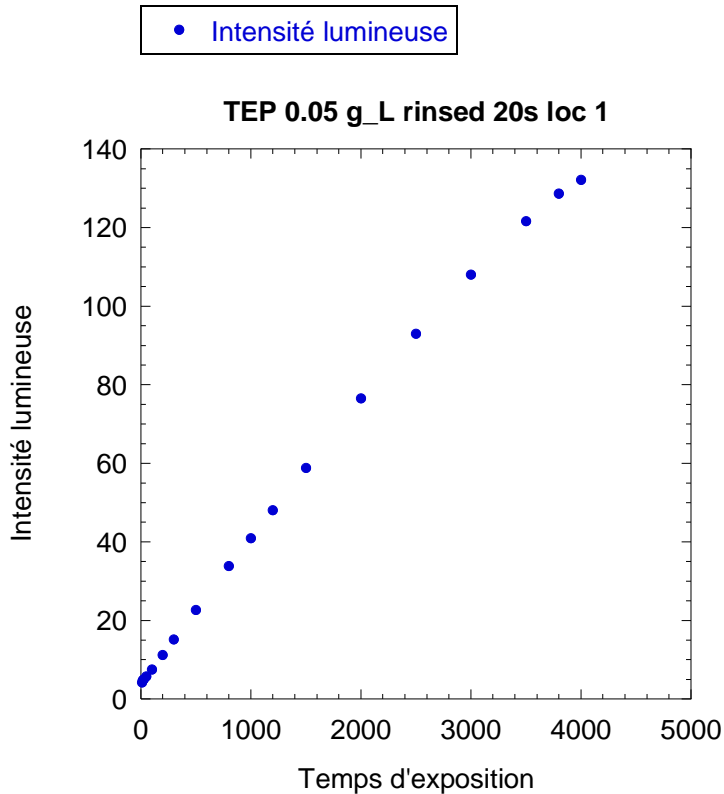


—•— Intensité lumineuse

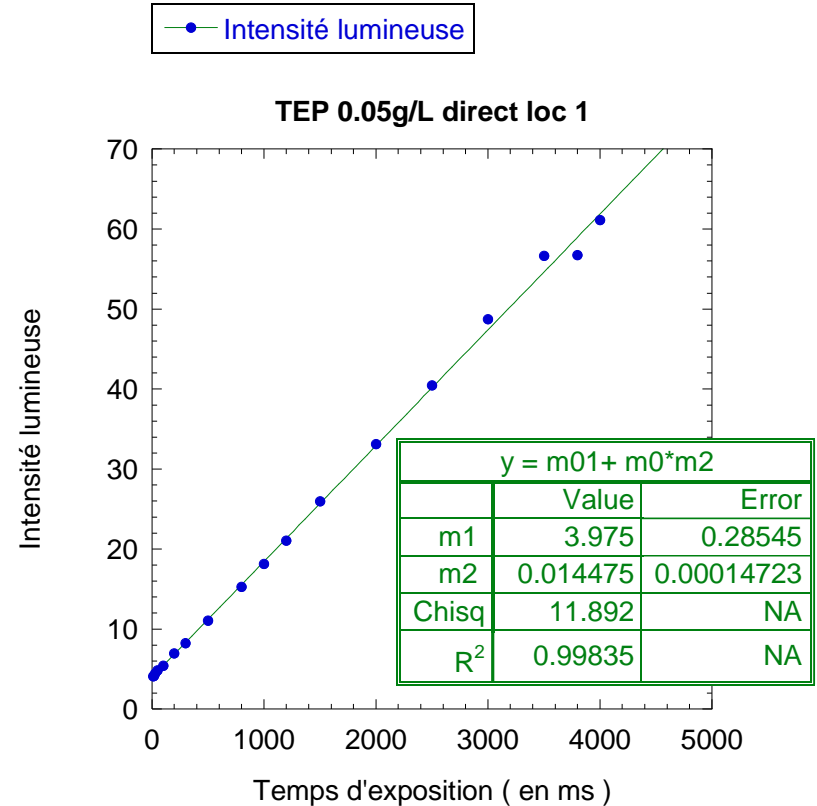
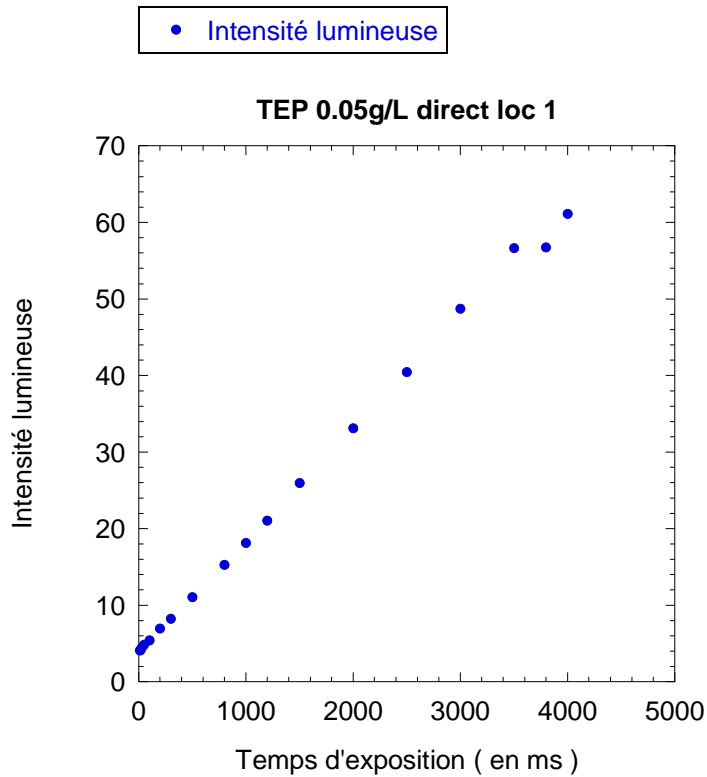
TEP 0.1 g/L direct loc 1



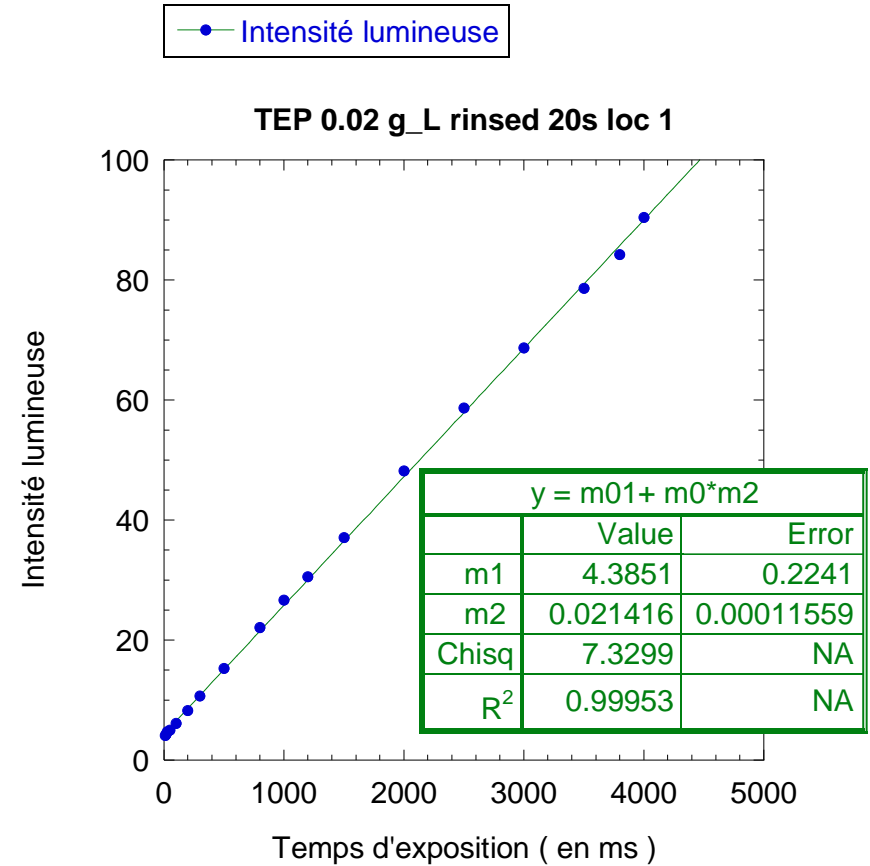
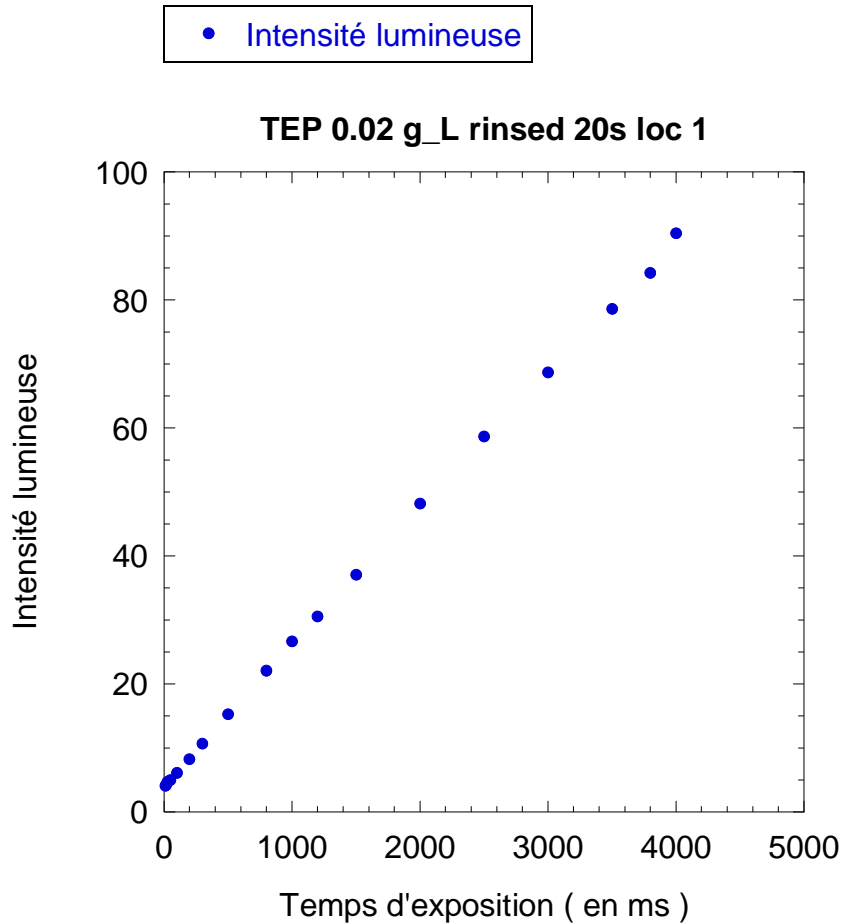
TEP 0.05 g/L rincé 20s



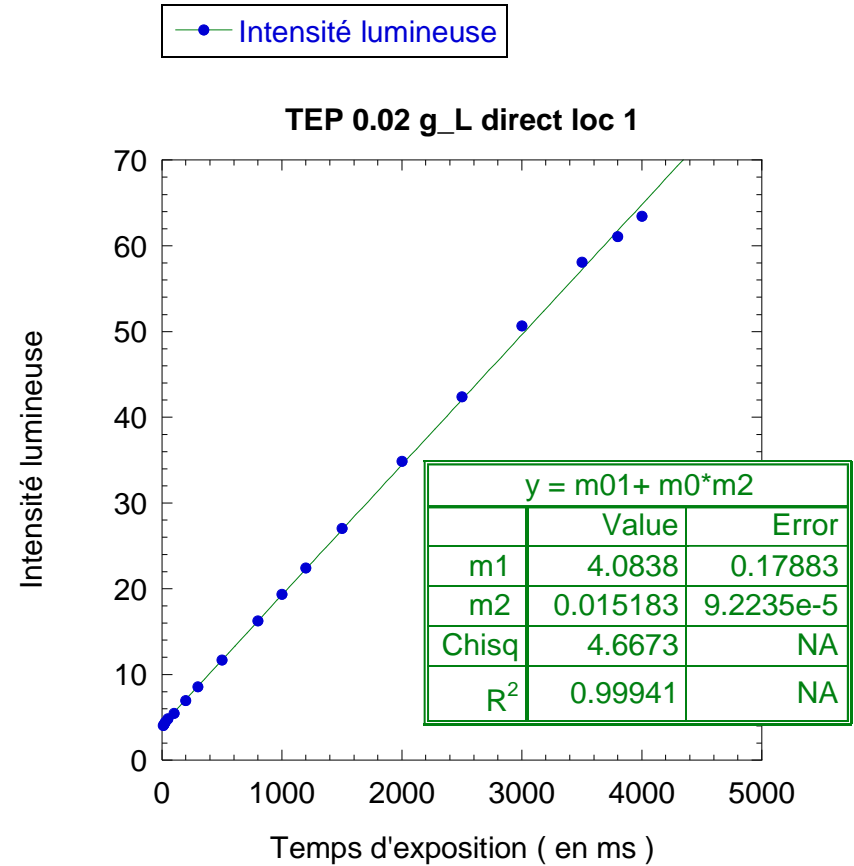
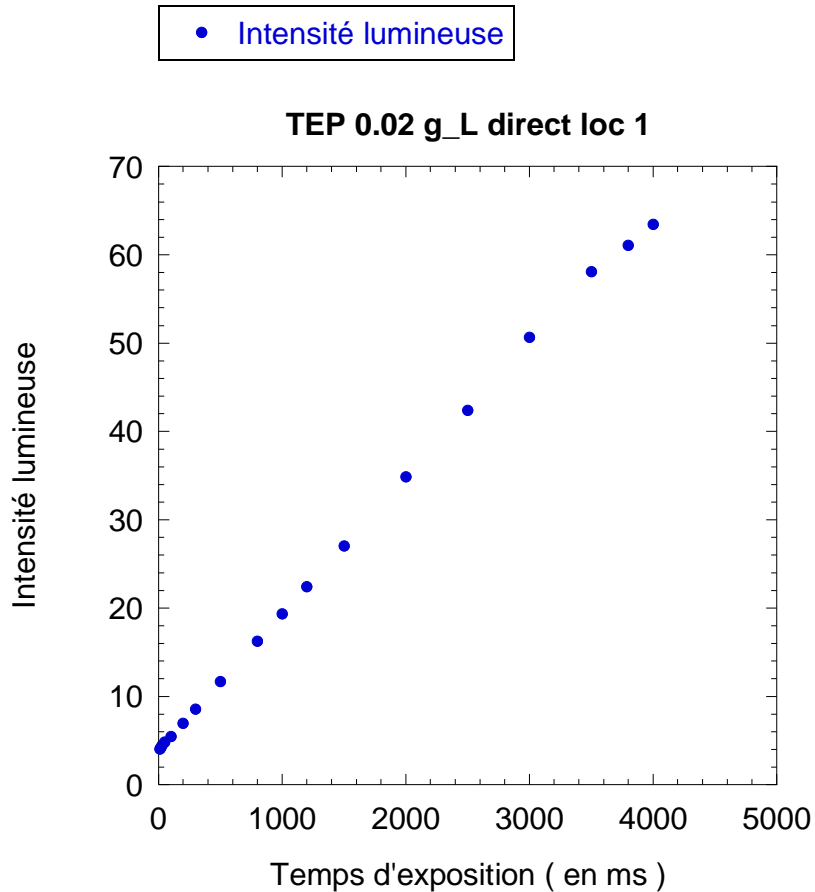
TEP 0.05 g/L direct



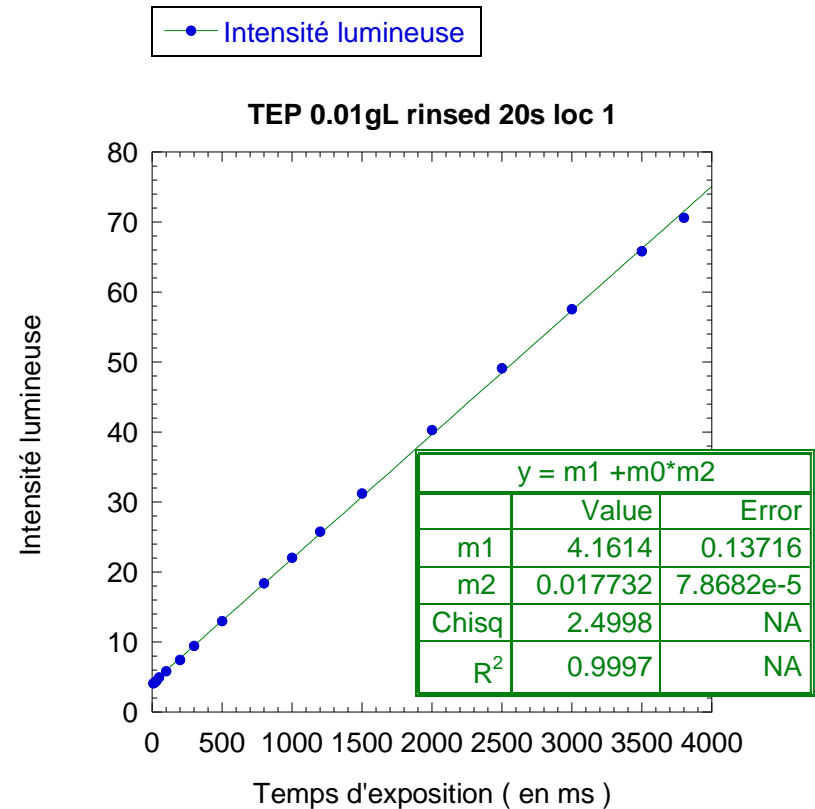
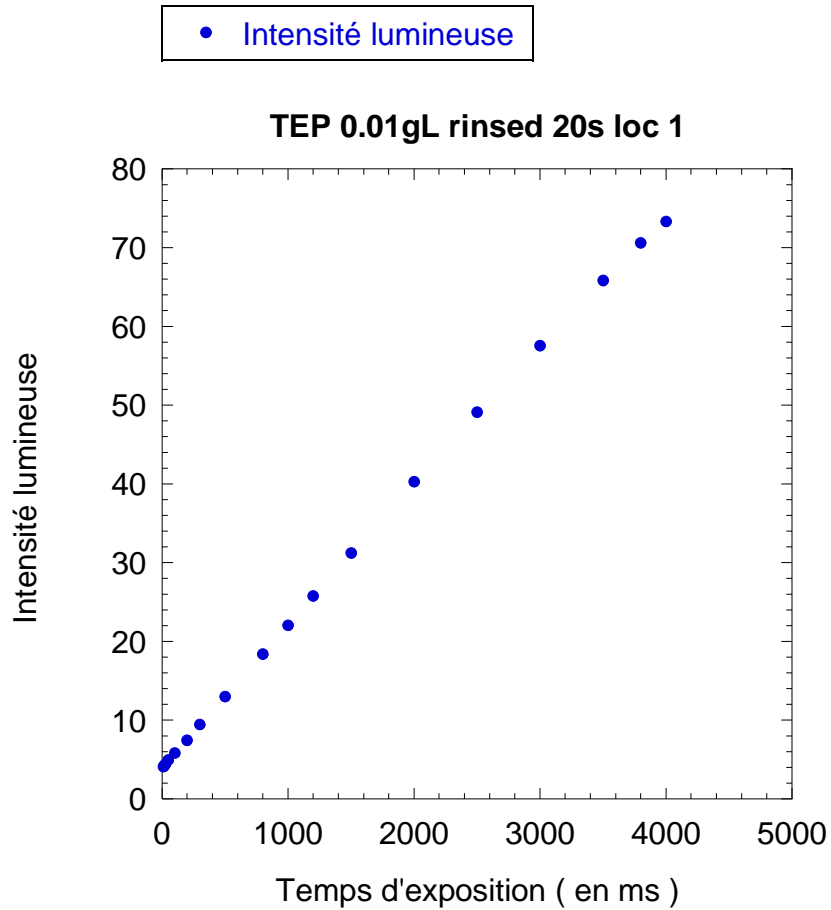
TEP 0.02 g/L rincé 20s



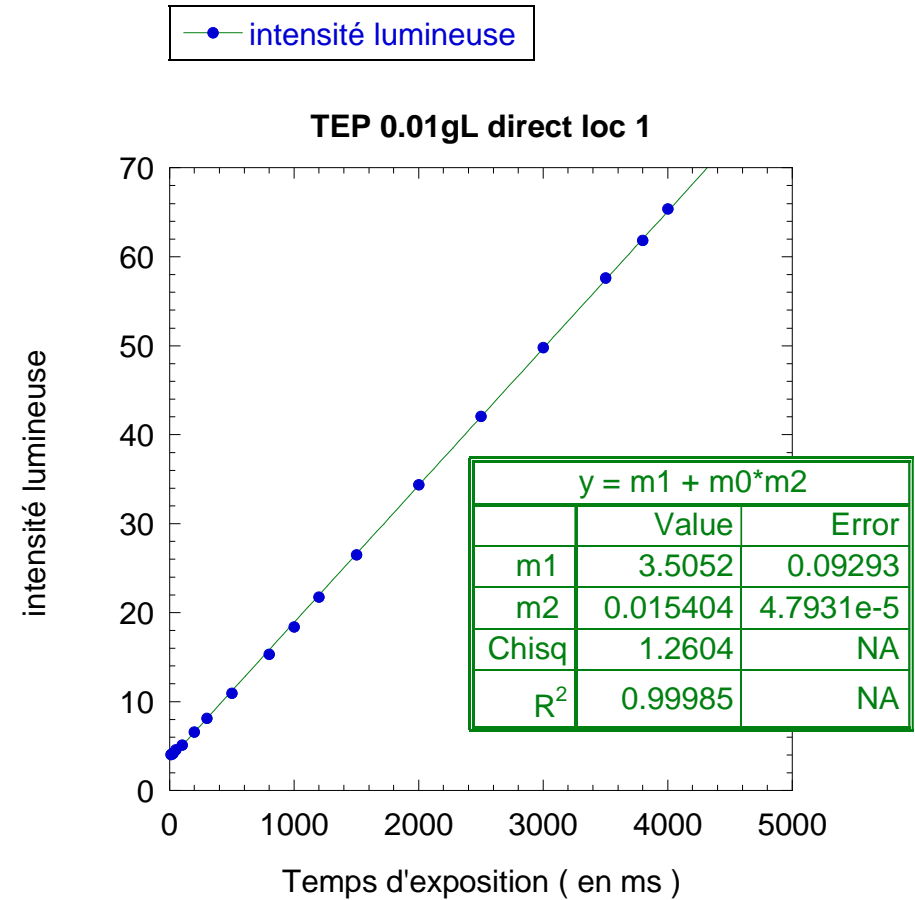
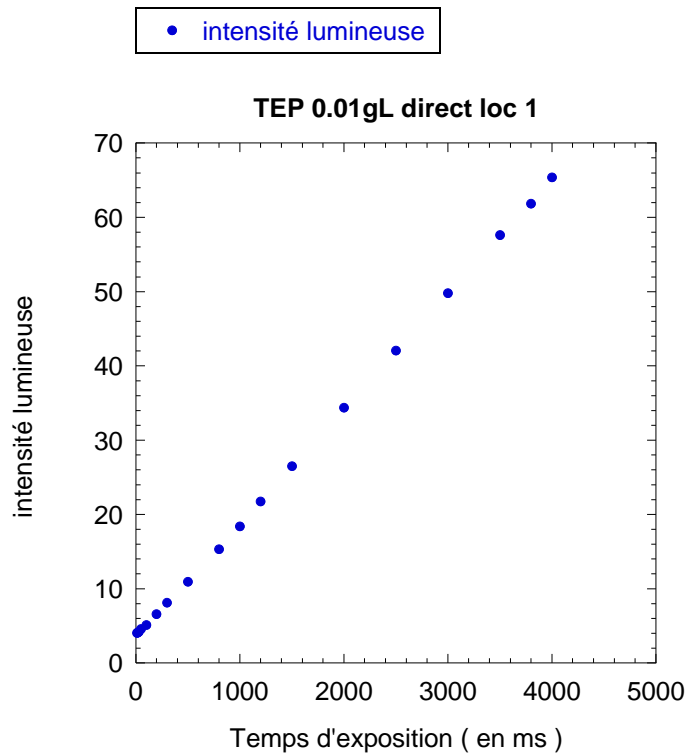
TEP 0.02 g/L direct

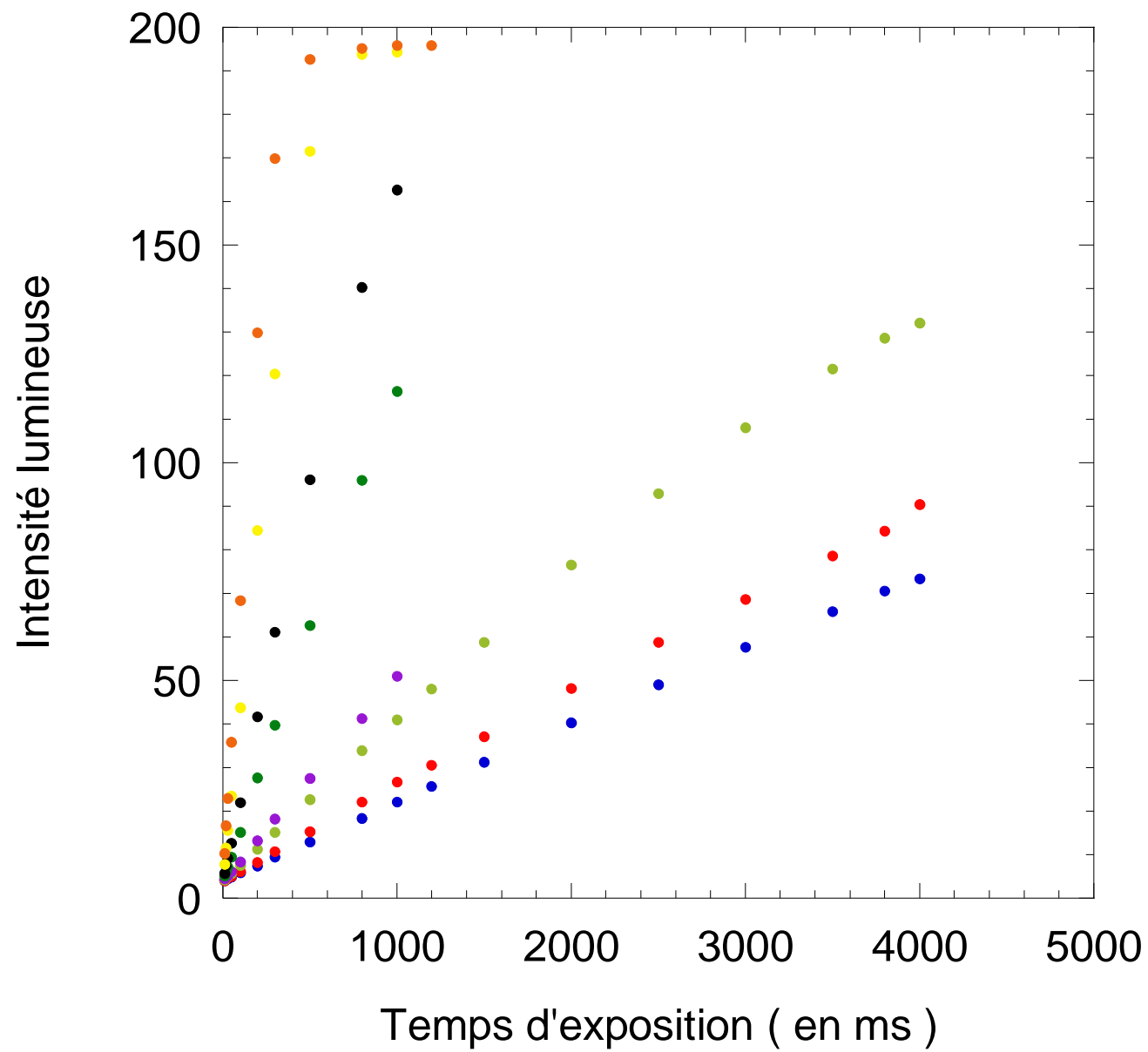


TEP 0.01g/L rincé 20s

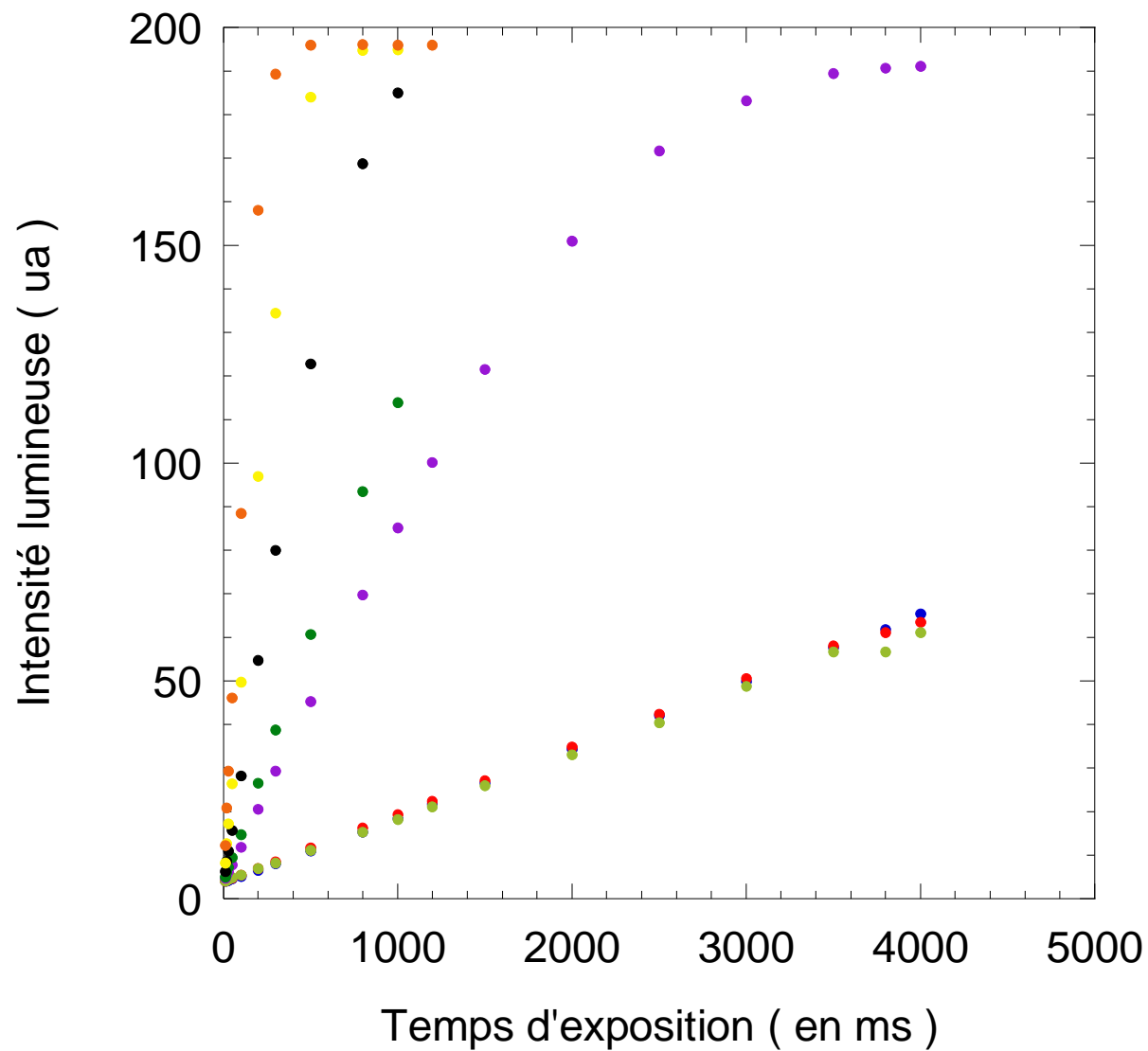


TEP 0.01g/L direct





tous les plots direct

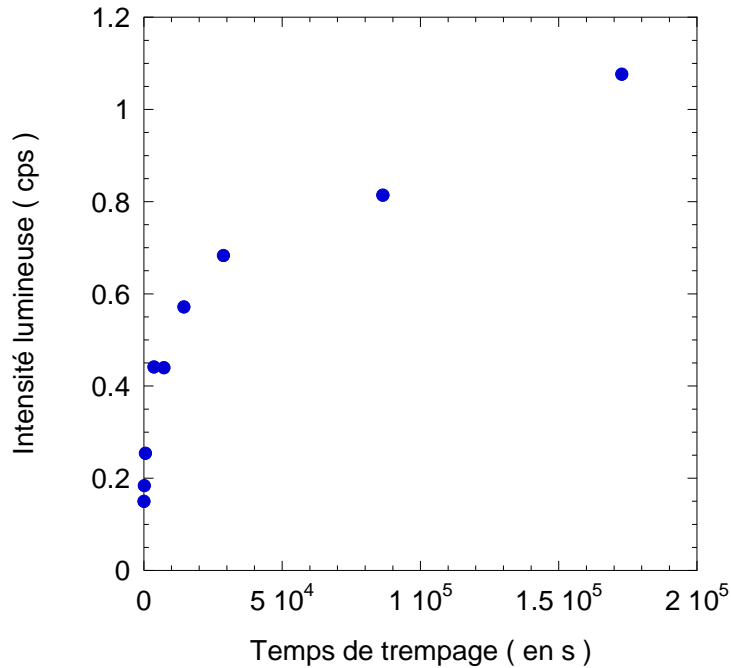


Cinétique TEP 0,05g/L et 0,5g/L

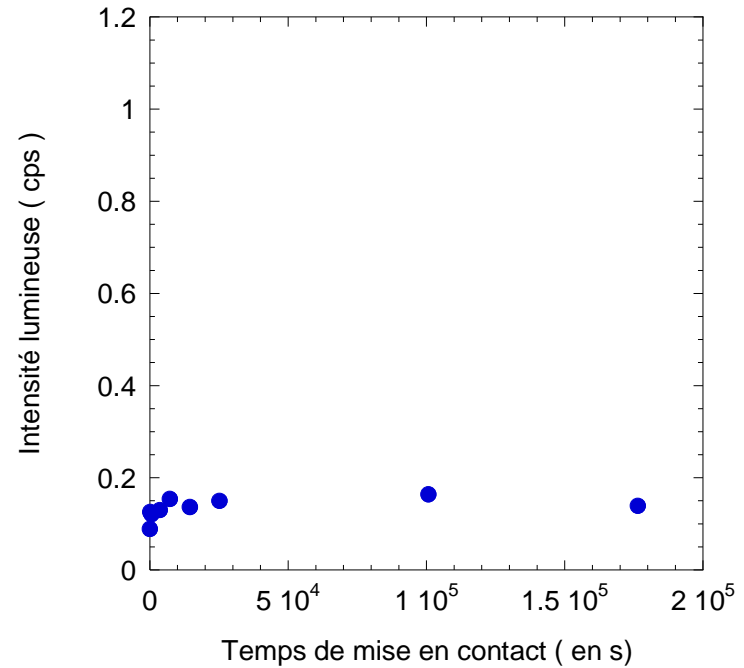
- 0,03g de coton dans 3mL TEP à 0,05g/L ou 0,5g/L pendant plusieurs temps
- Rincé 20s dans milliQ water
- Microscopie de fluorescence x10 intensité 1 gain 1 sur lamelle avec 40 μ L de milliQ water

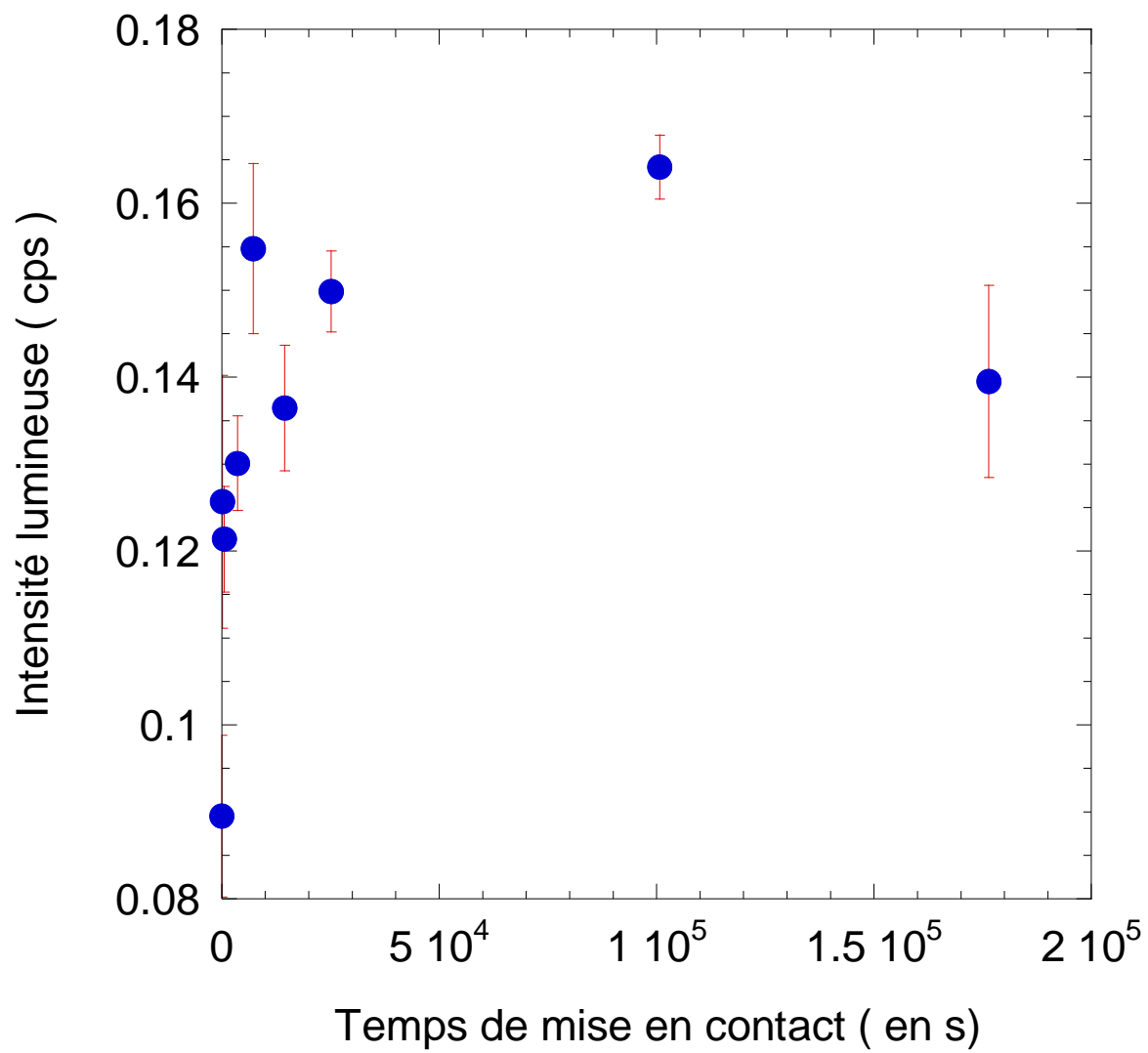
Comparaison cinétique TEP 0,05 et 0,5g/L

TEP 0,5g/L



TEP 0,05g/L



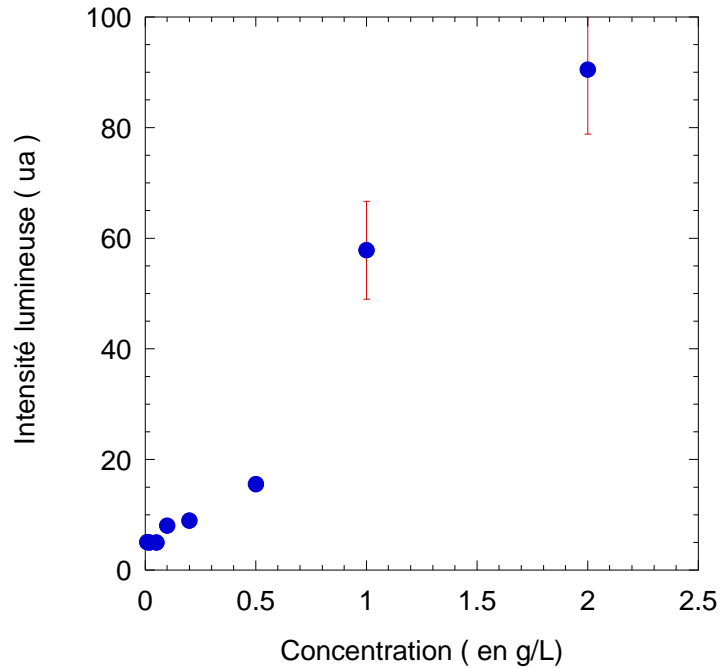


Analyse échantillons séchés

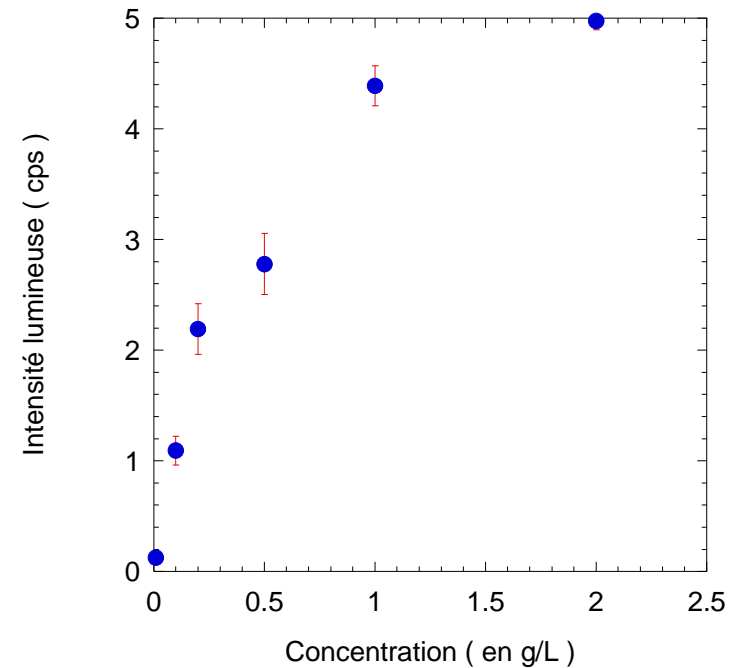
- 0,03g de coton trempé dans TEP et ES concentrés entre 0,01 et 2g/L mis à sécher.
- Rinçage 20s
- Observation sur lamelles sans eau
- Microscopie x10 intensité 1 gain 1

Comparaison ES et TEP séché

TEP rincé 20s séché

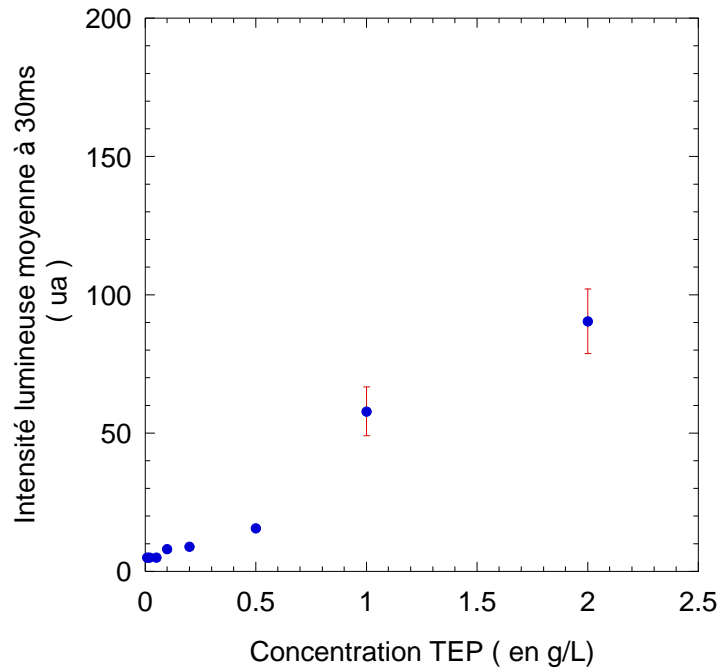


ES rincé 20s séché

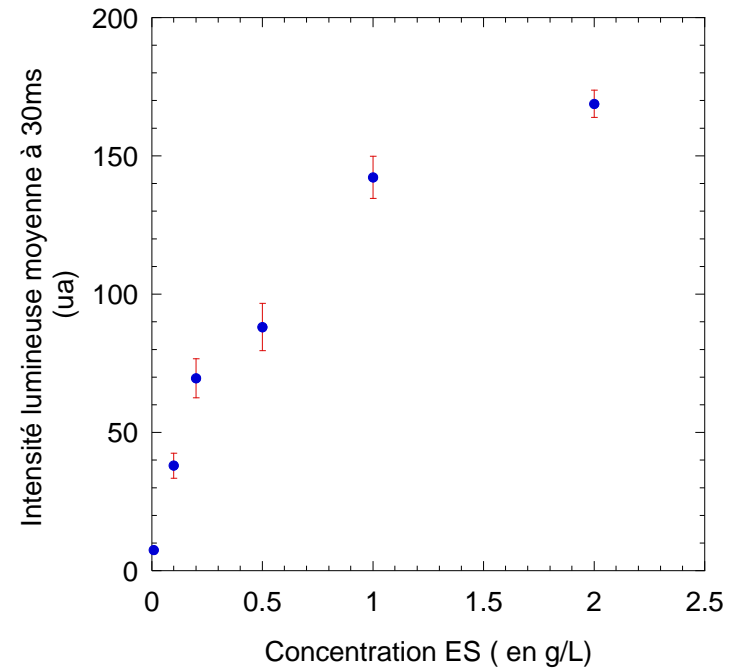


Comparaison à 30ms ES et TEP

TEP rincé 20s séché intensité à 30ms



ES rincé 20s séché intensité à 30ms



DEEDMAC 28/06/19

- Conditions expérimentales:
 - DEEDMAC concentré de 0,02 à 1g/L.
 - Echantillon 0,03g coton trempé 10min dans la dispersion
 - Soit rinçage 20s dans MilliQ Water, soit sans rinçage
 - Analyse au microscope fluo x10 Intensité 1 Gain 1
 - Fit avec isotherme de Langmuir

DEEDMAC rincé et non rincé

