**Running Head:** leaf chemistry and autotrophic microbes

**Title:** Genetic variation in tree leaf chemistry predicts the abundance and activity of autotrophic soil microorganisms

**Authors:** Paul C. Selmants\*1, Jennifer A. Schweitzer2,Karen L. Adair3, Liza M. Holeski4, Richard L. Lindroth5, Stephen C. Hart6 and Thomas G. Whitham4

1U.S. Geological Survey, Western Geographic Science Center, Menlo Park, CA USA

2Department of Ecology and Evolutionary Biology, University of Tennessee, Knoxville, TN USA

3Department of Entomology, Cornell University, Ithaca, NY USA

4Department of Biological Sciences, Northern Arizona University, Flagstaff, AZ USA

5Department of Entomology, University of Wisconsin-Madison, Madison, WI USA

6Department of Life & Environmental Sciences and Sierra Nevada Research Institute, University of California, Merced, CA USA

\*Corresponding author: pselmants@usgs.gov

**Abstract:**

Genetic variation in the chemistry of plant leaves can have ecosystem-level consequences. Here we address the hypothesis that genetic variation in foliar condensed tannins along a *Populus* hybridization gradient would influence soil ammonia oxidizers, autotrophic microorganisms that perform the first step of nitrification and are not dependent on carbon derived from plant photosynthesis. Evidence that genetically based plant traits influence the abundance and activity of autotrophic soil microbes would greatly expand the concept of extended plant phenotypes. We found that increasing foliar condensed tannin concentration reduced potential soil nitrification rates by ~ 75% and reduced the abundance of ammonia oxidizing archaea by ~ 66% but had no effect on ammonia oxidizing bacteria. Other indices that often drive nitrification rates, including soil total nitrogen, foliar nitrogen, and soil pH, were not significant predictors of either the activity or abundance of ammonia oxidizers, suggesting genetic variation in foliar condensed tannins may be the dominant regulating factor. These results demonstrate the condensed tannin phenotypes of two different tree species and their naturally occurring hybrids have extended effects on a key ecosystem process and provide evidence for indirect genetic linkages among autotrophs across at least two domains of life.

*Key words: Archaea*, *bacteria, community ecosystem phenotypes, condensed tannins, ammonia oxidizers*, *nitrification, autotrophic soil microorganisms.*

**Introduction:**

There is growing evidence that intra- and inter-specific genetic differences in plant chemistry can affect the structure and function of above- and belowground microbial communities (Silfver et al. 2007, Bailey et al. 2009, Madritch and Lindroth 2011, Lamit et al. 2015). For example, genotypic differences in foliar condensed tannins (CTs) exert a strong influence on both the composition of microbial communities (Schweitzer et al. 2008a, 2011) and key ecosystem processes they perform, including rates of leaf litter decomposition and nitrogen (N) mineralization in terrestrial (Schweitzer et al. 2004, Madritch et al. 2006) and aquatic ecosystems (Compson et al. 2018). This evidence supports the idea that genetic variation in plants can have far-reaching consequences on diverse biological communities (Whitham et al. 2012, Crutsinger 2016, Des Roches et al. 2018), yet the effects have thus far been restricted to heterotrophic organisms such as herbivores and decomposers that are directly dependent on carbon fixed by plants through photosynthesis.

Soil ammonia oxidizers are autotrophic microorganisms that perform the first and rate-limiting step in nitrification, the conversion of ammonia (NH3) to nitrate (NO3-), a process that regulates the retention of N in terrestrial ecosystems. At first glance, soil ammonia oxidizers should be insensitive to genetic variation in tree leaf chemistry because they are chemoautotrophs that fix carbon directly from CO2, and thus are not dependent on carbon derived from plant photosynthesis. However, plant chemistry may have an indirect effect on soil ammonia oxidizers. High concentrations of CTs in leaf litterfall reduce rates of soil N mineralization (Schweitzer et al. 2004), the conversion of organically bound N to inorganic NH3, which reduces substrate supply to soil ammonia oxidizers and could affect both their activity and abundance.

Several lines of evidence indicate that inputs of leaf litter with high concentrations of CTs can reduce soil nitrification rates (Hattenschwiler and Vitousek 2000, Kraus et al. 2003, Schweitzer et al. 2008b), yet the influence of foliar condensed tannins on the abundance of soil ammonia oxidizers remains unclear. This knowledge gap exists in part because, within the last decade, evidence has accumulated for a previously unknown large and nearly ubiquitous group of soil ammonia oxidizers in the domain Archaea (Francis et al. 2005, Nicol and Schleper 2006). Scientific consensus from the late 19th to the early 21st centuries held that a small group of bacteria within the phylum Proteobacteria were solely responsible for oxidizing NH3 to nitrite, the first step in nitrification (Nicol and Schleper 2006). More recent studies indicate that ammonia oxidizing archaea (AOA) in the Thaumarcheaota outnumber ammonia oxidizing bacteria (AOB) in most soils (Leininger et al. 2006, Adair and Schwartz 2008), and that AOA may dominate over AOB in controlling nitrification rates in terrestrial ecosystems with low to moderate N availability (Carey et al. 2016, Hink et al. 2018). However, the environmental factors controlling the relative abundance of AOA and AOB are only beginning to be understood, as are their relative contributions to rates of soil nitrification (Taylor et al. 2012, Lu et al. 2015, Hink et al. 2018).

Linkages between genetic variation in plants and the abundance and activity of autotrophic soil microorganisms remain largely unstudied and, if demonstrated, would extend the importance of plant gene expression as an organizing and predictive force governing the structure and function of terrestrial ecosystems. Here we examine the influence of genetic variation in tree foliar chemistry on the activity and abundance of ammonia oxidizers, a key functional group of soil autotrophic microorganisms, along a naturally occurring *Populus* hybridization gradient in northern Utah, USA. Previous work along this hybridization gradient and in nearby common gardens has demonstrated that foliar CT concentrations vary predictably with plant genotype and hybrid status, that foliar CTs reduce rates of leaf litter decomposition and soil net N mineralization, and that foliar CTs exert a strong control over the composition of heterotrophic soil microbial communities (Schweitzer et al. 2004, 2008a, 2011). Based upon this prior research, we hypothesized the abundance and activity of soil AOA and AOB would be lower in forest stands composed of *Populus* genotypes with high foliar CT concentrations. Support for this hypothesis would further extend the influence of plant genetics on ecosystem functioning and demonstrate for the first time that genetic variation in a foundation tree species can influence the structure and function of soil autotrophic microorganisms with no dependence on plant carbon inputs to soil.

**Methods:**

Hybridization gradients between *Populus fremontii* and *P. angustifolia* are common along rivers throughout the western United States, where stands at lower elevations are composed entirely of *P. fremontii* with generally low foliar CT concentrations, and higher elevation stands are dominated by *P. angustifolia* with up to four times higher concentrations of foliar CT (Rehill et al. 2006). These two species hybridize freely where their distributions overlap such that stands at middle elevations are composed of *P. fremontii, P. angustifolia*, their F1 hybrids, and complex, unidirectional backcrosses between F1 hybrids and *P. angustifolia*. Stands of trees within this hybrid zone have high genetic diversity (Whitham et al. 1999, Schweitzer et al. 2008b, 2011) and a wide range of foliar CT concentrations (Schweitzer et al. 2004, Rehill et al. 2006).

We collected foliage and soil samples from nine gallery forest stands along a naturally occurring *Populus* hybridization gradient within the Weber River drainage of northern Utah, USA (41.2˚ N, 112˚ W). The nine stands, three each in the Fremont, hybrid, and narrowleaf zones, were dominated by cottonwood trees of similar size (20 – 30 m tall) and density (~650 stems/ha; Fischer et al. 2007) and were separated from each other by at least one km, spanning a linear distance of ~100 km and an elevation gain of ~ 500 m (Schweitzer et al. 2004). We selected a central point in each stand and collected live foliage samples from the six nearest mature canopy trees to that point. Six 0-10 cm mineral soil samples were also collected from each stand within the area between the central point and the six sampled trees.

*Foliar condensed tannin and nitrogen concentrations*

Within each stand, fully expanded sun-lit leaves were collected mid-growing season from four cardinal directions on each of six trees with a pole pruner, flash frozen on dry ice, lyophilized, finely ground, and stored at -20 °C until chemical analysis. Foliar CT concentrations were determined by sequentially extracting finely ground leaf samples with 70% acetone + 10 mM ascorbic acid and then assaying the extracts with the butanol-HCl method (Porter et al. 1986) using purified CT standards from *P. angustifolia*. Foliar N concentrations were determined on subsamples of the same finely ground leaf samples by combustion with an elemental analyzer (Thermo Finnigan, San Jose, CA USA). We used CT concentrations from live leaves as a proxy for inputs of CT from leaf litterfall. Previous work along this hybridization gradient demonstrated that CT concentrations in green leaves explain nearly 90% of the variation in annual litterfall CT input to soil (Schweitzer et al. 2004).

*Potential nitrification rates and soil chemistry*

Mineral soil samples (n = 6 per stand) were subdivided upon collection with a portion of each sample flash frozen on dry ice in the field and stored at -80 °C until DNA extraction and analysis. The remaining portion of each soil sample was sieved to < 2 mm and stored at 4 °C until analyzed for nitrification potentials, pH, total organic carbon, and total N concentrations.

We conducted nitrification potentials (Hart et al. 1994) on subsets of the six mineral soil samples collected from each of the nine forest stands. We estimated maximum nitrification rates (*Vmax*) by aerobically incubating soil samples in 250-mL Erlenmeyer flasks on an orbital shaker at 180 rpm for 24 h at 23 °C with optimum water, NH4+-N, and PO43--P availability, removing 10 mL samples from each flask at 2, 4, 22, and 24 h (Hart et al. 1994). Solutions were analyzed for NO3--N with a Lachat Instruments flow-injection autoanalyzer (Loveland, CO USA).

Soil pH was determined in 1:2 (weight to volume) suspensions of air-dry soil to 0.01 M CaCl2 solution (Hendershot et al. 1993) using an Orion 720A pH meter (Allometrics Inc., Baton Rouge, LA USA). Soil organic C and soil total N concentrations of finely ground, oven-dried soil samples were determined by combustion at the Colorado Plateau Stable Isotope Facility (Northern Arizona University, Flagstaff, AZ USA).

*Archaeal and bacterial amoA gene abundance*

We estimated the abundance of AOA and AOB across the *Populus* hybridization gradient by quantifying the abundance of archaeal and bacterial versions of the *amoA* gene in soil samples collected from each of the nine stands. The *amoA* gene encodes the subunit containing the active site of ammonia monooxygenase, an enzyme essential for autotrophic ammonia oxidation. We isolated DNA from 0.5 g of frozen mineral soil using the PowerSoil DNA isolation kit (MoBio Laboratories, Carlsbad, CA USA) with an additional purification step via ethanol precipitation. All extractions were standardized to 5 ng DNA µL-1 by dilution in Tris-EDTA buffer. Gene copy number of archaeal and bacterial *amoA* per sample were determined with a DNA Engine Opticon Real-Time PCR system (Bio-Rad, Hercules, CA USA) by amplifying archaeal *amoA* with Arch-amoAF and Arch-amoAR primers

(Francis et al. 2005) and bacterial *amoA* with amoA1F and amoA2R primers (Rotthauwe et al. 1997). Samples were run in duplicate, with duplicate runs per sample averaged to yield final gene copy numbers. Standard curves of known archaeal or bacterial *amoA* concentrations (Adair and Schwartz 2008) were included in each qPCR run and purity of PCR product was verified with melting curves. Amplification efficiency for both archaeal and bacterial *amoA* primer sets was ~85%.

*Data analysis*

We used ordinary least-squares regression to determine whether stand-level potential nitrification, archaeal *amoA*, or bacterial *amoA* varied significantly as function of stand-level estimates of foliar CT concentrations. We also used ordinary least-squares regression to examine whether other soil and foliar characteristics were significant predictors of variation in potential nitrification, archaeal *amoA* or bacterial *amoA*. Our experimental unit for these analyses was the forest stand because tying the influence of a particular tree to an individual soil sample was not feasible, especially in hybrid zone stands. We used a significance level (α) of 0.10 due to small sample sizes (n = 9 forest stands) for all regression analyses.

We used one-way analysis of variance (ANOVA) to examine how soil samples from Fremont, hybrid, and narrowleaf zones differed in the ratio of archaeal to bacterial *amoA* gene abundance. Our experimental unit for this analysis was the individual soil sample, with zone (i.e., pure Fremont, hybrid, and pure narrowleaf) as the main effect and α = 0.10. All statistical tests were conducted using R version 3.2.4 (R Core Team 2018).

**Results:**

As shown in previous work along this *Populus* hybridization gradient (Schweitzer et al. 2004), foliar CT concentrations were much lower among trees in the Fremont zone (mean = 8.7 ± 0.4 mg g-1, dry mass), than among trees in either the hybrid zone (134.9 ± 22.8 mg g-1) or the narrowleaf zone (131.7 ± 15.2 mg g-1). Potential soil nitrification rates declined by ~ 75% (Fig. 1a) and the abundance of ammonia oxidizing archaea declined by ~ 66% (Fig. 1b) with increasing foliar condensed tannin concentration across the *Populus* hybridization gradient. In contrast, stand-level foliar CT concentrations were not a significant predictor of soil bacterial *amoA* gene abundance (Fig. 1c).

The ratio of archaeal-to-bacterial *amoA* in the Fremont zone (i.e., stands composed entirely of *P. fremontii*) was ~60% higher than in either the hybrid or narrowleaf zones on average, but differences across zones were not statistically significant (F = 1.24, p = 0.299, df = 2, 50) because of high within-zone variability, especially within the hybrid zone. Archaeal *amoA* was more abundant than bacterial *amoA* in ~85% of the 54 individual soil samples taken across the nine *Populus* forest stands. Bacterial *amoA* gene abundance exceeded that of archaeal *amoA* in one Fremont zone, three hybrid zone, and four narrowleaf zone soil samples (n = 18 soil samples per zone). Other stand-level environmental factors known to influence nitrification, including soil pH, foliar N concentrations, soil organic C concentrations, soil total N concentrations, and soil C:N, were relatively constant across the *Populus* hybridization gradient (Table 1) and were not significant predictors of potential nitrification rates, archaeal *amoA*, or bacterial *amoA* (p > 0.10 in all cases).

**Discussion:**

Genetic variation in plants has become increasingly recognized as an organizing and predictive force governing the structure and function of terrestrial ecosystems. We used a naturally occurring *Populus* hybridization gradient to test the hypothesis that genetic variation in a heritable plant trait, foliar CT concentrations, influences the activity and abundance of AOA and AOB, two phylogenetically distinct groups of soil autotrophic microorganisms that perform the first and rate-limiting step of nitrification, a key process regulating ecosystem N retention. The decline in potential nitrification rates with increasing foliar CT concentrations across the *Populus* hybridization gradient is consistent with our hypothesis and with previous findings from both field and laboratory studies (Hattenschwiler and Vitousek 2000, Kraus et al. 2003, Schweitzer et al. 2008b). By measuring potential rates of nitrification in tandem with quantifying AOA and AOB *amoA* gene abundance, our results suggest that long-term inputs of foliar CTs may decrease the functional capacity of ammonia oxidizing populations largely by reducing the abundance of AOA. These results further extend the influence of plant genetic variation on ecosystem processes, in this case nitrification, with the potential to feed back and affect both plant performance and ecosystem function. In addition, our results suggest for the first time that a heritable plant trait can influence the abundance of another group of autotrophic organisms, soil archaeal ammonia oxidizers, with no trophic linkage to plants as a source of carbon.

*Extended effects of foliar CTs on nitrification*

Although there is some evidence that CTs can directly inhibit nitrification (Kraus et al. 2003), we hypothesize that foliar CT inputs to soil indirectly reduce nitrification rates and AOA abundance by slowing rates of organic matter decomposition and net N mineralization. Condensed tannins reduce rates of decomposition and NH3/NH4+ release presumably by forming complexes with both proteins from decomposing leaf litter and with extracellular enzymes released into the soil solution by heterotrophic microorganisms (Hattenschwiler and Vitousek 2000), thereby reducing substrate supply to ammonia oxidizers. Previous work along the same *Populus* hybridization studied here demonstrated that increasing inputs of litterfall CTs to soil reduced annual rates of leaf litter decomposition, net N mineralization, and soil NH3/NH4+ availability (Schweitzer et al. 2004, Fischer et al. 2010). Litterfall CT inputs to soil were also positively correlated with an increase in *Populus* fine root production, presumably to compensate for the decline in soil N availability (Fischer et al. 2006). In addition, several laboratory experiments have demonstrated that CTs from other plant species added to soil reduce rates of net N mineralization (Schimel et al. 1998, Fierer et al. 2001, Kraus et al. 2003). Taken together, this evidence suggests the long-lived and cascading influence of genetic variation in foliar CTs extends to nitrification, a key ecosystem process that regulates ecosystem N retention. Because foliar CTs are heritable, they are subject to selection pressures, suggesting that different evolutionary trajectories of this plant trait within *Populus* species could alter the N status of entire ecosystems.

*Contrasting differences in AOA and AOB responses*

The discovery of ammonia oxidizers in the Thaumarchaeaota has spurred an ongoing search for key environmental and physiological factors leading to niche specialization and differentiation of soil AOA and AOB (Taylor et al. 2012, Lu et al. 2015, Carey et al. 2016, Hink et al. 2018). Our results indicate that genetic variation in the foliar chemistry of a foundation tree species may be an important factor influencing this niche differentiation. The linear decline in potential nitrification rates with increasing foliar CT concentrations across the *Populus* hybridization gradient mirrored the linear decline of AOA abundance but there was no trend in AOB abundance, suggesting AOA played a dominant role in nitrification in these forest ecosystems. Our results contrast with a recent meta-analysis showing that *amoA* gene abundance of AOB was more responsive to N additions than that of AOA, and that increased nitrification potential with N addition was only correlated with AOB (Carey et al. 2016). However, our study system differs from most of the studies included in the Carey et al. (2016) meta-analysis in two fundamental ways. First, changes in NH3/NH4+ availability across the *Populus* hybridization gradient were driven by foliar CT-induced declines in N mineralization from decaying organic matter, rather than from inorganic N additions. Several lines of evidence suggest that AOA are more responsive to N derived from organic matter mineralization than to inorganic N additions (Levičnik-Höfferle et al. 2012, Lu et al. 2015, Carey et al. 2016). Second, AOA are often the predominant contributors to nitrification in soils with neutral to slightly basic pH and low to moderate NH3/NH4+ availability (Levičnik-Höfferle et al. 2012, Lu et al. 2015, Hink et al. 2018), as is the case in our study. This evidence suggests that although AOA dominate nitrification potentials in low-N ecosystems, their activity and abundance are sensitive to heritable plant traits that reduce rates of N mineralization from organic matter.

*Implications*

Our results have several implications for research examining species interactions and the extended effects of plant genetic variation on the structure and functioning of terrestrial ecosystems. First, we have demonstrated that the biochemistry of an autotroph in one domain of life (Eukarya) can influence the abundance and activity of autotrophs in another domain of life (Archaea). This linkage of phylogenetically and functionally distinct autotrophs indicates that species interactions are not limited to direct trophic interactions or even to trophic cascades, because none of these organisms are directly or indirectly dependent on the others as a source of carbon. Second, although potential nitrification rates declined consistently with increasing foliar CT concentrations, the abundance of AOA and AOB responded very differently, suggesting a shift in competitive balance and possibly turnover of individual species of AOA and AOB in response to reductions in NH3/NH4+ supply (Fischer et al. 2010). Finally, our results have implications for plant-soil feedbacks by providing further evidence that plants may actively control terrestrial N cycling (Chapman et al. 2006). The conversion of NH3/NH4+ to NO3- via nitrification invariably leads to N losses from terrestrial ecosystems (Vitousek et al. 1982). Nitrate (NO3-) is highly mobile in the soil solution and thus easily lost from soils via leaching. Additionally, nitrification drives gaseous N losses to the atmosphere both directly and indirectly through denitrification that requires NO3- as a substrate. Our results provide further evidence that genetic predisposition toward high tannin production in *P. angustifolia*, F1, and backcross hybrids may be adaptive by serving as a N-retention mechanism in N-limited environments (Fischer et al. 2006).

**Acknowledgements:**

This research was supported by grants from NSF FIBR (DEB-0425908) and NSF Macrosystems (DEB-1340852) programs. Thanks to Clara Pregitzer and Morgan McLeod for assistance with field and lab work.

**Literature Cited:**

Adair, K. L., and E. Schwartz. 2008. Evidence that Ammonia-Oxidizing Archaea are More Abundant than Ammonia-Oxidizing Bacteria in Semiarid Soils of Northern Arizona, USA. Microbial Ecology 56:420–426.

Bailey, J. K., J. A. Schweitzer, F. Ubeda, J. Koricheva, C. J. LeRoy, M. D. Madritch, B. J. Rehill, R. K. Bangert, D. G. Fischer, G. J. Allan, and T. G. Whitham. 2009. From genes to ecosystems: a synthesis of the effects of plant genetic factors across levels of organization. Philosophical Transactions of the Royal Society B: Biological Sciences 364:1607–1616.

Carey, C. J., N. C. Dove, J. M. Beman, S. C. Hart, and E. L. Aronson. 2016. Meta-analysis reveals ammonia-oxidizing bacteria respond more strongly to nitrogen addition than ammonia-oxidizing archaea. Soil Biology and Biochemistry 99:158–166.

Chapman, S. K., J. A. Langley, S. C. Hart, and G. W. Koch. 2006. Plants actively control nitrogen cycling: uncorking the microbial bottleneck. New Phytologist 169:27–34.

Compson, Z. G., B. A. Hungate, T. G. Whitham, G. W. Koch, P. Dijkstra, A. C. Siders, T. Wojtowicz, R. Jacobs, D. N. Rakestraw, K. E. Allred, C. K. Sayer, and J. C. Marks. 2018. Linking tree genetics and stream consumers: isotopic tracers elucidate controls on carbon and nitrogen assimilation. Ecology 99:1759–1770.

Crutsinger, G. M. 2016. A community genetics perspective: opportunities for the coming decade. New Phytologist 210:65–70.

Des Roches, S., D. M. Post, N. E. Turley, J. K. Bailey, A. P. Hendry, M. T. Kinnison, J. A. Schweitzer, and E. P. Palkovacs. 2018. The ecological importance of intraspecific variation. Nature Ecology & Evolution 2:57–64.

Fierer, N., J. P. Schimel, R. G. Cates, and J. Zou. 2001. Influence of balsam poplar tannin fractions on carbon and nitrogen dynamics in Alaskan taiga floodplain soils. Soil Biology and Biochemistry 33:1827–1839.

Fischer, D. G., S. C. Hart, C. J. LeRoy, and T. G. Whitham. 2007. Variation in below-ground carbon fluxes along a Populus hybridization gradient. New Phytologist 176:415–425.

Fischer, D. G., S. C. Hart, B. J. Rehill, R. L. Lindroth, P. Keim, and T. G. Whitham. 2006. Do high-tannin leaves require more roots? Oecologia 149:668–675.

Fischer, D. G., S. C. Hart, J. A. Schweitzer, P. C. Selmants, and T. G. Whitham. 2010. Soil nitrogen availability varies with plant genetics across diverse river drainages. Plant and Soil 331:391–400.

Francis, C. A., K. J. Roberts, J. M. Beman, A. E. Santoro, and B. B. Oakley. 2005. Ubiquity and diversity of ammonia-oxidizing archaea in water columns and sediments of the ocean. Proceedings of the National Academy of Sciences of the United States of America 102:14683–14688.

Hart, S. C., J. M. Stark, E. A. Davidson, and M. K. Firestone. 1994. Nitrogen Mineralization, Immobilization, and Nitrification. Pages 985–1018 Methods of Soil Analysis: Part 2—Microbiological and Biochemical Properties. Soil Science Society of America, Madison, WI.

Hattenschwiler, S., and P. M. Vitousek. 2000. The role of polyphenols in terrestrial ecosystem nutrient cycling. Trends in Ecology and Evolution 15.

Hink, L., C. Gubry-Rangin, G. W. Nicol, and J. I. Prosser. 2018. The consequences of niche and physiological differentiation of archaeal and bacterial ammonia oxidisers for nitrous oxide emissions. The ISME Journal 12:1084–1093.

Kraus, T. E. C., R. A. Dahlgren, and R. J. Zasoski. 2003. Tannins in nutrient dynamics of forest ecosystems. Plant and Soil 256:41–66.

Lamit, L. J., P. E. Busby, M. K. Lau, Z. G. Compson, T. Wojtowicz, A. R. Keith, M. S. Zinkgraf, J. A. Schweitzer, S. M. Shuster, C. A. Gehring, T. G. Whitham, and R. Salguero‐Gómez. 2015. Tree genotype mediates covariance among communities from microbes to lichens and arthropods. Journal of Ecology 103:840–850.

Leininger, S., T. Urich, M. Schloter, L. Schwark, J. Qi, G. W. Nicol, J. I. Prosser, S. C. Schuster, and C. Schleper. 2006. Archaea predominate among ammonia-oxidizing prokaryotes in soils. Nature 442:806–809.

Levičnik-Höfferle, Š., G. W. Nicol, L. Ausec, I. Mandić-Mulec, and J. I. Prosser. 2012. Stimulation of thaumarchaeal ammonia oxidation by ammonia derived from organic nitrogen but not added inorganic nitrogen. FEMS Microbiology Ecology 80:114–123.

Lu, X., P. J. Bottomley, and D. D. Myrold. 2015. Contributions of ammonia-oxidizing archaea and bacteria to nitrification in Oregon forest soils. Soil Biology and Biochemistry 85:54–62.

Madritch, M. D., and R. L. Lindroth. 2011. Soil microbial communities adapt to genetic variation in leaf litter inputs. Oikos 120:1696–1704.

Nicol, G. W., and C. Schleper. 2006. Ammonia-oxidising Crenarchaeota: important players in the nitrogen cycle? Trends in Microbiology 14:207–212.

Porter, L. J., Liana N. Hrstich, and Bock G. Chan. 1986. The conversion of procyanidins and prodelphinidins to cyanidin and delphinidin. Phytochemistry 25:223–230.

R Core Team. 2018. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

Rehill, B. J., T. G. Whitham, G. D. Martinsen, J. A. Schweitzer, J. K. Bailey, and R. L. Lindroth. 2006. Developmental Trajectories in Cottonwood Phytochemistry. Journal of Chemical Ecology 32:2269–2285.

Rotthauwe, J.-H., K.-P. Witzel, and W. Liesack. 1997. The ammonia monooxygenase structural gene amoA as a functional marker: molecular fine-scale analysis of natural ammonia-oxidizing populations. Applied and environmental microbiology 63:4704–4712.

Schimel, J. P., R. G. Cates, and R. Ruess. 1998. The role of balsam poplar secondary chemicals in controlling soil nutrient dynamics through succession in the Alaskan taiga. Biogeochemistry 42:221–234.

Schweitzer, J. A., J. K. Bailey, D. G. Fischer, C. J. LeRoy, E. V. Lonsdorf, T. G. Whitham, and S. C. Hart. 2008a. Plant–soil–microorganism interactions: heritable relationship between plant genotype and associated soil microorganisms. Ecology 89:773–781.

Schweitzer, J. A., J. K. Bailey, B. J. Rehill, G. D. Martinsen, S. C. Hart, R. L. Lindroth, P. Keim, and T. G. Whitham. 2004. Genetically based trait in a dominant tree affects ecosystem processes: Plant genetics impact ecosystems. Ecology Letters 7:127–134.

Schweitzer, J. A., D. G. Fischer, B. J. Rehill, S. C. Wooley, S. A. Woolbright, R. L. Lindroth, T. G. Whitham, D. R. Zak, and S. C. Hart. 2011. Forest gene diversity is correlated with the composition and function of soil microbial communities. Population Ecology 53:35–46.

Schweitzer, J. A., M. D. Madritch, J. K. Bailey, C. J. LeRoy, D. G. Fischer, B. J. Rehill, R. L. Lindroth, A. E. Hagerman, S. C. Wooley, S. C. Hart, and T. G. Whitham. 2008b. From Genes to Ecosystems: The Genetic Basis of Condensed Tannins and Their Role in Nutrient Regulation in a Populus Model System. Ecosystems 11:1005–1020.

Silfver, T., J. Mikola, M. Rousi, H. Roininen, and E. Oksanen. 2007. Leaf litter decomposition differs among genotypes in a local Betula pendula population. Oecologia 152:707–714.

Taylor, A. E., L. H. Zeglin, T. A. Wanzek, D. D. Myrold, and P. J. Bottomley. 2012. Dynamics of ammonia-oxidizing archaea and bacteria populations and contributions to soil nitrification potentials. The ISME journal 6:2024–2032.

Vitousek, P. M., J. R. Gosz, C. C. Grier, J. M. Melillo, and W. A. Reiners. 1982. A Comparative Analysis of Potential Nitrification and Nitrate Mobility in Forest Ecosystems. Ecological Monographs 52:155–177.

Whitham, T. G., C. A. Gehring, L. J. Lamit, T. Wojtowicz, L. M. Evans, A. R. Keith, and D. S. Smith. 2012. Community specificity: life and afterlife effects of genes. Trends in Plant Science 17:271–281.

Whitham, T. G., G. D. Martinsen, P. Keim, K. D. Floate, H. S. Dungey, and B. M. Potts. 1999. Plant Hybrid Zones Affect Biodiversity: Tools for a Genetic-Based Understanding of Community Structure. Ecology 80:416–428.

**Table 1**. Biogeochemical characteristics of nine forest stands across a naturally occurring *Populus* hybridization gradient in northern Utah, USA. Values are means with standard error of the mean in parentheses (n = 6 per forest stand).

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Location** | **Foliar nitrogen (mg/g)** | **Soil pH** | **Soil organic carbon (g/kg)** | **Soil total nitrogen (g/kg)** | **Soil carbon to nitrogen mass ratio** |
| Fremont zone |  |  |  |  |  |
| Stand 1 | 21.0 (1.0) | 7.29 (0.05) | 40.9 (2.7) | 2.0 (0.2) | 21.13 (0.18) |
| Stand 2 | 18.4 (0.7) | 7.21 (0.05) | 28.7 (3.4) | 1.5 (0.2) | 19.61 (0.10) |
| Stand 3 | 18.1 (1.0) | 7.35 (0.04) | 39.0 (2.8) | 1.9 (0.2) | 21.07 (0.15) |
|  |  |  |  |  |  |
| Hybrid zone |  |  |  |  |  |
| Stand 4 | 18.6 (0.5) | 7.26 (0.06) | 39.5 (8.8) | 2.4 (0.6) | 18.46 (0.79) |
| Stand 5 | 17.7 (1.0) | 7.17 (0.04) | 45.2 (7.0) | 2.6 (0.4) | 17.90 (0.08) |
| Stand 6 | 16.7 (0.8) | 7.30 (0.02) | 27.0 (4.1) | 1.5 (0.2) | 17.80 (0.26) |
|  |  |  |  |  |  |
| Narrowleaf zone |  |  |  |  |  |
| Stand 7 | 16.2 (0.5) | 7.24 (0.03) | 81.1 (9.2) | 4.2 (0.5) | 19.59 (0.23) |
| Stand 8 | 18.1 (0.7) | 7.21 (0.10) | 20.4 (1.3) | 0.7 (0.1) | 31.05 (0.34) |
| Stand 9 | 19.0 (0.4) | 7.30 (0.06) | 30.7 (3.4) | 1.2 (0.1) | 26.47 (0.15) |

**Figure Caption:**

**Fig. 1.** Mean soil potential nitrification rates (a), mean soil gene abundance of archaeal *amoA* (b), and mean soil gene abundance of bacterial *amoA* (c) as a function of mean foliar condensed tannin concentrations of canopy trees in nine gallery forest stands across a naturally occurring *Populus* hybridization gradient. Horizontal and vertical error bars represent one standard error of the mean for each forest stand (*R*2 = 0.77, *p* = 0.002 for (a); *R*2 = 0.53, *p* = 0.03 for (b); p = 0.61 for (c); df = 7 for all three regressions).