



Whole Genome sequencing and Bioinformatics Training

4-7 March 2025 Lilongwe, Malawi

Day 1 – Introduction to WGS and Online Bioinformatics Basics

Focus: Setting the stage, understanding sequencing technologies, performing BLAST and exploring different online tools.

Morning Session 9:00-12:00: Welcome and Overview

• Introductions & Goals:

- o Welcome and participant introductions.
- o Outline training goals.

• WGS Overview:

- o Overview of whole genome sequencing (WGS) technologies and their laboratory roles.
- o Review basic bioinformatics concepts: file formats and sequence data types.

• Hands-On Exercise:

- BLAST Mystery Species Identification: Use BLAST to identify a mystery species.
- Emphasize effective communication between the lab and bioinformatics teams.

Lunch 12:00-13:00

Afternoon Session 13:00-17:00: Introduction to Online Tools

Demonstrations

- o Overview of browser-based platforms such as BLAST, Galaxy, Epi2me, PathogenWatch, CGE, "Assembly Server", Illumina Dragen, Terra Bio, etc.
- o Walk-through of each platform's interface and navigation tips.
- o Brief demo illustrating a command-line workflow (e.g., running an Epi2me workflow or assembly).
- o Showcase Marco's bacterial assembly pipeline.

Hands-On Exercise:

o Explore various online platforms through guided exercises.





Day 2 – Quality Control (QC) and Assembly

Focus: Understanding data quality, performing QC on raw reads, and exploring assembly theory and practice.

Morning Session 9:00-12:00: Quality Control (QC) of Raw Reads

• Presentation:

- o Concepts of data quality and its impact on downstream analysis.
- o Common issues: adapter contamination and low-quality reads.

• Demonstration:

o Review examples of good and poor run reports from Oxford Nanopore Technologies (ONT).

• Hands-On Exercises:

- o Use online QC tools (e.g., FastQC and ONT QC tool) to assess sample sequencing data.
- o Review pre-loaded sample datasets and interpret QC reports.

Lunch 12:00-13:00

Afternoon Session 13:00-17:00: Assembly Theory, Exercises, and QC

• Presentation:

- o Assembly Approaches:
 - Reference-based vs. de-novo assembly (including de Bruijn graphs).
 - Issues such as contamination and high contig counts.
- o Assembly Types:
 - Long, short, and hybrid assemblies.





- o Tools like BV-BRC for assembly.
- o Quality Assessment:
 - QC tools such as QUAST, MultiQC, CheckM, and BUCO.

• Hands-On Exercises:

- o Generate assemblies (ideally using participants' own data) via the BV-BRC Assembly Service.
- o Evaluate assembly quality using QC reports (reference QC reports from NICD as examples).
- o Run assemblies through BLAST to further assess their quality.

Day 3 - Typing and AMR Analysis Using Online Platforms

Focus: Understanding antimicrobial resistance (AMR) fundamentals and using online tools for bacterial typing and AMR gene detection.

Morning Session 9:00-12:00: Typing & AMR Fundamentals

• Presentation:

- o Compare traditional lab-based typing methods with bioinformatics approaches.
- o Typing Methods:
 - KmerFinder (including kmer theory), MLST (using tools like KMA), serotyping, PlasmidFinder, VirulenceFinder.
 - Additional tools: VirulenceFinder and PlasmidFinder.

Hands-On Exercise:

- Review Marco's slides on building a ResFinder/PlasmidFinder workflow.
- Build your own Resfinder/PlasmidFinder with MyDBFinder (upload section from a 16S reference database, download from NCBI).
- AMR package for R determining MDR

Lunch 12:00-13:00





Afternoon Session 13:00-17:00: Hands-On AMR Exercise

Review & Discussion:

o Revisit CGE results and discuss findings.

• AMR Overview Presentation:

- o Clinical significance and mechanisms of resistance (briefly).
- o Importance of surveillance and the genotype-to-phenotype correlation in AMR reporting (include Marco's story on cholera).

• Hands-On Exercises:

- o Use web-based tools for AMR gene detection (e.g., ResFinder, PathogenWatch, MEFinder, PlasmidFinder).
- o Discuss interpretation of results and the importance of consensus among tools (e.g., comparing ResFinder, AMRFinderPlus, CARD; demo Harmonize if possible).
- o Examine HTML reports generated from assemblies.

Day 4 – Bacterial Typing and Phylogenetic Analysis via Online Tools

Focus: Leveraging online platforms for bacterial typing and exploring phylogenetic relationships.

Morning Session 9:00-12:00: Bacterial Typing Overview

• Presentation - Overview of Typing Methods:

- o Introduction to methods such as MLST and cgMLST and their applications in tracking AMR epidemiologically.
- o Discussion on SNP and cgMLST analysis (kmer distance vs. SNPs vs genes/alles).

• Additional Topics:

- o Use of multiple sequence alignment (MSA) and tree-building tools.
- o Overview of tools such as CSI Phylogeny, MinTyper, and BEAST (including rooted trees, bootstrapping, and clock/time analysis).

Demonstration:

o Live demo using Enterobase.





Lunch 12:00-13:00

Afternoon Session 13:00-17:00: Hands-On Phylogenetic Analysis & Simulated Outbreak Scenario

Hands-On Exercise:

o Participants work with sample datasets to generate and visualize phylogenetic trees using online tools such as GrapeTree, Microreact, and iTOL.

• Theory & Discussion:

- o Interpreting tree topology and understanding outbreak clusters.
- o Discuss practical implications for AMR surveillance.

• Simulated Outbreak Scenario

- o Tasks:
 - Identify AMR genes using a web-based tool.
 - Perform bacterial typing and generate a phylogenetic tree.
 - Interpret the integrated results to propose potential clinical and epidemiological actions.

• Wrap-Up:

o Q&A session and final discussion.