ChIP Wash

1. Place sample plate on the magnet for ~1 min.
2. While still on the magnet, aspirate and discard supernate.
3. Wash beads with **NaCl 250 wash buffer.**
    a. Remove tubes from magnet, and add ~150 ul **NaCl 250** (4˚C) to each well.
    b. Cap the tubes and mix by inverting ten times.
    c. Centrifuge, 1 sec.
    d. Place sample plate on the magnet for ~1 min.
    e. Remove caps. *Keep them in order, and in same orientation.*
    f. While still on the magnet, aspirate and discard extract.
        i. *Take care to stay away from beads. No need to change tips. Quickly go to next step.*
4. Wash beads with **LiCl 250 wash buffer**.
    a. Repeat process described above.
5. Wash beads with 10 mM Tris-HCl.

ChIP Elution

1. Place sample against magnet and remove supernate.
2. Remove from magnet, and immediately add 40 µl **ChIP Elution Buffer**.
3. Incubate samples in thermomixer for 15 min at 65°C.
4. Spin, place tubes against magnet for 1 min, and **transfer supernatant to new tube.**
5. Add 1.5 µl of 20 mg/ml Proteinase K. Vortex to mix.
6. Incubate samples in heat block overnight at 65°C.

AMPure Purification

1. Remove aliquot of AMPure beads from refrigerator and warm them to room temperature before use. Prepare fresh 70% ethanol.
2. Add 1.8 volumes of AMPure beads (72 µl) to the sample. Mix 10 times with a pipette.
3. Incubate for 5 min at RT.
4. Place the tube in the magnetic rack for 1 min. Remove the supernatant.
5. Add 180 µl of 70% EtOH, incubate for 30 sec (do not disturb beads).
6. Remove the supernatant. Repeat EtOH was once more.
7. Place the tube in the magnetic rack. Remove the supernatant.
8. Dry the beads at room temp for a maximum 5 min.
9. Elute the DNA by adding 10 µl water. Mix 10 times with a pipette.
10. Place tube in the magnetic rack. Save the eluted sample in a 1.5 ml tube.