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Basement Membrane Increases G-Protein Levels and Follicle-Stimulating Hormone Responsiveness of Sertoli Cell Adenylyl Cyclase Activity*

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ABSTRACT. On a basement membrane substrate, Sertoli cells in culture have been shown to assume a phenotype similar to that of the in vivo differentiated cells. Sertoli cells from 10-dayold rats were cultured on plastic and on different extracellular matrix substrates [laminin, a 1- .onstituted basement membrane (Matrigel), and a synthetic laminin peptide containing the arginine-glycine-aspartic acid (RGD) tripeptide sequence] to investigate the effects of the extracellular matrix on FSH responsiveness. Both laminin and Matrigel markedly enhanced the cAMP response to FSH and cholera toxin, indicating modifications at the level of guanine nucleotide-binding regulatory (G) proteins. Furthermore, Sertoli cells grown on either of these two substrates responded to physiological levels of FSH (25-50 ng/ ml), whereas pharmacological levels of FSH (500 ng/ml) were required for cells grown on either plastic or on the RGDcontaining laminin peptide. Immunoblotting of Sertoli cell plasma membranes with antibodies directed against the a-sub-

unit of the stimulatory G-protein (Gsa) of adenylyl cyclase indicated that Sertoli cell culture on either laminin or Matrigel increased the amounts of Gsa. These results were further confirmed by immunoprecipitating the Gsa protein from the particulate fraction of [16S]methionine metabolically labeled Sertoli cells. However, Northern blot analysis using a cDNA probe for Gsa did not demonstrate changes in gene expression when Sertoli cells were grown on the various substrates. Immunofluorescent studies revealed that the Gs complex of adenylyl cyclase was preferentially located at the base of the Sertoli cells at the site of contact with the extracellular matrix. These data suggest that culture of epithelial Sertoli cells on basement membrane substrates enhances the Gs complex of adenylyl cyclase and the cAMP response to FSH, consistent with the more differentiated morphology and function of the cells. (Endocrinology 128: 1167-1176, 1991)

INCREASING evidence demonstrates that extracellular matrix can influence cell adhesion, shape, polarity, growth, migration, metabolism, and differentiation in vitro (1, 2). In the 10-day-old developing testis,
the seminiferous cords consist of early germ cells and
Sertoli cells (3), which are tall columnar cells with a
highly polarized cytoplasm (Fig. 1). Sertoli cells are in
contact at their basal surface with a basement membrane
matrix composed of laminin, type IV collagen, heparan
sulfate proteoglycan, and possibly entactin (4). When
Sertoli cells from 10-day-old rats are cultured on plastic,
they exhibit a squamous morphology and generally lack
the polarity observed in vivo. On a laminin substrate, the
Sertoli cells become somewhat more differentiated and

assume a cuboidal to low columnar shape (Fig. 1). However, when cultured on a Matrigel substrate, the Sertoli
cells adopt a phenotype very similar to that of the in vivo
10-day-old Sertoli cells. They appear tall and columnar
and are highly polarized with basally located nuclei and
abundant supranuclear cytoplasm (5) (Fig. 1). Sertoli
cells cultured on Matrigel also secrete more total protein
and more androgen-binding protein and transferrin than
cells grown on plastic (5). Little is known about the
intracellular signals involved in the Sertoli cell response
to basement membrane. In fact, the signal mechanisms
for cellular responses to extracellular matrix components
in most other cell systems have not been adequately
studied.

Sertoli cell function is under the control of the gonadotropin FSH (6, 7). FSH binding to plasma membrane receptors stimulates adenylyl cyclase and, subsequently, cAMP production, the postulated second messenger of FSH action in Sertoli cells (6, 7). Activation of adenylyl cyclase by the FSH-receptor complex is mediated by the Gs protein (8). The exchange of GDP to GTP at the

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