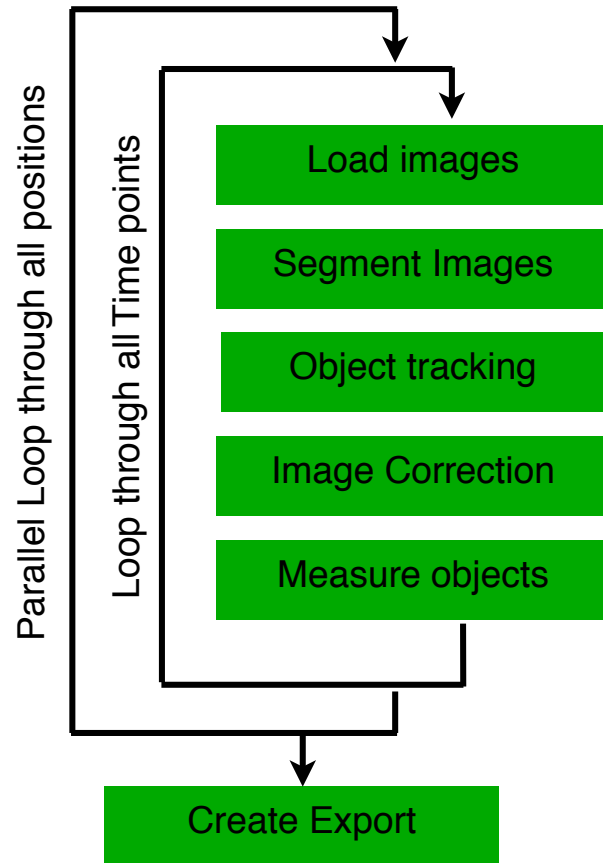


# YeastQuant 7 to 9



All positions are analyzed in parallel

For an analysis with 10 XY positions and 15 Time points  
10 parallel processes

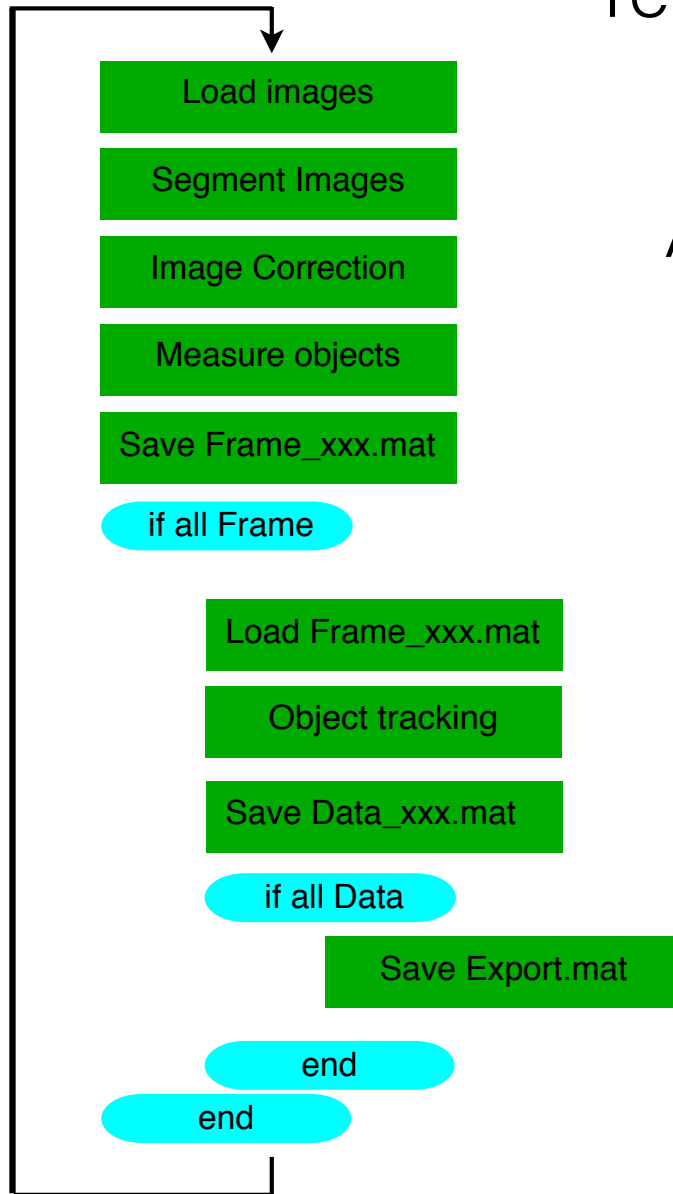
Export Files created when  
All positions are processed

# YeastQuant X

All frames are analyzed in parallel

For an analysis with 10 XY positions and 15 Time points  
150 parallel processes

Parallel loop through all frames



Export Files created  
as soon as all Data.mat from one  
experiment are present

Database entries

Same as before

BUT special analysis sheet

<b>Analysis</b>	Name	NuclBF_CFP_YFP_RFPX	# 58	<a href="#">GoTo</a>
	Segmentation	Nucl_BF		

Main difference in analysis sheet:

YQ v9

Tracked Obj	NuclInit
Linked Obj	
Tracking Distance	20

YQ X

Tracked Obj	Nucl
Linked Obj	
Tracking Distance	20

# Analysis preparation for signaling server

Same as before!

but:

Simpler call of prepare analysis

Platform

```
VarCell = PrepareAnalysis([2125:2127] 'Linux')
```

ExpNum number

removed one level of cell to structure to VarCell

YQ v9

```
VarCell{1}{1}.Analysis.xyz
```

YQ X

```
VarCell{1}.Analysis.xyz
```

# Running the analysis on Signaling server

Start screen!

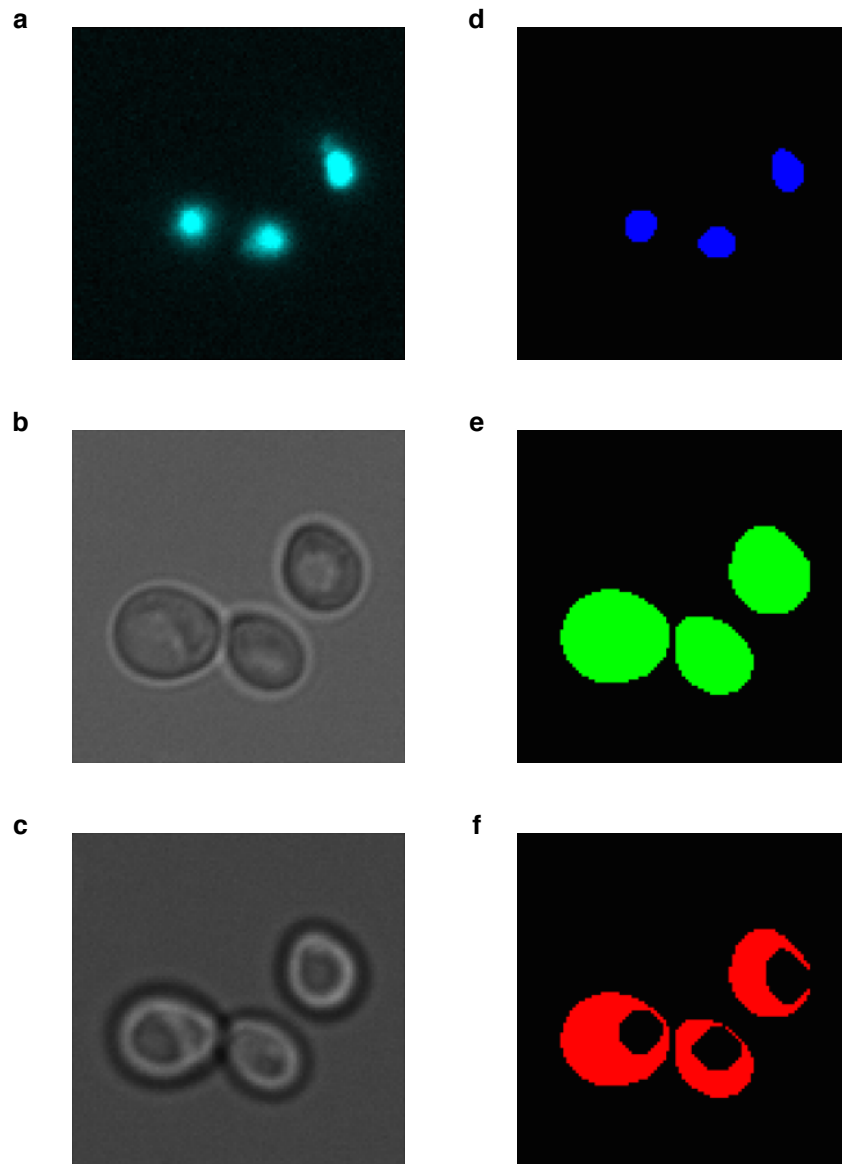
Start Matlab

load VarCell

Run ParallelFrameAnalysis

Main function: ParallelFrameAnalysis

```
function ParallelFrameAnalysis(VarCell, OverWrite)
%Overwrite = -1: Prepare only the VarFrame data
%OverWrite = 0: Perform segmentation only on missing Frame.mat files
%OverWrite = 1: Delete Data.mat file and reprocess the export file
%OverWrite = 2: Delete Frame.mat file and re-do the segmentation
```



### Supplementary Figure 2: Image segmentation process using YeastQuant

**a. - b. - c.** Microscopy images used for the segmentation: histone tag CFP (**a**), bright field image in the focal plane (**b**), and out of focus bright field image (**c**,  $z=-2.5\mu\text{m}$ ).

**d. - e. - f.** Different objects are defined by the segmentation process. First, the Nucleus is characterized using the CFP image (**d**). The two bright field images are used to find the cell contour and define the Cell object (**e**). Then, the Nucleus object, enlarged by 2 pixels, is subtracted to the Cell object, to define the Cytoplasm object (**f**).

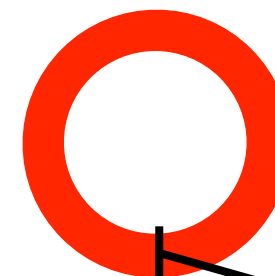
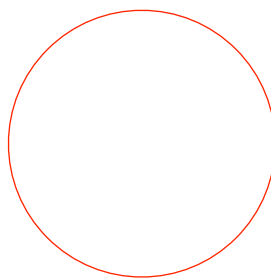
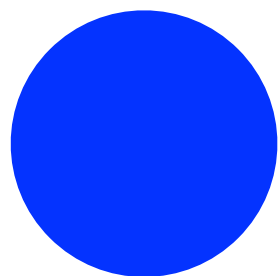
# Secondary object

Border definition:

get border pixels

Size = N  
Expand inside object by N # of pixels

CELL



BORDER

= N

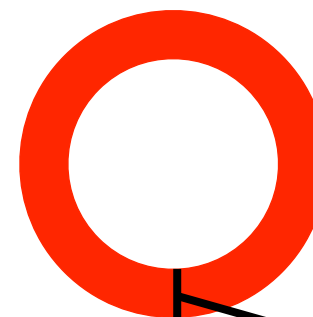
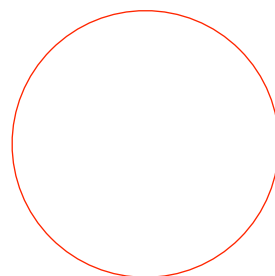
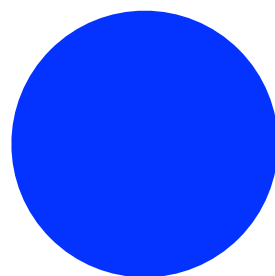
Size = N.M

Expand inside object by N # of pixels  
Expand outside object by M # of pixels

Perimeter definition

get border pixels

CELL



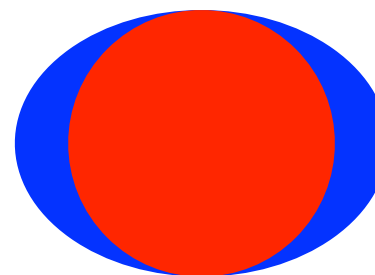
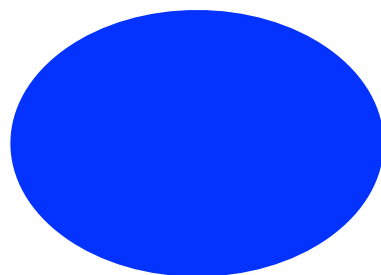
PERIMETER

= N+M

Circle definition

Match circle on cell shape  
Size argument is not used

CELL



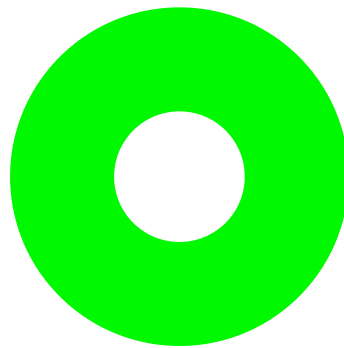
CIRCLE

Intensity image not required for those object definitions

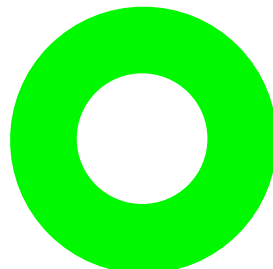
# Expand small objects



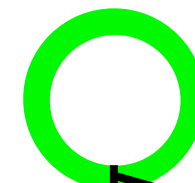
Grow **small object** by N  
pixels



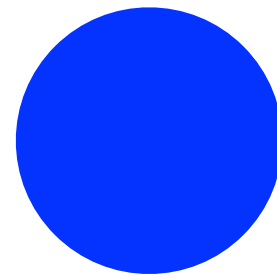
Exclude pixels outside  
**large object**



Exclude pix from small  
object with added  
border B



= N-B



**CELL**

