Transformations are essential to amplify the plasmid or express it.

Materials:

Plasmids to be transformed

Competent cells DH5 $\alpha$ (for cloning) or BL21(for expressing)

LB medium

## Procedure:

- 1. Label 1.5ml tubes with part name or well location. Fill lab ice bucket with ice, and pre-chill 1.5ml tubes in a floating foam tube rack.
- 2. Thaw competent cells on ice. Dispose of unused competent cells. Do not refreeze unused thawed cells, as it will drastically reduce transformation efficiency.
- 3. Pipette 50μl of competent cells into 1.5ml tube: 50μl in a 1.5ml tube per transformation. Keep all tubes on ice.
- 4. Pipette  $1\mu$ l of resuspended DNA or your plasmids of choice into 1.5ml tube. Gently pipette up and down a few times. Keep all tubes on ice.
- 5 Close 1.5ml tubes, incubate on ice for 30min: Tubes may be gently agitated/flicked to mix solution, but return to ice immediately.
- 6. Heat shock tubes at 42°C for 45 sec.
- 7. Incubate on ice for 5min: Return transformation tubes to ice bucket.
- 8. Pipette 950µl LB media to each transformation.

- 9. Incubate at 37°C for 1 hours, shaking at 180 rpm.
- $10\,{}^{\circ}$  Pipette  $100\mu L$  of each transformation onto petri plates Spread with sterilized spreader or glass beads immediately. This helps ensure that you will be able to pick out a single colony.
- 11. Incubate transformations overnight (14-18hr) at 37°C: Incubate the plates upside down (agar side up). If incubated for too long, colonies may overgrow and the antibiotics may start to break down; un-transformed cells will begin to grow.