Library clean-up and QC (SCI)

Note: Hold back 1uL of sample from the unpurified library to QC on the Bioanalyzer as well.

Ampure library clean-up and QC

1. Bring Ampure beads to room temperature and vortex to thoroughly resuspend the beads.

2. Hold back 1 uL of uncleaned-up library for QC.

3. Mix 20 uL library with 36 uL Ampure beads by pipetting up and down (or 1:1.8 ratio for library:beads).

4. Incubate 3-5 minutes at room temperature.

5. Place tube on magnet for 5-10 minutes to separate the beads from the solution.

    - While waiting, make up fresh 80% ethanol for wash steps 7 and 8.

6. When solution is clear, aspirate solution off beads and discard. Keep the tubes on the magnet.

7. Add 200 uL fresh 80% ethanol, incubate 30 seconds, and discard the ethanol (don't disturb pellet).

8. Repeat wash as in step 6 for a total of two washes.

9. Let pellet air dry for 5 minutes.

10. Take off magnet and resuspend beads in original volume of low TE.

11. Place on magnet and when the beads clear the magnet, elute the cleaned-up library a fresh tube.

12. Use HS Bioanalyzer kit following the manufacturer's directions to QC the 1 uL of uncleaned-up library and 1uL of cleaned-up library. This is a check for the quantity of library and its insert size, which should be recorded in the Single Cell Database under library information along with the volume remaining.

Data entry

Record QC details including quants and volume in Colossus in "Add Libraries" under "Library Quantification and Storage". Save the bioanalyzer file with a screencap of the electropherogram in region table (to show average insert size) to F:\SCI\04 Agilent Bioanalyzer and to Colossus.