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Candice A. Craig

Education

Graduate Education

University of Maryland, Baltimore County (UMBC)

Graduated: May 2014

Baltimore, MD GPA: 4.0/4.0

Degree: Master of Science in Applied Molecular Biology

Undergraduate Education

University of Maryland, Baltimore County (UMBC) Graduated: May 2013

Baltimore, MD GPA: 3.927/4.0

Degrees: Bachelor of Science in Biological Sciences and Bachelor of Arts in Modern Language, Linguistics and Intercultural Communication with a concentration in the Spanish language

Academic Honors: President's List, Dean's List, winner of the Barbara Simon Memorial Essay Contest, Member of the National Society of Collegiate Scholars, the Honor Society Sigma Alpha Lambda, Golden Key International, Omicron Delta Kappa, and Phi Beta Kappa.

Skills:

Molecular Biology

• DNA (plasmid and genomic) isolation and purification, Restriction Enzyme digestion, Gibson assembly, PCR (Long Range PCR, RT-PCR, colony PCR), Primer Design, Gel Electrophoresis, Gel Extraction and DNA Purification, bacterial transformation (electroporation and chemical transformation), subcloning, Sanger sequencing, transposon mutagenesis, bacterial competent cell preparation, and media and buffer preparation

Protein Biochemistry

 Protein induction and extraction, affinity chromatography, cell fractionation, renaturation of insoluble proteins, dialysis/buffer exchange, SDS-PAGE electrophoresis, and Western blot analysis

Animal work

• Proper rat and mouse care /handling, tail cutting/DNA extraction from murine tail tissue, designing and testing of behavioral assays in mice.

Languages: advanced reading, writing, and speaking skills in Spanish

Computer

- Word processing programs: Microsoft Office suite (Word, Powerpoint, Excel, Publisher, etc.)
- Data analysis software: NCBI tools, Vector NTI, Snapgene, and ImageJ

Communication: Experience in scientific writing as well as experience in verbal presentation of data

Biomedical Research Experience:

U.S. Food and Drug Administration CBER- Laboratory of Mucosal Pathogens and Cellular Immunology Silver Spring, Maryland 07/2014-Present

ORISE Post-baccalaureate Fellow

- Investigated a novel downstream repressive element (DRE) of the virulence factor fimbriae serotype 3 (fim3) of Bordetella pertussis via transposon mutagenesis of a fim3-luciferase fusion within B. pertussis. Such a fusion allowed for identification of factors which directly regulated the DRE of the fim3 promoter by observing for an increase in luciferase activity due to transposon knockout of DRE specific regulatory factors.
- Performed mutagenesis on a strain of *B. pertussis* which lacked a response regulator of the BvgAS two component system which is the essential for gene expression under modulating conditions. This mutagenesis was performed to identify mutants which are able to survive in modulating conditions in the absence of the primary response regulator in order to indicate genes which are involved with cell growth and survival of *B. pertussis* under modulating conditions.

University of Maryland, Baltimore County Applied Molecular Biology Laboratory, in affiliation with AthenaES Baltimore, Maryland 08/2013-05/2014

Graduate Researcher

Thesis: Development of a high-throughput glucose binding assay derived from the colorimetric detection of glucose binding proteins

Generated various fusions of the protein mCherry to a glucan-binding domain (GBD) in order to
investigate the use of such a protein in a colorimetric binding assay to observe for changes in
GBD activity under varying conditions. In addition, the potential of mCherry to be secreted to the
periplasm by various secretion pathways was examined with the ultimate goal of targeting the
mCherry-GBD fusion protein to the periplasm in order to increase active fusion protein
production.

University of Maryland, Baltimore County Developmental and Cancer Laboratory Baltimore, Maryland 09/2012-07/2013

Undergraduate Researcher

Thesis: Induction of inflammation leads to pelvic pain in a new mouse model of nonbacterial Chronic Pelvic Pain Syndrome/Chronic Prostatitis

• Optimized a behavioral assay for a mouse model of inducible prostatitis, where mice induced for prostatitis were tested for incontinence by observing for changes in urination patterns and for tactile allodynia through pressure tests using von frey filaments.

National Institute of Health

Bethesda, Maryland

DNA Diagnostic Laboratory, National Eye Institute

Summer Intern

07/2012-08/2012

- Verified the existence of mutations in patients diagnosed with Retinal Dystrophy, originally detected by next generation sequencing, by Sanger sequencing.
- Developed a screening method for mutations correlated to Retinal Dystrophy.

06/2011-08/2011

• Optimized Long Range PCR and Sanger sequencing conditions for detecting and defining the breakpoints of unique whole exon deletions found within the *RS1* gene in patients with X-Linked Juvenile Retinoschisis.

05/2010-08/2010

• Analyzed copy number defects of various exons within the *BEST1* gene in a monkey model for Viteliform Macular Dystrophy using RT-PCR and Sanger sequencing.

Publications:

D'Souza L, Cukras C, Antolik C, **Craig C**, Lee JY, He H, Li S, Smaoui N, Hejtmancik JF, Sieving PA, Wang X. Characterization of novel *RS1* exonic deletions in Juvenile X-Linked Retinoschisis. Mol Vis. 2013 Nov 7; 19:2209-16.

Academic Presentations:

Poster Presentation: Craig C., Isleen W., Julie W., and S. Broedel. "Assessing Solubility of Fusion Protein Dex-GBD, a Potential Dental Care Additive". Applied Molecular Biology Program at UMBC, Graduate Association of Biological Sciences Symposium, March 14, 2014.

Poster Presentation: **Craig C.**, Angela T., Matthew B., Harsha R., Kory J., Young F., Wadih Z., Anand S., and Wang X: "Verification of Mutations Found by High-Throughput Screening in Patients with Inherited Retinal Dystrophies". NIH Summer Research Program, Summer Intern Poster day, August 9, 2012.

Poster Presentation: **Craig C**, Hong He, Antolik C, Wang X: "Defining the Breakpoints of Whole Exon Deletions in the *RS1* Gene in Patients with X-Linked Juvenille Retinoschisis". NIH Summer Research Program, Summer Intern Poster day, August 4, 2011.

Poster Presentation: **Craig C**, Antolik C, Wang X: "Candidate Gene Analysis of a Potential Monkey Model of Viteliform Macular Dystrophy". NIH Summer Research Program, Summer Intern Poster day, August 5, 2010.

Volunteer Work:

 Assistant Coach of Spencerville Adventist Academy's Acro-Squad, a high school gymnastics team
 2010-2014

Leadership Roles:

•	Welcome Week and Orientation Leader at UMBC	2012-2013
•	Secretary of Alpha Sigma Kappa, Women in Technical Studies	2012-2013
•	Big/Little Chair of Alpha Sigma Kappa, Women in Technical Studies	Spring 2012

Membership:

•	Member-Pre-Medical Society of UMBC	2008-2009	
•	Member-Acro-Airs, collegiate level acrobatic team associated with	team associated with Washington	
	Adventist University (formerly Columbia Union College)	2008-2010	
•	Member-Alpha Sigma Kappa, Women in Technical Studies	2011-2013	

References

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