

# Decimation of EM Volume Mesh Objects with Pyvista

Examples from MICrONS and H01 Volumes



# Volume Examples

pinky

- MICrONS Layer 2/3 mouse visual cortex

minnie

- MICrONS 1mm<sup>3</sup> mouse visual cortex

H01

- Harvard/Google human temporal cortex

# Decimation Process

## Download

- Download mesh files to local disk (*or Google Drive for Google Colab*)

## Decimate

- Process and decimate mesh files (*creates a new ply file 95% smaller*)

## Visualize

- Visualize decimated mesh objects (*all examples shown here use vtk*)

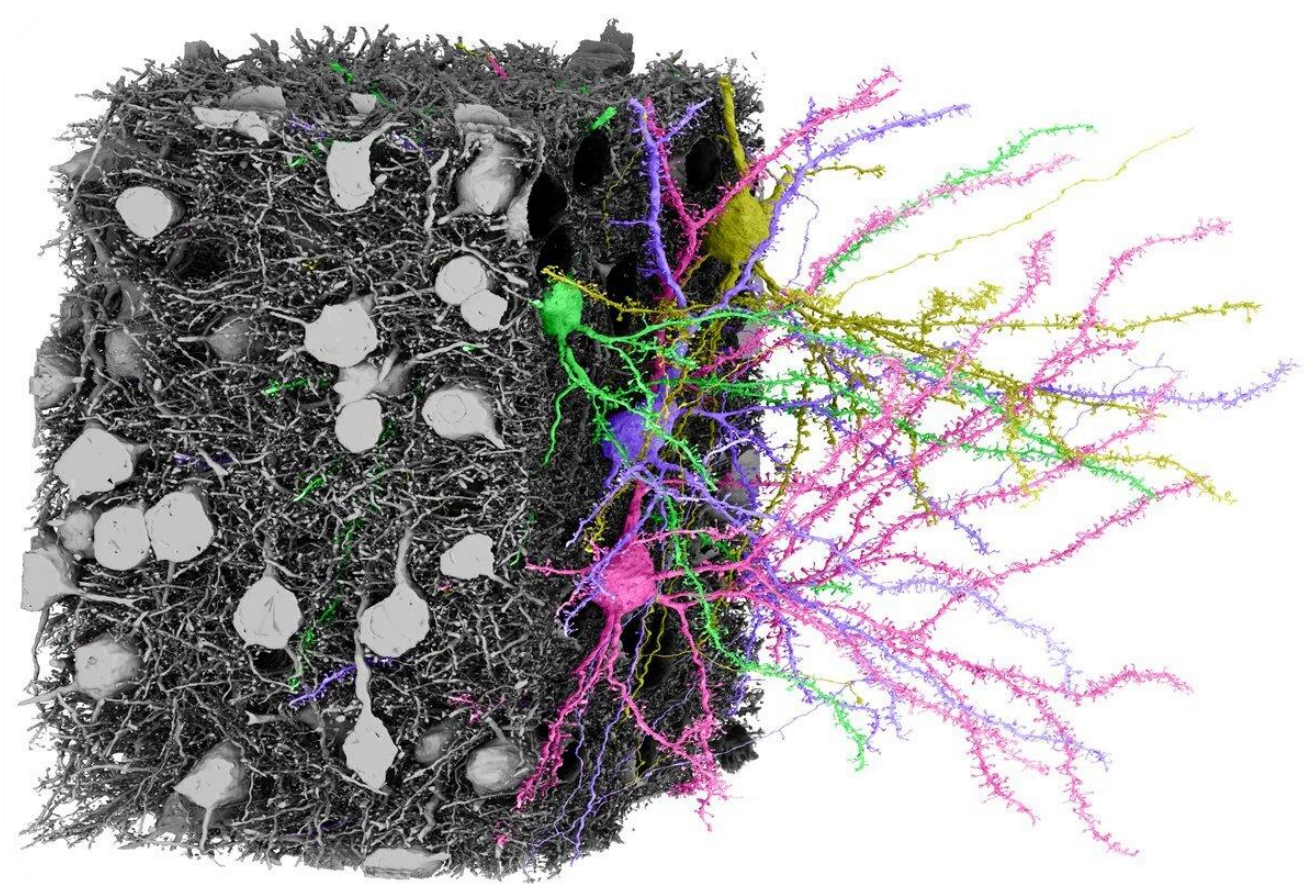
# Source: downloading meshes

- Mesh download code from Forrest Collman's [MeshExample.ipynb](#)
- Thank you Bethanny Danskin (Allen Institute) for providing a workaround for meshparty legacy file handling error
- See [Allen Institute Microns Binder github repository](#) for more details and additional notebooks for working with the MICrONS volume datasets

# Source: pyvista mesh decimation

- Mesh decimation code from Tyler Sloan's [Mesh Decimation Pipeline.ipynb](#)
- See [Quorumetrix github repository](#) for more details and additional scripts for working with volume meshes in Blender (however, all visualization examples shown here use vtk/OpenGL)

# MICrONS Layer 2/3 (pinky) volume



**Explore EM data  
from layer 2/3 of the  
mouse visual cortex**

# Mesh Types in Layer 2/3 Volume

Cell

- Cell soma meshes

Mito

- Mitochondria meshes

Nuc

- Nuclei meshes

# Layer 2/3: download meshes



Visit the [download\\_meshes](#) folder on github



Use the respective notebooks to download and view the cell, mitochondria, and nuclei meshes

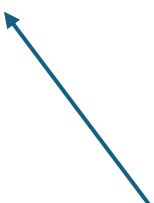


The cell body and nuclei notebooks also visualize the downloaded meshes with matplotlib and plotly plots



# Mesh download for cell bodies

```
# setup the mesh meta to handle downloads and  
    caching  
mesh_dir = 'data/neuron_meshes_v185/'  
seg_source =  
"precomputed://gs://microns_public_datasets/pinky100  
_v185/seg"  
cv_obj = cloudvolume.CloudVolume(seg_source,  
use_https=True)  
seg_id = [648518346349525821]
```



Workaround code for meshparty legacy issue with Layer 2/3 cell body mesh files

# Mesh download for cell bodies

*# Iterating through the seg\_id list*

```
for seg in seg_id:
```

```
    print(f"Processing seg_id: {seg}")
```

*# Retrieve the mesh object with tqdm progress bar*

```
    print("Retrieving mesh...")
```

```
    cv_mesh = cv_obj.mesh.get(seg,  
remove_duplicate_vertices=True)
```

# Mesh download for cell bodies

*# Extracting and reshaping faces if necessary*

```
print("Processing mesh faces...")
```

```
faces = np.array(cv_mesh.faces)
```

```
if len(faces.shape) == 1:
```

```
    faces = faces.reshape(-1, 3)
```

*# Creating the Mesh object*

```
print("Creating Mesh object...")
```

```
mesh = Mesh(vertices=cv_mesh.vertices, faces=faces)
```

# Mesh download for cell bodies

*# Constructing the file path*

```
mesh_file = os.path.join(mesh_dir, str(seg) + '.h5')
```

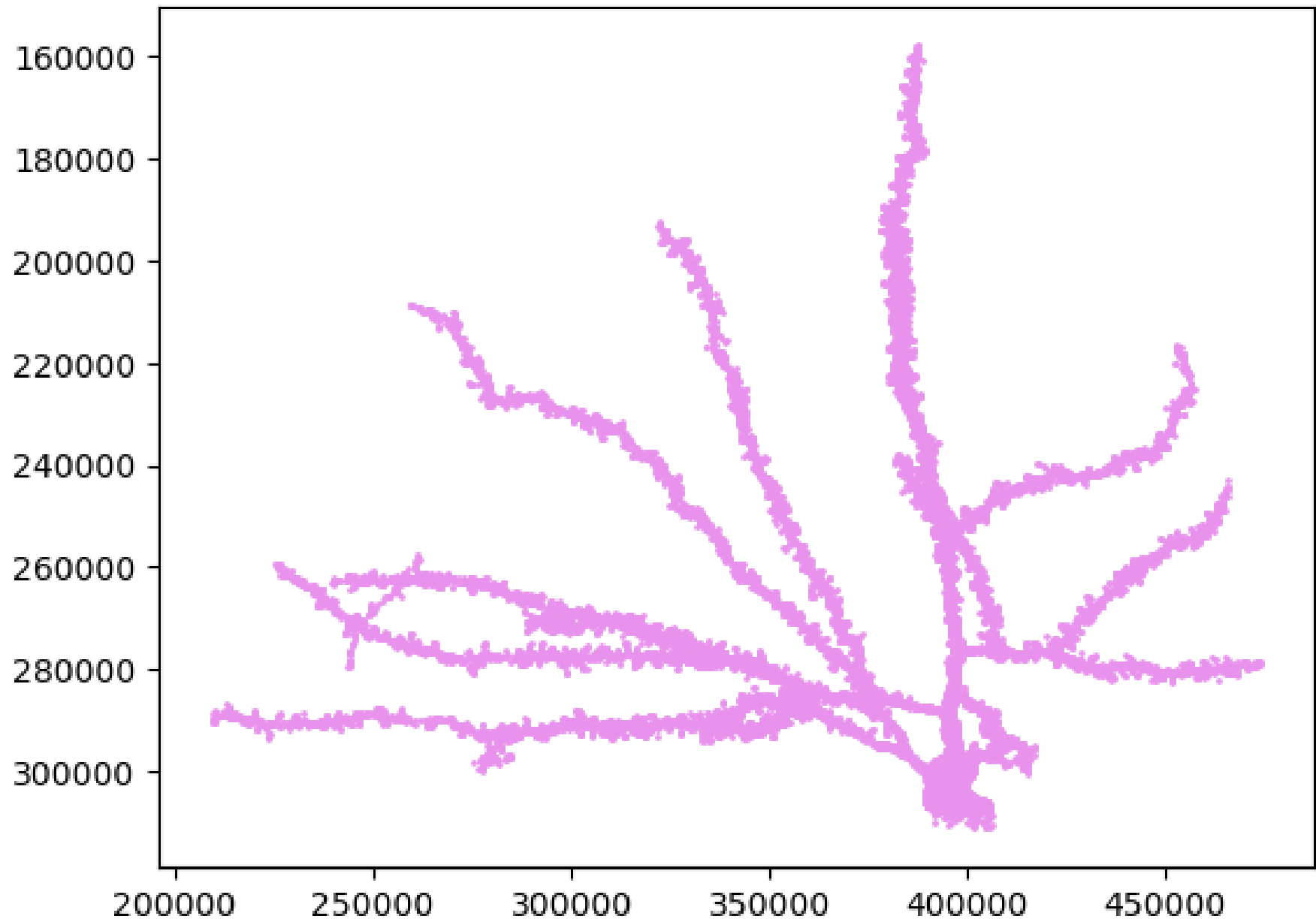
*# Writing the mesh to file*

```
print(f"Writing mesh to file: {mesh_file}")
```

```
mesh.write_to_file(mesh_file)
```

```
print(f"Finished processing seg_id: {seg}\n")
```

matplotlib  
visualization  
of a pyramidal  
cell mesh



# Mesh download for mitochondria

*# setup the mesh meta to handle downloads and caching*

```
mito_mesh_dir = 'data/meshes/'  
mito_source =  
"precomputed://https://td.princeton.edu/sseung-  
archive/pinky100-mito/seg_191220"  
mito_mm =  
trimesh_io.MeshMeta(cv_path=mito_source,  
  
disk_cache_path=mito_mesh_dir)
```

# Mesh download for mitochondria

```
cellid_list = [648518346349529031]
```

```
mitodf =  
mito[mito['cellid'].isin(cellid_list)]  
mitodf_mitoid_list = mitodf.mito_id.to_list()
```

# Mesh download for mitochondria

```
for i in range(len(mitodf_mitoid_list)):
    mito_id = mitodf_mitoid_list[i]
    mito_seg_id = mito_id
    mito_downloadmesh = mito_mm.mesh(seg_id =
mito_seg_id, remove_duplicate_vertices=True)

    print(f'completed ' + str(i) + ' of ' +
str(len(mitodf_mitoid_list)))
```



# Mesh download for nuclei

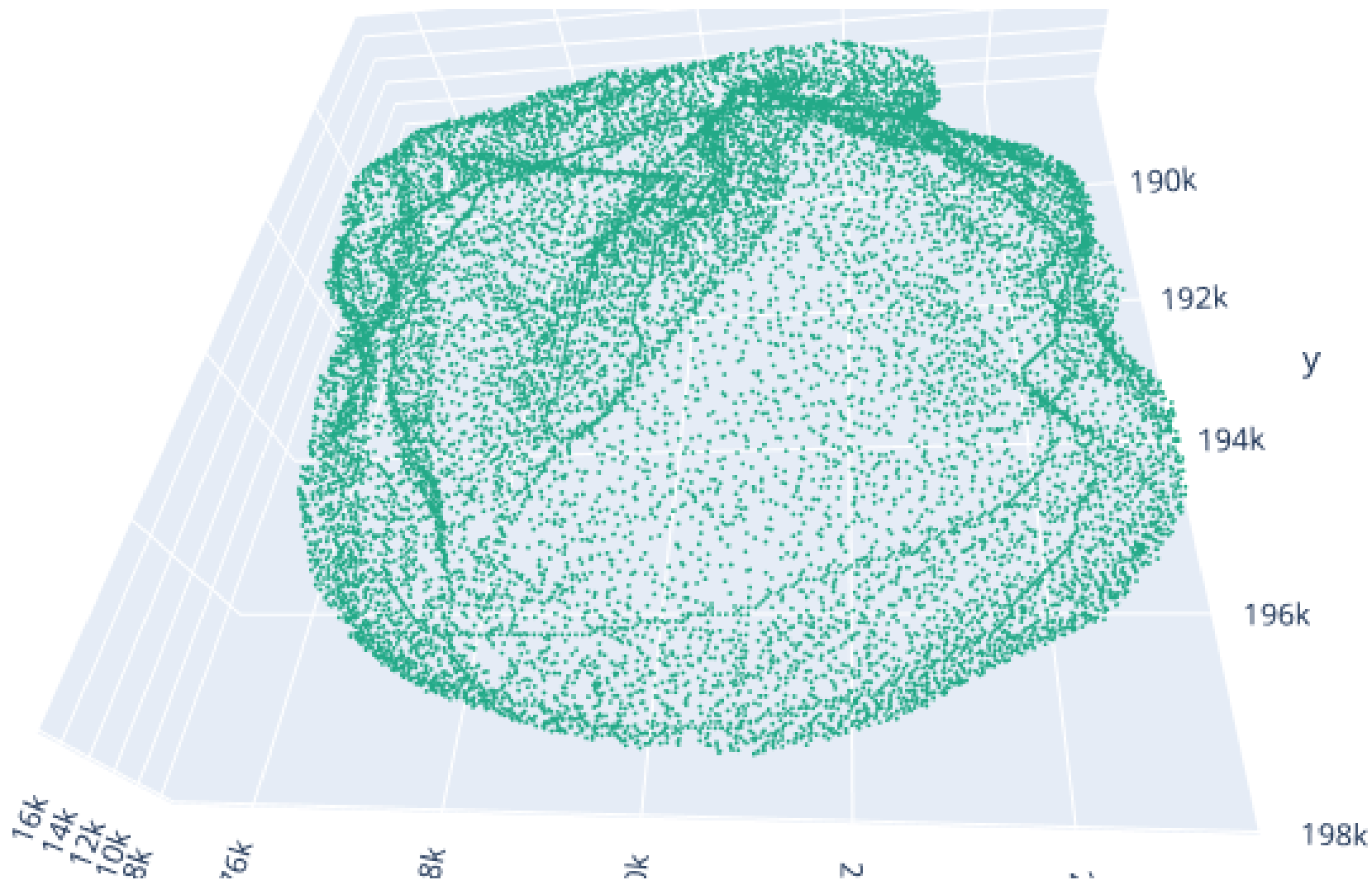
```
nuc_mesh_dir = 'data/nuclei_meshes/'  
nuc_source =  
"precomputed://https://td.princeton.edu/sseung  
-archive/pinky100-nuclei/seg"  
nuc_mm =  
trimesh_io.MeshMeta(cv_path=nuc_source,  
  
disk_cache_path=nuc_mesh_dir)
```

# Mesh download for nuclei

```
nuc_id = [5645]
```

```
for i in tqdm(range(len(nuc_id)),  
desc="Downloading Meshes"):  
    downloadmesh = nuc_mm.mesh(seg_id=nuc_id[i],  
remove_duplicate_vertices=True)
```

plotly  
visualization  
of a nucleus  
mesh



# Layer 2/3: decimate meshes



Visit the [decimated\\_meshes](#) folder on github



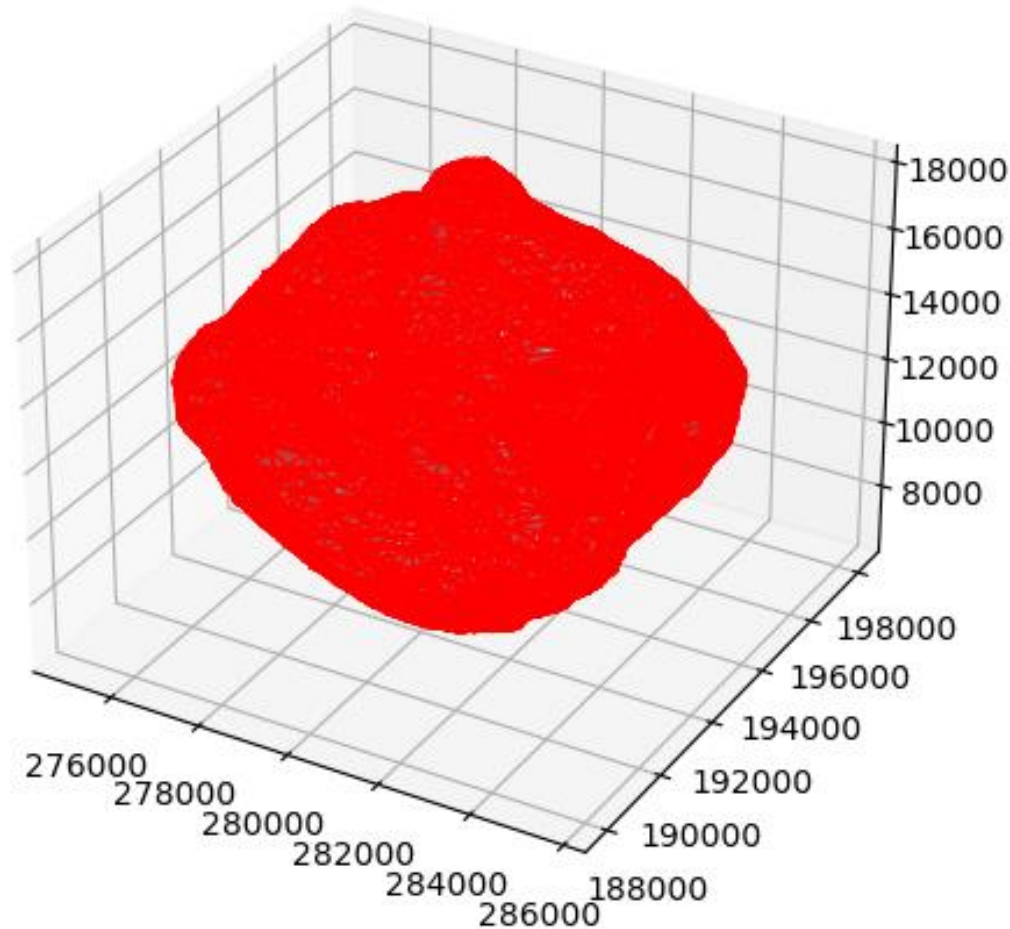
Use the respective notebooks to decimate the cell, mitochondria, and nuclei meshes



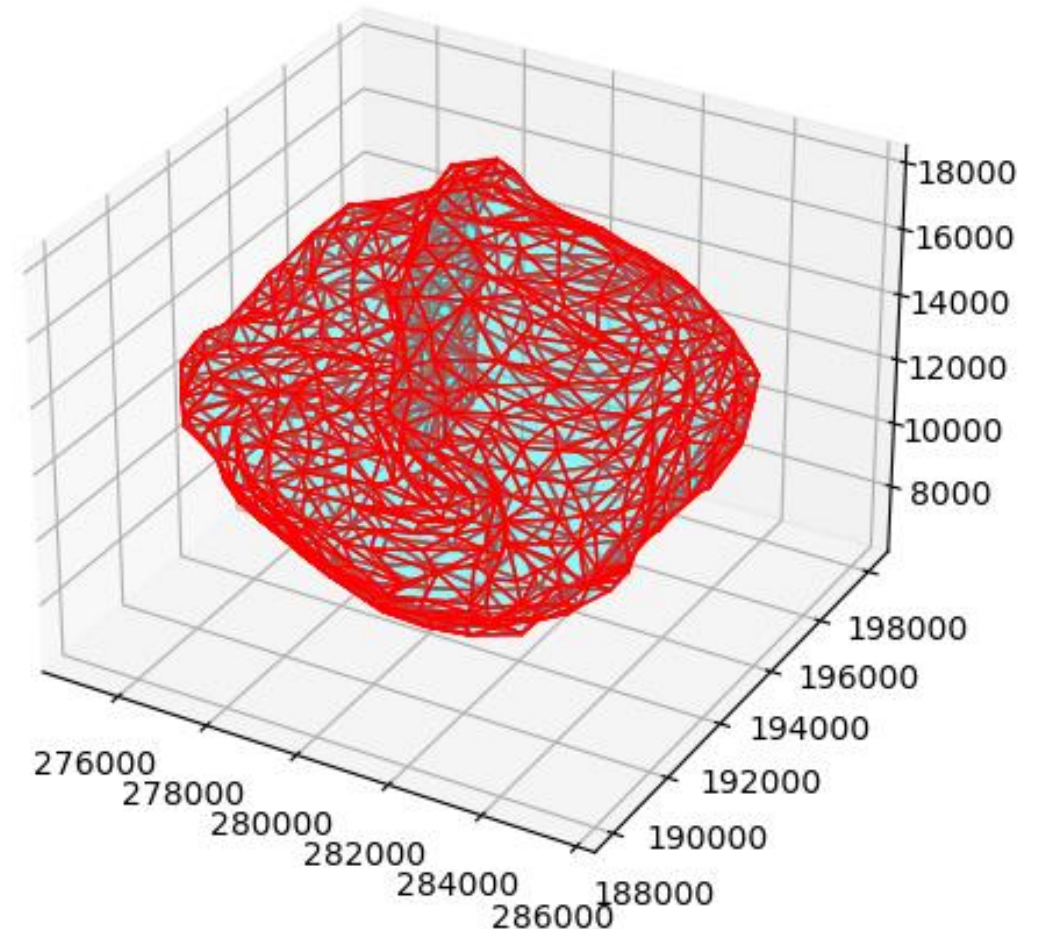
Options for single and batch processing meshes using lists

# Mesh decimation: nucleus example

Original Mesh



Decimated Mesh



# Layer 2/3 volume: visualize meshes

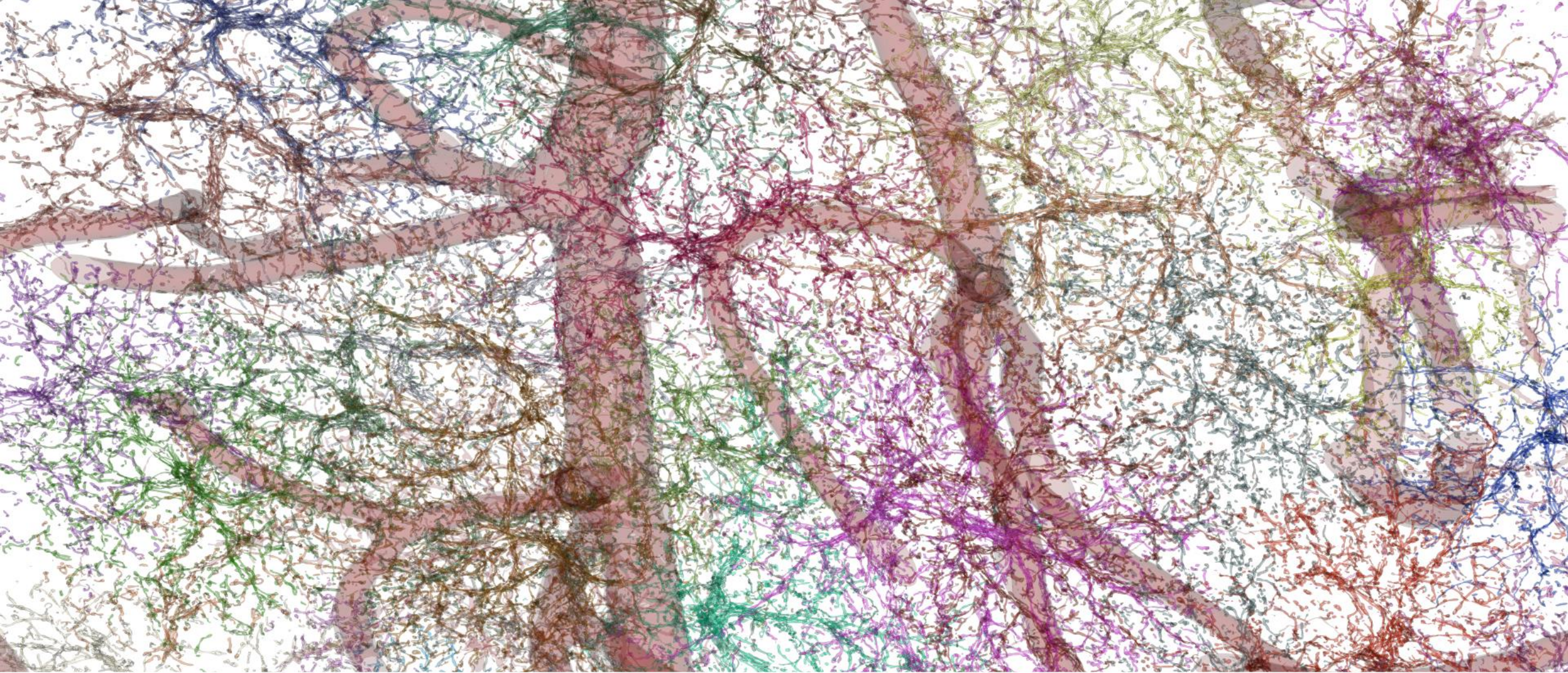


Visualize decimated meshes using a variety of methods with vtk with customized notebooks in the [decimated\\_meshes](#) directory



Additional options include adding pre- and post-synaptic sites, filtered for pre-synaptic sites for a neuron of interest, highlighted cells, highlighted mitochondria by size, and more

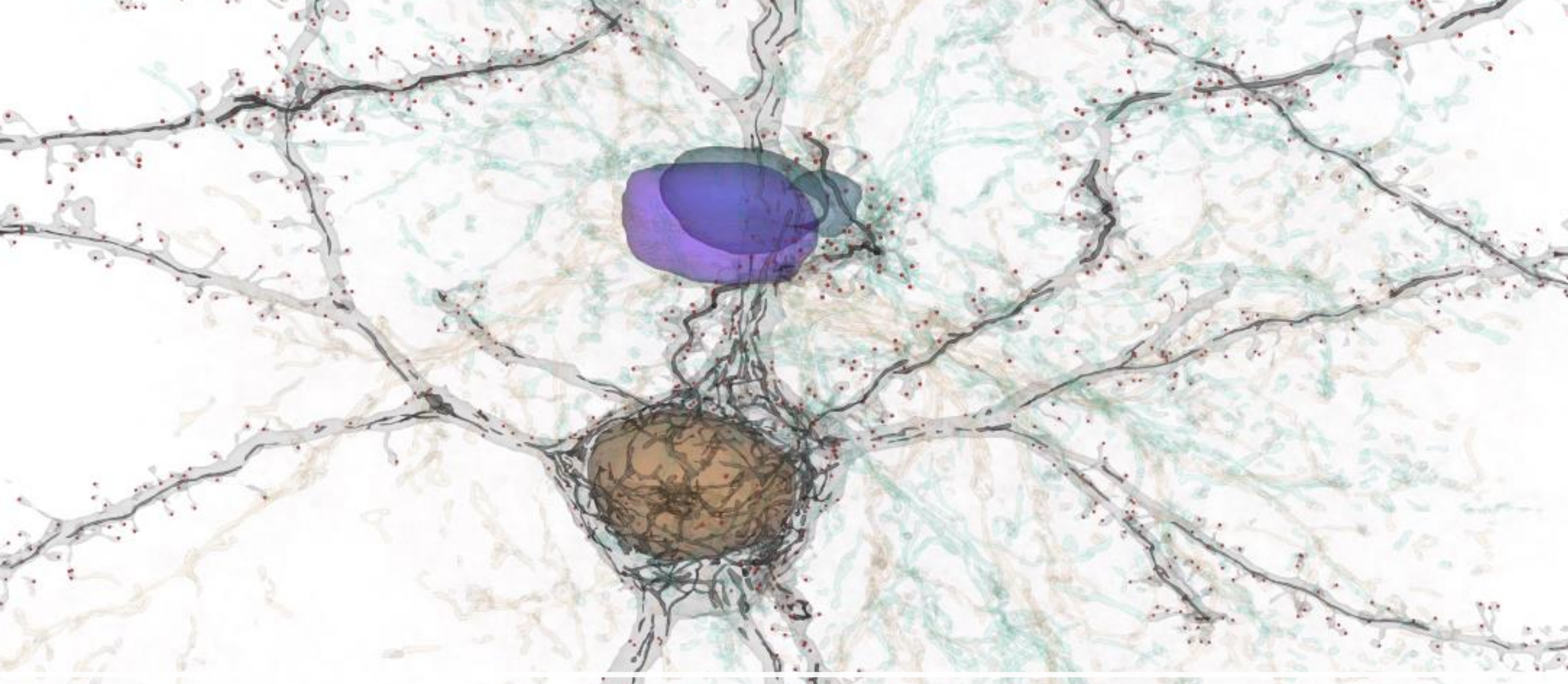




All mitochondria in 44 astrocytes in the Layer 2/3 volume with vasculature shown in red







Two astrocyte cell bodies distort the apical dendrite of a Layer 2/3 pyramidal neuron



# MICrONS 1 mm<sup>3</sup> (minnie) volume

MICrONS Explorer [Home](#) [Data](#) [Requests](#) [Tools](#) [Gallery](#) [About](#)



A functional connectome  
containing 200,000 cells,  
75,000 neurons with  
physiology, and 523 million  
synapses.

# minnie volume: download meshes



Visit the Jupyter Notebook folder on [github](#)



Use [minnie\\_plotly\\_and\\_vtk\\_visualizer.ipynb](#) to download and view cell meshes (list-based approach)

# Mesh download

*# setup the mesh meta to handle downloads and caching*

```
mesh_dir = 'data/cell_meshes'
```

```
seg_source =
```

```
"precomputed://gs://iarpa_microns/minnie/minnie65/  
seg"
```

```
mm = trimesh_io.MeshMeta(cv_path=seg_source,  
                           disk_cache_path=mesh_dir,  
                           cache_size=20)
```

# Mesh download

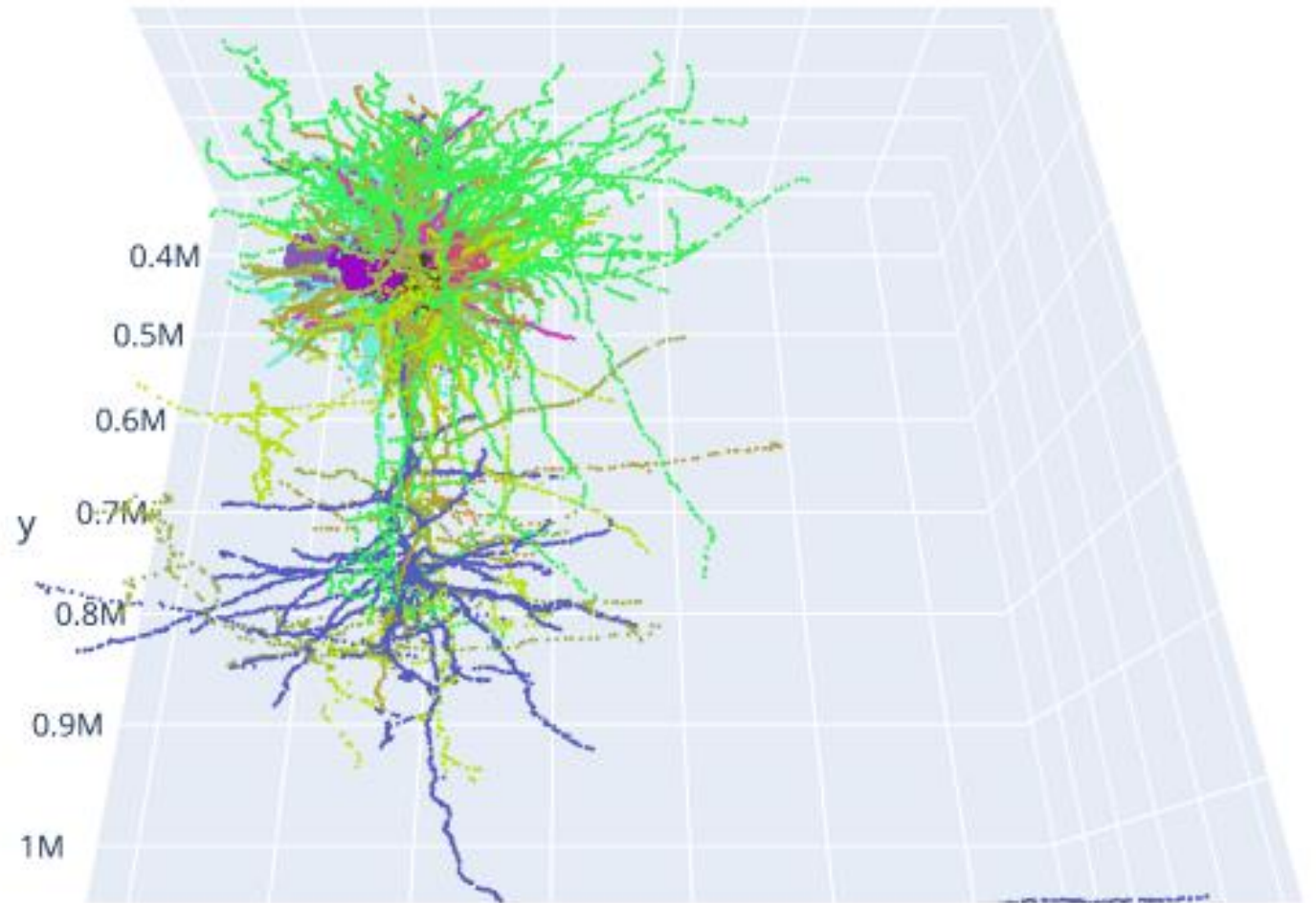
```
cell_id_list = [864691134964446239,  
864691135012524790, 864691135256138671,  
864691135269913253, 864691135337851366,  
864691135341421745, 864691135348239831,  
864691135386363265, 864691135497750291,  
864691135657783170, 864691135809446092,  
864691135837182867, 864691135880405261,  
864691136023933241, 864691136194008022,  
864691136330394007]
```

# Mesh download

```
for i in tqdm(range(len(cell_id_list)),  
desc="Downloading Meshes"):  
    downloadmesh = mm.mesh(seg_id=cell_id_list[i],  
remove_duplicate_vertices=True)
```



# Plotly visualization of mesh objects



# minnie volume: decimate meshes



Decimate meshes using a list of cellids of interest with [pyvista\\_decimate\\_cell\\_bodies\\_minnie.ipynb](#)

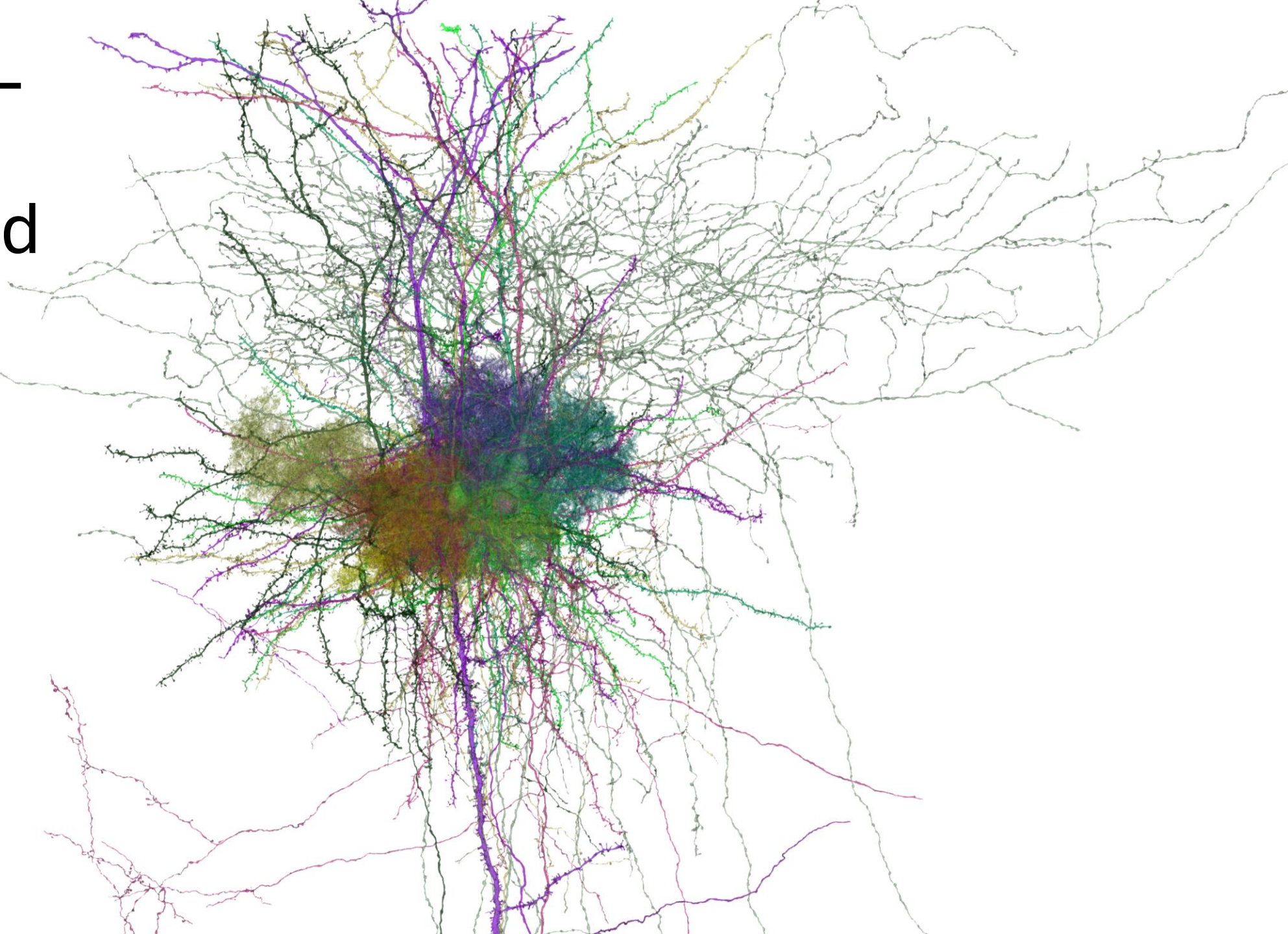
# minnie volume: visualize meshes



Visualize cell meshes by cell type  
(e.g., neurons, astrocytes,  
vasculature) with vtk



vtk OpenGL  
rendered  
neurons and  
astrocytes





# Harvard/Google H01 volume

**H01 Release**

[Explore](#) [Gallery](#) [Manuscript](#) [Layers](#) [SegCLR Embeddings](#) [CREST](#) [Proofreading](#) [Released Data](#) [Code](#)



**A Browsable Petascale Reconstruction of the Human  
Cortex**

# H01 volume: download meshes



Visit the Jupyter Notebook folder on [github](#)



Start with [h01\\_plotly\\_visualizer.ipynb](#) to download and view cell meshes



See [h01\\_cell\\_types](#) folder for a list of cellids of interest in the H01 volume (e.g., astrocytes, pyramidal neurons, etc.)

# Mesh download

*# setup the mesh meta to handle downloads and caching*

```
mesh_dir = 'data/c3_cell_meshes/' # or change to  
your desired folder
```

```
seg_source = "precomputed://gs://h01-  
release/data/20210601/c3"
```

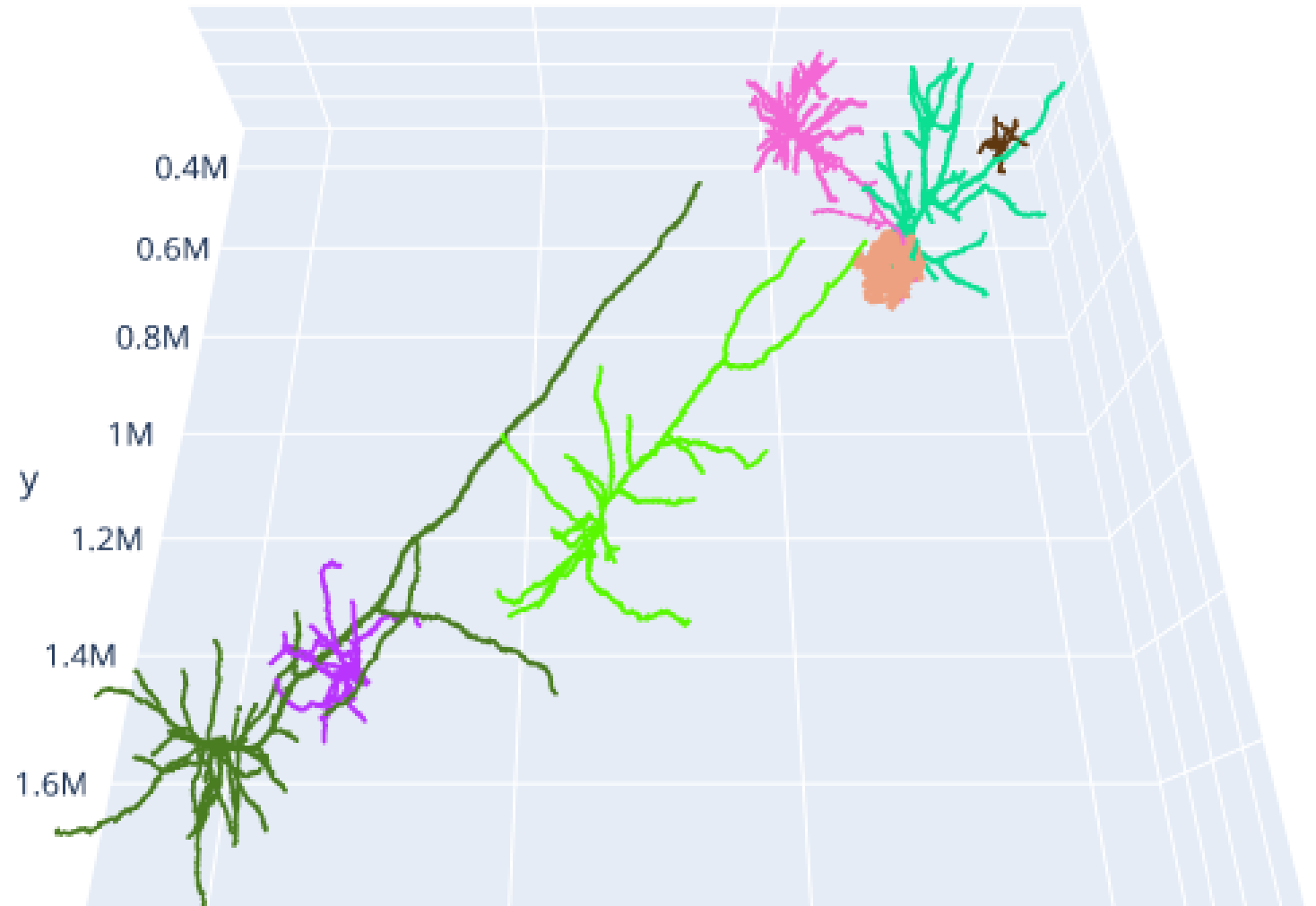
```
mm = trimesh_io.MeshMeta(cv_path=seg_source,  
                           disk_cache_path=mesh_dir,  
                           cache_size=20)
```

# Mesh download

```
cell_id_list = [28541451958, 27843438291,  
4970736617]
```

```
for i in tqdm(range(len(cell_id_list)),  
desc="Downloading Meshes"):  
    downloadmesh = mm.mesh(seg_id=cell_id_list[i],  
remove_duplicate_vertices=True)
```

# Plotly visualization of mesh objects



# H01 volume: decimate meshes



Decimate [astrocyte meshes](#) of interest as a list  
(can be used for other cell types too)



Complex meshes, such as astrocytes and vasculature, may take hours (~12 hours per astrocytes) or run out of memory on a typical computer



Use [Google Colab](#) with High-RAM (subscription to Google Colab Pro required) to allocate enough memory and reduce processing time (<10 min)

# H01 volume: visualize meshes



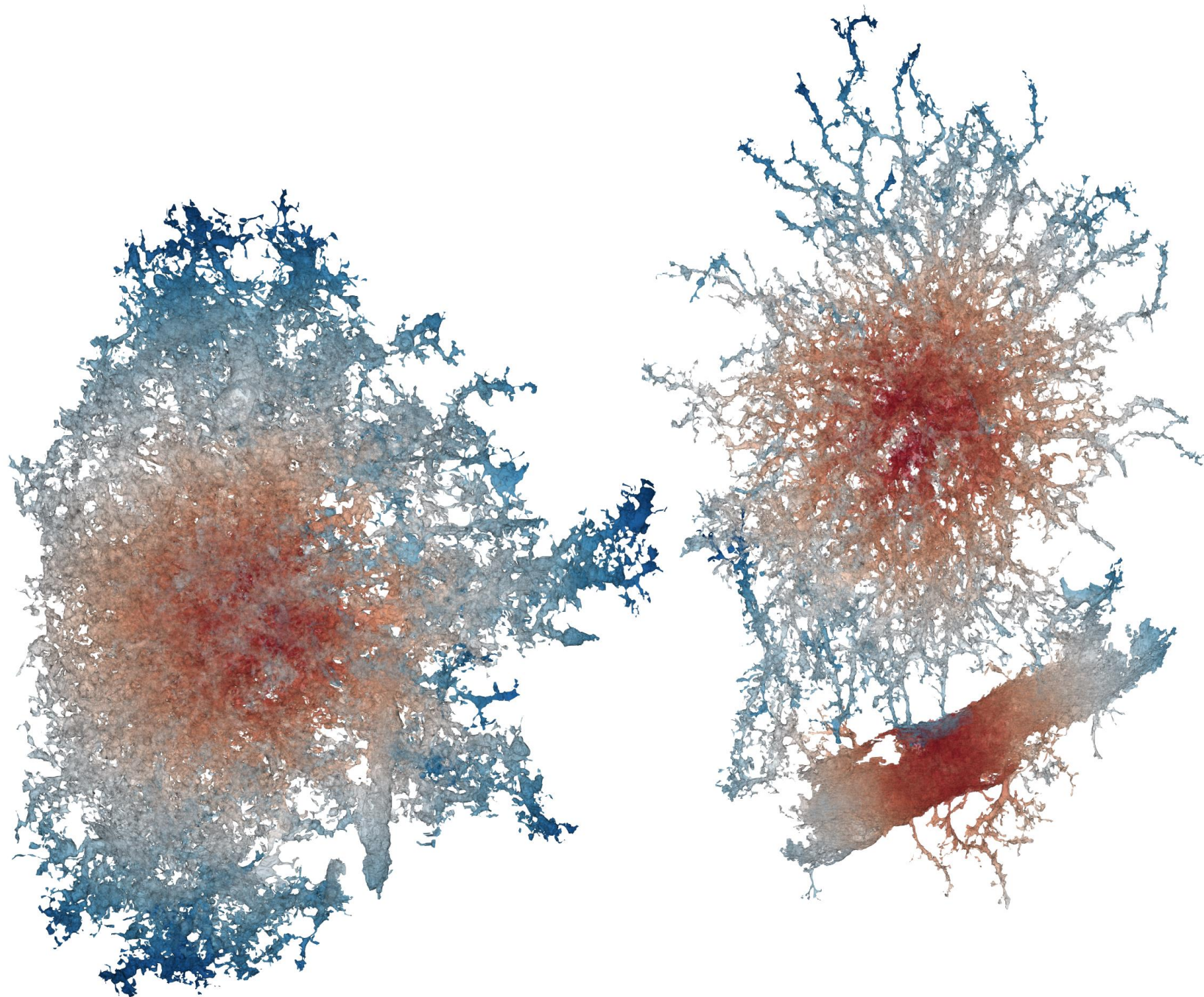
Visualize [decimated astrocyte meshes](#) using a gradient coloring method with vtk

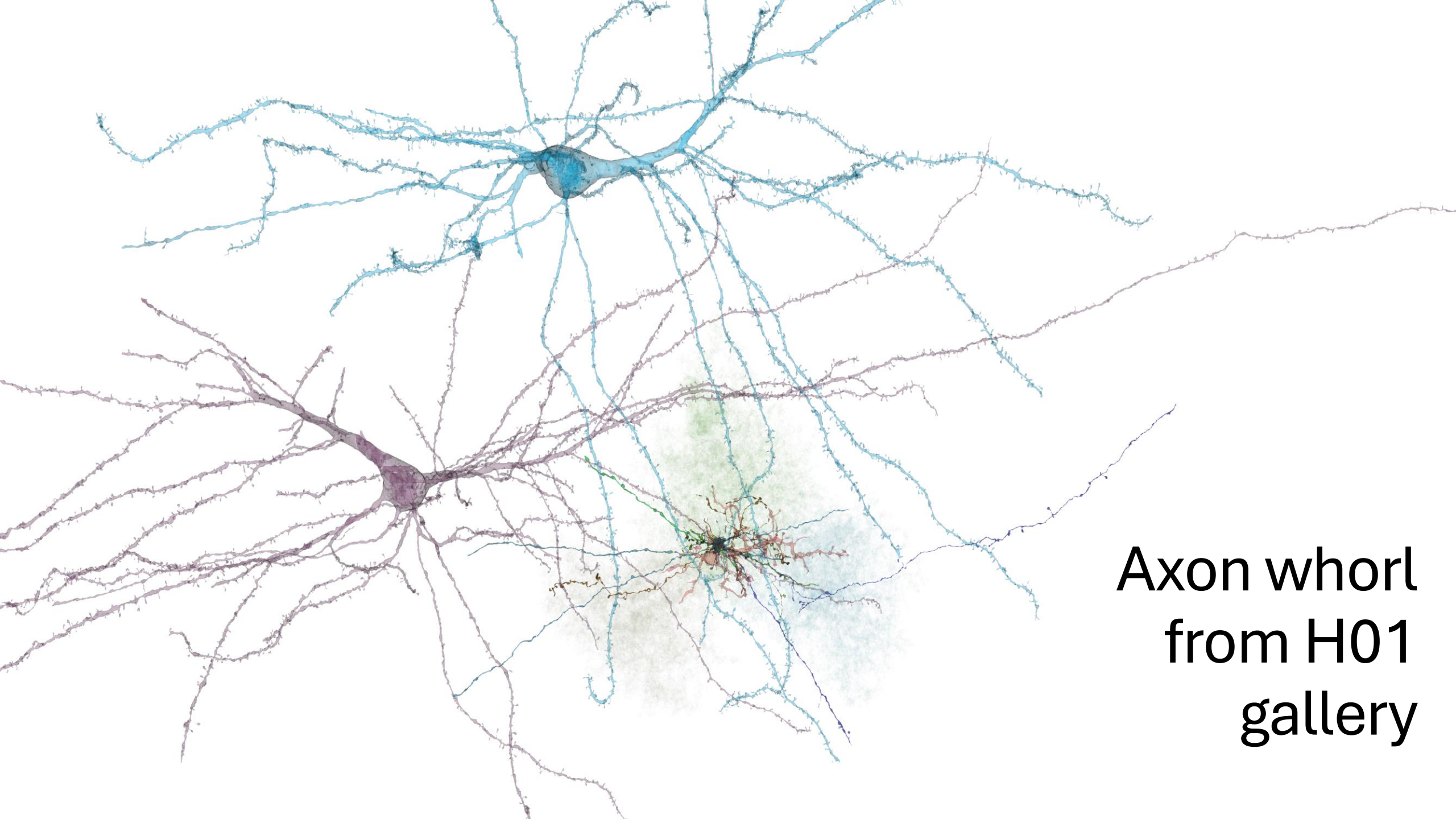


Visualize [decimated meshes by four cell categories](#) (neurons, astros, microglia, and unidentified segments) with vtk



# Gradient coloring of astrocytes





Axon whorl  
from H01  
gallery



# Acknowledgements: MICrONS



Read the original research papers on the [Citation](#) page at Allen Institute



Read the [Terms and Conditions](#) page



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# Acknowledgements: H01



Link to the original research paper on the [Manuscript](#) page of [H01 Release](#) website



Read the license on the [Released Data](#) page



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# Code Availability

Visit my github repositories  
for each volume:

- [layer23-volume](#)
- [minnie-volume](#)
- [h01-volume](#)