

Precision Immunotherapy to Improve Sepsis Outcomes

The ImmunoSep Randomized Clinical Trial

Evangelos J. Giannarellos-Bourboulis, MD; Antigone Kotsaki, MD; Ioanna Kotsamidi, MD; Aikaterini Efthymiou, MD; Vasiliki Koutsoukou, MD; Johannes Ehler, MD; Alexandra Paridou, MD; Frantzeska Frantzeskaki, MD; Marcella C. A. Müller, MD; Peter Pickkers, MD; Sylvain Meylan, PhD; Ioannis Nikolopoulos, MD; Mihaela Lupsie, MD; Alexandra Gavala, MD; Glykeria Vlachogianni, MD; Nicky Solomonidi, MD; Antonia Alevizou, MD; Eumorfia Kondili, MD; Eleni Antoniadou, MD; Maria Nakou, MD; Nikolaos Markou, MD; Erifili Hatzigelaki, MD; Athanassios Prekates, MD; Apostolos Komnos, MD; Lieke Bakkerus, MD; Marleen A. Slim, MD; George N. Dalekos, MD; Areti Karapanagiotou, MD; Sofia Ktena, BSc; Gennaro De Pascale, MD; Vassileios Koulouras, MD; Christos Psarrakis, MD; Eleni Massa, MD; Konstantina Dakou, MSc; Chrisoula Pavzanti, MD; Aikaterini Ioakeimidou, MD; Iraklis Tsangaris, MD; Nikolaos Antonakos, MD; Alexander P. J. Vlaar, MD; Souzana Anisoglou, MD; Thierry Calandra, MD; Vasilios Papaioannou, MD; Pavlos Myrianthefs, MD; Maria Patrani, MD; Ioannis Alamanos, MD; Massimo Antonelli, MD; Jos W. M. van der Meer, MD; Tom van der Poll, MD; W. Joost Wiersinga, MD; Maria Ntaganou, MD; Eleni Gkeka, MD; Michael Bauer, MD; Eleni Mouloudi, MD; Mihai G. Netea, MD; for the ImmunoSep Study Group

IMPORTANCE Sepsis is heterogeneous, and the optimal strategy for tailoring immunotherapy is uncertain.

OBJECTIVE To investigate whether precision immunotherapy guided by the presence of macrophage activation-like syndrome or sepsis-induced immunoparalysis improves organ dysfunction by day 9.

DESIGN, SETTING, AND PARTICIPANTS A randomized, double-blind, double-dummy, placebo-controlled clinical trial conducted in 6 countries. Patients with sepsis, defined by Sepsis-3, were included if they had community-acquired or hospital-acquired pneumonia or ventilator-associated pneumonia or bacteremia and sepsis and had displayed either macrophage activation-like syndrome (blood ferritin >4420 ng/mL) or sepsis-induced immunoparalysis (blood ferritin ≤4420 ng/mL and <5000 human leukocyte antigen DR receptors on CD45/CD14 monocytes). The first patient was enrolled August 5, 2021, and the last follow-up, April 29, 2024.

INTERVENTIONS Eligible patients were randomized to receive standard care and precision immunotherapy or standard care and placebo. Those in the precision immunotherapy group with macrophage activation-like syndrome received anakinra intravenously (IV) and placebo subcutaneously, and those with sepsis-induced immunoparalysis received subcutaneous recombinant human interferon gamma and IV placebo. Those in the placebo group received both IV and subcutaneous placebo. Treatment was administered for up to 15 days.

MAIN OUTCOMES AND MEASURES The primary end point was a decrease of at least 1.4 points in the mean Sequential Organ Failure Assessment (SOFA) score from baseline by day 9. The SOFA score evaluates 6 organ systems, ranging from 0, no dysfunction, to 4, failure, and the total score ranges from 0, normal, to 24, most severe form of multiorgan failure. Key secondary outcomes included 28-day mortality.

RESULTS Of 672 patients assessed for eligibility, 281 were randomized and 276 were included in the primary analysis population (mean [SD] age, 70 [13] years; 93 females [33.7%]; median baseline SOFA score, 9 [IQR, 7-11]). The SOFA decrease end point was attained by 46 of 131 patients (35.1%) in the precision immunotherapy group and by 26 of 145 patients (17.9%) in the placebo group (difference, 17.2% [95% CI, 6.8% to 27.2%]; $P = .002$). Mortality at 28 days was not statistically significantly different between groups. A total of 1069 serious treatment-emergent adverse events (88.8%) were reported; increased incidence of anemia was noted in the anakinra group; and hemorrhage in the recombinant human interferon gamma group.

CONCLUSIONS AND RELEVANCE Among patients with sepsis, precision immunotherapy targeting macrophage activation-like syndrome and sepsis-induced immunoparalysis improved organ dysfunction by day 9 compared with placebo.

TRIAL REGISTRATION ClinicalTrials.gov Identifier: [NCT04990232](#)

JAMA. doi:[10.1001/jama.2025.24175](#)

Published online December 8, 2025.

- + [Visual Abstract](#)
- + [Research Summary](#)
- + [Editorial](#)
- + [Supplemental content](#)

Author Affiliations: Author affiliations are listed at the end of this article.

Group Information: A list of the ImmunoSep Study collaborators appears at the end of the article and in *Supplement 3*.

Corresponding Author: Evangelos J. Giannarellos-Bourboulis, MD, PhD, Fourth Department of Internal Medicine, ATTIKON University Hospital, One Rimini St, 12462 Athens, Greece (egiamarel@med.uoa.gr).

Sepsis is a life-threatening organ dysfunction due to the dysregulated host response to an infection.¹ Many randomized clinical trials (RCTs) studied immunotherapies but failed to demonstrate an improvement in outcome. This is likely related to the heterogeneity of immune dysregulation, which varies from hyperinflammation to immunosuppression. As such, a precision approach to immunotherapy in sepsis may be necessary,² relying on rapid, accurate classification of the immune dysregulation using biomarkers.³

Recent data suggest that patients with sepsis with shock can be classified by using blood ferritin concentrations and the expression of the human leukocyte antigen (HLA) DR on monocytes into 3 strata of immune activation. These include, (1) the macrophage activation-like syndrome, which is a hyperinflammatory condition driven by excess production of interleukin 1 (IL-1) by tissue macrophages; (2) sepsis-induced immunoparalysis, which is a hypoinflammatory condition driven by exhaustion and apoptosis of lymphocytes and often of myeloid cells; and (3) unclassified patients with neither of these 2 types of immune dysregulation.⁴ Patients with macrophage activation-like syndrome who were randomized to receive a 7-day intravenous course of a recombinant human IL-1 receptor antagonist demonstrated a decrease in sequential Organ Dysfunction Score compared with placebo, although the effect was time-limited. Others suggest that the immunostimulating recombinant interferon gamma could reverse the effects of sepsis-induced immunoparalysis.⁵

This international multicenter phase 2b study aimed to assess if a precision immunotherapy strategy in sepsis tailored to the presence of macrophage activation-like syndrome or sepsis-induced immunoparalysis using either anakinra or recombinant human interferon gamma improved organ dysfunction compared with placebo.⁶

Methods

Trial Design

ImmunoSep was a prospective, multicenter, interventional, randomized, double-blind, double-dummy, placebo-controlled, clinical trial at 33 study sites in 6 countries (Greece, Germany, Italy, the Netherlands, Switzerland, and Romania; eTable 1 in *Supplement*). ImmunoSep is considered a phase 2b trial since the study drugs were tested in a precision approach for the first time. The study protocol and addenda were approved by the ethics committees and institutional review boards pursuant to country regulations (*Supplement 1*, eTable 2 in *Supplement 2*). The study was governed by the international standards for Good Clinical Practice, the Directive 2001/20/EC for Clinical trials and the General Data Protection Regulation 679/2016. The study was registered at EudraCT number: 2020-005768-74 and at Clinicaltrials.gov [NCT04990232](#). This trial followed the Consolidated Standards of Reporting Trials ([CONSORT](#)) reporting guideline.⁷

Participants

Prior to blood sampling and enrollment, written informed consent was provided by study participants or first-degree rela-

Key Points

Question Does immunotherapy tailored to the type of immune dysregulation improve outcomes among patients with sepsis?

Findings In this multicenter randomized clinical trial of 276 patients, precision immunotherapy, which included the administration of anakinra for patients with macrophage activation-like syndrome and recombinant human interferon gamma for patients with sepsis-induced immunoparalysis, decreased the Sequential Organ Failure Assessment score by at least 1.4 points by day 9 in 35.1% of patients in the immunotherapy group vs 17.9% in the placebo group.

Meaning A precision strategy of immunotherapy improved organ dysfunction at day 9 among patients with sepsis.

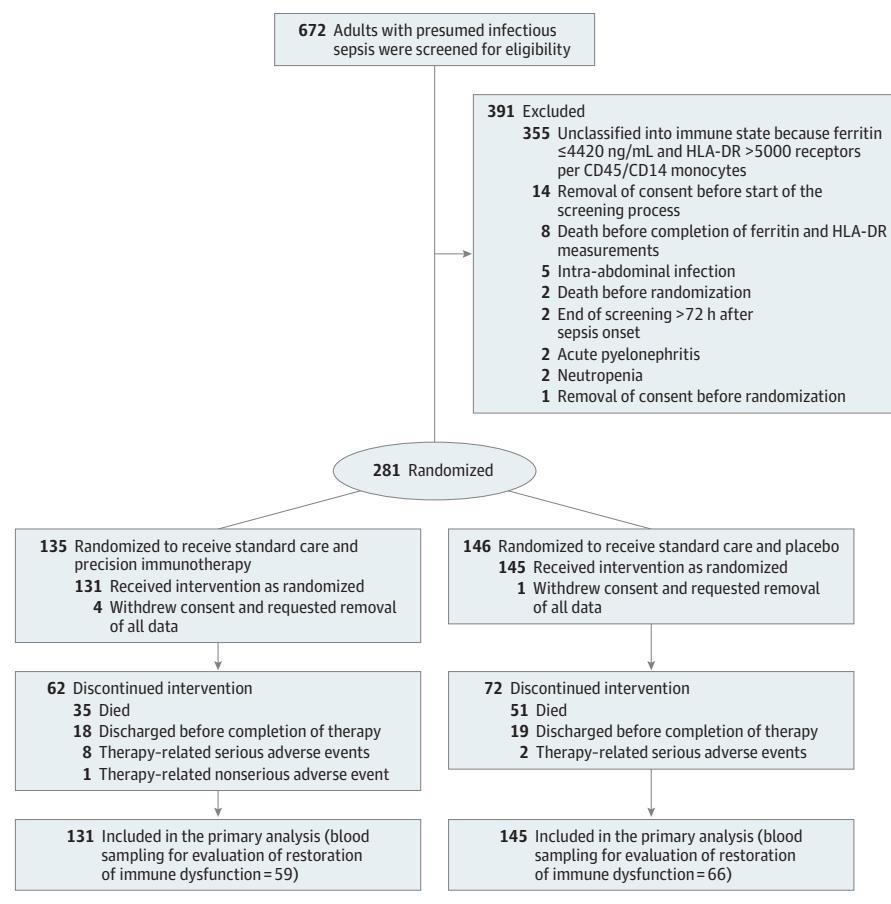
tives if patients could not consent. Patients were adults with community-acquired pneumonia, hospital-acquired pneumonia, ventilator-associated pneumonia, or primary bacteremia, with sepsis as defined by the Sepsis-3 consensus definitions¹ and macrophage activation-like syndrome or sepsis-induced immunoparalysis. The maximum time window allowed between sepsis onset and start of blinded intervention was less than 72 hours. Patients were excluded if the primary sepsis source was (1) abdominal infections due to confounding from the time to source control, (2) urinary tract infections because they usually have better outcomes than sepsis from other causes,⁸ or (3) SARS-CoV pneumonia in the absence of macrophage activation-like syndrome (details are available in eMethods and eTable 3 in *Supplement 2*). At enrollment, eligible patients had 6 mL of blood drawn into an ethylenediaminetetraacetic acid coated tube (Vacutainer, BD; Becton Dickinson). Biospecimens were shipped within 6 hours to the designated central laboratory in each participating country for measurements of ferritin and the number of HLA-DR receptors on CD45/CD14 monocytes (see detailed eMethods in *Supplement 2*). Patients with ferritin levels greater than 4420 ng/mL were classified as having macrophage activation-like syndrome⁴; and patients with ferritin levels of 4420 ng/mL or less and less than 5000 HLA-DR molecules on CD45/CD14-monocytes as having sepsis-induced immunoparalysis.⁴ Patients with ferritin levels of 4420 or less ng/mL and 5000 or more HLA-DR receptors on CD45/CD14 monocytes were considered to be in an “unclassified immune state” and were not enrolled in the study (eTable 4 in *Supplement 2*).

Randomization and Allocation

Enrolled patients were randomized 1:1 into groups: (1) intervention strategy: standard care and precision immunotherapy, and (2) control strategy: standard care and placebo immunotherapy, using separate randomization per study site with the aid of a computer-generated sequencing (*Figure 1*). Patients received 1 intravenous (IV) injection every 8 hours and 1 subcutaneous injection every 48 hours for 15 days.

In the precision immunotherapy group, the IV injection was 200 mg of anakinra (Kineret, Swedish Orphan BioVitrum) dissolved in 0.9% of normal saline to a final volume of 20 mL for patients with macrophage activation-like

Figure 1. Flow of Patients Through the ImmunoSep Study



HLA-DR indicates human leukocyte antigen DR.

syndrome; and the subcutaneous injection was 100 µg of recombinant human interferon gamma (Imukin, Clinigen) to a final volume of 0.5 mL for patients with sepsis-induced immunoparalysis. Patients with macrophage activation-like syndrome also received subcutaneous dummy treatment; and patients with sepsis-induced immunoparalysis received IV dummy treatment. In the placebo group, the IV injection was 20 mL of 0.9% normal saline and the subcutaneous injection 0.5 mL of 0.9% normal saline. Creatinine clearance was calculated daily by the Cockcroft and Gault equation. Patients with clearance less than 30 mL/min received half the IV dose.

The study drug was prepared by unblinded study pharmacists and identically labeled. Except for the unblinded pharmacists, the investigators, attending physicians, nursing personnel, and participants remained blinded to the assigned intervention. The data and safety monitoring board performed regular safety evaluations.

Study Procedures

Baseline data included demographics, comorbidities, severity scores (Acute Physiology and Chronic Health Evaluation [APACHE] II, Charlson Comorbidity Index [CCI], and Sequential Organ Failure Assessment [SOFA] score), underlying infection, laboratory results (complete blood cell count, biochem-

istry, and coagulation), and microbiology (Table 1). The maximum SOFA scores and organ support treatments were captured each day of patients' intensive care unit (ICU) stay until discharge, including the receipt of mechanical ventilation, vaso-pressors, or continuous kidney replacement therapy. All serious and nonserious treatment-emergent adverse events were captured and reported using the MeDRA 27.0 classification system. All data were uploaded to the electronic case report form system created by the Hellenic Institute for the Study of Sepsis trial sponsor (HISS; https://sepsisonlinesepsis.gr/W_Home) and monitored by blinded and unblinded monitors. Biospecimens were also obtained on days 2, 4, 7, 15, 21, and 28 for measurements of ferritin and HLA-DR on CD45/CD14-monocytes.

End Points

The primary end point was the proportion of patients in each group who attained a 1.4-point or more decrease in the mean SOFA score by day 9. The mean SOFA change was calculated after subtracting the mean value of SOFA scores between days 2 and 9 from the baseline SOFA score on day 1. The measurement window of 9 days for the assessment of the end point was based on previous data from the Efficacy of Volume Substitution and Insulin Therapy in Severe Sepsis (VISEP)⁹ and MAXSEP¹⁰ RCTs, which showed that reduction of the mean

Table 1. Baseline Characteristics of Participants in the ImmunoSep Trial

	No. (%) of patients	
	Precision immunotherapy (n = 131)	Placebo (n = 145)
Male	89 (67.9)	94 (64.8)
Female	42 (32.1)	51 (35.2)
Age, mean (SD), y	69 (13)	70 (14)
Race and ethnicity ^a		
African	1 (0.8)	0
Middle Eastern	1 (0.8)	0
White	129 (98.5)	145 (100.0)
Medical history		
Diabetes	45 (34.4)	48 (33.1)
COVID-19 ^b	34 (26.0)	35 (24.1)
Chronic obstructive pulmonary disease	26 (19.8)	39 (26.9)
Coronary heart disease	25 (19.1)	34 (23.4)
Atrial fibrillation	23 (17.6)	36 (24.8)
Heart failure	19 (14.5)	22 (15.2)
Cerebral stroke	18 (13.7)	18 (12.4)
Chronic kidney disease	13 (9.9)	16 (11.0)
Solid tumor malignancy without metastasis	12 (9.2)	9 (6.2)
Cerebral hemorrhage	9 (6.9)	9 (6.2)
Interventions		
Mechanical ventilation	116 (88.5)	129 (89.0)
Continuous venovenous hemofiltration	23 (17.6)	24 (16.6)
Underlying infection		
Ventilator-associated pneumonia	48 (36.6)	54 (37.2)
Hospital-acquired pneumonia	35 (26.7)	32 (22.1)
Community-acquired pneumonia	31 (23.7)	44 (30.3)
Primary bacteremia	16 (12.2)	12 (8.3)
Pneumonia by SARS-CoV-2	1 (0.8)	3 (2.1)
APACHE II score ^c		
Median (IQR)	20 (16-25)	19 (14-25)
≥25	38 (29.0)	40 (27.6)
<25	93 (71.0)	105 (72.4)
Charlson Comorbidity Index ^d		
Median (IQR)	4 (3-6)	5 (3-6)
≥5	54 (41.2)	76 (52.4)
<5	77 (58.8)	69 (47.6)
SOFA score ^e		
Median (IQR)	10 (7-12)	9 (6-11)
≥10	67 (51.1)	68 (46.9)
<10	64 (48.9)	77 (53.1)
Po ₂ :FiO ₂ ratio, median (IQR)	3 (2-3)	3 (2-3)
Platelets, median (IQR)	0 (0-1)	0 (0-1)
Glasgow coma scale, median (IQR)	1 (0-3)	1 (0-3)
Cardiovascular, median (IQR)	4 (3-4)	4 (3-4)
Bilirubin, median (IQR)	0 (0-1)	0 (0-0)
Creatinine or urine output, median (IQR)	0.5 (0-2)	0 (0-1)
Baseline laboratory		
Macrophage activation-like syndrome	25 (19.1)	23 (15.9)
Sepsis-induced immunoparalysis	106 (80.9)	122 (84.1)
Ferritin, median (IQR), ng/mL (reference: 30-350 ng/mL for men, 20-250 ng/mL for women)		
Macrophage activation-like syndrome	5438 (5000-7009)	6221 (5000-7500)
Sepsis-induced immunoparalysis	633 (353-1149)	703 (340-1488)

(continued)

Table 1. Baseline Characteristics of Participants in the ImmunoSep Trial (continued)

	No. (%) of patients	
	Precision immunotherapy (n = 131)	Placebo (n = 145)
mHLA-DR, median (IQR), antibodies per CD45/14 cell (reference: 15 000-40 000 Abs per CD-14 cell)		
Macrophage activation-like syndrome	3576 (2124-6205)	4990 (3068-7811)
Immunoparalysis	3043 (2297-4135)	3078 (1762-4004)
Total white blood cell count, mean (SD), per μ L (reference: 4000-11 000 per μ L)	17 003.8 (9075)	16 904.6 (9582.3)
Hemoglobin, mean (SD), g/dL (reference: 13.5-17.5 g/dL for men, 12.0-15.5 g/dL for women)	10.39 (6.88)	9.83 (1.85)
Platelets, mean (SD), $\times 10^3/\mu$ L (reference: 150 000-400 000 $\times 10^3/\mu$ L)	201.4 (147.1)	209.5 (138.4)
International normalized ratio, median (IQR) (reference: 0.8-1.2)	1.2 (1.1-1.4)	1.2 (1.0-1.4)
Fibrinogen, mean (SD), mg/dL (reference: 200-400 mg/dL)	582.5 (231.5)	629.7 (924.7)
D-dimer, median (IQR), μ g/dL (reference: <0.5 μ g/dL)	14.3 (0.8-2301.0)	16.3 (0.7-2001.1)
Creatinine, median (IQR), mg/dL (reference: 0.7-1.2 mg/dL)	1.3 (0.8-2.0)	1.0 (0.7-1.8)
Aspartate aminotransferase, median (IQR), U/L (reference: 10-40 U/L)	41 (27-101)	44 (21-92)
Alanine aminotransaminase, median (IQR), U/L (reference: 7-56 U/L)	36 (20-77)	35 (17-75)
Total bilirubin, median (IQR), mg/dL (reference: 0.3-1.2 mg/dL)	0.7 (0.4-1.3)	0.6 (0.4-1.1)
$\text{PaO}_2:\text{FiO}_2$ ratio, median (IQR), mm Hg (reference: >400 mm Hg)	161.6 (127.0-247.0)	171.9 (127.8-232.8)
Lactate, median (IQR), mg/dL (reference: 4.5-19.8 mg/dL)	15.3 (9.9-21.6)	15.3 (9.2-28.4)
C-reactive protein, median (IQR), mg/dL (reference: <5 mg/dL)	2.7 (1.4-19.0)	3.3 (1.5-13.9)
Procalcitonin, median (IQR), ng/mL (reference: <0.03 ng/mL)	1.3 (0.5-5.9)	1.3 (0.5-11.5)
Time between sepsis onset and initiation of the study drug, median (IQR), h	36.0 (24.0-48.0)	47.0 (24.0-48.0)

Abbreviations: APACHE, Acute Physiology and Chronic Health Evaluation; mHLA-DR, monocytic human leukocyte antigen DR; $\text{PaO}_2:\text{FiO}_2$, partial pressure of arterial oxygen to fraction of inspired oxygen; SOFA, Sequential Organ Failure Assessment.

SI conversion factors: To convert alanine aminotransferase and aspartate aminotransferase from U/L to pkat/L, multiply by 0.0167; bilirubin, from mg/dL to μ mol/L, multiply by 17.104; creatinine from μ g/dL to μ mol/L, multiply by 88.4; lactate from mg/dL to mmol/L, multiply by 0.111.

^a Self-reported from a list of options.

^b Refers to history of resolved COVID-19 infection.

^c The APACHE II is calculated using a weighted sum of 12 physiological variables (such as temperature, blood pressure, heart rate, respiratory rate, oxygenation, arterial pH, electrolytes and hematocrit levels, white blood cell

count, and Glasgow Coma Scale), along with age and chronic health conditions. The score ranges from 0 to 71, with higher values indicating more severe illness and a higher risk of mortality.

^d The Charlson's Comorbidity Index is determined by assigning weighted scores (1-6) to 19 predefined comorbid conditions based on their mortality risk. Scores range from 0 to 37, with higher scores representing greater comorbidity burden and increased risk of mortality.

^e The SOFA score is calculated by assigning 0-4 points for the degree of dysfunction in the respiratory, cardiovascular, hepatic, coagulation, renal, neurological organ systems, based on specific clinical and laboratory criteria. Total score ranges from 0 to 24. Higher scores indicate more severe organ dysfunction and higher mortality risk.

SOFA score by 1.4 points was associated with reduced 28-day mortality.

There were 6 prespecified secondary end points. These included, (1) the attainment of a 1.4-point or more decrease of the mean SOFA score by day 9 in patients with either macrophage activation-like syndrome or sepsis-induced immunoparalysis, (2) 28-day mortality, (3) 90-day mortality, (4) attainment of at least a 1.4-point decrease of the mean SOFA score by day 15, (5) the reversal of sepsis-induced immune dysfunction (SIDF), and (6) the infection resolution by day 15. For patients with macrophage activation-like syndrome, the reversal of SIDF was a decrease of baseline ferritin by at least 15%; for patients with sepsis-induced immunoparalysis, an increase of the absolute number of HLA-DR receptors on CD45/CD14 monocytes of more than 8000 per cell. On day 15, the state of infection was classified as resolved, intermediate, treatment failure, or superinfection by the site investigators using predefined criteria (eTable 3 in *Supplement 2*).

One sensitivity analysis of the primary end point was done excluding patients who were discharged earlier than day 9. Exploratory post hoc analyses included (1) the time to reversal of SIDF, (2) baseline APACHE II score, (3) baseline CCI, and

(4) baseline SOFA score subgroups based on the likelihood of their predicting 28-day mortality. The cutoff points of these severity scores were defined by the Youden index of the respective receiver operator characteristics curves. Significant interactions were defined as $P < .05$.

Sample Size Calculation

The study was powered for the primary end point anticipated to be attained in 40% in the intervention group and in 20% in the control group. At the significance level of 5% and a power of 90%, the calculation yielded 117 patients per trial group. Considering a dropout rate of about 15%, 280 were needed.⁶

Statistical Analyses

The primary analysis included all patients randomized, who did not withdraw their consent or requested removal of all data. For patients who died before day 9, all values from the death day until day 15 were replaced by 24 (the maximum SOFA score). For patients discharged home or lost to follow-up before day 9, the last recorded SOFA score was carried forward. The SOFA score was parameterized as a continuous variable for all analyses.

Table 2. Primary and Main Secondary Outcomes of the ImmunoSep Trial

End points	No./total (%) of patients		Difference, % (95% CI)	OR unadjusted (95% CI)	P value
	Precision immunotherapy	Placebo			
Primary end point					
≥1.4-Point decrease of mean SOFA score d 2 to 9 ^a	46/131 (35.1)	26/145 (17.9)	17.2 (6.8 to 27.2)	2.48 (1.42 to 4.32)	.002
Main secondary outcomes					
28-d Mortality	57/131 (43.5)	72/145 (49.7)	6.1 (-5.6 to 17.6)	0.78 (0.49 to 1.26)	.34
90-d Mortality	90/131 (68.7)	98/145 (67.6)	1.1 (-9.8 to 11.9)	1.05 (0.63 to 1.75)	.90
≥1.4-Point decrease of mean SOFA score d 2 to 15 ^a	52/131 (39.7)	34/145 (23.4)	16.3 (5.3 to 26.8)	2.15 (1.28 to 3.61)	.004
Reversal of sepsis-induced immune dysfunction ^b	46/59 (78.0)	32/66 (48.5)	29.5 (12.6 to 44.0)	3.76 (1.72 to 8.22)	.001
Assessment of infection by d 15					
Resolution	58/131 (44.3)	46/145 (31.7)			
Intermediate	11/131 (8.4)	9/145 (6.2)			
Failure	32/131 (24.4)	44/145 (30.3)	NA	0.59 (0.38 to 0.91) ^c	.02
Superinfection	30/131 (22.9)	46/145 (31.7)			

Abbreviations: OR, odds ratio; NA, not applicable; SOFA, Sequential Organ Failure Assessment.

^aThe SOFA score is calculated by assigning 0 to 4 points for the degree of dysfunction of the respiratory, cardiovascular, hepatic, coagulation, kidney, and neurological organ systems, based on specific clinical and laboratory criteria. Total score ranges from 0 to 24. Higher scores indicate more severe organ dysfunction and higher mortality risk.

^bDefined as at least a 15% decrease of ferritin for patients with macrophage activation-like syndrome remaining decreased over follow-up time blood

draws; and increase of the absolute number of human leukocyte antigen DR receptors to more than 8000 per CD45/CD14 monocyte remaining higher than these values over follow-up time blood draws. Patients not having at least 2 serial blood draws to confirm the permanence of the changes were considered not to have attained reversal of sepsis-induced organ dysfunction.

^cUsing predefined criteria (eTable 3 in *Supplement 2*) any infection at day 15 was characterized as resolved, intermediate, treatment failure, or superinfection. The OR expresses the risk of worse outcome of the precision immunotherapy group vs the placebo group.

Comparisons for primary and all secondary end points were calculated by the Fisher exact test; the odds ratios (ORs) and 95% CIs by the Mantel-Haenszel test. A confirmatory model for the primary end point included a stepwise logistic regression for the proportion who achieved at least a 1.4-point decrease in SOFA score by day 9 as the dependent variable, with the blinded intervention, baseline CCI, SOFA score, and study site as the independent variables. Kaplan-Meier and Cox regression analyses reporting hazard ratios (HRs) with 95% CIs were used to illustrate survival. The time to reversal of SIDF was compared by Cox regression analysis. Comparisons of the state of infection by day 15—defined as resolved, intermediate, treatment failure, or superinfection—were conducted by ordinal regression analysis. $P < .05$ was considered statistically significant. Analysis was conducted using SPSS version 29.0.2.0 (IBM). All analyses followed those defined in the statistical analysis plan (*Supplement 1*).

Results

Patients

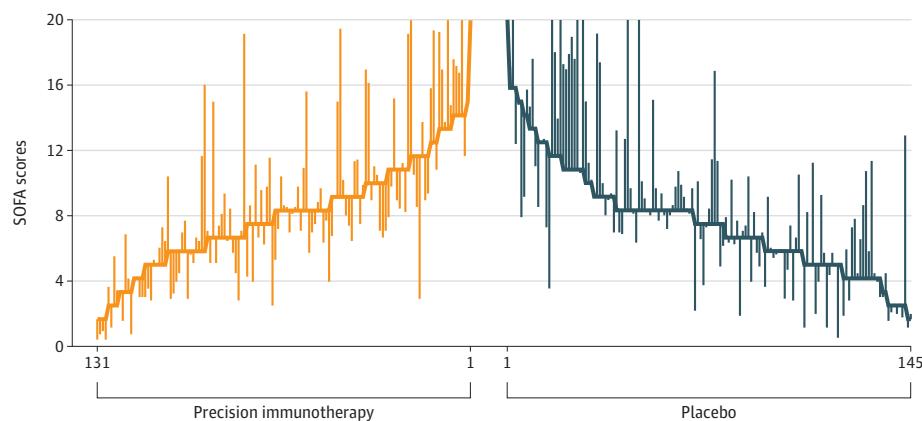
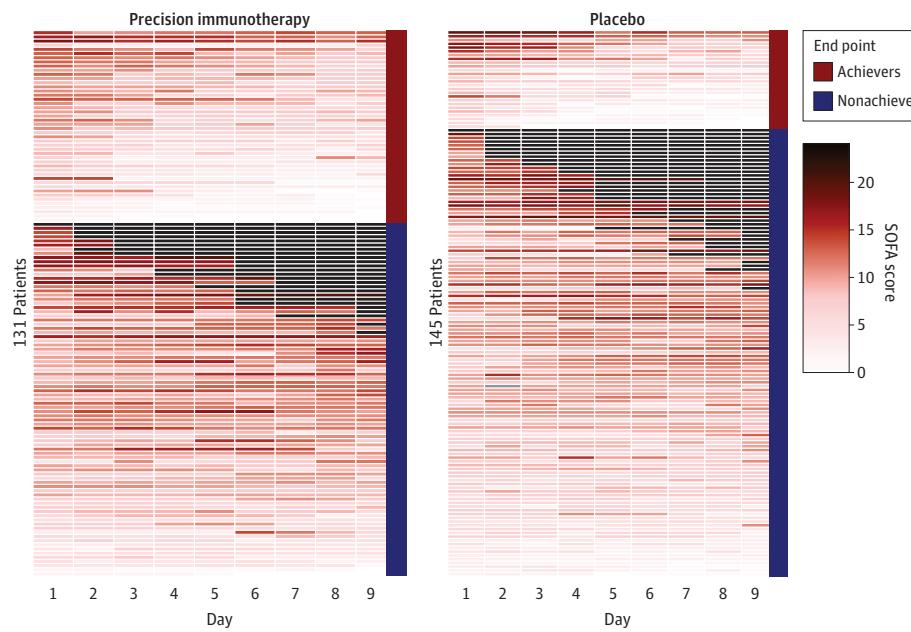
Among 672 patients screened for eligibility, 281 patients were randomized. The first patient was enrolled on August 5, 2021, and the follow-up of the last patient was completed on April 29, 2024. Five patients withdrew consent and requested removal of all their data, 4 in the precision immunotherapy group and 1 in the placebo group, leaving 276 patients (mean [SD] age, 70 [13] years; 93 females [33.7%]) in the primary analysis

population (Figure 1). Baseline characteristics were comparable between groups (Table 1; eTables 5 and 6 in *Supplement 2*). Forty-eight patients (17.4%) were classified with macrophage activation-like syndrome (25 allocated to anakinra treatment and 23 to placebo) and 228 patients (82.6%) with sepsis-induced immunoparalysis (106 allocated to recombinant human interferon gamma treatment and 122 allocated to placebo; eTable 7 in *Supplement 2*). In the precision immunotherapy group, 21 patients (16.0%) died before day 9 compared with 29 (20.0%) in the control group. Cumulatively, 8.3% of patients had SOFA scores carried forward because of ICU discharge earlier than day 9 (eTable 8 in *Supplement 2*).

Primary End Point

The primary end point (≥1.4-point decrease of mean SOFA score by day 9) was present in 46 of 131 patients (35.1%) in the precision immunotherapy group and 26 of 145 patients (17.9%) in the placebo group ($P = .002$; **Table 2** and **Figure 2A**). In a confirmatory model adjusted for baseline illness severity, the precision strategy was associated with greater odds of attaining the prespecified decrease in the SOFA score by day 9 (adjusted OR, 2.49; 95% CI, 1.42-4.36; eTable 9 in *Supplement 2*). To illustrate the change in SOFA score after randomization, heat maps reveal the total SOFA score was lower among the precision immunotherapy group than among the placebo group (Figure 2B and eFigure 1 in *Supplement 2*). A sensitivity analysis excluding patients who were discharged prior to day 9 was consistent with the primary analysis (eFigure 2 in *Supplement 2*).

Figure 2. Primary End Point

A Change from the baseline SOFA score**B** Heat map distribution of SOFA scores

A. The parallel line plot represents change for each participant in the trial, from the baseline Sequential Organ Failure Assessment (SOFA) score measured before initiating the study drug to the mean SOFA score for days 2 through 9. Each single line represents this change for each participant and is ordered by baseline SOFA score.

B. The SOFA scores are presented for each treatment group over the 9 days of follow-up. Treatment started on day 1 (baseline). Each group is separated into achievers and nonachievers of the primary end point. Achievers were defined as patients reaching at least a 1.4-point decrease of the mean SOFA score between days 2 and 9 from the baseline SOFA score.

The SOFA score is calculated by assigning 0 to 4 points for the degree of dysfunction in respiratory, cardiovascular, hepatic, coagulation, kidney, and neurological organ systems, based on specific clinical and laboratory criteria. Total score ranges from 0 to 24. Higher scores indicate more severe organ dysfunction and higher mortality risk.

Secondary End Points

There were 6 prespecified secondary end points. First, the decrease of mean SOFA score of 1.4-points or more by day 9 was attained in both states of immune dysregulation (eTable 10 in *Supplement 2*). In patients with macrophage activation-like syndrome, the decrease of mean SOFA score by day 9 was attained by 12 of 25 patients (48.0%) in the precision immunotherapy group and 4 of 23 patients (17.4%) in the placebo group ($P = .04$). Similarly, among patients with sepsis-induced immunoparalysis, 34 of 106 patients (32.1%) in the precision immunotherapy group and 22 of 122 (18.0) in the placebo group yielded the prespecified reduction of mean SOFA score by day 9 ($P = .02$). Second, 28-day mortality occurred in 57 of 131 patients (43.5%) in the precision immunotherapy group and 72 of 145 patients in the placebo group (49.7%), a difference that did not reach statistical significance ($P = .34$, Table 2; eFigure 3 in *Supplement 2*). Third, no differences in 90-day mor-

tality were found (Table 2; eFigure 4 in *Supplement 2*). Fourth, the attainment of at least a 1.4-point decrease of the mean SOFA score by day 15 was greater in the group of precision immunotherapy (39.7%; 51 of 131) than in the placebo group (23.4%; 34 of 145; $P = .004$). Fifth, in a subset of 125 patients, the reversal of SIDF by day 28 was found in 46 of 59 patients (78.0%) of the precision immunotherapy group and in 32 of 66 patients (48.5%) of the placebo group ($P < .001$, Table 2). Sixth, in ordinal regression models, the odds for a worse infection outcome by day 15 were significantly lower in the precision immunotherapy group (OR, 0.59; 95%CI, 0.38-0.91; $P = .02$) than in the placebo group (eFigure 5 in *Supplement 2*).

Post Hoc and Subgroup Analyses

The time to reversal of SIDF was significantly shorter in the precision immunotherapy group (eFigure 6 in *Supplement 2*). Interaction tests showed statistical significance of

Table 3. Most Common Serious Treatment-Emergent Adverse Events by System-Organ Class and Preferred Term, Classified by Relationship to Study Drug

Category	Probably or possibly related, No. (%)		Probably not related or unrelated, No. (%)	
	Precision immunotherapy (n = 131)	Placebo (n = 145)	Precision immunotherapy (n = 131)	Placebo (n = 145)
Any serious adverse event ^{a,b}	8 (6.1)	3 (2.1)	108 (82.4)	126 (86.9)
Blood and lymphatic system disorders ^b				
Anemia	0	0	47 (35.9)	38 (26.2)
Thrombocytopenia	5 (3.8)	1 (0.7)	9 (6.9)	8 (5.5)
Cardiac disorders ^b				
Atrial fibrillation	0	0	19 (14.5)	14 (9.7)
Bradycardia	0	0	6 (4.6)	6 (4.1)
Cardiac arrest	0	1 (0.7)	9 (6.9)	15 (10.3)
General disorders and administration site conditions ^b				
Multiple organ dysfunction syndrome ^c	2 (1.5)	0	51 (38.9)	58 (40.0)
Infections and infestations ^b				
Bacteraemia	0	0	36 (27.5)	27 (18.6)
Device-related infection	0	0	2 (1.5)	4 (2.8)
Fungaemia	0	0	17 (13.0)	12 (8.3)
Fungal infection	0	0	7 (5.3)	5 (3.4)
Pneumonia	0	0	35 (26.7)	32 (22.1)
Sepsis ^d	0	0	9 (6.9)	12 (8.3)
Septic shock ^d	0	0	29 (22.1)	17 (11.7)
Urinary tract infection	0	0	4 (3.1)	7 (4.8)
Kidney and urinary disorders ^b				
Acute kidney injury	0	0	21 (16.0)	19 (13.1)
Respiratory, thoracic, and mediastinal disorders ^b				
Bronchial obstruction	0	0	3 (2.3)	3 (2.1)
Hypoxia	0	0	4 (3.1)	7 (4.8)
Pleural effusion	0	0	5 (3.8)	3 (2.1)
Pneumothorax	0	0	10 (7.6)	7 (4.8)
Respiratory failure	0	0	2 (1.5)	7 (4.8)
Tracheal stenosis	0	0	7 (5.3)	8 (5.5)
Vascular disorders ^b				
Hemorrhage	0	0	6 (4.6)	0

interaction of the precision immunotherapy group with a CCI of 5 or higher and a SOFA score 10 or higher for the achievement of primary end point (eFigures 7 in *Supplement 2*); and of the precision immunotherapy group with a CCI of 5 or higher and a SOFA score 10 or higher for the decrease of 28-day mortality (eFigure 8 in *Supplement 2*). No significant interactions were found between any subgroup and precision immunotherapy for an effect on 90-day mortality (eFigure 9 in *Supplement 2*).

Adverse Events

A total of 1069 serious treatment-emergent adverse events were reported in 245 patients (88.8%); 13 were serious adverse reactions probably (n = 3) or possibly (n = 10) related to the study drug; 2 serious adverse reactions were reported in the anakinra group, 7 in the recombinant human interferon gamma

group, and 4 in the placebo group. Six serious adverse reactions were considered suspected unexpected serious adverse reactions for 5 patients; 3 patients (2.5%) received placebo and 2 patients (1.9%) recombinant human interferon gamma (eTable 12 in *Supplement 2*). Two significant differences were found in the incidence of serious treatment-emergent adverse events: increase of anemia in the anakinra group; and hemorrhage in the recombinant human interferon gamma group. Notably, 4 of the hemorrhagic cases in the recombinant human interferon gamma group occurred in patients with thrombocytopenia; however, no differences in the incidence of thrombocytopenia were found between the 2 groups (Table 3; eTable 13 in *Supplement 2*).

Comparisons of the nonserious treatment-emergent adverse events indicated decreased incidence of bradycardia in the recombinant human interferon gamma group; decreased signs

^a The serious treatment-emergent adverse events were classified by relationship to the study drug using Medra 27.0 for events captured during the 90 days of follow-up for the enrolled 276 patients. In the analysis, these adverse events were graded as *probably related*, which signifies strong time relationship to the drug or relapse if reintroduced, and another etiology is improbable or clearly less probable; *possibly related*, strong time relationship to the drug and an alternative etiology is as probable or less probable; *probably not related*, slight or no time relationship to the drug and/or a more probable alternative etiology; and *unrelated*, causality by an underlying or concomitant disease or another pharmaceutical product, with no time relationship and a much more probable alternative etiology. Site investigator graded treatment-emergent adverse events.

^b These data represent the number of patients with at least 1 event, whereas all other data in the table correspond to the number of events.

^c Represents the progression to Multiple organ dysfunction syndrome of the initial sepsis or septic shock for which the patient was enrolled in the study.

^d Represents a new episode of sepsis or septic shock after the resolution of the initial sepsis or septic shock for which the patient was enrolled in the study.

of ileus in the anakinra group; and more creatinine increase, fibrinogen decrease, and γ -glutamyl transferase increase in the recombinant human interferon gamma group (eTable 14 in *Supplement 2*). The number of patients needed to treat and to harm were calculated (eTable 15 in *Supplement 2*).

Discussion

In this multicenter RCT, a precision immunotherapy strategy targeting macrophage activation-like syndrome and sepsis-induced immunoparalysis improved organ dysfunction by day 9 compared with placebo. Although no significant effect was found for mortality, significant differences were found for the other secondary end points including overall improvement of organ dysfunction by day 15, reversal of SIDF, and resolution of the underlying infection. No safety concerns were raised.

These findings are consistent with previous work targeting immune dysregulation in sepsis. First, precision immunotherapy for severe infections has been increasingly considered since the COVID-19 pandemic: a strategy of anakinra treatment guided by soluble urokinase plasminogen activator receptor concentrations as a biomarker of lung hyperinflammation has been registered by the European Medicines Agency and for Emergency Use Authorization by the US Food and Drug Administration for treatment of patients with COVID-19 with pneumonia.¹¹ Second, in the 1990s, continuous IV anakinra infusion was studied in 2 large-scale phase 3 RCTs that failed to demonstrate survival benefit in sepsis.^{12,13} Interestingly, however, a post hoc reanalysis of the first trial based on hepatobiliary dysfunction and disseminated intravascular coagulation as criteria for macrophage activation-like syndrome showed that patients treated with anakinra had a significantly lower 28-day mortality than patients treated with placebo (35% vs 65%).¹⁴

Similarly, recombinant human interferon gamma has been proposed as a treatment for sepsis due to its immunostimulatory mechanisms. Recombinant human interferon gamma restored the low HLA-DR levels on blood monocytes of healthy volunteers who had been previously exposed to experimental endotoxemia.¹⁵ In a single-arm open-label trial, recombinant human interferon gamma administered subcutaneously to 9 patients with septic shock and low monocytic HLA-DR levels rapidly increased HLA-DR levels and the capacity of circulating monocytes to produce tumor necrosis factor α .⁵ Favorable clinical responses with reciprocal increases of monocytic HLA-DR expression are also reported in a small-series study of patients with invasive *Candida* or *Aspergillus* infections.¹⁶ Notably, our

current findings differ from a recent multicenter study of the prevention of health care-associated pneumonia with recombinant human interferon gamma vs placebo in critically ill patients.¹⁷ This prevention study was prematurely stopped due to an increased incidence of pneumonia with recombinant human interferon gamma treatment (34.5% vs 15.1%, respectively), although 28-day mortality was lower among patients receiving recombinant human interferon gamma than among those receiving placebo (12.7% vs 17%, respectively). However, enrollment was not limited to patients with sepsis-induced immunoparalysis.

The current work demonstrates an improvement in organ function measured by the SOFA score. This end point may be considered a surrogate marker of improved outcome and has been used by others to demonstrate the efficacy of drugs such as nangibotide¹⁸ and adrecizumab.¹⁹

The majority of screened patients were excluded from the current trial because their immune state was unclassified. This suggests that ferritin and HLR-DR levels, alone, do not capture the full extent of immune dysregulation. Others report that latent classes may be present or that a profile of dysregulation could be derived using machine learning and multiomic approaches.²⁰⁻²² A comparison of how these immunotypes, individually or combined, can be used for enrichment in clinical trials of immunotherapy agents is an urgent priority.

Limitations

This trial has several limitations. First, the primary end point of a change in SOFA score is a surrogate outcome, so future adequately powered trials should test these strategies on patient-centered outcomes. Second, the measurement of ferritin and HLA-DR measures may not be feasible across hospitals, and the complexity and assay time could limit scaling and implementation. It is notable that average time from sepsis onset to when the study drug was first administered was 36 to 48 hours. Third, the stability of macrophage activation-like syndrome and sepsis-induced immunoparalysis subtypes over time deserves further study. Fourth, the study findings are generalizable only to patients with sepsis with pneumonia or bacteremia, and other etiologies of sepsis were excluded.

Conclusions

Among patients with sepsis, a precision immunotherapy strategy targeting macrophage activation-like syndrome and sepsis-induced immunoparalysis improved organ dysfunction by day 9 compared with placebo.

ARTICLE INFORMATION

Accepted for Publication: November 18, 2025.

Published Online: December 8, 2025.

doi:10.1001/jama.2025.24175

Author Affiliations: Fourth Department of Internal Medicine, National and Kapodistrian University of Athens, Medical School, Greece (Giambarellos-Bourboulis, Kotsaki, Alevizou, Ktena, Dakou, Antonakos); Department of Anesthesiology and Intensive Care, AHEPA Thessaloniki University General Hospital, Thessaloniki, Greece (Efthymiou, Gkeka); Polyvalent Intensive Care Unit, Sotiria Athens General Hospital of Chest Diseases, Athens, Greece (Koutsoukou, Ntaganou); Department of

Sepsis, Athens, Greece (Giambarellos-Bourboulis, Kotsaki, Alevizou, Ktena, Dakou, Antonakos); Intensive Care Unit, Ippokrateion General Hospital of Thessaloniki, Thessaloniki, Greece (Kotsamidi, Massa, Mouloudi); Department of Anesthesiology and Intensive Care, Jena University Hospital, Jena, Germany (Ehler, Bauer); Intensive Care Unit, Asklepieion General Hospital, Voula, Greece (Paridou, Ioakeimidiou); Second Department of Critical Care Medicine, National and Kapodistrian University of Athens, Greece (Frantzeskaki, Tsangaris); Department of Intensive Care Medicine, Amsterdam Medical University Center, Amsterdam, the Netherlands (Müller, Vlaar); Intensive Care Unit, Radboud University Medical Center, Nijmegen, the

Netherlands (Pickkers); Infectious Diseases Service, Department of Medicine, Lausanne University Hospital, and University of Lausanne, Lausanne, Switzerland (Meylan); Intensive Care Unit of Center for Respiratory Failure, Sotiria Chest Diseases Athens General Hospital, Greece (Nikolopoulos); Department of Infectious Diseases, Clinical Hospital for Infectious Diseases Cluj-Napoca, Romania (Lupse); Clinic of Intensive Care and Pulmonary Diseases, Faculty of Nursing, School of Health Sciences, National and Kapodistrian University of Athens, Athens, Greece (Gavala, Myrianthefs); Intensive Care Unit, Aghios Dimitrios General Hospital, Thessaloniki, Greece (Vlachogianni); Intensive Care Unit, Korgialeneio-Benakeio General Hospital, Athens, Greece (Solomonidi, Patrani); Department of Intensive Care Medicine, University Hospital of Heraklion, University of Crete, Heraklion, Greece (Kondili); Intensive Care Unit, Georgios Gennimatas General Hospital, Thessaloniki, Greece (Antoniadou, Karapanagiotou); Department of Critical Care Medicine, Democritus University of Thrace, Alexandroupolis, Greece (Nakou, Papaioannou); Intensive Care Unit of Latseion Burn Center, Thriasio General Hospital, Eleusis, Greece (Markou); Second Department of Propedeutic Medicine, National and Kapodistrian University of Athens, Medical School, Greece (Hatzigelaki); Intensive Care Unit, Tzaneio General Hospital, Piraeus, Greece (Prekates); Intensive Care Unit, Koutlibaneion and Triantafylleion General Hospital, Larissa, Greece (Komnos); Department of Internal Medicine and Radboud Center for Infectious Diseases, Radboud University Medical Center, Nijmegen, the Netherlands (Bakkerus, van der Meer, Netea); Center of Infection and Molecular Medicine, Division of Infectious Diseases, Amsterdam Medical University Center, University of Amsterdam, Amsterdam, the Netherlands (Slim, van der Poll, Wiersinga); Department of Medicine and Research Laboratory of Internal Medicine, Expertise Center of Greece in Autoimmune Liver Diseases, European Reference Network on Hepatological Diseases (ERN RARE-LIVER), General University Hospital of Larissa, Larissa, Greece (Dalekos); Dipartimento di Scienze Biotecnologiche di Base, Cliniche Intensivologiche e Perioperatorie, Università Cattolica del Sacro Cuore, Rome, Italy (De Pascale, Antonelli); Dipartimento di Scienze dell'Emergenza, Anestesiologiche e della Rianimazione, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy (De Pascale, Antonelli); Department of Critical Care Medicine, University of Ioannina, Ioannina, Greece (Koulouras); Intensive Care Unit, 424 Military General Hospital, Thessaloniki, Greece (Pavzanti); Intensive Care Unit, Theageneion Anticancer Hospital, Thessaloniki, Greece (Anisoglou); Service of Immunology and Allergy, Center Human Immunology Lausanne, Department of Medicine, Department of Laboratory Medicine and Pathology, Lausanne University Hospital, University of Lausanne, Lausanne Switzerland (Calandra); Intensive Care Unit, KAT General Hospital, Kifissia, Greece (Alamanos); Department of Immunology and Metabolism, Life and Medical Sciences Institute, University of Bonn, Bonn, Germany (Netea).

Author Contributions: Dr Giamarellos-Bourboulis had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Concept and design: Giamarellos-Bourboulis, Efthymiou, Pickkers, Solomonidi, Kondili, Antoniadou, Komnos, Bakkerus, De Pascale, Massa, Vlaar, van der Meer, van der Poll, Wiersinga, Bauer, Netea.

Acquisition, analysis, or interpretation of data: Giamarellos-Bourboulis, Kotsaki, Kotsamidi, Koutsoukou, Ehler, Paridou, Frantzeskaki, Müller, Pickkers, Meylan, Nikolopoulos, Lupse, Gavala, Vlachogianni, Alevizou, Antoniadou, Nakou, Markou, Hatzigelaki, Prekates, Bakkerus, Slim, Dalekos, Karapanagiotou, Ktena, Koulouras, Psarrakis, Dakou, Pazvanti, loakeimidou, Tsangaris, Antonakos, Vlaar, Anisoglou, Calandra, Papaioannou, Myrianthefs, Patrani, Alamanos, Antonelli, van der Poll, Ntaganou, Gkeka, Bauer, Mouloudi, Netea.

Drafting of the manuscript: Giamarellos-Bourboulis, Kotsaki, Paridou, Frantzeskaki, Vlachogianni, Solomonidi, Markou, Bakkerus, Psarrakis, Massa, Myrianthefs, Patrani, van der Poll.

Critical review of the manuscript for important intellectual content: Kotsaki, Kotsamidi, Efthymiou, Koutsoukou, Ehler, Müller, Pickkers, Meylan, Nikolopoulos, Lupse, Gavala, Vlachogianni, Solomonidi, Alevizou, Kondili, Antoniadou, Nakou, Markou, Hatzigelaki, Prekates, Komnos, Slim, Dalekos, Karapanagiotou, Ktena, De Pascale, Koulouras, Massa, Dakou, Pazvanti, loakeimidou, Tsangaris, Antonakos, Vlaar, Anisoglou, Calandra, Papaioannou, Myrianthefs, Alamanos, Antonelli, van der Meer, van der Poll, Wiersinga, Ntaganou, Gkeka, Bauer, Mouloudi, Netea.

Statistical analysis: Giamarellos-Bourboulis, Pickkers, Bakkerus, Dakou.

Obtained funding: Pickkers, van der Poll, Bauer, Netea.

Administrative, technical, or material support: Giamarellos-Bourboulis, Ehler, Paridou, Frantzeskaki, Pickkers, Meylan, Lupse, Bakkerus, Karapanagiotou, De Pascale, Dakou, Papaioannou, van der Poll, Wiersinga, Bauer, Netea.

Supervision: Giamarellos-Bourboulis, Müller, Pickkers, Meylan, Solomonidi, De Pascale, Koulouras, Vlaar, Calandra, Papaioannou, Myrianthefs, Antonelli, van der Meer, van der Poll, Wiersinga, Bauer, Netea.

Conflict of Interest Disclosures:

Dr Giamarellos-Bourboulis reported receiving personal fees from Abbott Products Operations, BioMerieux, Brahms GmbH, Swedish Orphan BioVitrum, AbbVie, UCB, Sanofi, and Novartis to the National and Kapodistrian University of Athens and grants from Abbott Products Operations, Swedish Orphan BioVitrum, and Horizon Health grants ImmunoSep, Epic Crown-2, Risk in COVID, POINT, and Homi-Lung studies to the Hellenic Institute outside the submitted work. Dr Ehler reported receiving personal fees from B Braun Melsungen AG for attending study meetings of the GENIUS trial outside the submitted work. Dr Dalekos reported receiving grants from Gilead and Ipsen; and personal fees from Ipsen, Gilead, Genesis, Pfizer, Sanofi, and Sobi; and other from Amyndas Pharmaceuticals, Intercept Pharma, CymaBay Therapeutics, Genkyotex, Novo Nordisk, Pfizer, Regulus Therapeutics, Sobi, and Tiziana Life Sciences outside the submitted work. Dr Vlaar reported receiving European Horizon grants during the conduct of the study and personal fees from InflaRx and CSL Behring to his institution outside the submitted work. Dr Calandra reported serving on the executive committee and council of the

International Sepsis Forum; serving on the data and safety monitoring boards of Basilea, Gilead Sciences, MSD, Moderna, Pfizer, and Shionogi, the European Infection in Leukemia conference organizing committee, and the national advisory board of the Swiss Sepsis Program; and receiving travel and conference participation fees from the International Symposium on Intensive Care and Emergency Medicine outside the submitted work. Dr Antonelli reported receiving personal fees from Menarini and Novartis and grants from Fisher & Paykel and serving on the board of Astra Zeneca outside the submitted work. Dr van der Poll reported receiving grants from the European Horizon commission, serving on the advisory board of Matisse to his institution during the conduct of the study; grants from Ministry of Economic Affairs & Health Holland; and serving on the data and safety monitoring board of the REMAP-CAP study outside the submitted work. Dr Wiersinga reported receiving grants from the Netherlands Organisation for Health Research and Development (ZonMw) and COVID-19-related research and consultancy fees from AstraZeneca, all fees paid to host institution (Amsterdam Medical Center) outside the submitted work. Dr Bauer reported receiving speaker fees from Sobi and personal fees from La Jolla Pharmaceutical Company, SNIPR Biome Denmark, CytoSorbents GmbH, Thermo Fisher Scientific (BRAHMS GmbH), Roche Diagnostics International, Bayer, ArtCline, Baxter, and deepull and nonfinancial support from the International Sepsis Forum outside the submitted work. No other disclosures were reported.

Funding/Support: The ImmunoSep trial has received funding from the European Union Horizon 2020 research and innovation programme under grant agreement 847422. Dr Netea was supported by grant OCENW.XL.23.055 from NWO-XL and by a Spinoza grant of the Netherlands Organization for Scientific Research.

Role of the Funder/Sponsor: The funders had no role in the study design and collection, management, analysis, and interpretation of the data. The sponsor of the study, Hellenic Institute for the Study of Sepsis, was responsible for study design, study conduct, data analysis, data interpretation, and the decision to submit for publication.

Members of the ImmunoSep Study Group:

Hellenic Institute for the Study of Sepsis: Evangelos J. Giamarellos-Bourboulis (principal investigator), Antonia Alevizou, Artemis Bogosian, Vasiliki Bourika, Athanasia Chatzianastasiou, Konstantina Dakou, Effrosyni Dimopoulou, George Dimopoulos, Leda Efstratiou, Eleni Florou, Eleni Katsigianni, Paraskevi Kanellakopoulou, Myrto Kolia, Elli Konstantinou, Antigone Kotsaki, Miltiades Kyprianou, Sofia Ktena, Onisiphoros Neophytou, Melina Nikolakea, Elisavet Olntasi, Dimitra Papatheodosiou, Varvara Perraki, Christina Maria Rimpa, Christina Sideratou, Chrysanthi Sidiropoulou, Emmanuel Sofianopoulos, Emmanouil Stylianakis, Georgios Tavoulareas, and Ioannis Theodorou; **Fourth Department of Internal Medicine, National and Kapodistrian University of Athens, Greece:** Antonios Papadopoulos (principal investigator), Nikolaos Antonakos, Amalia Bolanou, Georgia Damoraki, Theologia Gkavogianni, Konstantina Katrini, Panagiotis Koufarygis, Konstantinos Leventogiannis, Styliani Michalakopoulou, Christos Psarrakis, Asimina Safarika, and Athanasios

Siampanos; Second Department of Propaedeutic Internal Medicine, National and Kapodistrian University of Athens, Greece: Eirifil Hatzigelaki (principal investigator), and Sokratis Katopodis; Second Critical Care Department, National and Kapodistrian University of Athens, Greece: Iraklis Tsagkaris (principal investigator), Apostolos Armananidis, Frantzeska Franteskaki, Eleni Karakike, Evodia Kyriazopoulou, and Michail Rizos; Department of Medicine and Research Laboratory of Internal Medicine, Expertise Center of Greece in Autoimmune Liver Diseases, European Reference Network on Hepatological Diseases (ERN RARE-LIVER), General University Hospital of Larissa, Larissa, Greece: George Dalekos (principal investigator), Nikolaos Gatselis, Sara Georgiadou, Georgios Giannoulis, Vasiliki Lygoura, Georgios Perifanos, Anna Samakidou, Angelos Stefos, Dafni Sveroni, and Georgios Vatidis; Second Intensive Care Unit, KAT Attica General Hospital: Ioannis Alamanos (principal investigator), Petros Boukas, Konstantinos Mantis, Dimitra Markopoulou, and Eleni Papadaki; Department of Critical Care Medicine, University of Ioannina, Ioannina, Greece: Vasileios Koulouras (principal investigator), Eirini Demertzis, Konstantina Dimou, Eirini Ioannou, Dimitrios Kantas, Nikolaos Lagos, Georgios Papathanakos, Athanasios Papathanasiou, Maria Paschali, Sofia Peristeri, Maria Saranti, Nikoletta Stefanatou, and Maria Vagia; Department of Critical Care Medicine, Democritus University of Thrace, Alexandroupolis, Greece: Vasilios Papaioannou (principal investigator), Maria Nakou, Konstantina Nikolaou, and Vasiliki Theodorou; Intensive Care Unit, G. Gennimatas General Hospital, Thessaloniki, Greece: Eleni Antoniadou (principal investigator), Elli Antypa, Eleftheria Chassou, Areti Karapanagiotou, Kosmas Kosmidis, Violetta Mourouzidou, Evangelia Panagiotidou, Achilles Pitsoulis, Polychronis Tassioudis, and Maria Vasileiou; Intensive Care Unit, Aghios Dimitrios General Hospital, Thessaloniki, Greece: Glykeria Vlachogianni (principal investigator), Eleni Aimoniou-Georgiou, Eirini Apostolidou, Apostolos Bakas, Eirini Fylaki, Eleni Papandreou, Maria Papathanasiou, and Konstantinos Psaroulis; Department of Anesthesiology and Intensive Care, AHEPA Thessaloniki University General Hospital, Thessaloniki, Greece: Eleni Gkekla (principal investigator), Zoi Adoni, Pinelopi Amoiridou, Vasiliki Birmpa, Aikaterini Efthymiou, Marios Karvouniaris, Konstantinos Pagioulas, Stella-Niki Primikeyi, Ioanna Soultati, Alexandros Tsakalidis, Stella Tsianta, Christos Tsiantas, Iliana Tryfonidou, Nopi Veliki, and Fotini Veroniki; Intensive Care Unit, Ippokrateion General Hospital of Thessaloniki, Thessaloniki, Greece: Eleni Mouloudi (principal investigator), Ioannis Alevroudis, Triantafyllenia Bargiota, Zoi Bosmou, Ioannis Kafantaris, Damianos Karapiperidis, Ioanna Kotsamidi, Eleni Massa, Evangelia Michailidou, Chrysanthi Nakou, Savvas Papadopoulos, Maria Passakiotou, Sotiris Patsatzakis, Panagiotis Roumelis, Styliani Soundoulounaki, and Dafni Stamou; Intensive Care Unit, Theageneion Anticancer Hospital, Thessaloniki, Greece: Souzana Anisoglu (principal investigator), Christos Charatsis, Panagiotis Chaloulis, Periklis Faneromenos, Ali Fanti, Fotis Geroussou, Dimitris Kokaridas, Eirini Papageorgiou, Dimitris Rimarev, and Konstantina Vryza; Intensive Care Unit, Tzaneio General Hospital, Piraeus, Greece: Athanasios Prekates (principal investigator), Konstantinos Athanasiou,

Konstantinos Chatzigeorgiou, Varvara Grammatikopoulou, Athanasios Gravos, Sofia Kanakaki, Konstantina Katsifa, Evangelia Sklavou, Charilaos Tsapas, and Paraskevi Tselioti; Intensive Care Unit, Koutlibaneio & Triantafylleio General Hospital, Larissa, Greece: Apostolos Komnos (principal investigator), Periklis Katsiafylloudis, Dimitrios Papadopoulos, Asimina Valsamaki, and Maria Xanthoudaki; Intensive Care Unit of Latseion Burn Center, Thriasio General Hospital, Eleusis, Greece: Nikolaos Markou (principal investigator), Paraskevi Alexandropoulou, Ioannis Koutsodimitropoulos, Konstantinos Ouranos, Aikaterini Panagiotakopoulou, and Eleni Stefanatou; Polyvalent Intensive Care Unit, Sotiria Athens General Hospital of Chest Diseases, Athens, Greece: Maria Ntaganou (principal investigator), Sofia Avgeri, Lampros Balaskas, Ourania Belagia, Eirini Bourgani, Aikaterini Flevari, Styliani Giannakaki, Georgia Katsagani, Theodora Katsarou, Clementine Bostantzoglou, Olga Kouniaki, Eleni Koutsogianni, Vasiliki Koutsoukou, Despoina-Anastasia Markantonaki, Evgenia Nanou, Maria Poulopou, Konstantina Stathopoulou, Panagiotis Vasileiou, and Efstratia Vrettou; Intensive Care Unit, 424 Military General Hospital, Thessaloniki, Greece: Chrisoula Pasvant (principal investigator), Konstantinos Dimitroulakis, Ioannis Kafantaris, Dimitrios Karapiperis, Ioannis Katsiadramis, and Anna Vemvetsou; Intensive Care Unit, Asklepieion General Hospital, Voula, Greece: Aikaterini Ioakeimidou (principal investigator), Ioannis Giagtzoglou, Yorgos Nikolaidis, Christos Papadas, Alexandra Paridou, and Panagiotis Patsaouras; Intensive Care Unit of Center for Respiratory Failure, Sotiria Chest Diseases Athens General Hospital, Greece: Ioannis Nikolopoulos (principal investigator), Theoni Agapitou, Nikolaos Garmpis, Anastasia Kosmidou, Vasiliki Nanou, Panagiotis Nikolopoulos, Sofia Pouriki, Asimina Raftopoulou, and Giannoula Tsekoura; Intensive Care Unit, Korgialeneio-Benakeio Athens General Hospital, Athens, Greece: Maria Patrani (principal investigator), Georgios Kassianidis, Georgios Marinakis, and Nikolitsa Solomonidi; Department of Intensive Care Medicine, University Hospital of Heraklion, University of Crete, Heraklion, Greece: Eumorfia Kondili (principal investigator), Sofia Kokkini, Eleftherios Papadakis, Georgios Prinianakis, and Athanasia Proklou; Clinic of Intensive Care and Pulmonary Diseases, Faculty of Nursing, School of Health Sciences, National and Kapodistrian University of Athens, Athens, Greece: Pavlos Myriantheis (principal investigator), Alexandra Gavala, Anna Korompeli, Panagiota Makrygianni, and Evangelia Tsigkou; Department of Anesthesiology and Intensive Care, Jena University Hospital, Jena, Germany: Michael Bauer (principal investigator), Sebastian Weis (deputy principal investigator), Frank Bloos, Petra Bloos, Martin Brauer, Karen Dlubatz, Johannes Ehler, Tobias Fischer, Anja Haucke, Carsten Herzog, Michael Hofmann, Karina Knuhr-Kohlberg, Andres Kortgen, Carolin Neumann, Franziska Röstel, Hendrik Rüddel, Katrin Schwope, Anna Seidel, Mark Simon, Helga Skupin, Oliver Sommerfeld, Daniel Thomas-Rüddel, Isabella Westermann, and Johannes Winning; Departments of Intensive Care Medicine and Infectious Diseases, Amsterdam Medical University Center, Amsterdam, the Netherlands: Alexander P.J. Vlaar (principal investigator), Mirjam Dijkstra, Maurice Kroon, Marcella C.A. Müller, Marleen Slim, Rombout

Van Amstel, Tom Van der Poll, Niels Van Mourik, Lonneke Van Vugt, and W. Joost Wiersinga; Intensive Care Unit and Department of Internal Medicine and Infectious Diseases, Radboud University Medical Center, Nijmegen, the Netherlands: Peter Pickkers (principal investigator), Lieke Bakkerus, Aline De Nooijer, Konstantin Fohse, Jacobien Hoogerwerf, Matthijs Kox, Jaap ten Oever, Mihai G. Netea, and Jos W. M. Van der Meer; Department of Infectious Diseases, Clinical Hospital for Infectious Diseases Cluj-Napoca, Romania: Mihaela Lupșe (principal investigator), Mirela Flonta, Lucia Herbel, Zsuzsa Kalmar, and Monica Muntean; Service of Immunology and Allergy, Infectious Diseases and Adult Intensive Care Unit, Lausanne University Hospital, University of Lausanne, Lausanne, Switzerland: Thierry Calandra (principal investigator), Jean-Daniel Chiche, Isabelle Cristiani, Gilda Filipe De Castro Rebelo, Aurelie Fayet, Clemence Ferlay, Charly Gilbert, Anouk Grandjean, Didier Le Roy, Alessia Marino, Sylvain Meylan, Marion Ohl, Thierry Roger, Tia Snaka, Isabelle Sommer, Monica Vieira Da Silva, and Aline Voidey; Dipartimento di Scienze Biotecnologiche di Base, Cliniche Intensivologiche e Perioperatorie, Università Cattolica del Sacro Cuore; Dipartimento di Scienze dell'Emergenza, Anestesiologiche e della Rianimazione, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy: Massimo Antonelli (principal investigator), Alessandro Caroli, Salvatore Lucio Cutuli, Gennaro De Pascale, Valentina Di Gravio, Chiara Iacovelli, Marilena La Sorda, Gianmarco Lombardi, Luca Montini, and Eloisa Sofia Tanzarella.

Data Sharing Statement: See Supplement 4.

REFERENCES

- Singer M, Deutschman CS, Seymour CW, et al. The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA*. 2016;315(8):801-810. doi:[10.1001/jama.2016.0287](https://doi.org/10.1001/jama.2016.0287)
- van de Veerdonk FL, Giamarellos-Bourboulis E, Pickkers P, et al. A guide to immunotherapy for COVID-19. *Nat Med*. 2022;28(1):39-50. doi:[10.1038/s41591-021-01643-9](https://doi.org/10.1038/s41591-021-01643-9)
- Giamarellos-Bourboulis EJ, Aschenbrenner AC, Bauer M, et al. The pathophysiology of sepsis and precision-medicine-based immunotherapy. *Nat Immunol*. 2024;25(1):19-28. doi:[10.1038/s41590-023-01660-5](https://doi.org/10.1038/s41590-023-01660-5)
- Leventogiannis K, Kyriazopoulou E, Antonakos N, et al. Toward personalized immunotherapy in sepsis: the PROVIDE randomized clinical trial. *Cell Rep Med*. 2022;3(11):100817. doi:[10.1016/j.xcrm.2022.100817](https://doi.org/10.1016/j.xcrm.2022.100817)
- Döcke WD, Rando F, Syrbe U, et al. Monocyte deactivation in septic patients: restoration by IFN-gamma treatment. *Nat Med*. 1997;3(6):678-681. doi:[10.1038/nm0697-678](https://doi.org/10.1038/nm0697-678)
- Kotsaki A, Pickkers P, Bauer M, et al. ImmunoSep (Personalised Immunotherapy in Sepsis) international double-blind, double-dummy, placebo-controlled randomised clinical trial: study protocol. *BMJ Open*. 2022;12(12):e067251. doi:[10.1136/bmjopen-2022-067251](https://doi.org/10.1136/bmjopen-2022-067251)
- Hopewell S, Chan AW, Collins GS, et al. CONSORT 2025 statement: updated guideline for reporting randomized trials. *JAMA*. 2025;333(22):1998-2005. doi:[10.1001/jama.2025.4347](https://doi.org/10.1001/jama.2025.4347)

- 8.** Tandogdu Z, Koves B, Ristovski S, et al; SERPENS Investigators. Urosepsis 30-day mortality, morbidity, and their risk factors: SERPENS study, a prospective, observational multi-center study. *World J Urol.* 2024;42(1):314. doi:[10.1007/s00345-024-04979-2](https://doi.org/10.1007/s00345-024-04979-2)
- 9.** Brunkhorst FM, Engel C, Bloos F, et al; German Competence Network Sepsis (SepNet). Intensive insulin therapy and pentastarch resuscitation in severe sepsis. *N Engl J Med.* 2008;358(2):125-139. doi:[10.1056/NEJMoa070716](https://doi.org/10.1056/NEJMoa070716)
- 10.** Brunkhorst FM, Oppert M, Marx G, et al; German Study Group Competence Network Sepsis (SepNet). Effect of empirical treatment with moxifloxacin and meropenem vs meropenem on sepsis-related organ dysfunction in patients with severe sepsis: a randomized trial. *JAMA.* 2012;307(22):2390-2399. doi:[10.1001/jama.2012.5833](https://doi.org/10.1001/jama.2012.5833)
- 11.** Kyriazopoulou E, Poulakou G, Milionis H, et al. Early treatment of COVID-19 with anakinra guided by soluble urokinase plasminogen receptor plasma levels: a double-blind, randomized controlled phase 3 trial. *Nat Med.* 2021;27(10):1752-1760. doi:[10.1038/s41591-021-01499-z](https://doi.org/10.1038/s41591-021-01499-z)
- 12.** Opal SM, Fisher CJ Jr, Dhainaut JF, et al; The Interleukin-1 Receptor Antagonist Sepsis Investigator Group. Confirmatory interleukin-1 receptor antagonist trial in severe sepsis: a phase III, randomized, double-blind, placebo-controlled, multicenter trial. *Crit Care Med.* 1997;25(7):1115-1124. doi:[10.1097/00003246-199707000-00010](https://doi.org/10.1097/00003246-199707000-00010)
- 13.** Fisher CJ Jr, Dhainaut JF, Opal SM, et al; Phase III rhIL-1 α Sepsis Syndrome Study Group. Recombinant human interleukin 1 receptor antagonist in the treatment of patients with sepsis syndrome: results from a randomized, double-blind, placebo-controlled trial. *JAMA.* 1994;271(23):1836-1843. doi:[10.1001/jama.1994.03510470040032](https://doi.org/10.1001/jama.1994.03510470040032)
- 14.** Shakoory B, Carcillo JA, Chatham WW, et al. Interleukin-1 receptor blockade is associated with reduced mortality in sepsis patients with features of macrophage activation syndrome: reanalysis of a prior phase III trial. *Crit Care Med.* 2016;44(2):275-281. doi:[10.1097/CCM.00000000000001402](https://doi.org/10.1097/CCM.00000000000001402)
- 15.** Leentjens J, Kox M, Koch RM, et al. Reversal of immunoparalysis in humans in vivo: a double-blind, placebo-controlled, randomized pilot study. *Am J Respir Crit Care Med.* 2012;186(9):838-845. doi:[10.1164/rccm.201204-0645OC](https://doi.org/10.1164/rccm.201204-0645OC)
- 16.** Delsing CE, Gresnigt MS, Leentjens J, et al. Interferon-gamma as adjunctive immunotherapy for invasive fungal infections: a case series. *BMC Infect Dis.* 2014;14:166. doi:[10.1186/1471-2334-14-166](https://doi.org/10.1186/1471-2334-14-166)
- 17.** Roquilly A, Francois B, Huet O, et al; Atlanrea study group and the Société Française d'Anesthésie Réanimation (SFAR) Research Network. Interferon gamma-1b for the prevention of hospital-acquired pneumonia in critically ill patients: a phase 2, placebo-controlled randomized clinical trial. *Intensive Care Med.* 2023;49(5):530-544. doi:[10.1007/s00134-023-07065-0](https://doi.org/10.1007/s00134-023-07065-0)
- 18.** François B, Lambden S, Fivez T, et al; ASTONISH investigators. Prospective evaluation of the efficacy, safety, and optimal biomarker enrichment strategy for nangibotide, a TREM-1 inhibitor, in patients with septic shock (ASTONISH): a double-blind, randomised, controlled, phase 2b trial. *Lancet Respir Med.* 2023;11(10):894-904. doi:[10.1016/S2213-2600\(23\)00158-3](https://doi.org/10.1016/S2213-2600(23)00158-3)
- 19.** Laterre PF, Pickkers P, Marx G, et al; AdrenOSS-2 study participants. Safety and tolerability of non-neutralizing adrenomedullin antibody adrecizumab (HAM8101) in septic shock patients: the AdrenOSS-2 phase 2a biomarker-guided trial. *Intensive Care Med.* 2021;47(11):1284-1294. doi:[10.1007/s00134-021-06537-5](https://doi.org/10.1007/s00134-021-06537-5)
- 20.** Davenport EE, Burnham KL, Radhakrishnan J, et al. Genomic landscape of the individual host response and outcomes in sepsis: a prospective cohort study. *Lancet Respir Med.* 2016;4(4):259-271. doi:[10.1016/S2213-2600\(16\)00046-1](https://doi.org/10.1016/S2213-2600(16)00046-1)
- 21.** Scicluna BP, van Vught LA, Zwinderman AH, et al; MARS consortium. Classification of patients with sepsis according to blood genomic endotype: a prospective cohort study. *Lancet Respir Med.* 2017;5(10):816-826. doi:[10.1016/S2213-2600\(17\)30294-1](https://doi.org/10.1016/S2213-2600(17)30294-1)
- 22.** Sweeney TE, Perumal TM, Henao R, et al. A community approach to mortality prediction in sepsis via gene expression analysis. *Nat Commun.* 2018;9(1):694. doi:[10.1038/s41467-018-03078-2](https://doi.org/10.1038/s41467-018-03078-2)