report

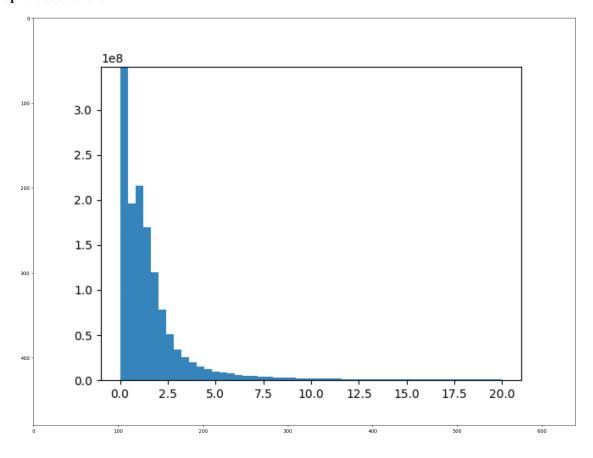
December 15, 2022

0.0.1 Number of observations / variables in the dataset

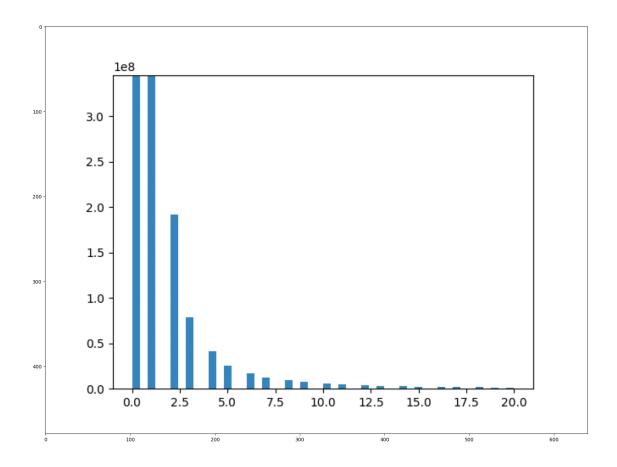
	num obs	num	variables
train	72208		5000
test	18052		5000

0.0.2 Histograms of the data (capped)

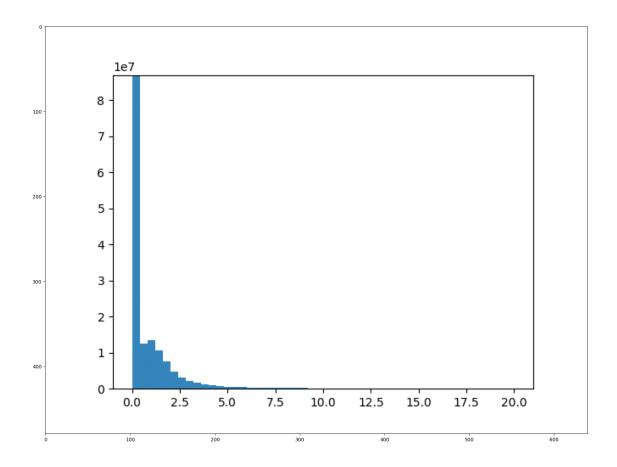
preprocessed train



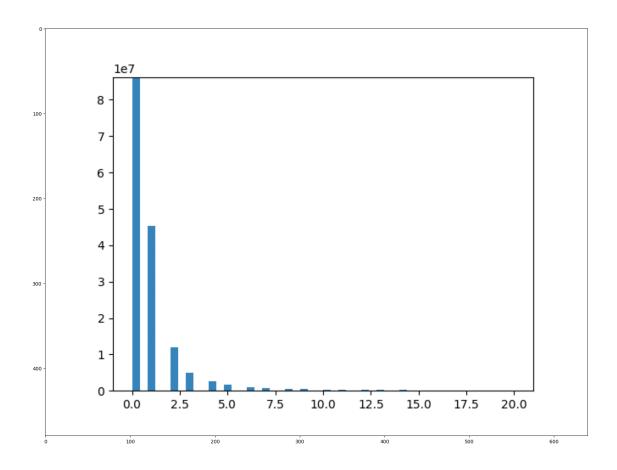
raw train



preprocessed test



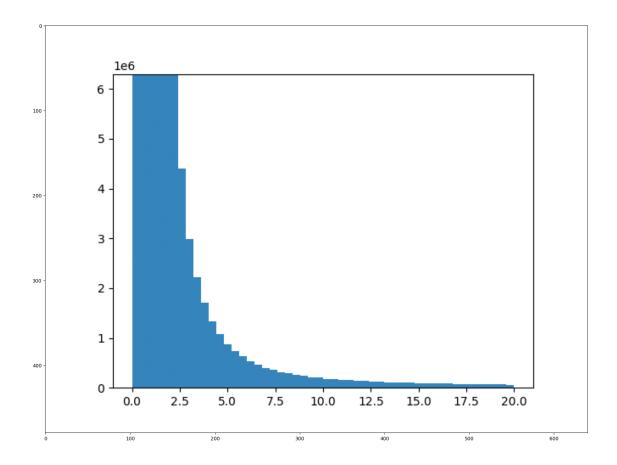
 $\mathbf{raw} \ \mathbf{test}$



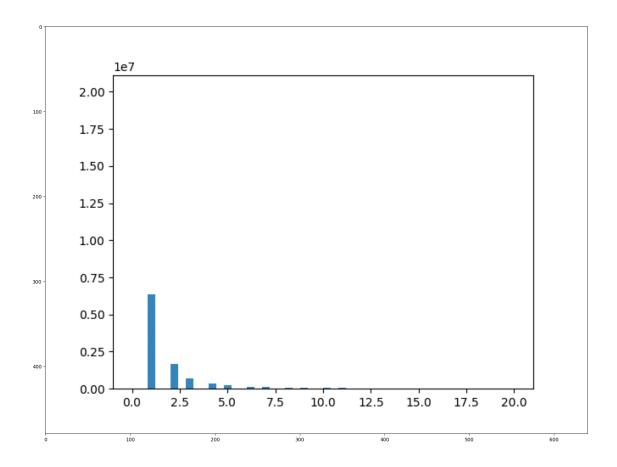
0.0.3 Data preprocessing

Data was preprocessed by dividing it by the GEX_size_factors variable in the obs

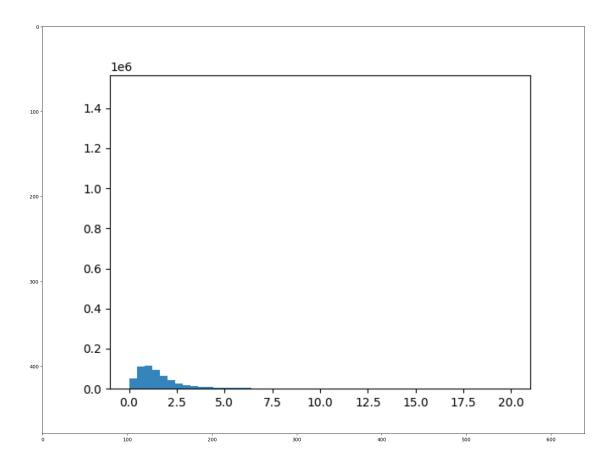
0.0.4 Histograms with zeros left out (capped as well) preprocessed train



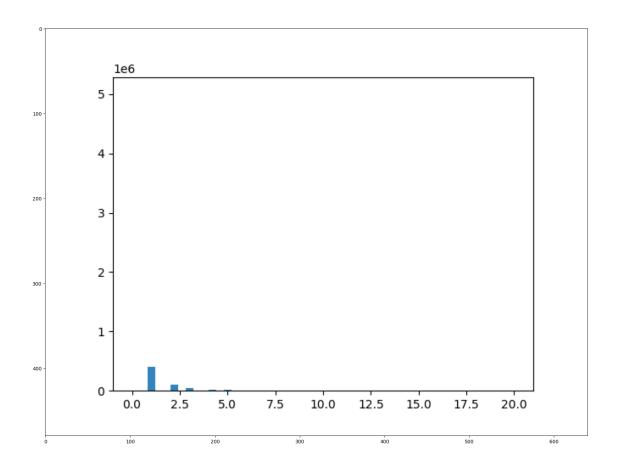
raw train



preprocessed test



 $\mathbf{raw} \ \mathbf{test}$



0.0.5 What is the distribution of the data

At first it seems that a poisson distribution would be sufficient. (which is common in RNA modelling, according to my almost nonexistent biological knowledge).

However due to the variance being larger than the mean (and a lot of zeros present) it is wise to suggest a negative binomial distribution (which is tightly connected to poisson distribution)

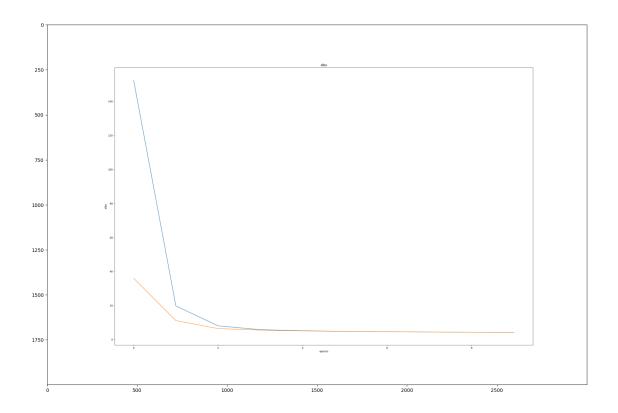
0.0.6 What are the contexts of the adata.obs

additional, explanatory information for each observation. Biological details about RNA sequences and the patients

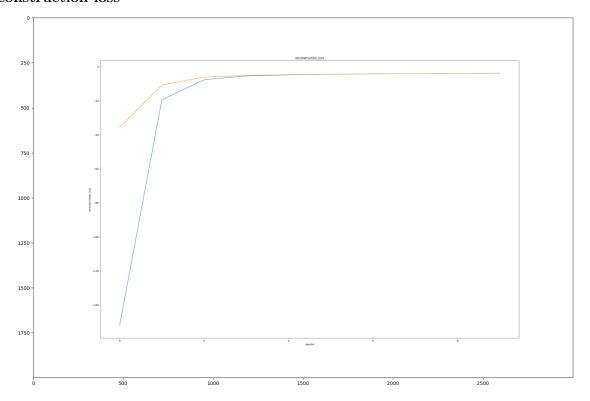
0.0.7 number of categories (donors, sites, cell types)

0.0.8 Training of the vanilla VAE (latent size of 32, 10 epochs, 128 batch size, 4 hidden layers of width 256 in encoder/decoder)

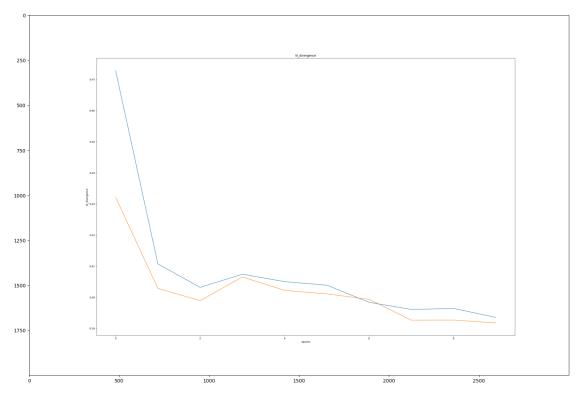
ELBO



reconstruction loss



KL divergence

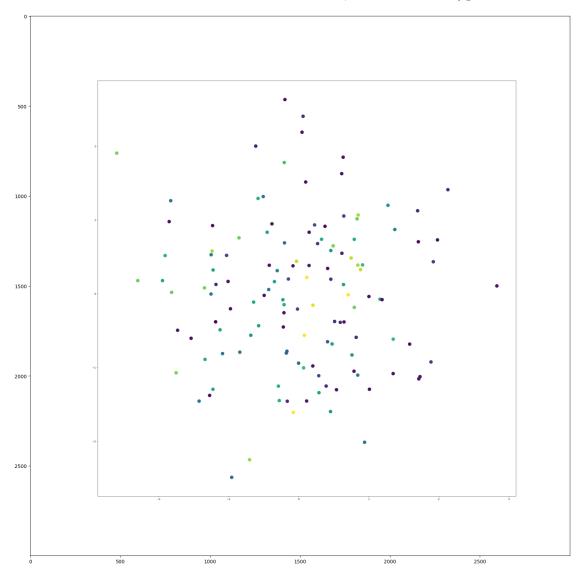


0.0.9~ 3 runs with different latent sizes (and minimum number of PCA components to reach 95% variance explained)

latent_size	elbo	kl divergence	recon loss	min components	\
32	4.220623	0.391793	3.828830	31	
16	4.306129	0.421133	3.884996	16	
8	4.416090	0.460025	3.956065	8	
	32 16	32 4.220623 16 4.306129	32 4.220623 0.391793 16 4.306129 0.421133	32 4.220623 0.391793 3.828830 16 4.306129 0.421133 3.884996	16 4.306129 0.421133 3.884996 16

explained_variance
0 0.971123
1 1.000000
2 1.000000

0.0.10 2-PCA for the 32-dimensional latent_size, coloured cell types



0.0.11 Overall thoughts of the training

I chose to use the raw dataset, since there was no clear empirical benefit of using the preprocessed dataset. I'll be using the raw data (integers) for the custom decoder as well. That way it'll be easier to compare both methods.

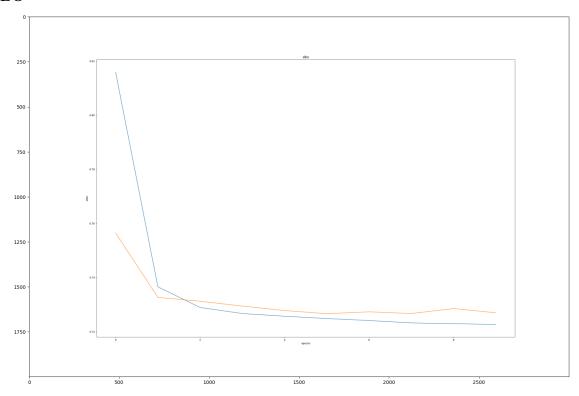
I used latent_size of 32, since it performed better on the test set than 128, 64, 16 or 8. I used 10 epochs and such latent sizes for the training times to not be too long (while training on cpu).

0.0.12 Custom decoder

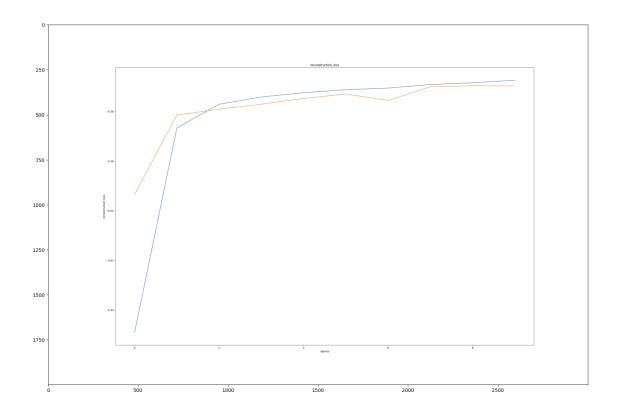
Negative binomial decoder results in few times lower ELBO, reconstruction loss is much lower and KL divergence is slightly lower as well.

I chose that distribution mainly because the data resembles it a lot.

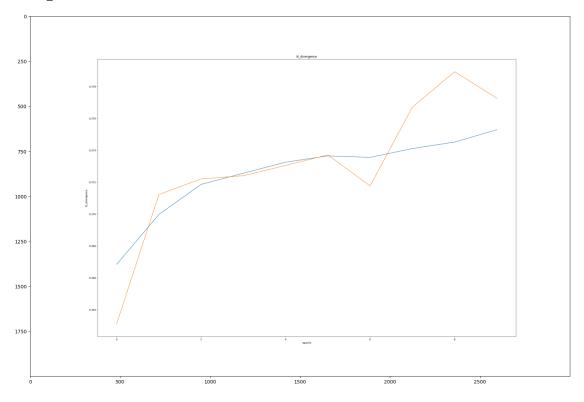
ELBO



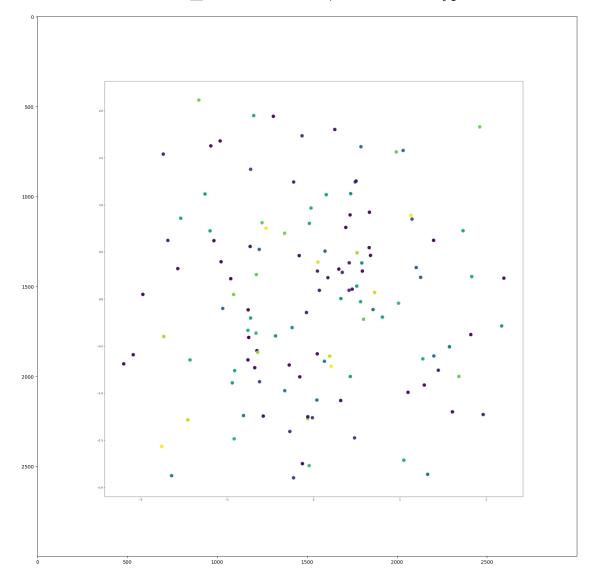
reconstruction loss



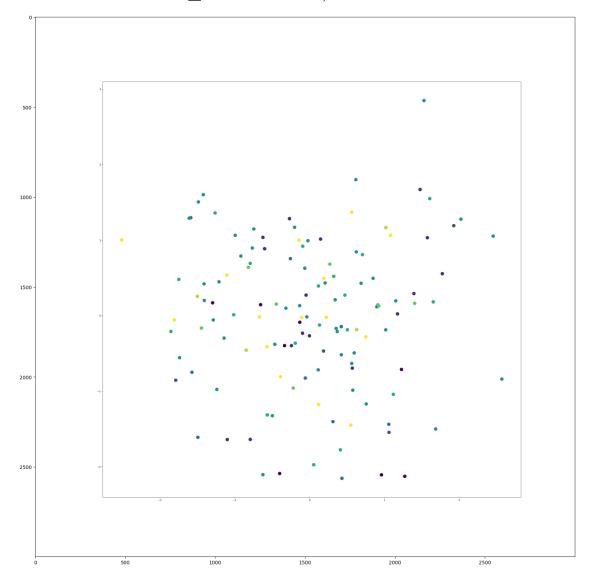
KL divergence



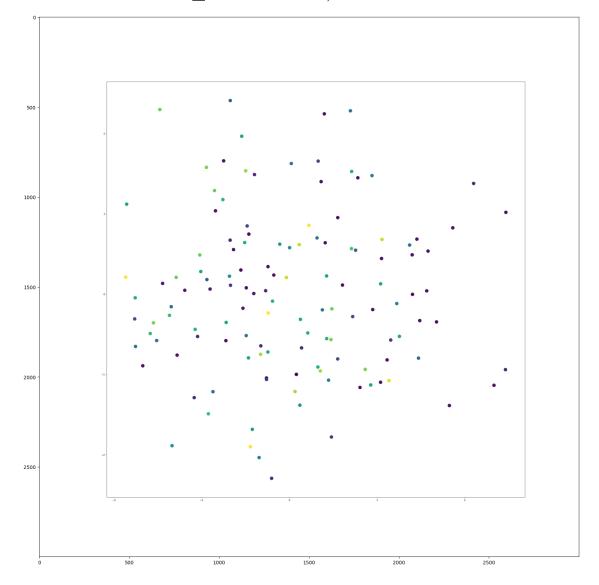
 $0.0.13 \quad \hbox{2-PCA for 32-latent_dim custom VAE, coloured cell type}$



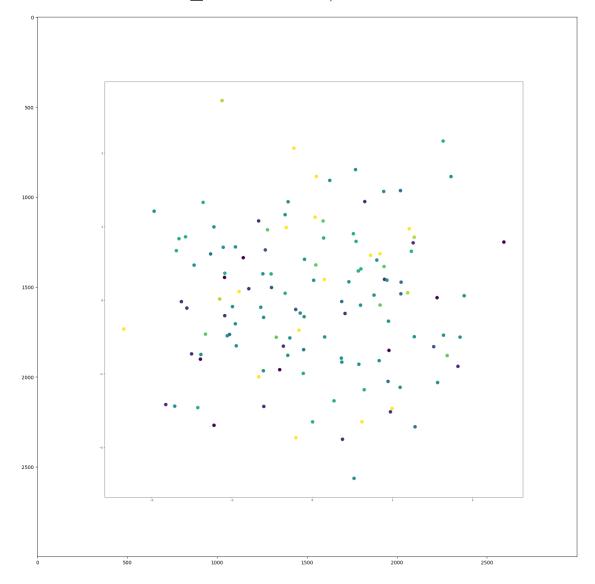
0.0.14 2-PCA for 32-latent_dim vanilla VAE, coloured donor id



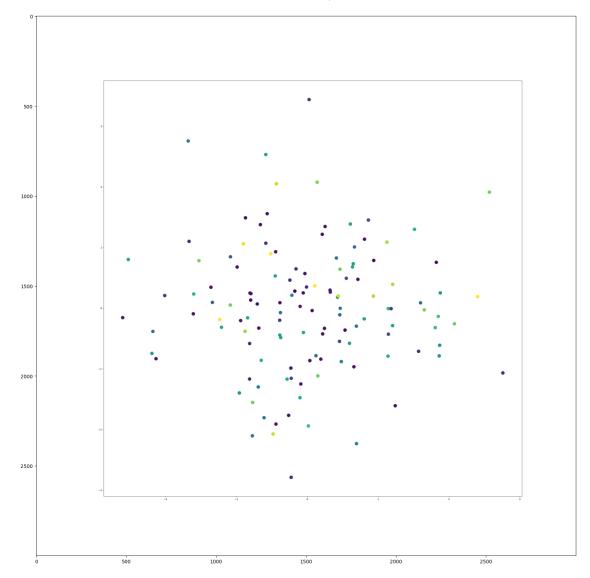
0.0.15 2-PCA for 32-latent_dim vanilla VAE, coloured site



 $0.0.16 \quad \hbox{2-PCA for 32-latent_dim custom VAE, coloured donor id}$

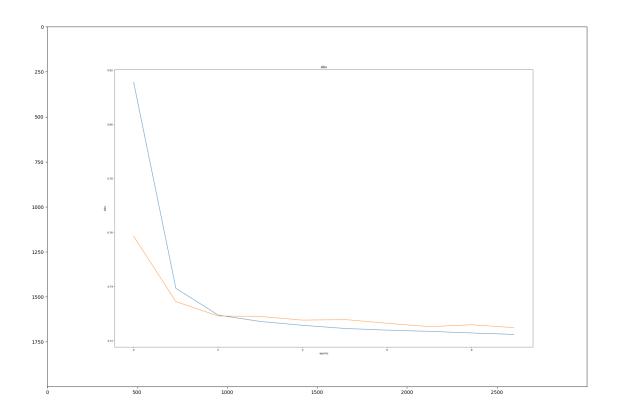


0.0.17 2-PCA for 32-latent_dim custom VAE, coloured site

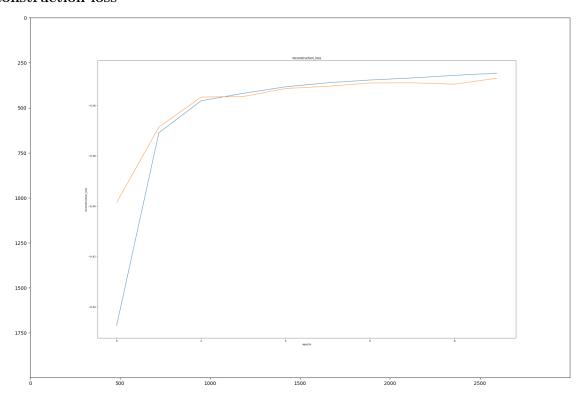


0.0.18 I took one-hot encoded site and concatenated it to the latent variables, fed into the decoder

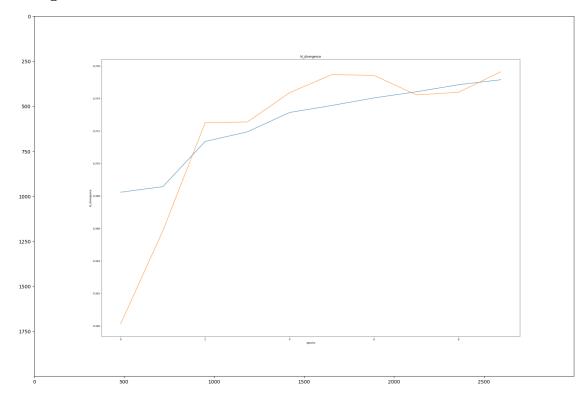
ELBO



reconstruction loss



KL divergence



0.0.19 2-PCA, coloured cell type

