Benchmark Dataset Construction | GSA Tools Comparison

Shaurya Jauhari, PhD

Mora Lab.

shauryajauhari@gzhmu.edu.cn

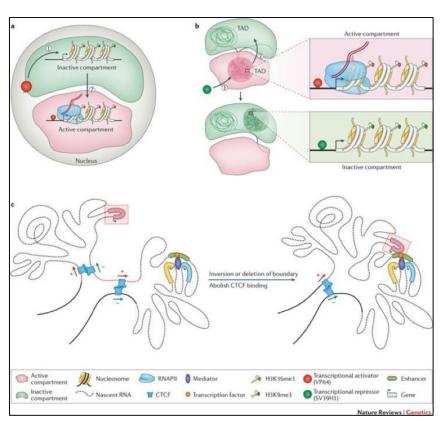
2019/01/15

Outline

- Core Idea
- Benchmarking Data | "Gold Standard" dataset
- Testing
- Caveat(s) hitherto
- Files/ Snapshots

Core Idea

- GSA Pipeline: Sequences/ Peaks/
 Genomic Regions -> Genes (Gene List)
 -> Pathways (Gene Sets)
- BED files are the major input form that manifest genomic regions for variegated interactions.
- Genome organization is a complex structuring in 3D space; loops, domains, sub-compartments aren't just plainly to accommodate ~ 2 meter long DNA into the nucleus, but also have biological meaning, i.e. to enact the genomic code in an organism.



Organization and function of the 3D genome, Boyan Bonev and Giacomo Cavalli, *Nature* Reviews Genetics **volume 17**, pages 661–678 (2016)

Core Idea ... Contd.

- Current suite of GSA tools typically overlook this standard of 3D genome organization. They assume a linear window around the gene (TSS) to be the ideal regulatory region.
- GREAT, CompGO, Seq2pathway, Enrichr, Chipenrich, Broadenrich, Polyenrich are the contenders.



Coding Regions

Non-Coding Regions Straightforward mapping to the known "working" definitions of genes.

• "Speculation" on gene association.



Benchmarking Data | "Gold Standard" dataset

- Tabulation of Genomic Regions (DNA) Associated TF/ Histone Mark (Protein) interactions, pertaining to a physiological state.
- Data chosen on the grounds of:
 - Study/ Publication impact (Journal, Year, Citations)
 - Coverage (KEGG, GO, Cistrome)
- Omitting of Input/Normal assays.
- Experiment (GSE) Samples (GSM) form the key denominations of the benchmark dataset.
- In compliance to Bioconductor standards, a major criterion is to make use of existing packages to scale reproducibility. **GRanges** has been used as a proforma to store BED files for the R data objects.
- To apply to the tools, the samples (data objects) are again converted to BED files via export.bed().

Granges object

```
## GRanges object with 3 ranges and 0 metadata columns:
##
                 ranges strand
       segnames
          <Rle> <IRanges> <Rle>
##
       chr1 [1, 3]
    [1]
##
    [2] chr2 [3, 5]
##
    [3] chr1 [5, 7] +
##
##
    seqinfo: 2 sequences from hg19 genome
##
```

Testing

- The tracking of the comparative analysis is being maintained as a markdown document (code and output).
- Currently underway with GREAT (Web interface) and Seq2pathway (R package)
 - Still some samples remaining
- GO terms for Biological Processes, and Disease Ontology terms have been considered from GREAT.
- The standard window size has been derived from Seq2pathway's default value, i.e. 150 kBP. This shall be maintained as a basis of comparison, for now.

Caveat(s) Hitherto

MEMORY INSUFFICIENCY

- Samples over 100 MB disk file size fail to run
- The MAC system has 32 GB RAM.
- The **BioInfoServer** (IP Address: 10.168.122.47) has internet connectivity issues that impede package installation and system upgradation.
- The NAS in the lab cannot be a substitute for extended system storage; it's an auxiliary storage device.

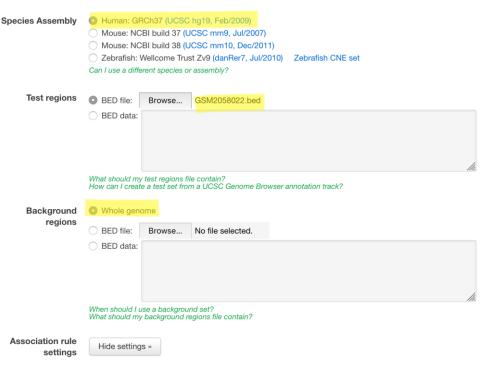
R Markdown

Loading Benchmark dataset

```
```{r sourcing the compiled gold standard benchmark dataset }
 ∰ ▼ ▶
install.packages("~/Desktop/Labwork/GSAChIPSeqComparitiveAnalysis/GSAChIPSeqGold_0.1.0.tar.qz", repos = NULL, type =
"source")
 * installing *source* package 'GSAChIPSeqGold' ...
 ** data
 *** moving datasets to lazyload DB
 ** help
No man pages found in package 'GSAChIPSeqGold'
*** installing help indices
** building package indices
** testing if installed package can be loaded
* DONE (GSAChIPSeqGold)
```{r Iterative variable naming from file names}
                                                                                                            # ≥ ▶
for (i in 1:length(grl)){
 export.bed(grl[i],format="bed",con=paste(eval(parse(text='names(grl[i])')),"data.bed",sep=""))
 paste(eval(parse(text='names(grl[i])')), "data", sep="") =
read_bed(paste(eval(parse(text='names(grl[i])')),"data.bed",sep=""))
paste(eval(parse(text='names(grl[i])')), "seq2pathway", sep="_")=runseq2pathway(paste(eval(parse(text='names(grl[i])'))
),"data",sep=""))
## Testing individual data in the benchmark dataset with
## each GSA tool package
## Seq2Pathway
```{r}
 ∰ ▼ ▶
library(GSAChIPSeqGold)
library(seq2pathway)
library(rtracklayer)
library(chipenrich)
export.bed(GSM1847178,format="bed",con="./regen/GSM1847178.bed")
GSM1847178data <- read_bed("./regen/GSM1847178.bed")</pre>
GSM1847178_seq2pathway=runseq2pathway(GSM1847178data,genome = "hg19")
export.bed(GSM2058015,format="bed",con="./regen/GSM2058015.bed")
GSM2058015data <- read_bed("./regen/GSM2058015.bed")</pre>
GSM2058015_seq2pathway=runseq2pathway(GSM2058015data, genome = "hg19")
Loading Benchmark dataset $
 R Markdown $
```

# GREAT





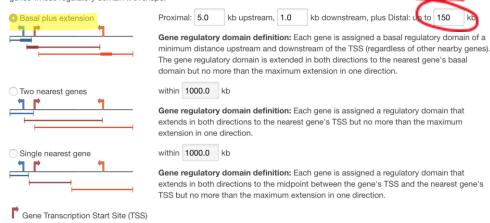
### Associating genomic regions with genes

✓ Include curated regulatory domains
What are curated regulatory domains?

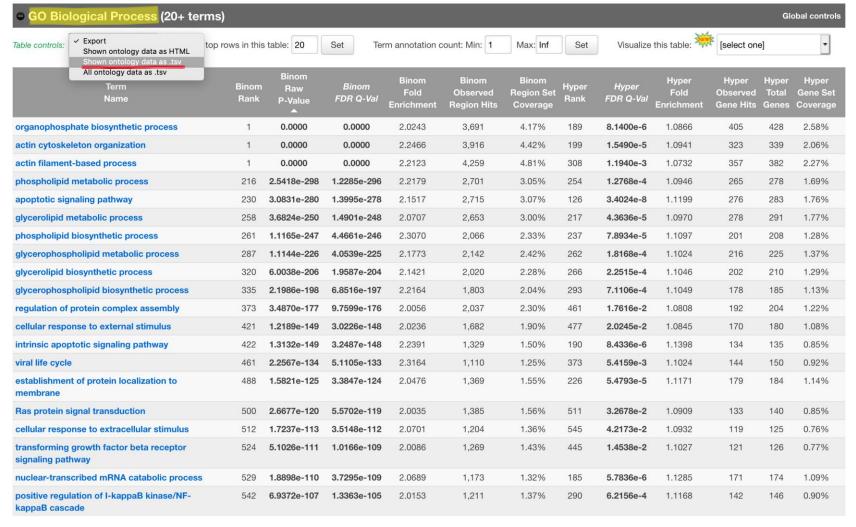
Reset

GREAT calculates statistics by associating genomic regions with nearby genes and applying the gene annotations to the regions.

Association is a two step process. First, every gene is assigned a regulatory domain. Then, each genomic region is associated with all genes whose regulatory domain it overlaps.



# GREAT . . . Contd.

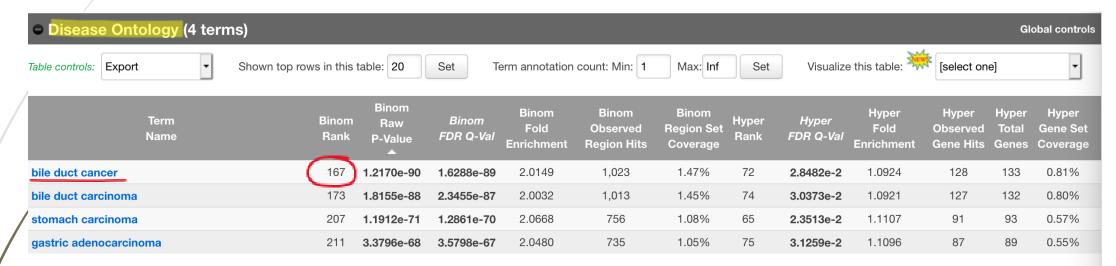


The test set of 88,556 genomic regions picked 15,711 (87%) of all 18,041 genes.

GO Biological Process has 10,440 terms covering 15,441 (86%) of all 18,041 genes, and 950,065 term - gene associations.

<sup>10,440</sup> ontology terms (100%) were tested using an annotation count range of [1, Inf].

# GREAT . . . Contd.



The test set of 69,699 genomic regions picked 15,894 (88%) of all 18,041 genes.

Disease Ontology has 2,235 terms covering 7,886 (44%) of all 18,041 genes, and 232,324 term - gene associations.

2,235 ontology terms (100%) were tested using an annotation count range of [1, Inf].



runseq2pathway 13

4. more columns: Other custom defined information (optional)

search\_radius

A non-negative integer, with which the input genomic regions can be assigned not only to the matched or nearest gene, but also with all genes within a search radius for some genomic region type. This parameter works only when the parameter "SNP" is FALSE. Default is 150000.

oromoter\_radius

A non-negative integer. Default is 200. Promoters are here defined as upstream regions of the transcription start sites (TSS). User can assign the promoter radius, a suggested value is between 200 to 2000.

promoter\_radius2

A non-negative integer. Default is 100. Promoters are here defined as down-

stream regions after the transcription start sites (TSS).

genome A character specifies the genome type. Currently, choice of "hg38", "hg19",

"mm10", and "mm9" is supported.

adjacent A Boolean. Default is FALSE to search all genes within the search\_radius.

Using "TRUE" to find the adjacent genes only and ignore the parameters "SNP"

and "search\_radius".

SNP A Boolean specifies the input object type. FALSE by default to keep on search-

ing for intron and neighboring genes. Otherwise, runseq2gene stops searching

when the input genomic region is residing on exon of a coding gene.

PromoterStop A Boolean, "FALSE" by default to keep on searching neighboring genes using

the parameter "search\_radius". Otherwise, runseq2gene stops searching neighboring genes. This parameter has function only if an input genomic region maps

to promoter of coding gene(s).

NearestTwoDirection

A boolean, "TRUE" by default to output the closest left and closest right coding genes with directions. Otherwise, output only the nearest coding gene regardless of direction.

of direction.

UTR3 A boolean, "FALSE" by defalt to calculate the distance from genes' 5UTR. Oth-

erwsie, calculate the distance from genes' 3UTR.

DataBase A character string assigns an R GSA.genesets object to define gene-set. User

can call GSA.read.gmt to load customized gene-sets with a .gmt format. If not specified, a character "GOterm" by default, three categories of GO-defined gene sets (BP,MF,CC) will be used. Alternatively, user can specify a category by the

choice of "BP", "MF", "CC".

FAIMETest A boolean values. By default is FALSE. When true, executes function of gene2pathway

test using the FAIME method, which only functions when the fifth column of

input file exsists and is a vector of scores or values.

FisherTest A Boolean value. By default is TRUE to excute the function of the Fisher's

exact test. Otherwise, only excutes the function of gene2pathway test.

collapsemethod A character for determining which method to use when call the function col-

lapseRows in package WGCNA. The function "collapsemethod" uses this paramter

to call the collapseRows() function in package "WGCNA".

alpha A positive integer, 5 by default. This is a FAIME-specific parameter. A higher

value puts more weights on the most highly-expressed ranks than the lower ex-

pressed ranks.

logCheck A Boolean value. By default is FALSE. When true, the function takes the log-

transformed values of gene if the maximum value of sample profile is larger than

20.

# Seq2pathway

- <variable\_name>\$seq2gene\_result.FET\$seq2gene\_CodingGeneOnlyResult
- <variable\_name>\$seq2gene\_result.FET\$seq2gene\_FullResult

- <variable\_name>\$gene2pathway\_result.FET\$GO.MF
- <variable\_name>\$gene2pathway\_result.FET\$GO.BP
- <variable\_name>\$gene2pathway\_result.FET\$GO.CC
- In R, the results can be directed to a dataframe format to extract useful heads, eg. Gene Names, Ensemble IDs, GO terms, etc.