

WGCNA Demo

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Installing the package and setting up the options.

```
install.packages("BiocManager",
                 repos='http://cran.us.r-project.org',
                 dependencies = TRUE)

## Installing package into '/Users/mei/Library/R/3.6/library'
## (as 'lib' is unspecified)

## Warning: dependency 'BiocStyle' is not available
##
##   There is a binary version available but the source version is
##   later:
##           binary source needs_compilation
## BiocManager 1.30.7 1.30.9                FALSE
## installing the source package 'BiocManager'
BiocManager::install("WGCNA")

## Bioconductor version 3.9 (BiocManager 1.30.9), R 3.6.0 (2019-04-26)
## Installing package(s) 'WGCNA'

## Package which is only available in source form, and may need
##   compilation of C/C++/Fortran: 'WGCNA'

## installing the source package 'WGCNA'

## Warning in install.packages(...): installation of package 'WGCNA' had non-
## zero exit status

## Old packages: 'digest', 'jpeg', 'JuliaCall', 'mgcv', 'plotmo', 'rlang'
install.packages("ggdendro",
                 repos='http://cran.us.r-project.org',
                 dependencies = TRUE)

## Installing package into '/Users/mei/Library/R/3.6/library'
## (as 'lib' is unspecified)
##
## The downloaded binary packages are in
## /var/folders/hm/c3_fjypn62v5xh5b5ygv267m0000gn/T//Rtmp59rqut/downloaded_packages
## Setting options

options(stringsAsFactors = FALSE)

library(WGCNA)

## Loading required package: dynamicTreeCut
## Loading required package: fastcluster
```

```
##
## Attaching package: 'fastcluster'

## The following object is masked from 'package:stats':
##
##      hclust
##
##
## Attaching package: 'WGCNA'

## The following object is masked from 'package:stats':
##
##      cor
library(ggdendro)
library(ggplot2)
library(cowplot)

##
## *****

## Note: As of version 1.0.0, cowplot does not change the
##      default ggplot2 theme anymore. To recover the previous
##      behavior, execute:
##      theme_set(theme_cowplot())

## *****

Importing data files from female and male liver tissues from mice, and exploring them.
mydataf <- read.csv("../FemaleLiver-Data/LiverFemale3600.csv", header = TRUE)
colnames(mydataf)

##      [1] "substanceBXH" "gene_symbol" "LocusLinkID" "ProteomeID"
##      [5] "cytogeneticLoc" "CHROMOSOME" "StartPosition" "EndPosition"
##      [9] "F2_2" "F2_3" "F2_14" "F2_15"
##     [13] "F2_19" "F2_20" "F2_23" "F2_24"
##     [17] "F2_26" "F2_37" "F2_42" "F2_43"
##     [21] "F2_45" "F2_46" "F2_47" "F2_48"
##     [25] "F2_51" "F2_52" "F2_54" "F2_63"
##     [29] "F2_65" "F2_66" "F2_68" "F2_69"
##     [33] "F2_70" "F2_71" "F2_72" "F2_78"
##     [37] "F2_79" "F2_80" "F2_81" "F2_83"
##     [41] "F2_86" "F2_87" "F2_88" "F2_89"
##     [45] "F2_107" "F2_108" "F2_109" "F2_110"
##     [49] "F2_111" "F2_112" "F2_117" "F2_119"
##     [53] "F2_125" "F2_126" "F2_127" "F2_141"
##     [57] "F2_142" "F2_143" "F2_144" "F2_145"
##     [61] "F2_154" "F2_155" "F2_156" "F2_157"
##     [65] "F2_162" "F2_163" "F2_164" "F2_165"
##     [69] "F2_166" "F2_167" "F2_169" "F2_180"
##     [73] "F2_181" "F2_182" "F2_187" "F2_188"
##     [77] "F2_189" "F2_190" "F2_191" "F2_192"
##     [81] "F2_194" "F2_195" "F2_200" "F2_201"
##     [85] "F2_212" "F2_213" "F2_214" "F2_215"
##     [89] "F2_221" "F2_222" "F2_223" "F2_224"
```

```
## [93] "F2_225"      "F2_226"      "F2_227"      "F2_228"
## [97] "F2_241"      "F2_242"      "F2_243"      "F2_244"
## [101] "F2_245"      "F2_247"      "F2_248"      "F2_261"
## [105] "F2_263"      "F2_264"      "F2_270"      "F2_271"
## [109] "F2_272"      "F2_278"      "F2_287"      "F2_288"
## [113] "F2_289"      "F2_290"      "F2_291"      "F2_296"
## [117] "F2_298"      "F2_299"      "F2_300"      "F2_302"
## [121] "F2_303"      "F2_304"      "F2_305"      "F2_306"
## [125] "F2_307"      "F2_308"      "F2_309"      "F2_310"
## [129] "F2_311"      "F2_312"      "F2_320"      "F2_321"
## [133] "F2_323"      "F2_324"      "F2_325"      "F2_326"
## [137] "F2_327"      "F2_328"      "F2_329"      "F2_330"
## [141] "F2_332"      "F2_355"      "F2_357"
```

```
head(mydataf)
```

```
## substanceBXH gene_symbol LocusLinkID ProteomeID cytogeneticLoc
## 1 MMT000000044 1700007N18Rik 69339 286025 0
## 2 MMT000000046 Mast2 17776 157466 0
## 3 MMT000000051 Ankrd32 105377 321939 0
## 4 MMT000000076 0 383154 0 0
## 5 MMT000000080 Ldb2 16826 157383 0
## 6 MMT000000102 Rdhs 216453 0 10_70.0_cM
## CHROMOSOME StartPosition EndPosition F2_2 F2_3 F2_14 F2_15
## 1 16 50911260 50912491 -0.01810 0.0642 6.44e-05 -0.05800
## 2 4 115215318 115372404 -0.07730 -0.0297 1.12e-01 -0.05890
## 3 13 74940309 74982847 -0.02260 0.0617 -1.29e-01 0.08710
## 4 16 49345114 49477048 -0.00924 -0.1450 2.87e-02 -0.04390
## 5 5 43546124 43613704 -0.04870 0.0582 -4.83e-02 -0.03710
## 6 10 1337265 1347607 0.17600 -0.1890 -6.50e-02 -0.00846
## F2_19 F2_20 F2_23 F2_24 F2_26 F2_37 F2_42
## 1 0.04830 -0.15197410 -0.00129 -0.23600 -0.0307 -0.02610 0.073705890
## 2 0.04430 -0.09380000 0.09340 0.02690 -0.1330 0.07570 -0.009193803
## 3 -0.11500 -0.06502607 0.00249 -0.10200 0.1420 -0.10200 0.064289290
## 4 0.00425 -0.23610000 -0.06900 0.01440 0.0363 -0.01820 0.477874600
## 5 0.02510 0.08504274 0.04450 0.00167 -0.0680 0.00567 -0.075348680
## 6 -0.00574 -0.01807182 -0.12500 -0.06820 0.1250 0.00998 -0.037366600
## F2_43 F2_45 F2_46 F2_47 F2_48 F2_51 F2_52 F2_54
## 1 -0.0466 -0.00673 -0.0193 0.09040 0.0290 0.0356 -0.0388 -0.0360
## 2 -0.0075 0.01700 0.0722 -0.08390 0.0273 -0.0784 -0.0178 0.1120
## 3 0.0169 -0.01590 -0.1430 -0.00492 -0.0735 0.0657 -0.0197 -0.1290
## 4 0.1440 0.11100 0.0113 0.11900 0.0225 0.0932 0.1430 0.2640
## 5 -0.0673 -0.04720 0.0701 -0.08790 -0.0180 -0.1290 -0.0469 -0.0352
## 6 -0.0402 -0.02190 0.0269 0.13300 0.0732 0.1070 -0.0362 -0.0696
## F2_63 F2_65 F2_66 F2_68 F2_69 F2_70 F2_71 F2_72
## 1 -0.05600 0.009840 -0.0261 0.00856 -0.01180 -0.03350 -0.08310 -0.0471
## 2 0.12300 0.051700 0.0731 0.08670 0.05710 0.00693 -0.00606 -0.0390
## 3 -0.14300 -0.061600 0.0419 -0.29000 -0.10800 -0.09950 -0.00315 0.0975
## 4 -0.09280 -0.000635 -0.0126 0.06910 0.02260 -0.08630 -0.22900 0.0178
## 5 -0.00166 0.058700 -0.0206 -0.13000 0.00392 0.05450 -0.11200 0.1070
## 6 -0.19400 -0.117000 -0.0400 0.06890 0.04320 -0.00338 -0.05270 -0.0416
## F2_78 F2_79 F2_80 F2_81 F2_83 F2_86 F2_87 F2_88
## 1 -0.02820 0.047264410 0.0296 0.0114 0.0498 -0.0249 -0.00264 -0.02050
## 2 0.01870 0.008471275 -0.0687 -0.0114 -0.0262 -0.0215 -0.09580 -0.01930
## 3 0.01030 -0.134271000 0.1010 0.0521 -0.0607 -0.0285 0.02560 -0.01350
```

```

## 4  0.00166  0.064096960  0.0103 -0.0258 -0.0837  0.1880  0.03310 -0.00652
## 5  0.01190  0.008985630 -0.1030 -0.1400 -0.0282 -0.1090  0.02070 -0.01370
## 6 -0.03040  0.025920240  0.0697  0.1150  0.0953  0.0127  0.05490  0.00311
##      F2_89 F2_107 F2_108 F2_109 F2_110 F2_111 F2_112 F2_117
## 1  0.0826 -0.0421  0.0663  0.03620  0.0808 -0.0404  0.0877  0.07240
## 2 -0.1140  0.0815  0.0285  0.00299 -0.0407 -0.0657  0.0643 -0.00022
## 3  0.0796  0.0553 -0.0380  0.12900 -0.0361  0.0441 -0.1640 -0.01420
## 4  0.1550  0.0458  0.0752  0.12200 -0.0104  0.0914 -0.0355  0.06520
## 5 -0.0288 -0.1220  0.1270 -0.09390  0.1200 -0.0850  0.1400  0.00867
## 6  0.0955 -0.1520 -0.0670 -0.00599 -0.0438  0.0634  0.1380 -0.04010
##      F2_119 F2_125 F2_126 F2_127 F2_141 F2_142 F2_143 F2_144
## 1 -0.0210  0.04540 -0.03220 -0.00654  0.03490 -0.0315 -0.02170  0.00370
## 2 -0.0877  0.00167  0.00321 -0.01260 -0.04530 -0.0579  0.05920  0.00239
## 3 -0.0279  0.00677  0.07360  0.01750  0.10900 -0.0216 -0.01250  0.05460
## 4  0.1280  0.05940  0.01630  0.00292  0.00714 -0.0565  0.10200  0.03480
## 5  0.1440  0.08710 -0.03360  0.17300  0.08270  0.0594 -0.00317 -0.06750
## 6  0.1310 -0.12600  0.00484 -0.00256 -0.06800  0.0941 -0.04220  0.12000
##      F2_145      F2_154      F2_155 F2_156 F2_157 F2_162 F2_163 F2_164
## 1  0.0322 -0.02150730 -0.000958 -0.0850  0.00462  0.03990  0.0716 -0.0923
## 2 -0.0383  0.02457782 -0.030300 -0.1260 -0.06670 -0.00637 -0.0161 -0.2340
## 3  0.0403 -0.01674888  0.059900  0.0311 -0.05190  0.01890  0.0207  0.0929
## 4  0.0245  0.06776892  0.016500 -0.0382  0.02120  0.06690  0.0512 -0.2450
## 5  0.0495  0.13520570  0.016500  0.0832  0.04350  0.19300  0.0586 -0.0768
## 6  0.1080 -0.05128296 -0.005590  0.0136  0.09910  0.06770 -0.0520  0.1550
##      F2_165 F2_166 F2_167 F2_169 F2_180      F2_181 F2_182 F2_187
## 1  0.10900  0.0102  0.0337  0.00911  0.03210  0.03144772  0.0543  0.01120
## 2 -0.09610 -0.1290 -0.0109 -0.11300 -0.00677 -0.16704700 -0.0239  0.00304
## 3  0.00917  0.0874 -0.1260 -0.00949 -0.09900  0.02700180 -0.0570 -0.05160
## 4  1.23000 -0.0402 -0.0635  0.06880  0.03790 -0.02058180  0.0227  0.04180
## 5  0.04600  0.0484  0.2810  0.07210 -0.00630  0.37074790  0.0618  0.10800
## 6  0.07890  0.0336  0.0648  0.14400  0.02770  0.09297908  0.0601  0.02960
##      F2_188 F2_189 F2_190 F2_191 F2_192 F2_194 F2_195 F2_200
## 1  0.01060  0.1130 -0.03960 -0.0504  0.0877 -0.0563 -0.00557 -0.0484
## 2 -0.03580 -0.1330 -0.01830 -0.0623 -0.0648 -0.0652  0.05020 -0.0912
## 3 -0.04970  0.1660  0.05000  0.0498  0.0431 -0.0224 -0.10700  0.0715
## 4  0.01010  0.2170  0.00206 -0.0155  0.6550  0.2820 -0.01310 -0.0387
## 5  0.12100  0.0237  0.02960  0.1130  0.0839  0.1050  0.15500  0.0823
## 6  0.00198  0.0251  0.00059 -0.0282  0.0429  0.0697  0.04930  0.0414
##      F2_201      F2_212 F2_213 F2_214 F2_215 F2_221 F2_222 F2_223
## 1 -0.0273 -0.10816380 -0.0183 -0.0132 -0.00432 -0.6630  0.01440  0.0310
## 2 -0.0180  0.05682362 -0.0238  0.0721  0.03910  0.1070  0.00923 -0.0397
## 3  0.0432 -0.13217820  0.0205 -0.0411  0.07670 -0.0783 -0.06860 -0.0254
## 4 -0.0667 -0.32395020 -0.0245  0.0865  0.06470 -2.0000  0.00874  0.0847
## 5  0.1140  0.03542023 -0.2020  0.0822  0.04260  0.1030 -0.10100  0.1630
## 6 -0.0708 -0.10881230  0.0359 -0.0678 -0.11000 -0.1420  0.08430 -0.0610
##      F2_224 F2_225 F2_226 F2_227 F2_228 F2_241 F2_242 F2_243
## 1  0.00818 -0.00892 -0.08710  0.0129  0.0937  0.0313  0.0821  0.00621
## 2 -0.06400  0.06300 -0.00152  0.0555  0.0947 -0.0387  0.0592 -0.00636
## 3 -0.05680 -0.13300 -0.07560 -0.0557 -0.0890 -0.1460 -0.0739 -0.01120
## 4 -0.09720  0.00746 -0.55200  0.0415  0.0733  0.0815  0.1100  0.21400
## 5  0.07410 -0.01640  0.08700 -0.0557 -0.1910  0.0219  0.0913  0.01120
## 6  0.08760 -0.03960  0.10200  0.0190 -0.1190  0.0687 -0.0525 -0.00716
##      F2_244 F2_245 F2_247      F2_248 F2_261 F2_263 F2_264
## 1  0.0307 -0.13700  0.075300 -0.096881950 -0.01670 -0.0928 -0.00957

```

```

## 2  0.0614  0.02850 -0.000633  0.001598228 -0.00267 -0.0198  0.16300
## 3 -0.0528  0.05050  0.027700 -0.067933370 -0.02220 -0.0684 -0.04930
## 4  0.0135 -0.13500 -0.003100  0.072318780  0.01030 -0.3150  0.08420
## 5  0.1190  0.00383  0.041700 -0.038618510  0.11800  0.0123  0.03700
## 6 -0.1460 -0.14500  0.029400  0.035281240 -0.05660  0.0917 -0.08080
##      F2_270  F2_271  F2_272  F2_278  F2_287      F2_288  F2_289  F2_290
## 1  0.0287 -0.01300 -0.0292 -0.03810 -0.0488  0.17361240 -0.097900  0.0383
## 2 -0.1310 -0.04260 -0.0514  0.07260 -0.0481 -0.16211430 -0.123000 -0.1370
## 3  0.0328  0.00537 -0.0259 -0.14400  0.0170  0.25924220 -0.041400 -0.0229
## 4  0.0351      NA  0.0730  0.00914  0.0556  0.18311140  0.051700  0.1780
## 5 -0.0142  0.00563 -0.0504 -0.05970 -0.0871  0.20897910 -0.000188 -0.0328
## 6  0.0362  0.00790 -0.0246 -0.07330  0.0125 -0.04778892  0.082500  0.1360
##      F2_291      F2_296  F2_298  F2_299      F2_300  F2_302  F2_303  F2_304
## 1  0.01850 -0.08937784  0.0230 -0.06250 -0.000142  0.0344  0.0358 -0.0139
## 2 -0.05720 -0.07416870 -0.0688 -0.06540 -0.102000 -0.0780 -0.0820 -0.1830
## 3 -0.00664 -0.05915232 -0.0134  0.09740  0.015500 -0.0934  0.1780  0.0842
## 4  0.05250 -0.21653720 -0.2210 -0.00266  0.545000  0.0127  0.0273 -0.0928
## 5 -0.16600 -0.07897525  0.1410 -0.12900  0.090600 -0.1330 -0.2120 -0.0797
## 6  0.04620  0.03811979 -0.0346  0.04690 -0.034800  0.0110  0.0323  0.1660
##      F2_305      F2_306  F2_307  F2_308  F2_309  F2_310  F2_311  F2_312
## 1  0.0134 -0.03145069  0.02780 -0.01190 -0.0744  0.00197 -0.0151 -0.0721
## 2 -0.0270 -0.09822316 -0.07890 -0.05480 -0.1320 -0.11000 -0.1130 -0.0805
## 3  0.0870  0.15520470  0.03410 -0.06830  0.0555 -0.04060  0.0835  0.0514
## 4  0.0469  0.10038160 -2.00000  0.05240  0.1260  0.07280  0.0600 -0.0455
## 5 -0.0191 -0.11958500  0.00294 -0.10600 -0.0518 -0.13200  0.0494  0.0221
## 6 -0.0866  0.05385017  0.09570 -0.00949  0.1120  0.20800  0.0872 -0.0555
##      F2_320  F2_321  F2_323      F2_324  F2_325  F2_326  F2_327      F2_328
## 1 -0.0118  0.0200  0.0222  0.047700 -0.0488  0.0168 -0.0309  0.02740
## 2 -0.1200  0.0101 -0.1610 -0.049200 -0.0350 -0.0738 -0.1730 -0.07380
## 3  0.0713 -0.1130  0.0466  0.000612  0.1210  0.0996  0.1090  0.02730
## 4 -0.0464  0.0667 -0.1850 -0.270000  0.0803  0.0424  0.1610  0.05120
## 5  0.0272 -0.0938  0.1020  0.113000 -0.0859 -0.1340  0.0639  0.00731
## 6  0.0748 -0.1420  0.0590 -0.080000 -0.1200  0.1230  0.1870  0.05410
##      F2_329  F2_330  F2_332  F2_355      F2_357
## 1 -0.0310  0.0660 -0.0199 -0.0146  0.065000
## 2 -0.2010 -0.0820 -0.0939  0.0192 -0.049900
## 3  0.1200 -0.0629 -0.0395  0.1090  0.000253
## 4  0.2410  0.3890  0.0251 -0.0348  0.114000
## 5  0.1240 -0.0212  0.0870  0.0512  0.024300
## 6  0.0699  0.0708  0.1450 -0.0399  0.037500

```

```

## LocusLinkID and ProteomeID are annotations from the said databases
## http://www.ncbi.nlm.nih.gov/LocusLink/

```

Moving on, we extract expression data from the master dataframe. Recall that the rows represent genes and the columns represent different samples (mice) in the original data. WGCNA requires that genes be given in columns.

```

exprdata = as.data.frame(t(mydataf[, -c(1:8)]))
names(exprdata) = mydataf$gene_symbol
rownames(exprdata) = names(mydataf)[-c(1:8)]

```

```

## Let us consider a subset of data for this demonstration. We'll use first 500 features.

```

```

exprdata <- exprdata[,1:500]

gsg = goodSamplesGenes(exprdata, verbose = 3)

## Flagging genes and samples with too many missing values...
## ..step 1

gsg$allOK

## [1] TRUE

```

Scale Free Topology

A scale free network topology is the one where all nodes's degree distribution is in abundance to power law ,i.e. $P(k) \sim k^{-\beta}$. If any nodes have to be added to this connected network, the degrees are accordingly adjusted.

```

trial_powers <- c(c(1:10), seq(from=12, to=20, by=2))
sft_thresh <- pickSoftThreshold (exprdata,
                                dataIsExpr = TRUE,
                                powerVector = trial_powers,
                                corFnc = cor,
                                corOptions = list(use = 'p'),
                                networkType = "unsigned")

## Warning: executing %dopar% sequentially: no parallel backend registered

##      Power SFT.R.sq   slope truncated.R.sq mean.k. median.k. max.k.
## 1      1 0.000592 -0.0489      0.283000 100.000  1.03e+02 161.00
## 2      2 0.273000 -0.8680      0.809000  33.800  3.34e+01  75.60
## 3      3 0.484000 -1.4100      0.942000  14.800  1.36e+01  42.40
## 4      4 0.626000 -1.6100      0.955000   7.550  6.30e+00  26.20
## 5      5 0.752000 -1.6600      0.941000   4.320  3.39e+00  17.20
## 6      6 0.859000 -1.4900      0.877000   2.690  2.03e+00  12.00
## 7      7 0.232000 -3.0500      0.044400   1.800  1.19e+00  10.60
## 8      8 0.256000 -3.0400      0.059600   1.270  7.33e-01   9.62
## 9      9 0.219000 -2.6300      0.000234   0.934  4.68e-01   8.88
## 10     10 0.205000 -3.2000      0.002140   0.715  3.09e-01   8.29
## 11     12 0.144000 -2.4600      0.028900   0.459  1.43e-01   7.35
## 12     14 0.150000 -2.3200      0.021300   0.323  6.62e-02   6.62
## 13     16 0.152000 -1.6800      0.026400   0.242  3.18e-02   6.02
## 14     18 0.159000 -1.6300      0.027800   0.190  1.60e-02   5.52
## 15     20 0.872000 -1.2500      0.911000   0.155  8.18e-03   5.09

cat("The best estimate for use is", sft_thresh$powerEstimate, ".")

## The best estimate for use is 6 .

print(sft_thresh$fitIndices)

```

```

##      Power      SFT.R.sq      slope truncated.R.sq      mean.k.      median.k.
## 1      1 0.000591956 -0.04892954  0.282584079 100.3934169 1.032604e+02
## 2      2 0.272783992 -0.86849966  0.808785217  33.8373469 3.341476e+01
## 3      3 0.483755802 -1.41438909  0.941534027  14.7534022 1.362163e+01
## 4      4 0.626177009 -1.61216723  0.954882010   7.5478368 6.299250e+00
## 5      5 0.751600155 -1.66289137  0.941321384   4.3206844 3.386659e+00
## 6      6 0.858673907 -1.49250031  0.877183575   2.6934134 2.028749e+00

```

```
## 7      7 0.232232581 -3.05120613    0.044428656    1.7965152 1.186614e+00
## 8      8 0.256337946 -3.04292255    0.059632912    1.2661169 7.334995e-01
## 9      9 0.219405670 -2.63191579    0.000233961    0.9338283 4.684453e-01
## 10     10 0.204590608 -3.20415305    0.002141256    0.7153326 3.091732e-01
## 11     12 0.144177374 -2.46273973    0.028862575    0.4593058 1.427019e-01
## 12     14 0.150084643 -2.32214905    0.021330651    0.3228727 6.622747e-02
## 13     16 0.151669038 -1.67666388    0.026396129    0.2421974 3.178078e-02
## 14     18 0.159074112 -1.63426374    0.027829208    0.1904629 1.595709e-02
## 15     20 0.872145990 -1.24994141    0.910750187    0.1550880 8.177421e-03
##      max.k.
## 1    160.589081
## 2     75.613728
## 3     42.390932
## 4     26.155630
## 5     17.211457
## 6     12.023663
## 7     10.570718
## 8      9.616364
## 9      8.884697
## 10     8.288647
## 11     7.348466
## 12     6.617885
## 13     6.021118
## 14     5.518548
## 15     5.086358
```

It has been determined that $\beta=6$ is the best exponent for unsigned networks, and $\beta=12$ is the optimal one for signed networks.

Typically, the degrees per node will “steadily” decrease with the increase in the number of nodes of the network cluster. This is another way of understanding the notion of scale-free topology.

```
# Scale-free topology fit index as a function of the soft-thresholding power

p1 <- ggplot(data = as.data.frame(sft_thresh),
  aes (x = sft_thresh$fitIndices[,1],
    y = (-sign(sft_thresh$fitIndices[,3]) * sft_thresh$fitIndices[,2]),
    label = trial_powers)) +
  geom_label()+

  # the sign expression returns the vector of signs of estimated slopes, multiplied by the correlation

  labs( x="Soft Threshold (power)",
    y="Scale Free Topology Model Fit, signed R^2", title = "Scale Independence", tag = "A")+
  theme_classic()+
  geom_vline(xintercept = 8, color="red") # Red line corresponds to using an R^2 cut-off

# Mean connectivity as a function of the soft-thresholding power

p2 <- ggplot(data = as.data.frame(sft_thresh),
  aes (x = sft_thresh$fitIndices[,1],
    y = sft_thresh$fitIndices[,5],
    label = trial_powers)) +
  geom_label()+
```

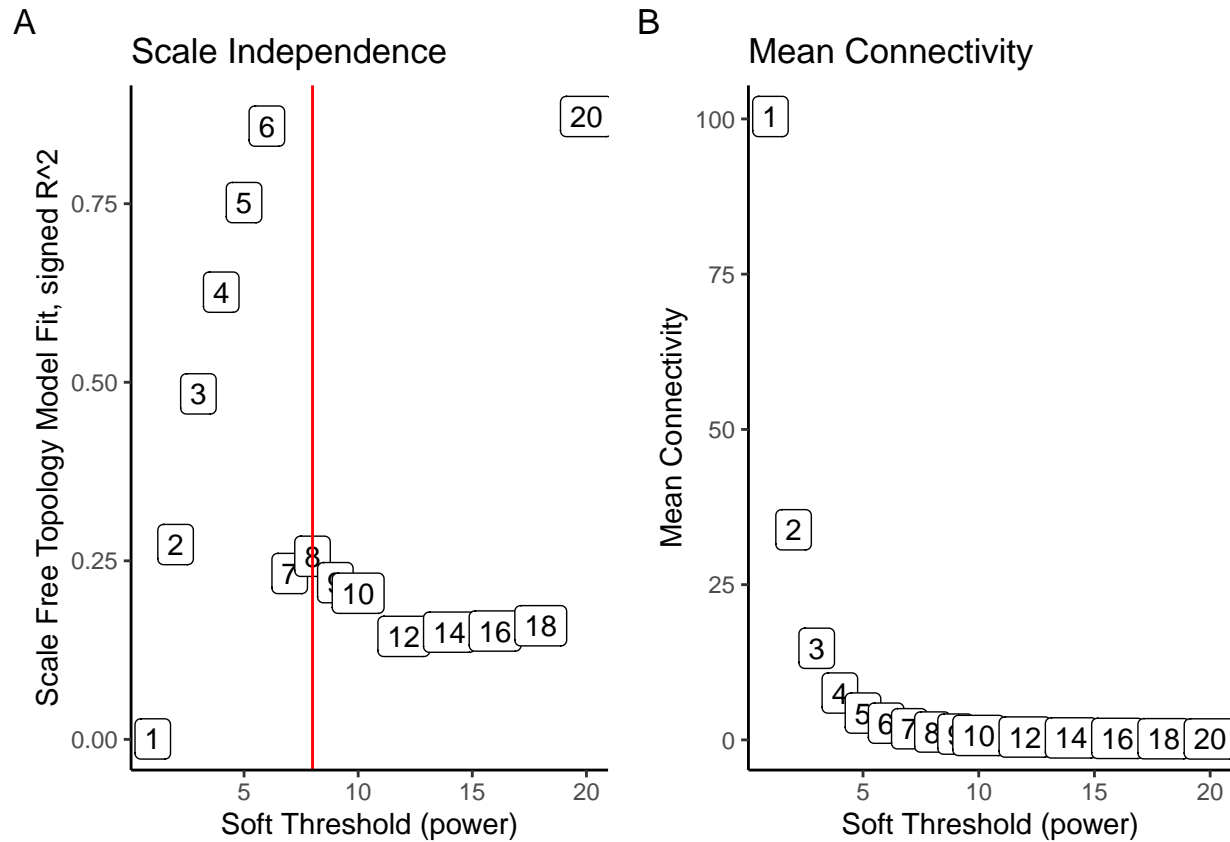
```

# the sign expression returns the vector of signs of estimated slopes, multiplied by the correlation

labs( x="Soft Threshold (power)",
      y="Mean Connectivity", title = "Mean Connectivity", tag = "B")+
theme_classic()

plot_grid(p1,p2,ncol=2)

```



Topological Overlap Measure

The next steps is to calibrate the **Topological Overlap Measure (TOM)**. It is premised over the notion of shared relationships between two clusters.

Now we generate adjacency values and TOM similarity matrices on the basis of selected softpower.

```

softPower = 7;

#calclute the adjacency matrix
adj= adjacency(exprdata,
              type = "unsigned",
              power = softPower);

#turn adjacency matrix into topological overlap to minimize the effects of noise and spurious associati
TOM = TOMsimilarityFromExpr(exprdata,
                           networkType = "unsigned",
                           TOMType = "unsigned",

```



```

        power = softPower);

## TOM calculation: adjacency..
## ..will not use multithreading.
## Fraction of slow calculations: 0.437876
## ..connectivity..
## ..matrix multiplication (system BLAS)..
## ..normalization..
## ..done.

colnames(TOM) = rownames(TOM) = names(exprdata) # same rows as columns, therefore adjacency.
dissTOM=1-TOM # "distance/ dissimilarity = 1 - similarity"

```

Module Detection

```

#hierarchical clustering of the genes based on the TOM dissimilarity measure

library(flashClust)

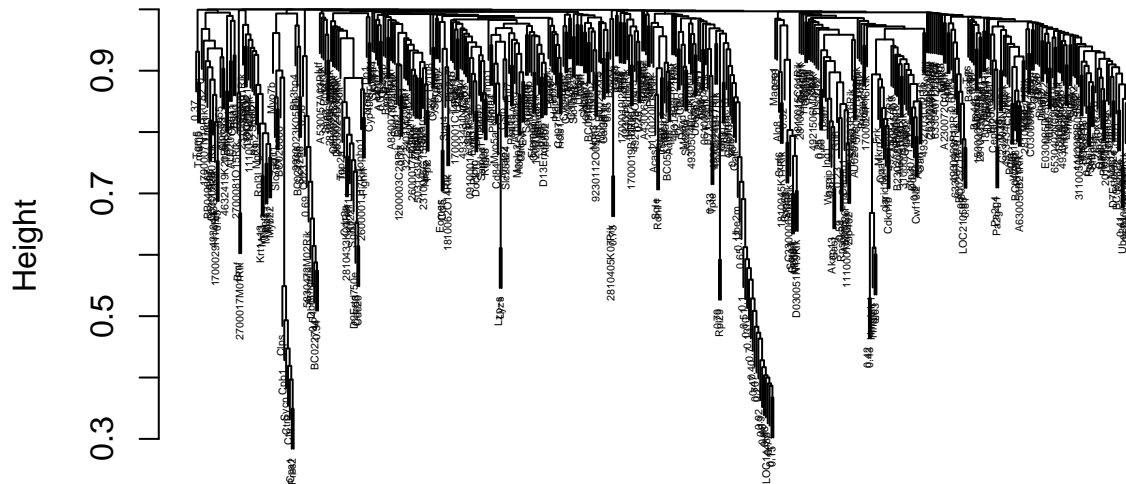
##
## Attaching package: 'flashClust'
## The following object is masked from 'package:fastcluster':
##
##      hclust
## The following object is masked from 'package:stats':
##
##      hclust

gene_tree = flashClust(as.dist(dissTOM),
                       method="average")

#plot the resulting clustering tree (dendrogram)
plot(gene_tree,
     xlab="",
     sub="",
     cex=0.3)

```

Cluster Dendrogram

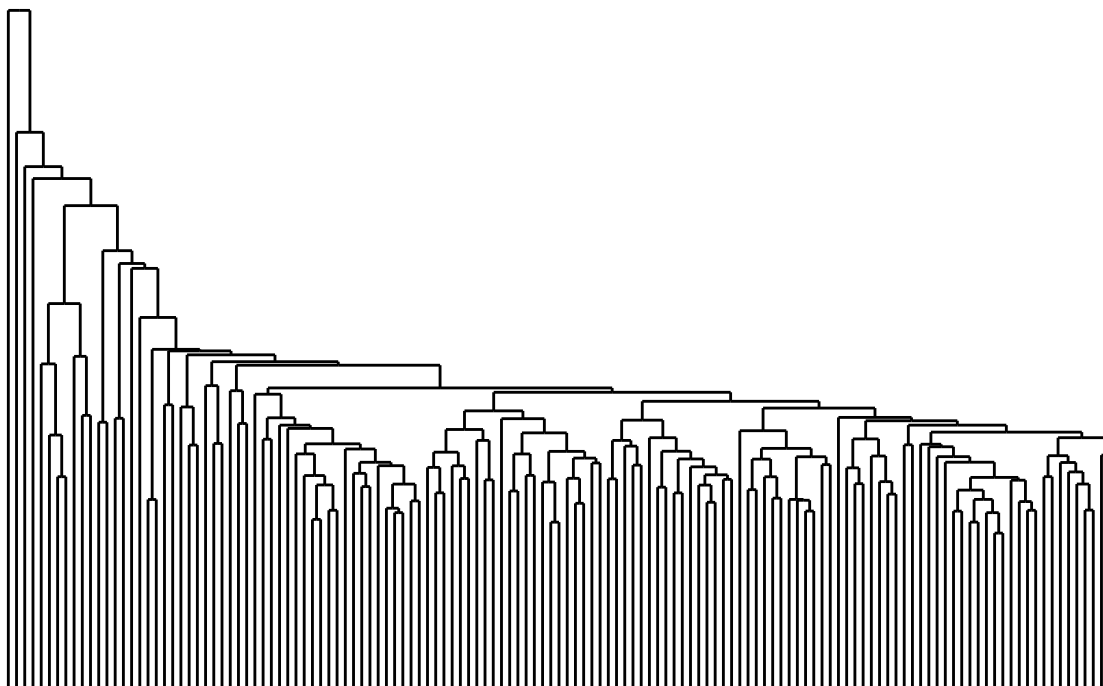


```
sample_tree <- as.dendrogram(hclust(dist(exprdata), method = "average"))

dplot <- gg dendrogram(data= sample_tree, rotate = FALSE)+
  theme_dendro()+
  ggtitle("Sample clustering to detect outliers")+
  theme(plot.title = element_text(hjust = 0.5))+
  xlab("Samples")+
  geom_label()

print(dplot)
```

Sample clustering to detect outliers



```
## Set the minimum module size
```

```
min_mod_size = 20
```

```
## Module identification using dynamic tree cut
```

```
dynamic_mods = cutreeDynamic(dendro = gene_tree,  
                             method="tree",  
                             minClusterSize = min_mod_size)
```

```
dynamic_mods1 = cutreeDynamic(dendro = gene_tree,  
                              distM = dissTOM,  
                              method="hybrid",  
                              deepSplit = 2,  
                              pamRespectsDendro = FALSE,  
                              minClusterSize = min_mod_size)
```

```
## ..cutHeight not given, setting it to 0.996 ==> 99% of the (truncated) height range in dendro.  
## ..done.
```

```
## we could highlight module labels and the size(number of nodes) of each module. Label 0 is reserved for independent nodes
```

```
table(dynamic_mods)
```

```
## dynamic_mods  
## 0 1 2 3 4 5 6 7 8  
## 203 57 53 40 37 31 29 25 25
```

```
table(dynamic_mods1)
```

```
## dynamic_mods1  
## 0 1 2 3 4 5  
## 168 145 72 45 44 26
```

It is always better to have an enhanced, color-coded visualization of modules for precise interpretation.

```
## we can conveniently assign colors to these labels. "Grey" is assigned to label 0.
```

```
dynamic_mod_colors = labels2colors(dynamic_mods)  
table(dynamic_mod_colors)
```

```
## dynamic_mod_colors  
##      black      blue      brown      green      grey      pink      red  
##        25        53        40        31        203        25        29  
## turquoise    yellow  
##         57         37
```

```
dynamic_mod_colors1 = labels2colors(dynamic_mods1)  
table(dynamic_mod_colors1)
```

```
## dynamic_mod_colors1  
##      blue      brown      green      grey turquoise      yellow  
##        72        45        26        168        145        44
```

We can also choose to discard the independent nodes (since they do not belong to any network) and just map the coherent modules.

```

## discard the unassigned genes, and focus on the rest

rest_genes= (dynamic_mod_colors != "grey")
dissTOM1 = 1 - TOMsimilarityFromExpr(exprdata[,rest_genes],
                                     power = softPower)

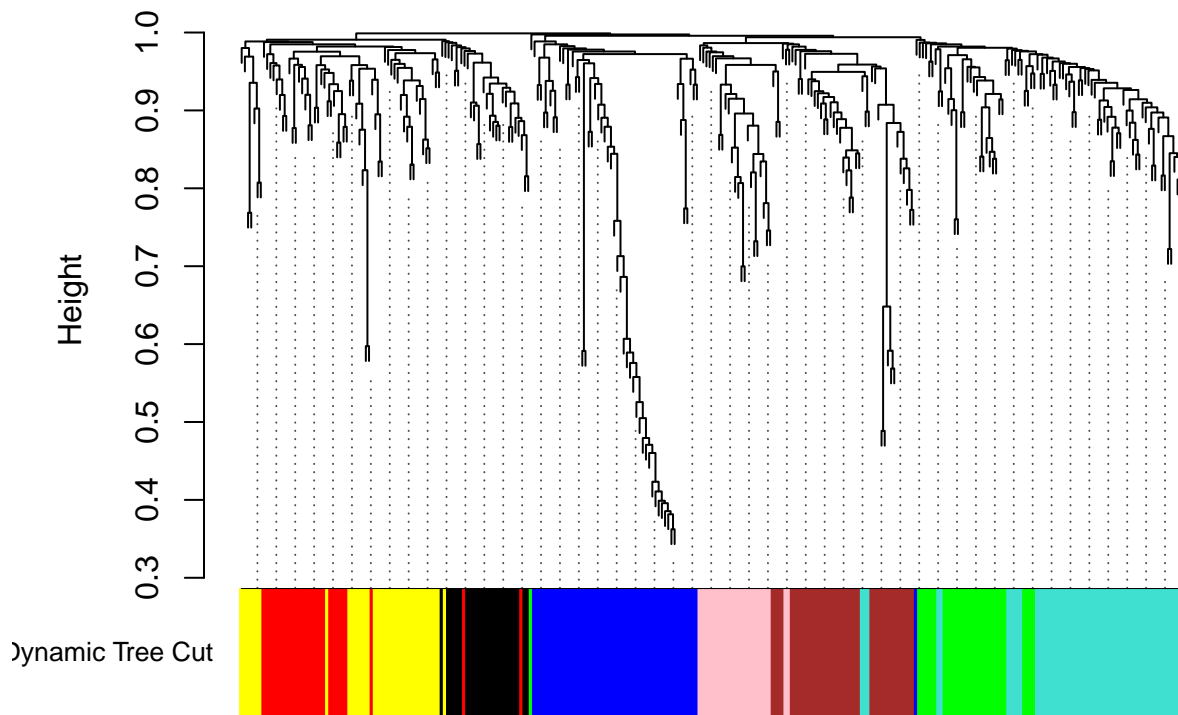
## TOM calculation: adjacency..
## ..will not use multithreading.
## Fraction of slow calculations: 0.431795
## ..connectivity..
## ..matrix multiplication (system BLAS)..
## ..normalization..
## ..done.

## Let's plot the tree now.

colnames(dissTOM1) = rownames(dissTOM1) = names(exprdata[rest_genes])
hier_tree = flashClust(as.dist(dissTOM1), method="average" )
plotDendroAndColors(hier_tree,
                    dynamic_mod_colors[rest_genes],
                    "Dynamic Tree Cut",
                    dendroLabels = FALSE,
                    hang = 0.03,
                    addGuide = TRUE,
                    guideHang = 0.05,
                    main = "Gene dendrogram and module colors")

```

Gene dendrogram and module colors



```

## set the diagonal of the dissimilarity to NA

```

```
diag(dissTOM1) = NA;

## Visualize the TOM plot. Raise the dissimilarity matrix to the power of 4 to bring out the module str

sizeGrWindow(7,7)
TOMplot(dissTOM1, hier_tree, as.character(dynamic_mod_colors[rest_genes]))
```

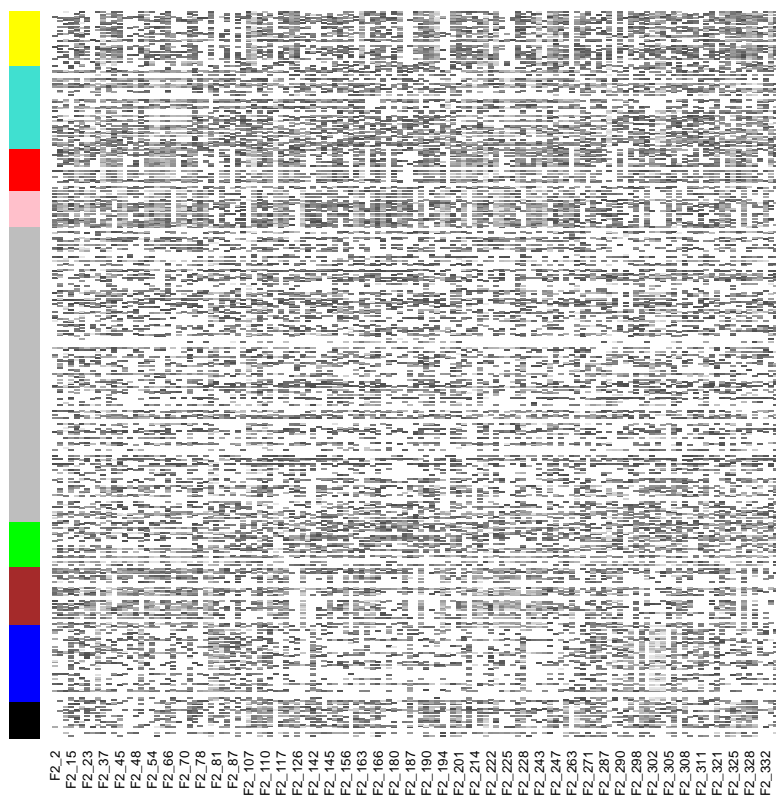
Let us extract the modules now. The modules could also be envisioned as communities or colonies of like entities.

```
module_colors= setdiff(unique(dynamic_mod_colors), "grey")
for (color in module_colors){
  module = names(exprdata)[which(dynamic_mod_colors == color)]
  write.table(module,
              paste0("module_",color, ".txt"),
              sep="\t",
              row.names=FALSE,
              col.names=FALSE,
              quote=FALSE)
}
```

Let us now visualize the expression patterns of these genes, as they're clustered. There ought to be distinct pattern cues inside and outside of a cluster.

```
module.order <- unlist(tapply(1:ncol(exprdata),
                             as.factor(dynamic_mod_colors),
                             I))
m <- t(t(exprdata[,module.order])/apply(exprdata[,module.order],2,max))

heatmap (t(m),
         zlim = c(0,1),
         col = gray.colors(100),
         Rowv = NA,
         Colv = NA,
         labRow = NA,
         scale = "none",
         RowSideColors = dynamic_mod_colors[module.order])
```



We can now look at the module gene listings and try to interpret their functions .. for instance using <http://amigo.geneontology.org/rte>)

Quantify module similarity by eigengene correlation. Eigengenes: Module representatives, putatively (by definition) the rank 1 vectors that span the module expression.

```
MEList <- moduleEigengenes(exprdata, colors = dynamic_mod_colors)
MEs <- MEList$eigengenes
plotEigengeneNetworks(MEs, "",
                      marDendro = c(0,4,1,2),
                      marHeatmap = c(3,4,1,2))
```

