## Lab 8 R Source Code

## Code:

```
1 #
2 # Shawn Schwartz, 2019
_3 # EEB C174 UCLA Spring 2019
 # Lab 8 HW - BAMM and FiSSE
  #
6
  rm(list=ls())
  #imports
  library (BAMMtools)
  library (coda)
12
  cwd <- "~/Developer/EEB-C174-Labs/Lab8/cetacean BAMM"
13
  setwd (cwd)
14
15
  tree <- read.tree("whaleTree.tre")
16
  tree <- ladderize (tree)
17
  plot(x = tree, cex = 0.2, no.margin = T)
18
  axisPhylo()
19
20
  #check bamm assumptions
21
  is . ultrametric (tree)
  is.binary(tree)
23
  \min(\text{tree\$edge.length}) > 0
24
25
  #generate bamm control file
  # estimate priors and create control files
  priors <- setBAMMpriors(phy = tree, traits = "whaleSize.txt",
     outfile = NULL)
  generateControlFile(file = "cetacean bamm controlfile.txt", type =
30
      "trait", params = list(
     treefile = "whaleTree.tre";
31
     traitfile = "whaleSize.txt",
32
     seed = sample(1:1000000, 1),
33
     overwrite = 0,
34
     expectedNumberOfShifts = "2",
35
     betaInitPrior = as.numeric(priors["betaInitPrior"]),
36
```

```
betaShiftPrior = as.numeric(priors["betaShiftPrior"]),
37
    numberOfGenerations = "120000000",
38
    mcmcWriteFreq = "30000",
39
    eventDataWriteFreq = "30000",
40
    printFreq = "15000",
41
    acceptanceResetFreq = "30000",
42
    outName = "cetacean bamm homework",
43
    numberOfChains = "8"
44
  ))
45
46
  edata <- getEventData(tree, eventdata = "2_expected_shifts/
     cetacean bamm homework event data.txt", burnin = 0.1, type = "
     trait")
  summary (edata)
48
49
  ##### Assess MCMC convergence #####
50
  mcmcout <- read.csv("2 expected shifts/cetacean bamm homework mcmc
     out.txt")
  plot (mcmcout$logLik ~ mcmcout$generation)
52
53
  ##### Test for convergence of the MCMC chains with the coda package
      for R ####
  burnstart <- floor (0.1 * nrow (mcmcout))
  postburn <- mcmcout[burnstart:nrow(mcmcout), ]</pre>
  effectiveSize(postburn$N shifts)
57
  effectiveSize (postburn$logLik)
59
  ##### Analysis of rate shifts #####
  plotPrior("2_expected_shifts/cetacean_bamm homework mcmc out.txt",
61
      expectedNumberOfShifts = 2
  s <- plot.bammdata(edata, labels = T, font = 3, cex = 0.1,
62
     logcolor = T)
  title (main = "Mean phenotypic rate", sub = "time before present")
63
  addBAMMlegend(s, location = "left", nTicks = 1)
  axisPhylo()
65
66
  css <- credibleShiftSet (edata, expectedNumberOfShifts = 30,
67
     threshold = 5, set.limit = 0.95)
  css$number.distinct
68
  summary (css)
69
70
  sss <- plot.credibleshiftset(css, border = F, logcolor = T)
```

```
72
   plot.new()
   best <- getBestShiftConfiguration(edata, expectedNumberOfShifts =
75
   ss <- plot.bammdata(best, labels = T, font = 3, cex = 0.1,
76
      logcolor = T
   title (main = "Best shift configuration", sub = "time before
      present")
   addBAMMlegend(ss, location = "left", nTicks = 1)
   addBAMMshifts(best, cex = 3, pch = 1)
   axisPhylo()
80
81
  #### scaled branch lengths #####
82
   par(font = 1)
83
   marg probs <- marginalShiftProbsTree(edata)
84
   plot.phylo(marg probs, cex = 0.45, no.margin = T)
   title (sub = "Marginal shift probability")
86
   add.scale.bar(x = 0.5, y = 0.5, font = 1)
87
88
89
  #### rate through time analysis #####
   par(font = 1)
91
   plotRateThroughTime(edata,
                         ratetype = "auto",
93
                         avgCol = "black", intervalCol = "gray80",
94
                            intervals = c(0.05, 0.95), opacity = 1)
95
  #### FiSSE ####
96
   library (phangorn)
   library (diversitree)
98
   library (ape)
99
100
   source ("trait Dependent functions .R")
101
102
   diet <- read.csv(file = "whaleDiet.csv", stringsAsFactors = F)
103
   generalist <- diet$generalist
104
   names (generalist) <- diet $ species
105
   generalist <- generalist [tree$tip.label]
106
   colvec <- rep("white", length(generalist))</pre>
107
   colvec [generalist == 1] <- "black"
   plot.phylo(tree, show.tip.label = F, cex = 0.4, no.margin = T)
```

```
tiplabels (pch = 21, bg = colvec, cex = 0.8)
   add.scale.bar(x = 0.5, y = 0.5, font = 1)
   title (sub = "time before present (my)")
113
   res <- FISSE.binary(tree, generalist)
   res
115
116
  # two-tailed pvalue is obtained as
117
   pval \ 2tailed \leftarrow min(res pval, 1 - res pval) * 2
   pval 2 tailed
119
120
  #two planel plot for lab report
121
   par(mfrow = c(1,2))
   par(mar=c(5, 1.0, 2, 0.0))
  #plotPrior("2 expected shifts/cetacean bamm homework mcmc out.txt
      ", expectedNumberOfShifts = 2)
  s <- plot.bammdata(edata, labels = T, font = 3, cex = 0.5,
125
      logcolor = T)
   title (main = "Mean phenotypic rate", sub = "time before present (
126
   addBAMMlegend(s, location = "left", nTicks = 1)
127
   axisPhylo()
129
   par(font = 1)
   marg probs <- marginalShiftProbsTree(edata)
   plot.phylo(marg probs, cex = 0.5, no.margin = F)
132
   title (main = "Marginal shift probability")
133
   add.scale.bar(x = 0.75, y = 0.5, font = 1)
134
135
136
   library(geiger)
137
   library (apTreeshape)
138
   library (phytools)
139
140
   ltt.plot(tree, \log = "y")
141
   lines(c(0, 1), c(log(2), log(100)), lty = "dashed", lwd = 2, col =
142
       "red")
  gammaStat(tree)
```