

Metagenome Report

Project: ibdmdb_test

Date: 07/31/2020

Contents

| | |
|--|-----------|
| Introduction | 1 |
| Quality Control | 2 |
| DNA Samples Quality Control | 3 |
| DNA Samples Tables of Filtered Reads | 3 |
| DNA Samples Plots of Filtered Reads | 6 |
| Taxonomic Profiling of Metagenomic Reads | 7 |
| Taxonomic Count Table | 8 |
| Ordination | 9 |
| Species | 9 |
| Genera | 12 |
| Heatmaps | 14 |
| Species | 15 |
| Genera | 17 |
| Barplot | 19 |
| Species | 19 |
| Genera | 22 |
| Functional Profiling of Metagenomic Reads | 25 |
| Pathway and ECs Abundance | 25 |
| Features | 31 |

Introduction

The data was run through the standard workflow for whole metagenome shotgun sequencing.

Quality Control

This report section contains information about the quality control processing for all 6 paired-end fastq input files. These files were run through the [KneadData](#) QC pipeline. Reads were first trimmed then filtered against contaminate reference databases: rRNA, hg37dec_v0.1 and mRNA. Reads were filtered sequentially with those reads passing the first filtering step used as input to the next filtering step. This chain of filtering removes reads from all references in serial. Data is organized by paired and orphan reads. When one read in a pair passes a filtering step and the other does not the surviving read is an orphan. The tables and plots are annotated as follows:

- raw : Untouched fastq reads.
- trim : Number of reads remaining after trimming bases with Phred score < 20. If the trimmed reads is < 50% of original length then it is removed altogether.
- rRNA : Number of reads after depleting against reference database rRNA. The SILVA (rRNA) database is used to remove small and large subunit ribosomal RNA.
- hg37dec_v0.1 : Number of reads after depleting against reference database rRNA and hg37dec_v0.1.
- mRNA : Number of reads after depleting against reference database rRNA and hg37dec_v0.1 and mRNA. The human transcriptome (hg38 mRNA) database is used to remove reads originating from host gene isoforms.

DNA Samples Quality Control

DNA Samples Tables of Filtered Reads

| DNA Paired end reads | | | | | |
|----------------------|------------|------------|------------|--------------|------------|
| | Raw | Trim | rRNA | hg37dec_v0.1 | mRNA |
| CSM9X23N | 10,529,590 | 10,529,590 | 10,482,541 | 10,429,946 | 10,482,541 |
| HSM6XRQY | 8,655,985 | 8,655,985 | 8,622,357 | 8,573,730 | 8,622,359 |
| HSM7J4NY | 7,241,429 | 7,241,425 | 7,192,958 | 5,764,053 | 7,192,959 |
| HSM33KE | 10,869,261 | 10,869,261 | 10,831,951 | 10,770,433 | 10,831,951 |
| HSM33OT | 8,036,913 | 8,036,911 | 7,863,474 | 5,584,651 | 7,863,540 |
| MSM6J2QD | 3,845,089 | 3,845,087 | 3,783,951 | 1,900,300 | 3,783,951 |

A data file exists of this table: [qc_counts_pairs_table.tsv](#)

| DNA Orphan reads | | | | | | |
|------------------|--------------|--------------|----------------------|----------------------|--------------|--------------|
| | rRNA orphan1 | rRNA orphan2 | hg37dec_v0.1 orphan1 | hg37dec_v0.1 orphan2 | mRNA orphan1 | mRNA orphan2 |
| CSM9X23N | 16,559 | 16,158 | 19,029 | 18,559 | 16,560 | 16,158 |
| #SM6XRO | 9,642 | 9,009 | 11,519 | 11,078 | 9,642 | 9,009 |
| HSM7J4NY | 22,026 | 21,765 | 775,171 | 46,603 | 22,027 | 21,764 |
| H5MA33KE | 12,675 | 12,939 | 15,492 | 15,755 | 12,677 | 12,939 |
| #SMA33OT | 10,547 | 9,595 | 143,022 | 38,658 | 10,546 | 9,555 |
| #SM6J2QD | 40,396 | 17,525 | 979,627 | 31,368 | 40,396 | 17,525 |

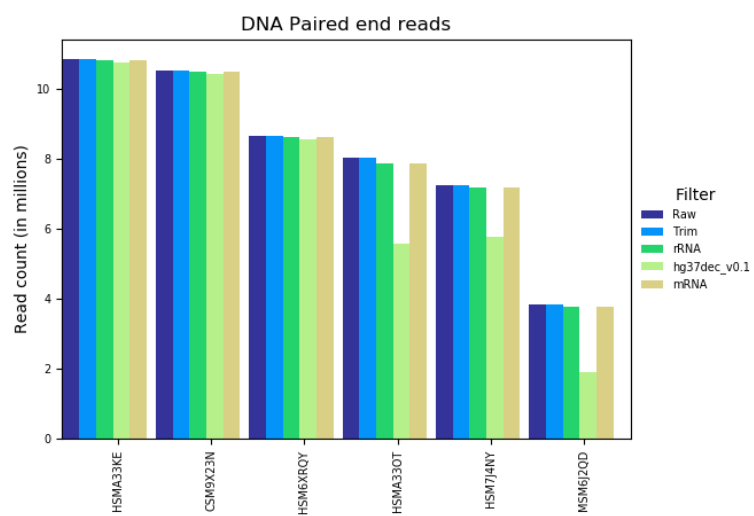
A data file exists of this table: [qc_counts_orphans_table.tsv](#)

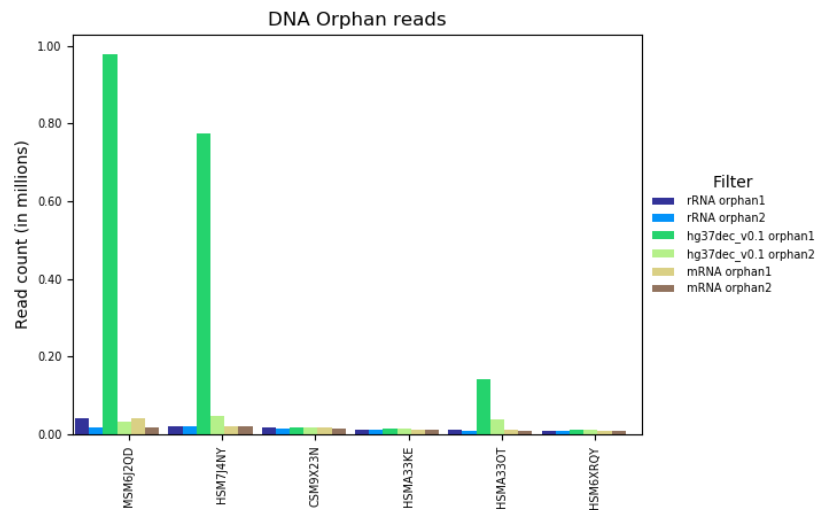
| DNA microbial read proportion | | | | | | |
|-------------------------------|-------------|------------|---------------------|--------------------|-------------|------------|
| | rRNA / Trim | rRNA / Raw | hg37dec_v0.1 / Trim | hg37dec_v0.1 / Raw | mRNA / Trim | mRNA / Raw |
| CSM9X23N | 0.99566 | 0.99720 | 0.99065 | 0.99219 | 0.99554 | 0.99709 |
| HSM6XRQY | 0.99623 | 0.99730 | 0.99059 | 0.99166 | 0.99612 | 0.99719 |
| HSM7J4NY | 1.04517 | 1.04833 | 0.79829 | 0.80070 | 0.99333 | 0.99633 |
| HMA33KE | 0.99670 | 0.99788 | 0.99106 | 0.99223 | 0.99657 | 0.99775 |
| HMA33OT | 0.98668 | 0.98791 | 0.69700 | 0.69788 | 0.97846 | 0.97968 |
| MSM6J2QD | 1.10544 | 1.11377 | 0.49683 | 0.50057 | 0.98422 | 0.99163 |

Proportion of reads remaining after removing host reads relative to the number of: i) quality-trimmed reads, and ii) raw unfiltered reads.

A data file exists of this table: [microbial_counts_table.tsv](#)

DNA Samples Plots of Filtered Reads





Taxonomic Profiling of Metagenomic Reads

This report section contains information about the taxonomy for all DNA samples. These samples were run through [MetaPhlAn2](#).

Taxonomic abundances are passed through a basic filter requiring each species or genus to have at least 0.01 % abundance in at least 10 % of all samples.

A total of 88 species and 51 genera were identified. After basic filtering 79 species and 47 genera remained.

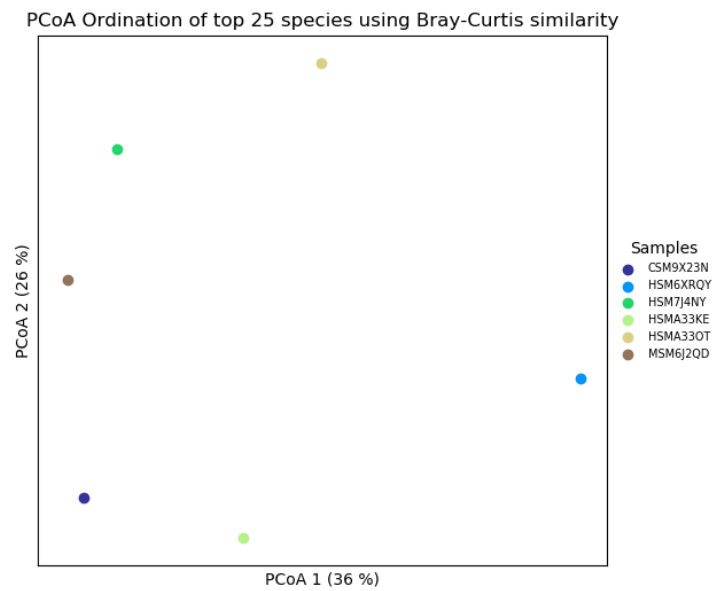
Taxonomic Count Table

| Total taxa per sample | | | | |
|-----------------------|---------|------------------|--------|-----------------|
| | Species | Species filtered | Genera | Genera filtered |
| CSM9X23N | 34 | 33 | 19 | 19 |
| HSM6XRQY | 33 | 30 | 22 | 21 |
| HSM7J4NY | 11 | 11 | 6 | 6 |
| HSM33KE | 58 | 52 | 37 | 34 |
| HSM33OT | 26 | 26 | 16 | 16 |
| MSM6J2QD | 10 | 10 | 6 | 6 |

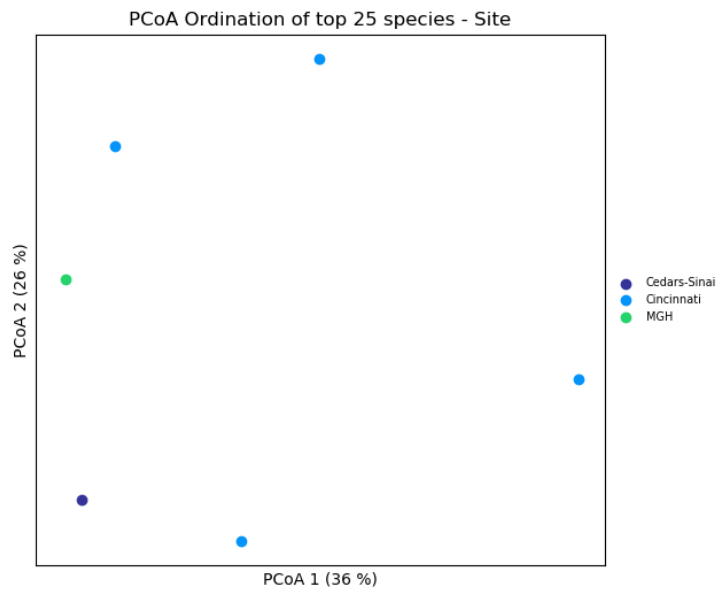
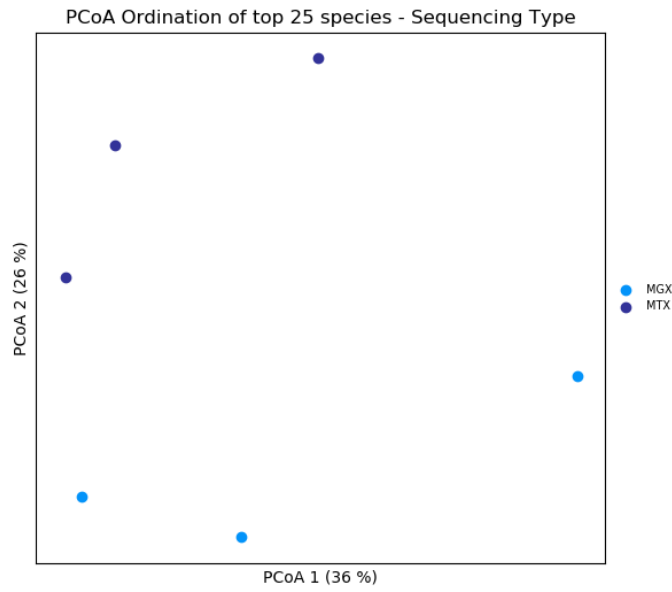
A data file exists of this table: [taxa_counts_table.tsv](#)

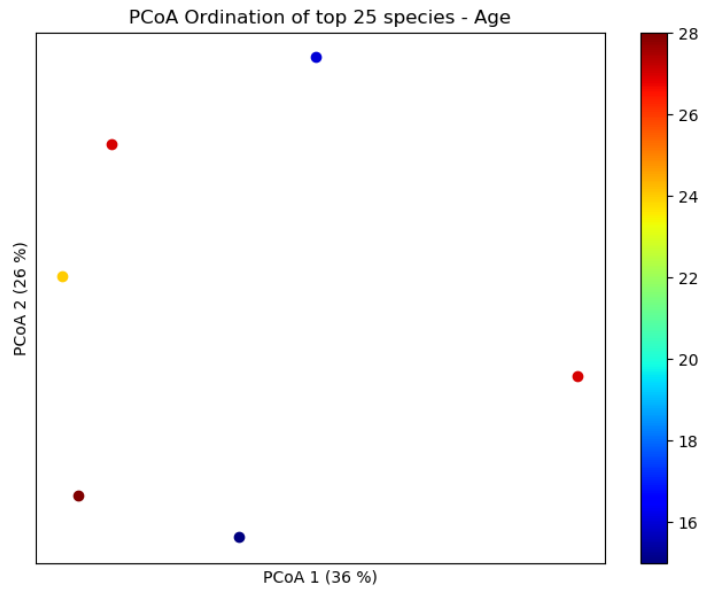
Ordination

Species

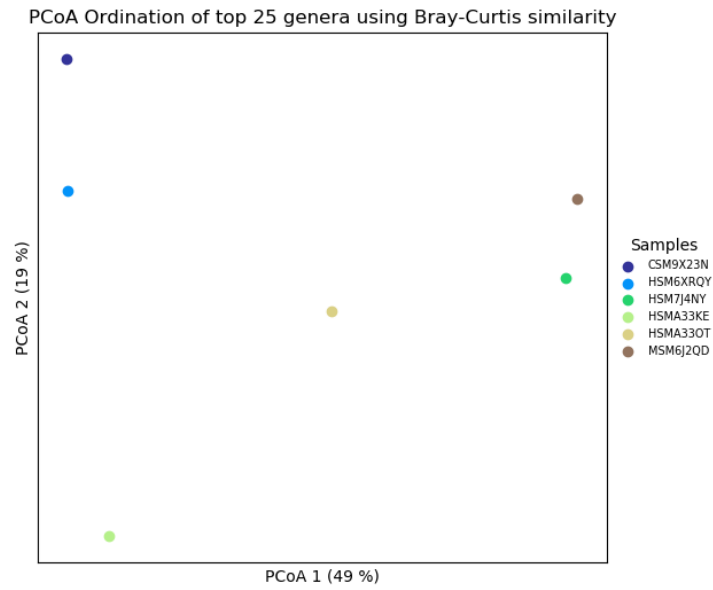


Principal coordinate analysis of variance among samples, based on Bray-Curtis dissimilarities between species profiles of samples. Numbers in parenthesis on each axis represent the amount of variance explained by that axis.

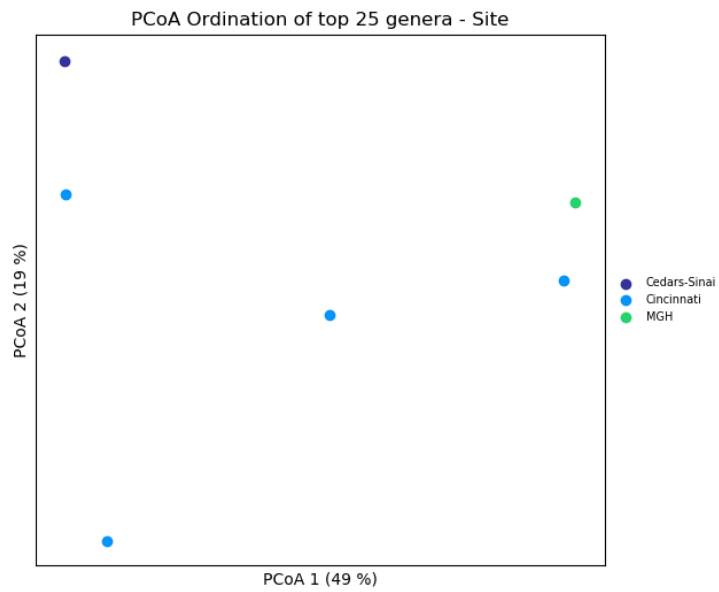
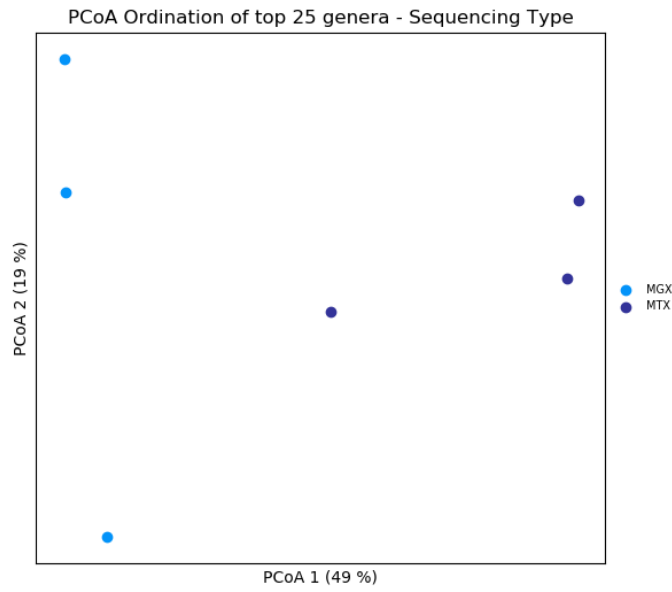


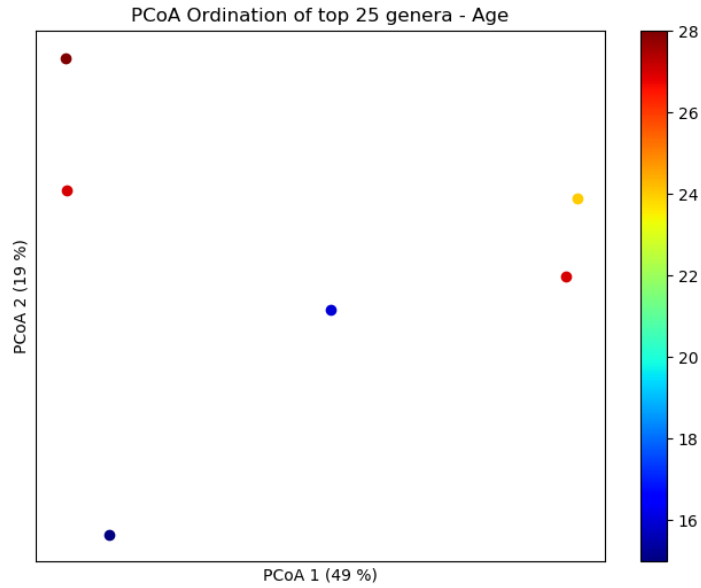


Genera



Principal coordinate analysis of variance among samples, based on Bray-Curtis dissimilarities between genera profiles of samples. Numbers in parenthesis on each axis represent the amount of variance explained by that axis.



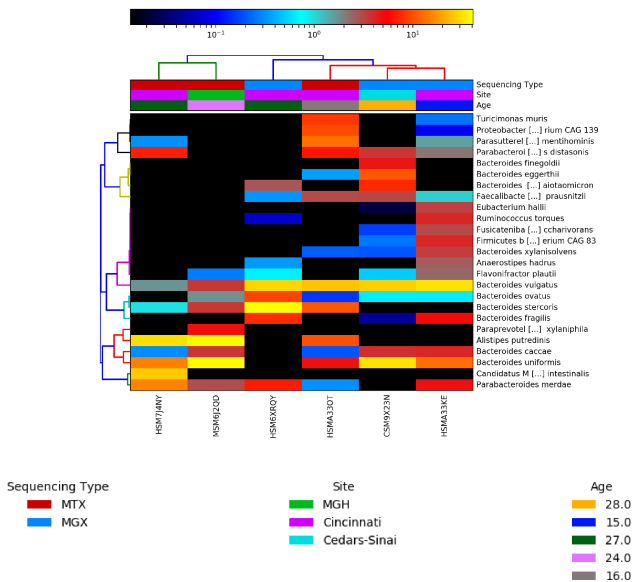


Heatmaps

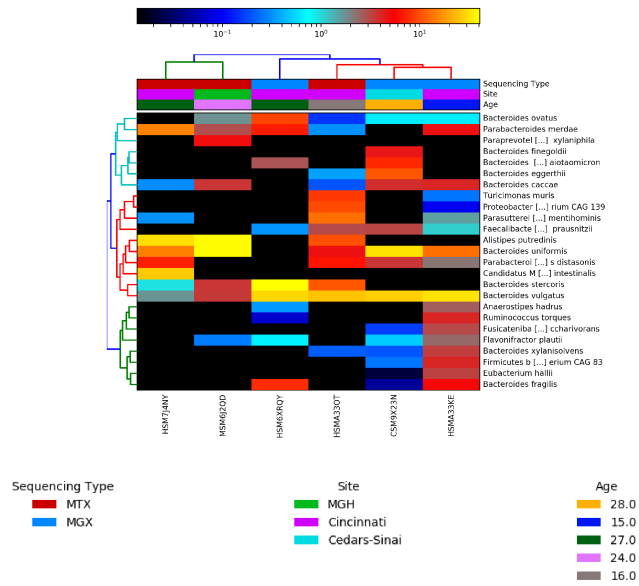
Hierarchical clustering of samples and species and genera, using top 25 species and genera with highest mean relative abundance among samples. The 'average linkage' clustering on the Euclidean distance metric was used to cluster samples. The species and genera dendrogram is based on pairwise (Spearman and Bray-Curtis) correlation between pathways. Samples are columns and pathway are rows. The heatmaps were generated with [Hclust2](#).

Species

Top 25 species by average abundance (Spearman)

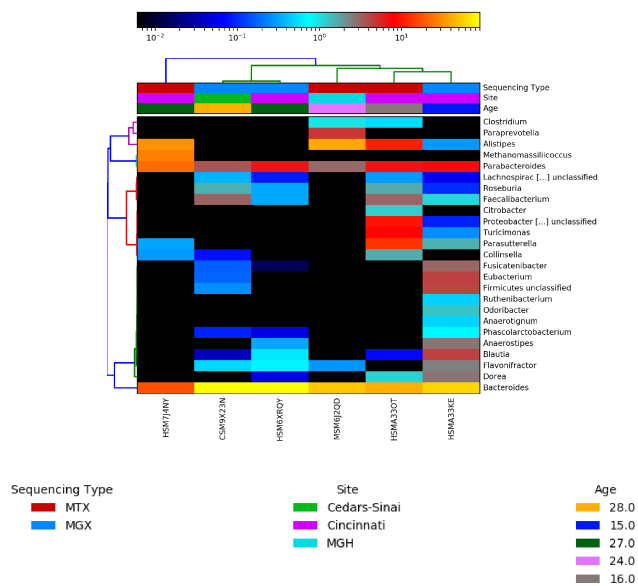


Top 25 species by average abundance (Bray-Curtis)

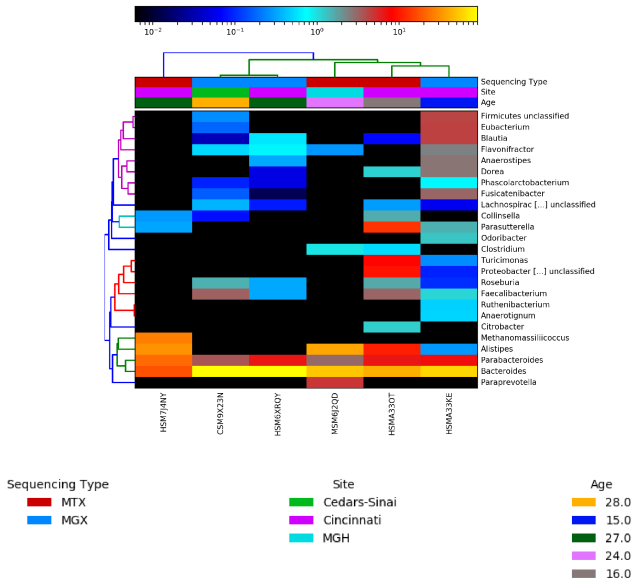


Genera

Top 25 genera by average abundance (Spearman)

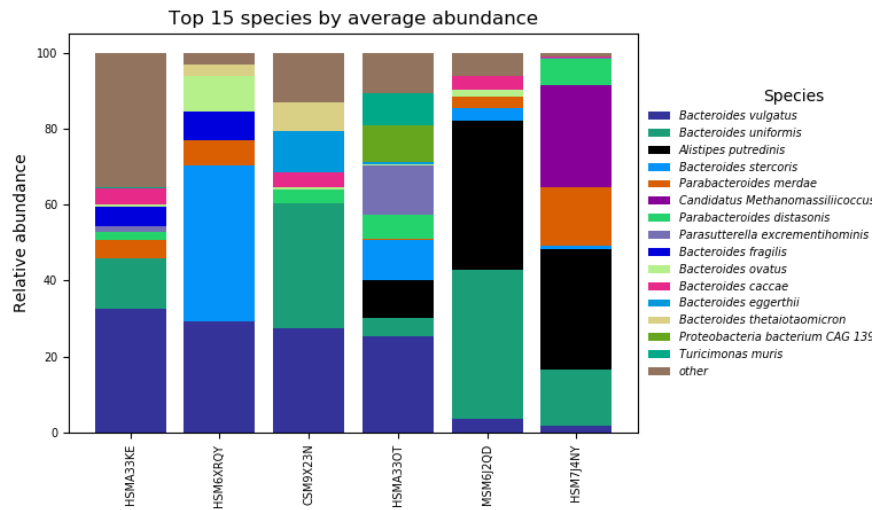


Top 25 genera by average abundance (Bray-Curtis)



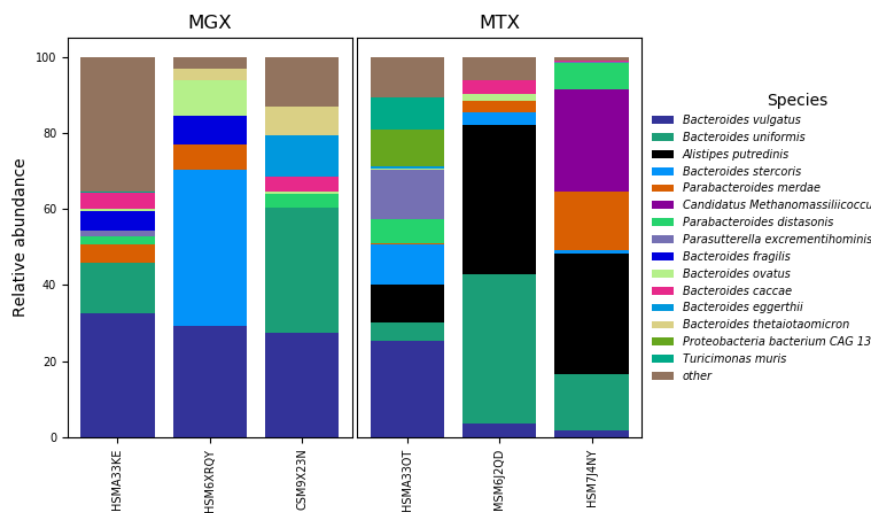
Barplot

Species

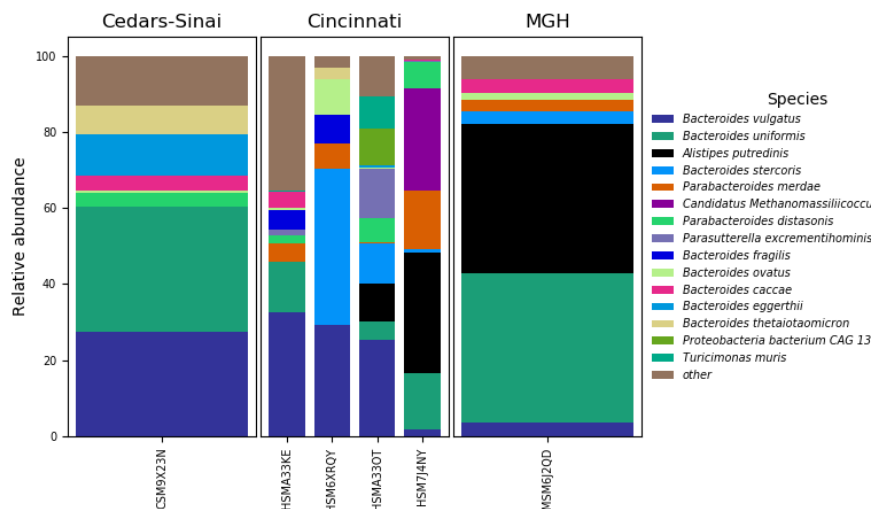


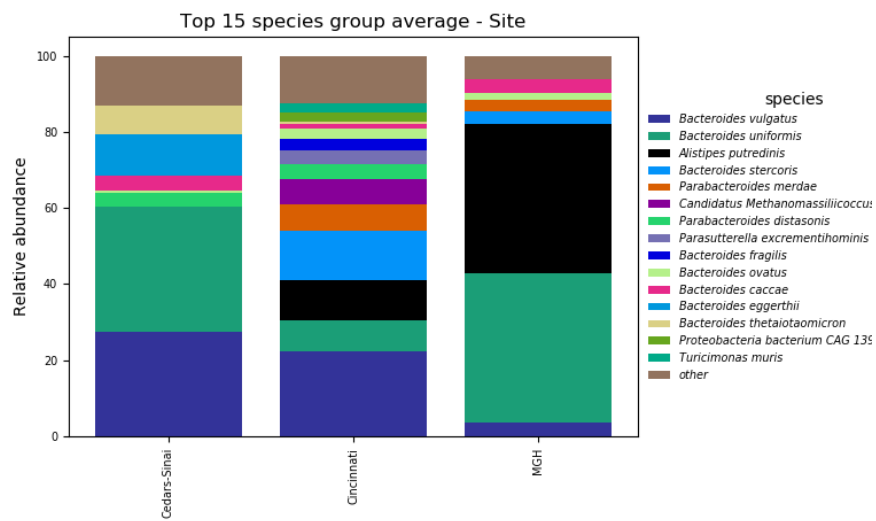
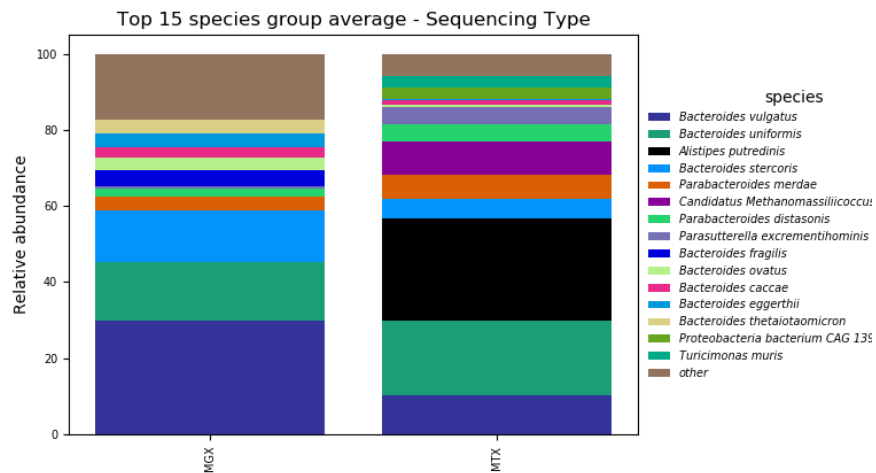
Stacked barplot of 15 most abundant species among samples. Samples in the plot were sorted on the species with the highest mean abundances among samples, in decreasing order.

Top 15 species by average abundance - Sequencing Type



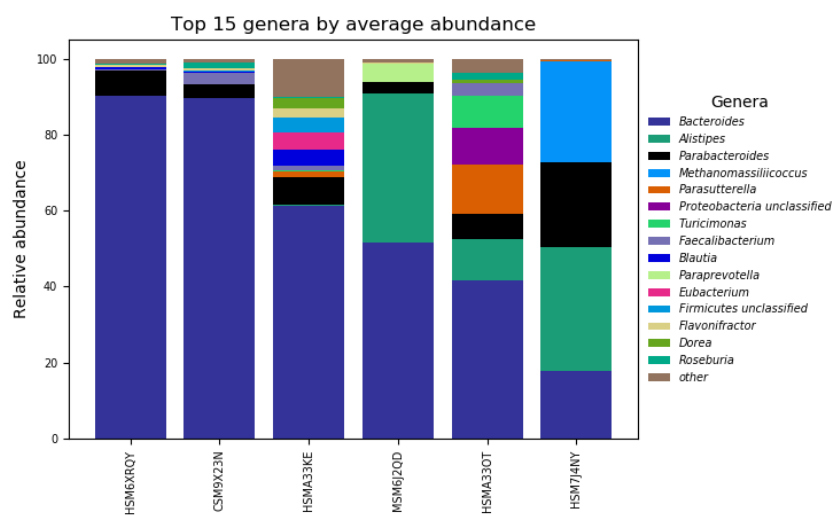
Top 15 species by average abundance - Site Cedars-Sinai to MGH





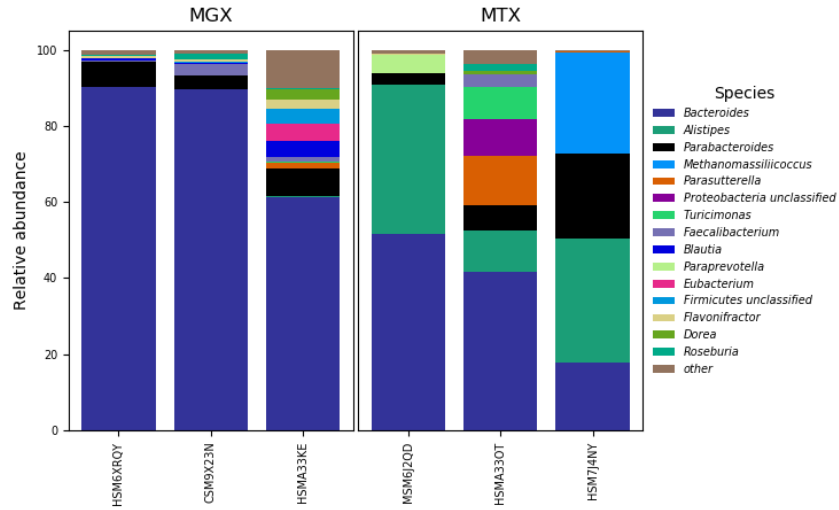
Stacked barplot of species average abundance grouped by metadata.

Genera

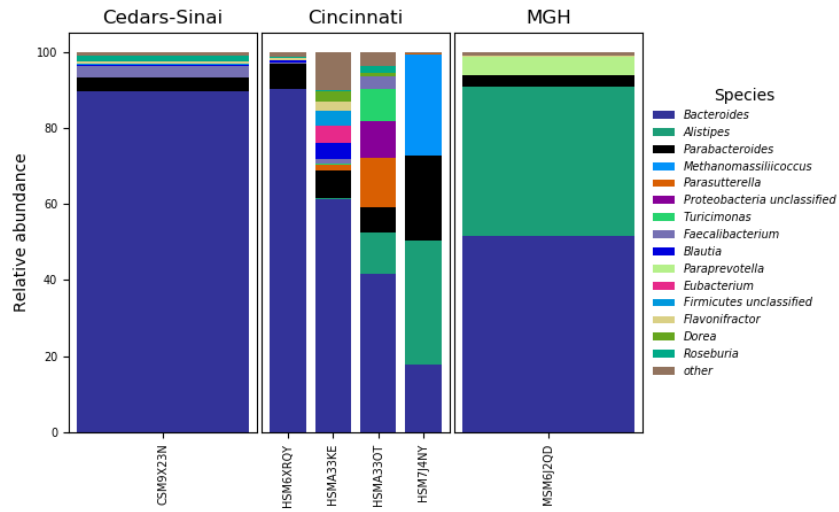


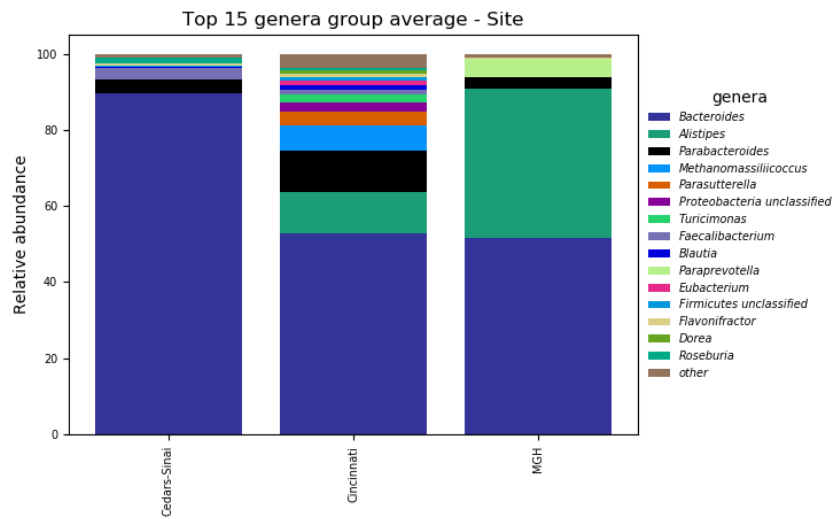
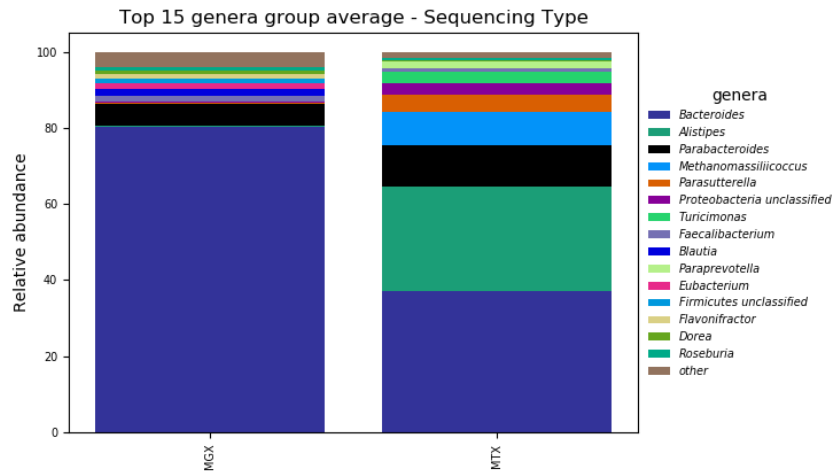
Stacked barplot of 15 most abundant genera among samples. Samples in the plot were sorted on the genera with the highest mean abundances among samples, in decreasing order.

Top 15 genera by average abundance - Sequencing Type



Top 15 genera by average abundance - Site Cedars-Sinai to MGH





Stacked barplot of genera average abundance grouped by metadata.

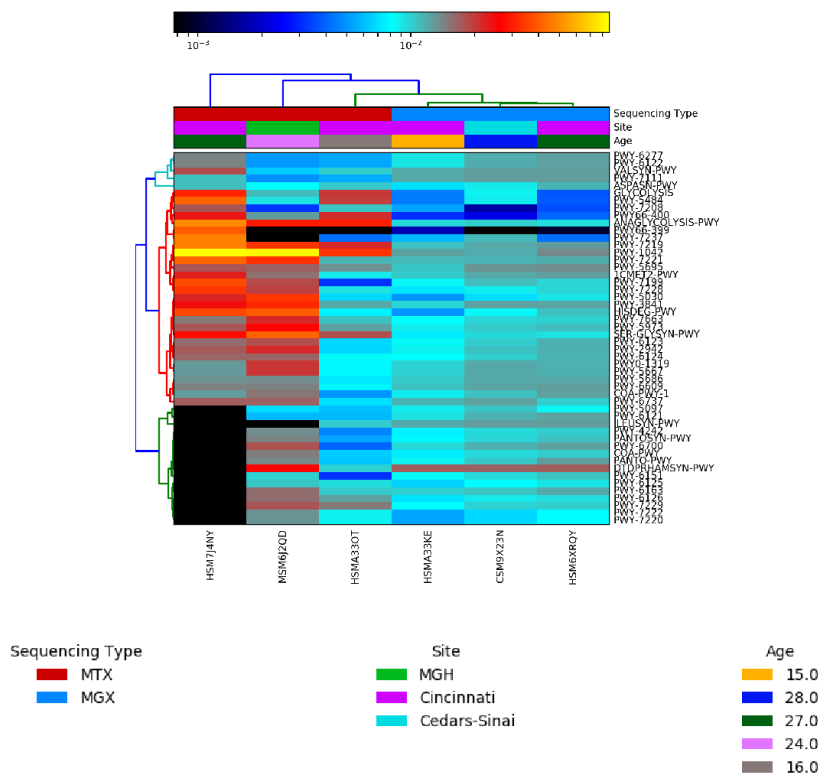
Functional Profiling of Metagenomic Reads

This report section contains preliminary exploratory figures that summarize HUMAnN2 functional profiling of all samples. HUMAnN2 performs species-specific and species-agnostic quantification of gene families, EC enzyme modules, and pathways, using the UniRef and MetaCyc databases. For more information on functional profiling and the databases used, see websites for [HUMAnN2](#), [UniRef](#), and [MetaCyc](#).

Pathway and ECs Abundance

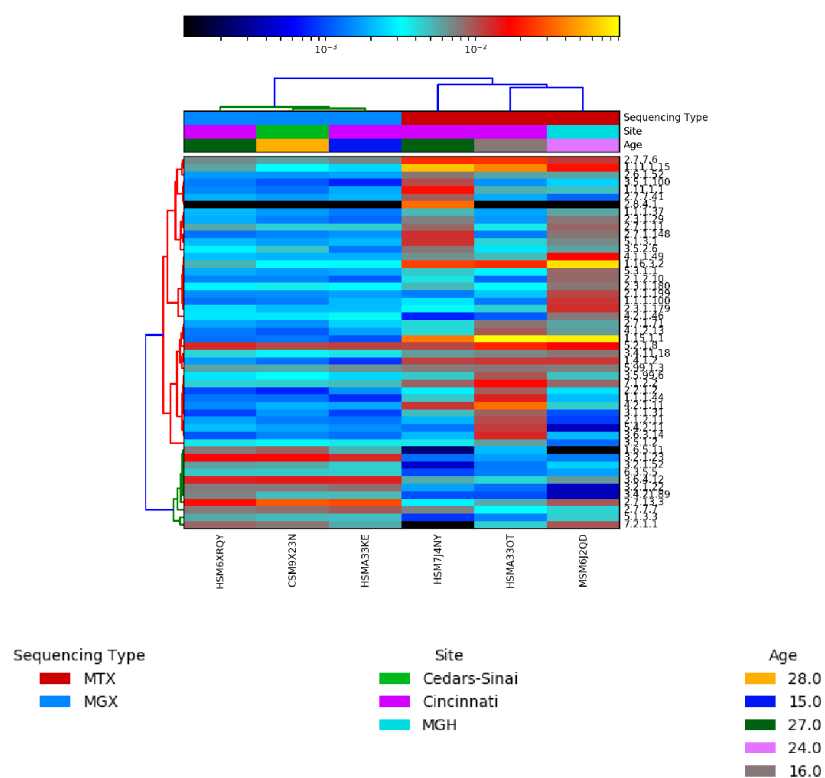
Hierarchical clustering of samples and pathways, using top 50 pathways with highest mean relative abundance among samples. The ‘average linkage’ clustering on the Euclidean distance metric was used to cluster samples. The pathways dendrogram is based on pairwise (Spearman) correlation between pathways. Samples are columns and pathway are rows. The heatmaps were generated with [Hclust2](#).

Top 50 pathways by average abundance

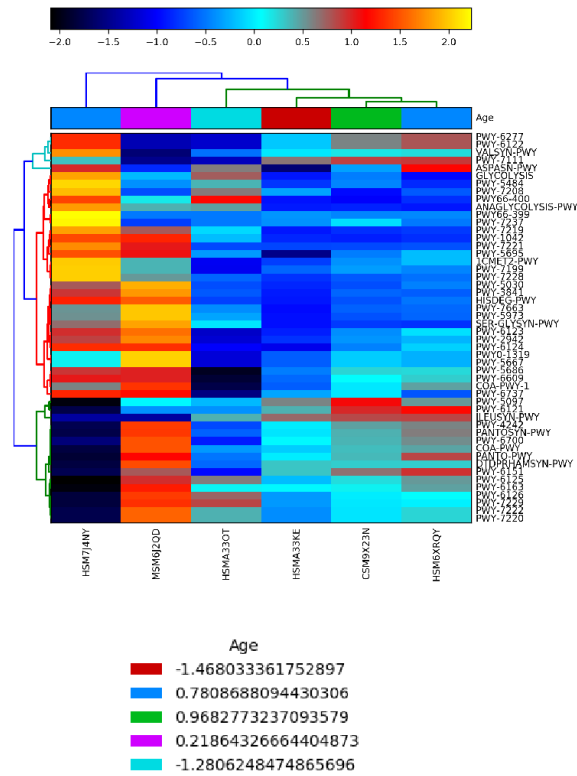


Abundances were log10 transformed prior to clustering. The color bar represents relative abundances on a log10 scale.

Top 50 ecs by average abundance

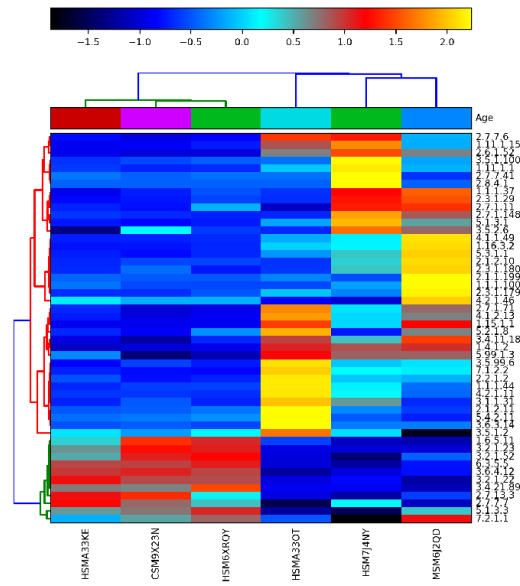


Top 50 pathways by average abundance



Abundances were z-score transformed prior to clustering. The color bar represents relative abundances on a z-score scale.

Top 50 ecs by average abundance



Age

- 1.468033361752897
- 0.21864326664404873
- 0.7808688094430306
- 0.9682773237093579
- 1.2806248474865696

Top 50 pathways by average abundance (partial table)

| | Average | Variance |
|--|---------|----------|
| PWY-1042: glycolysis IV (plant cytosol) | 0.0399 | 0.00102 |
| ANAGLYCOLYSIS-PWY: glycolysis III (from glucose) | 0.0232 | 0.000225 |
| PWY-7219: adenosine ribonucleotides de novo biosynthesis | 0.0231 | 0.00018 |
| PWY-7221: guanosine ribonucleotides de novo biosynthesis | 0.0201 | 0.000151 |
| SER-GLYSYN-PWY: superpathway of L-serine and glycine biosynthesis I | 0.019 | 0.000147 |
| HISDEG-PWY: L-histidine degradation I | 0.0181 | 0.000209 |
| PWY-3841: folate transformations II | 0.0172 | 6.59e-05 |
| PWY-7228: superpathway of guanosine nucleotides de novo biosynthesis I | 0.0147 | 8.82e-05 |
| PWY-7199: pyrimidine deoxyribonucleosides salvage | 0.0147 | 0.00012 |
| DTDPRHAMSYN-PWY: dTDP-L-rhamnose biosynthesis I | 0.0147 | 6.58e-05 |
| PWY-5484: glycolysis II (from fructose 6-phosphate) | 0.0146 | 0.000173 |
| PWY-5973: cis-vaccenate biosynthesis | 0.0146 | 3.66e-05 |
| PWY-5695: urate biosynthesis/inosine 5'-phosphate degradation | 0.0143 | 5.81e-06 |
| PWY-5030: L-histidine degradation III | 0.014 | 0.000111 |
| 1CMET2-PWY: N10-formyl-tetrahydrofolate biosynthesis | 0.0138 | 2.12e-05 |
| GLYCOLYSIS: glycolysis I (from glucose 6-phosphate) | 0.0132 | 9.25e-05 |
| PWY-2942: L-lysine biosynthesis III | 0.0129 | 2.74e-05 |
| PWY-7663: gondoate biosynthesis (anaerobic) | 0.0127 | 2.11e-05 |
| PWY-5667: CDP-diacylglycerol biosynthesis I | 0.0126 | 1.62e-05 |
| PWY0-1319: CDP-diacylglycerol biosynthesis II | 0.0126 | 1.62e-05 |

The table is too large to include the full table in this document. A partial table is shown which includes only 20 rows. Please see the data file for the full table: [top_average_pathways_names.tsv](#)

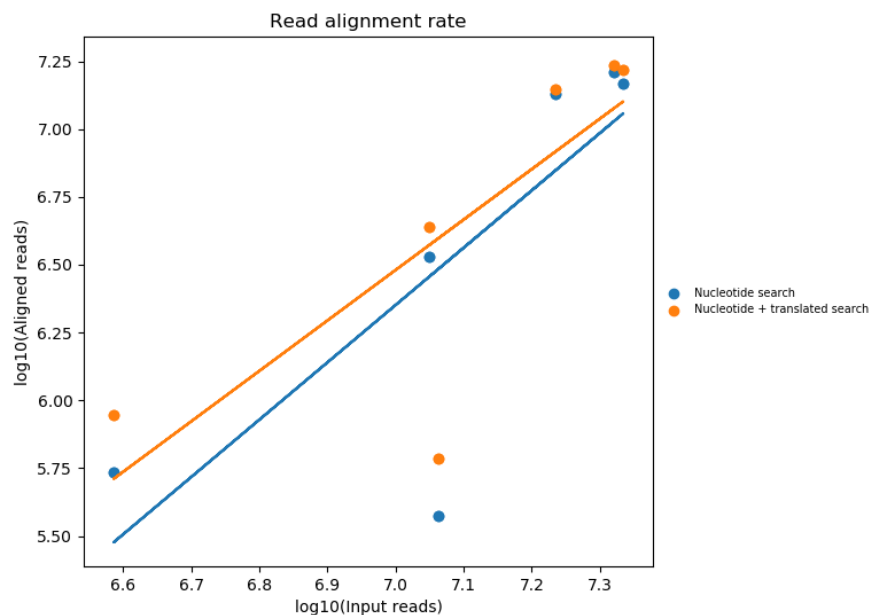
Detailed functions of the top 3 pathways can be found on the following MetaCyc pages:

- * [PWY-1042: glycolysis IV \(plant cytosol\)](#)
- * [ANAGLYCOLYSIS-PWY: glycolysis III \(from glucose\)](#)
- * [PWY-7219: adenosine ribonucleotides de novo biosynthesis](#)

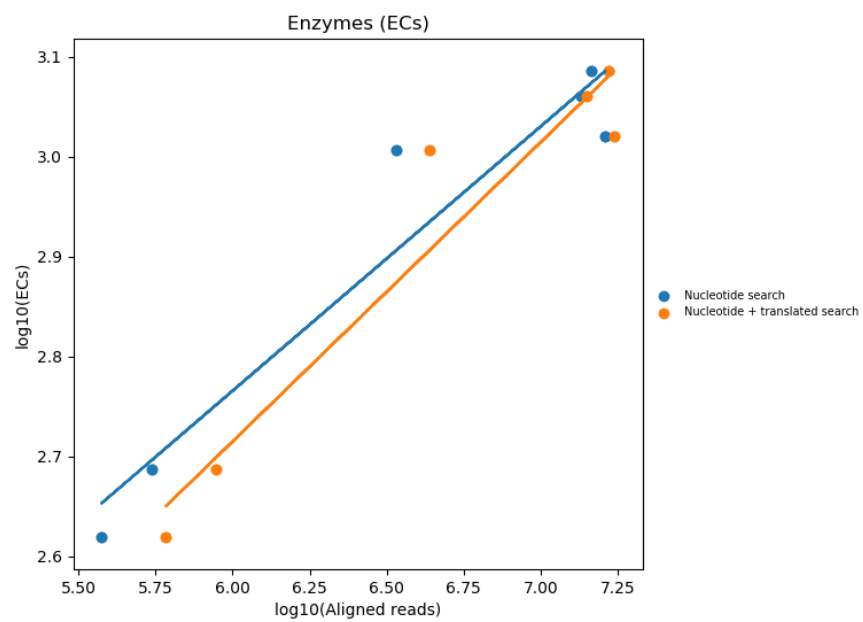
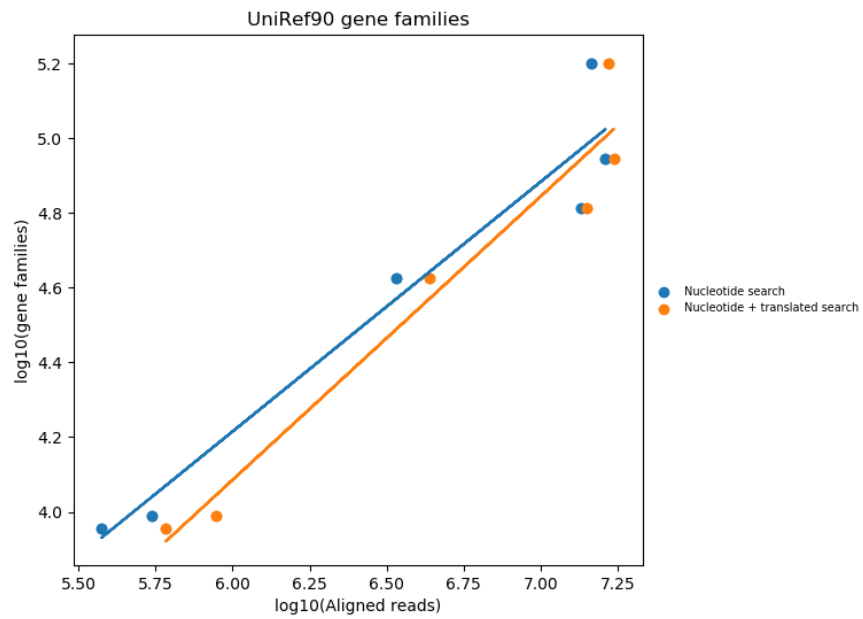
To learn more about other pathways, search for the pathway by name on the [MetaCyc](#) website.

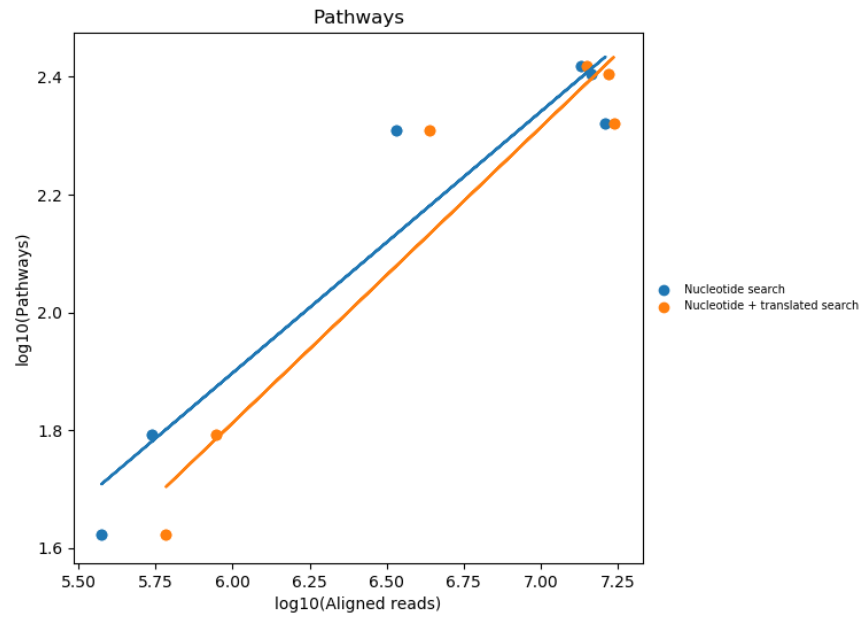
Features

Feature detection as a function of sequencing depth. Effect of sample sequencing depth on the ability to detect microbiome functional features in metagenomic sequence data. HUMAnN2 functional profiling of DNA quality filtered reads was performed on individual samples in species-specific mode (blue), i.e. nucleotide alignment against pangenomes of species identified in the sample with MetaPhlAn2, and in combined species-specific and -agnostic (orange) mode, in which reads not matching any pangenome reference sequences were subjected to translated searching against the UniRef90 database. Each profiled sample is represented by a orange and blue point in each plot. Linear regression fit is represented by straight lines in each plot.



Number of aligned reads in species-specific (nucleotide search) and species-agnostic (translated search) HUMAnN2 mode as a function of input reads.





Detection of UniRef90 gene families, enzyme modules, and pathways as a function of aligned reads.