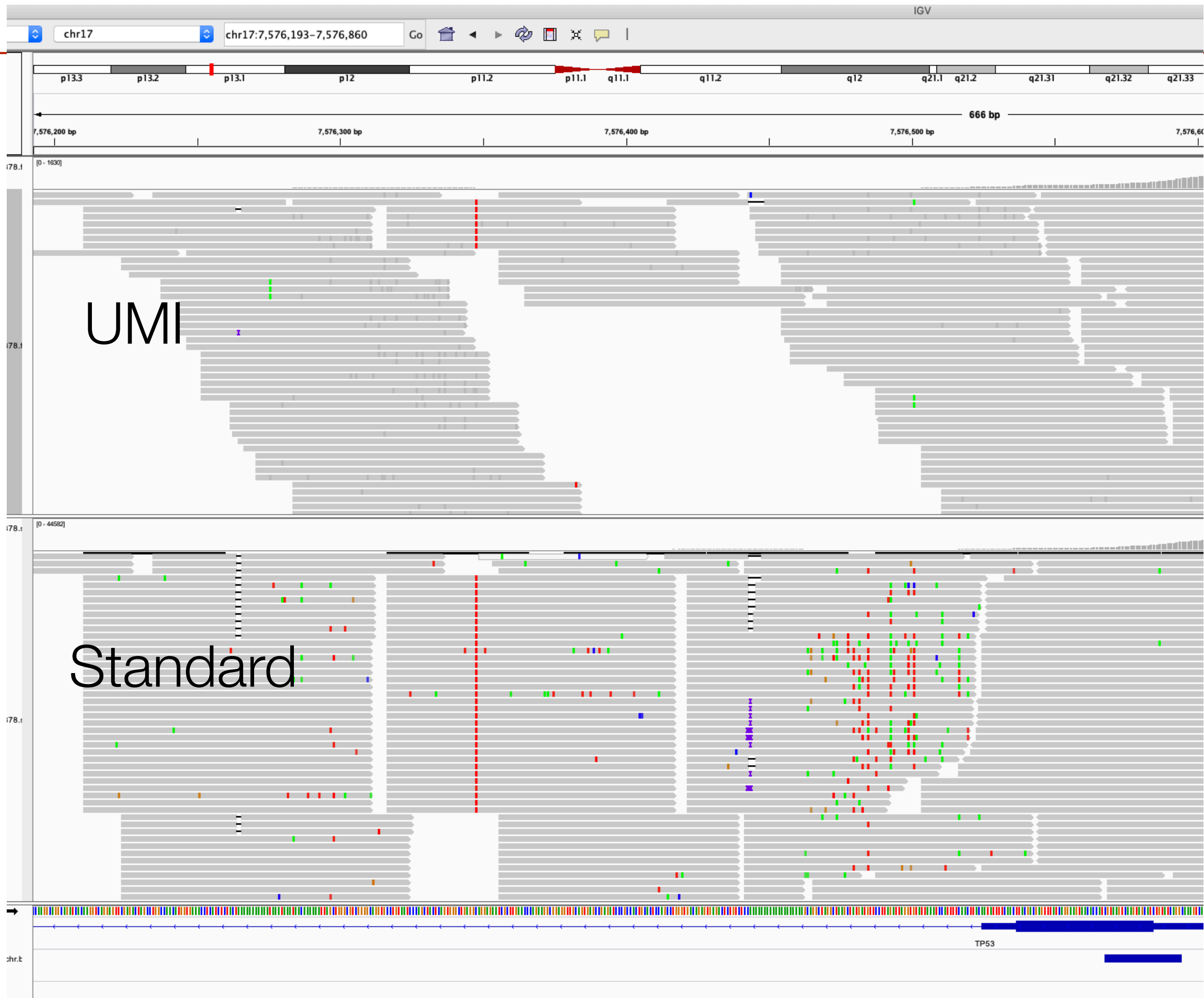
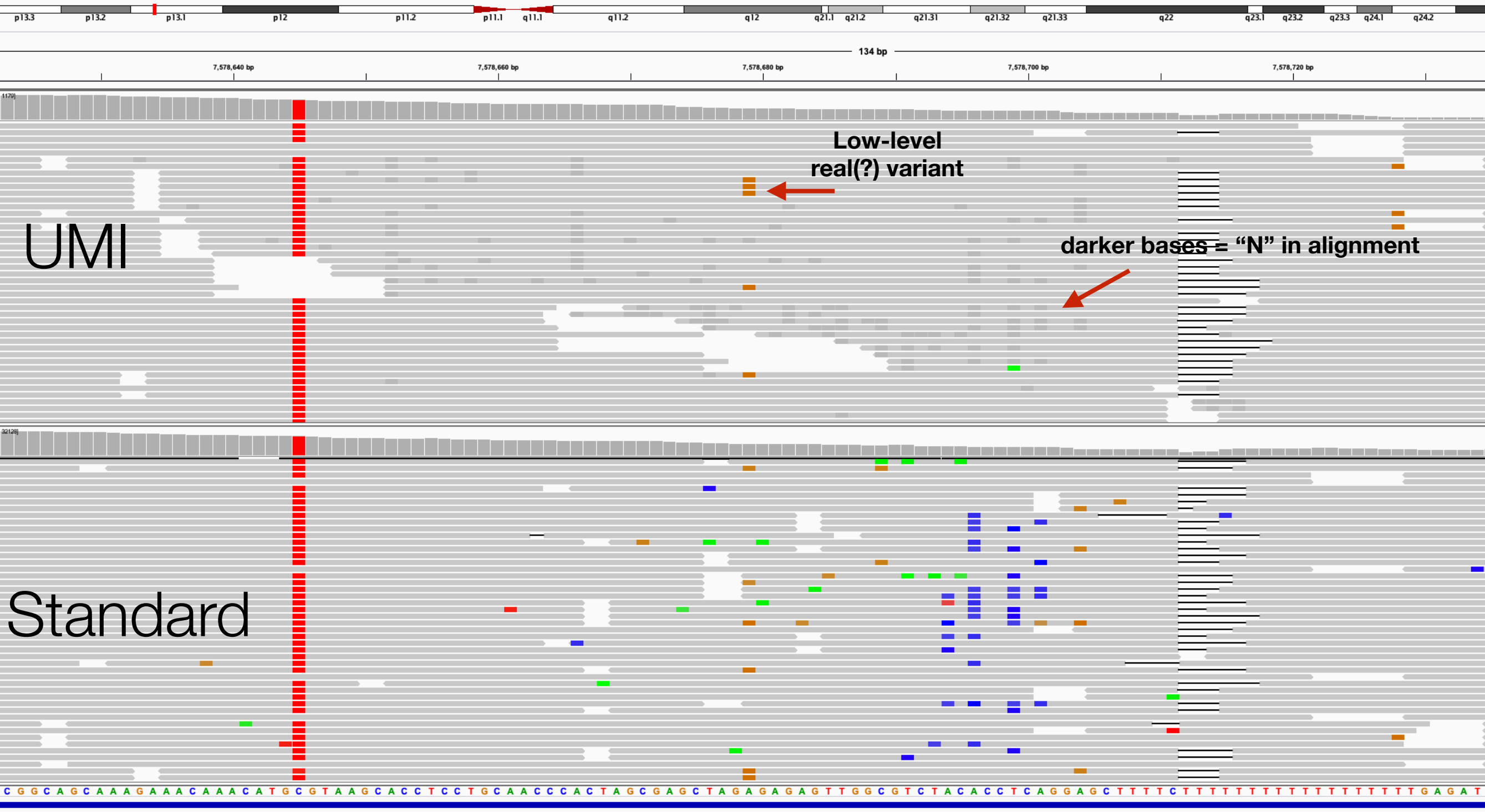

UMI downsampling QC reports

First, a qualitative view... UMI does work!





263R

<https://batchman.labmed.uw.edu/workflows/fa984b8e-5b77-4b5a-af67-c44dab886e3f>

[illegible]

263R stats

Table 1

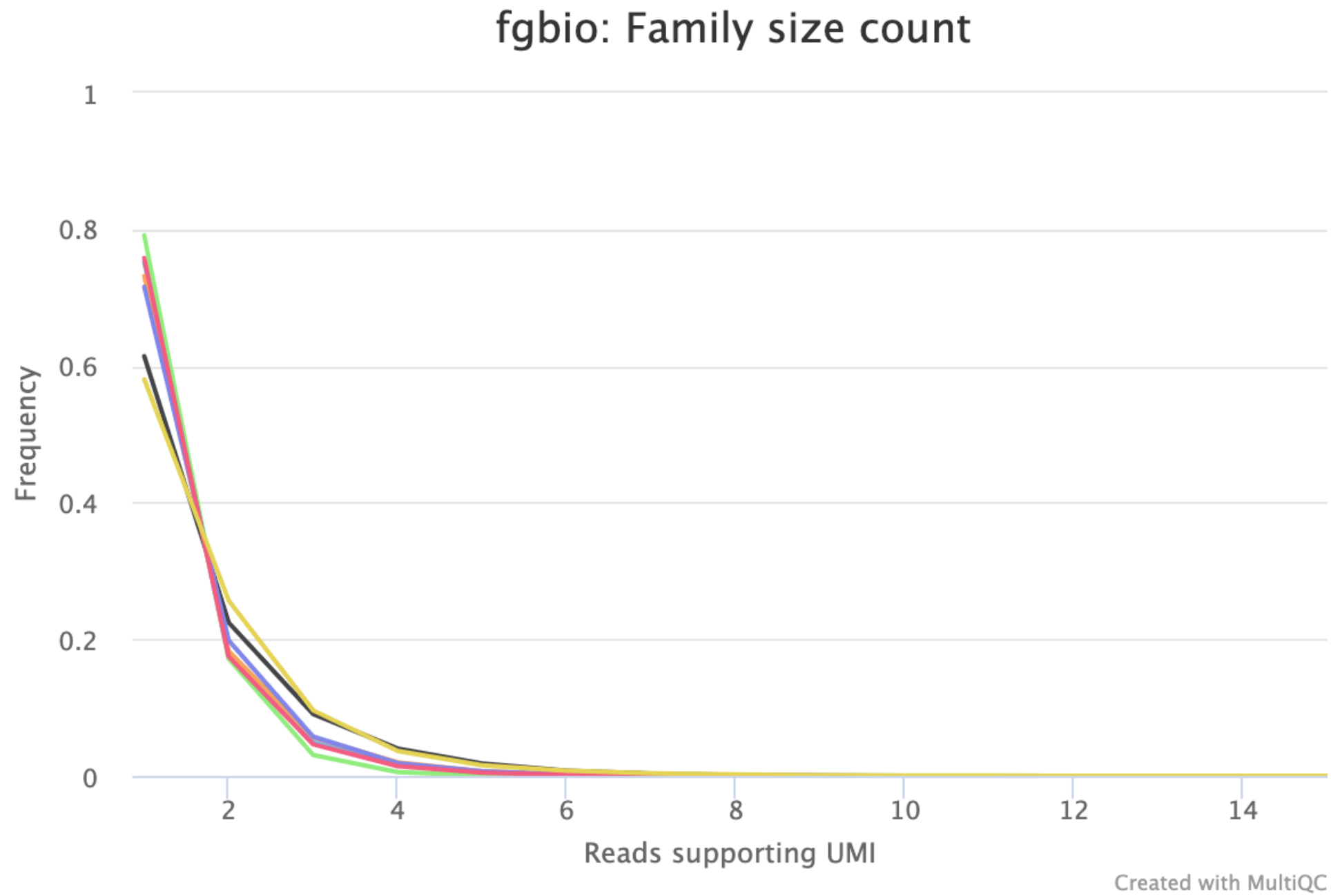
Sample Name		Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x		% UMI ≥2
		standard	final	standard	final	standard	final	standard	final	standard	final	
263R01		50,000,000	7,926,802	12,513	2,454	79%	68%	41%	33%	100%	99.9%	25%
263R02		50,000,000	10,050,168	12,482	3,029	79%	69%	39%	33%	100%	99.9%	39%
263R03		50,000,000	7,223,314	12,390	2,116	77%	69%	38%	33%	100%	99.8%	21%
263R04		50,000,000	7,900,734	12,330	2,472	79%	67%	41%	33%	100%	99.9%	27%
263R05		50,000,000	8,276,458	11,871	2,432	77%	66%	39%	32%	100%	100%	28%
263R06		50,000,000	7,083,300	12,104	2,184	77%	64%	41%	33%	100%	100%	24%
263R07		26,327,000	5,655,734	6,392	1,571	75%	68%	37%	33%	100%	100%	42%

Table 1-1

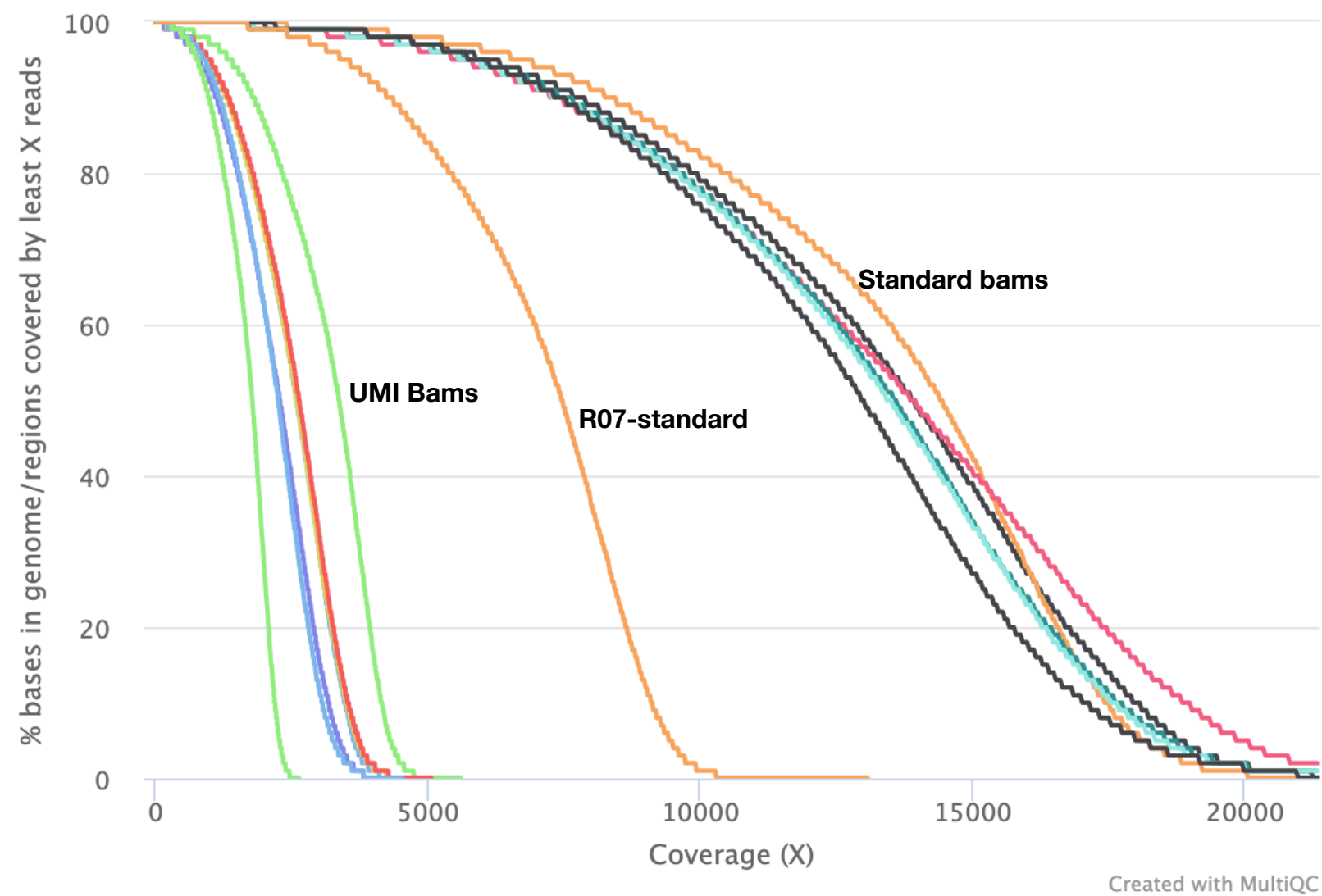
Means (w/o R07)		50,000,000	8,076,796	12,282	2,448	78%	67%	40%	33%	100%	100%	27%
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- ▶ R07 failed — note poor insert size distribution + low read count

263 UMI family size



Coverage of standard + final bam



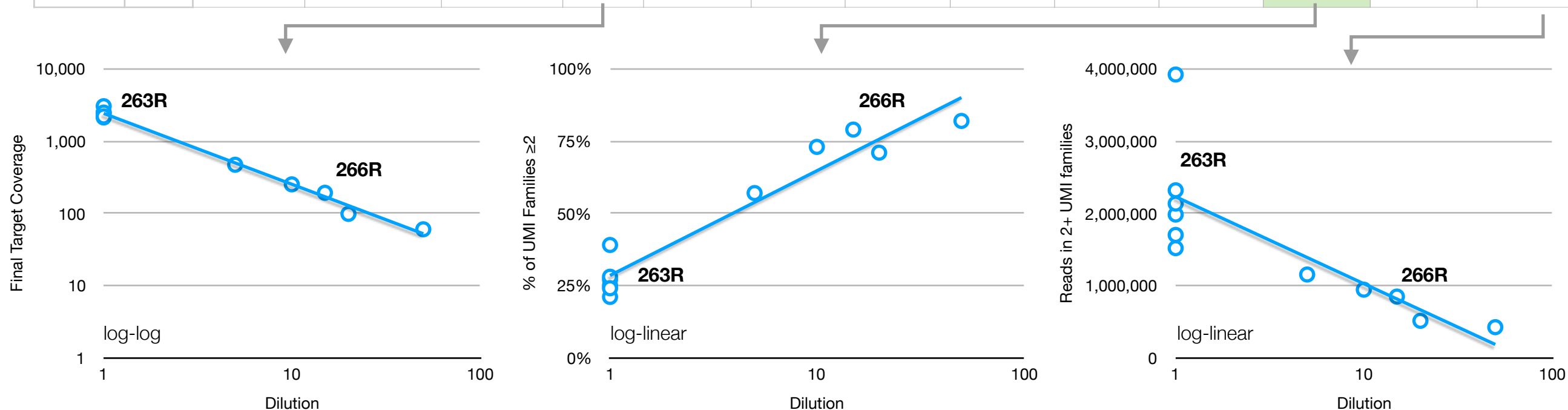
Important!

263 vs 266

Conclusion: Even though dilutions of 266R result in bigger UMI families, coverage drops dramatically and erases this benefit.

Table 1

Sample		Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x		% UMI ≥2	# singleton reads	# reads in 2+ Families
		standard	final	standard	final	standard	final	standard	final	standard	final			
263R01	1:1	50,000,000	7,926,802	12,513	2,454	79%	68%	41%	33%	100%	99.9%	25%	5,945,102	1,981,701
263R02	1:1	50,000,000	10,050,168	12,482	3,029	79%	69%	39%	33%	100%	99.9%	39%	6,130,602	3,919,566
263R03	1:1	50,000,000	7,223,314	12,390	2,116	77%	69%	38%	33%	100%	99.8%	21%	5,706,418	1,516,896
263R04	1:1	50,000,000	7,900,734	12,330	2,472	79%	67%	41%	33%	100%	99.9%	27%	5,767,536	2,133,198
263R05	1:1	50,000,000	8,276,458	11,871	2,432	77%	66%	39%	32%	100%	100%	28%	5,959,050	2,317,408
263R06	1:1	50,000,000	7,083,300	12,104	2,184	77%	64%	41%	33%	100%	100%	24%	5,383,308	1,699,992
266R01	1:5	50,000,000	2,021,550	13,477	469	73%	63%	36%	31%	100%	99%	57%	869,267	1,152,284
266R02	1:10	50,000,000	1,291,048	13,597	251	74%	53%	37%	26%	100%	98%	73%	348,583	942,465
266R03	1:15	50,000,000	1,072,306	13,481	192	74%	49%	37%	24%	100%	96%	79%	225,184	847,122
266R04	1:20	50,000,000	720,616	13,537	98	74%	38%	37%	18%	100%	47%	71%	208,979	511,637
266R05	1:50	50,000,000	516,344	13,222	60	74%	33%	37%	16%	100%	3%	82%	92,942	423,402



266R

266R

Sample				Pre-PCR			Post-PCR		Pooling prior to hybridization	
Name	size	ng/uL	Fetal fraction	WellId	Input	Theoretical max coverage	Avg. Size [bp]	Conc. [ng/μl]	Mass added to pre-hyb pool (ng)	Moles added to pre-hyb pool
266R01	188	0.194ng/uL	—	A7	1:5	400x	361	11.9	125	5.33E-13
266R02	188	0.194ng/uL	—	B7	1:10	193x	374	29.9	125	5.14E-13
266R03	188	0.194ng/uL	—	C7	1:15	130x	372	33.5	125	5.17E-13
266R04	188	0.194ng/uL	—	D7	1:20	96.7x	370	30.6	125	5.2E-13
266R05	188	0.194ng/uL	—	E7	1:50	40x	367	31.9	125	5.24E-13

- Limit the amount of cell-free DNA carried forward from adaptor ligation into pre-capture PCR, to:
 - hopefully increase the number of duplicate reads
 - determine which dilution would be most effective in maximizing the fraction of UMI families with a family size of at least 2.
- Five remnant cell-free DNAs from the PDNAS assay (due to having few to zero flagged bins and no detected aneuploidies), pooled together.
- Pool-level quality control
(tape station): 0.194ng/ul = 5.82ng in 30ul

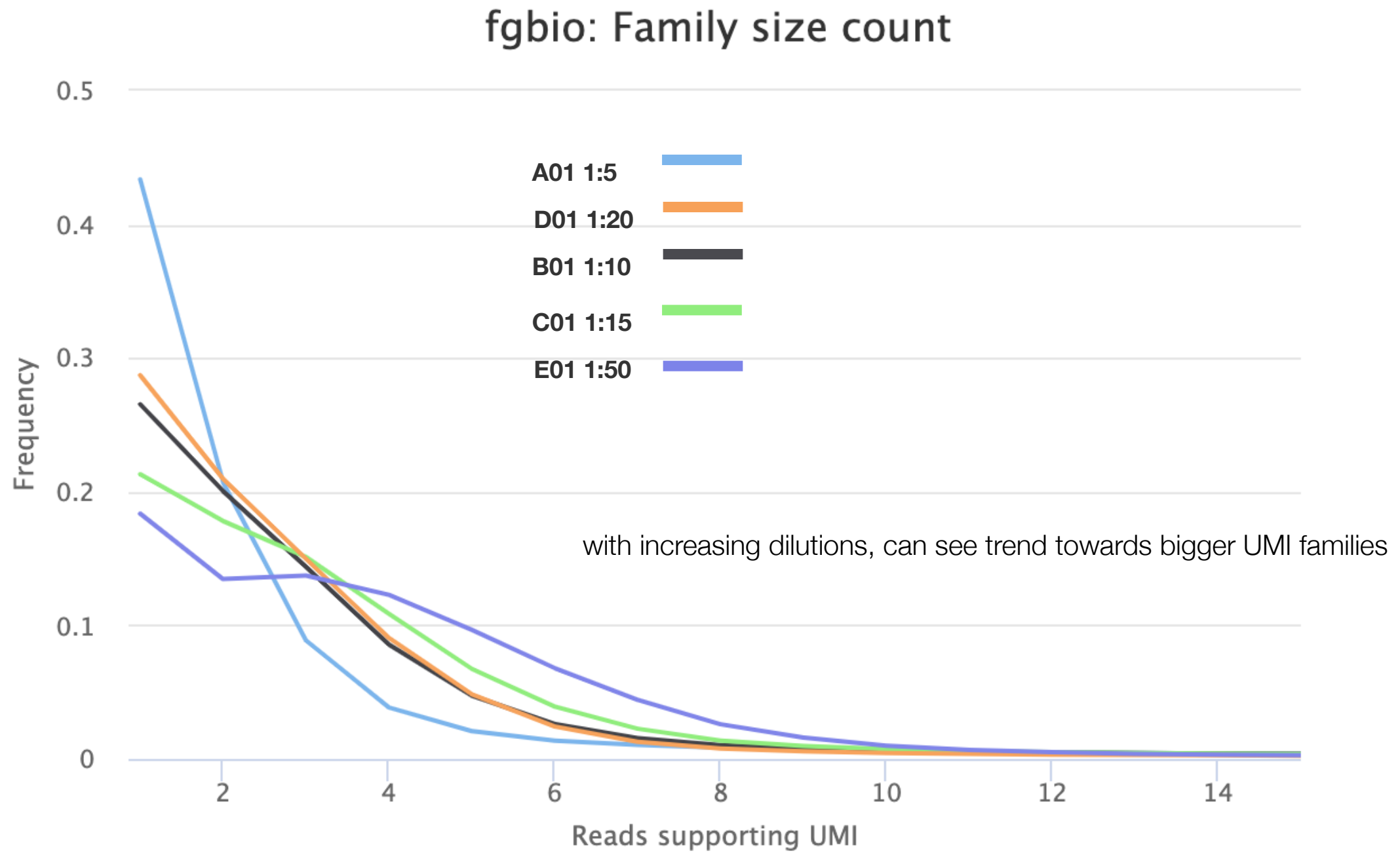
266R

Table 1

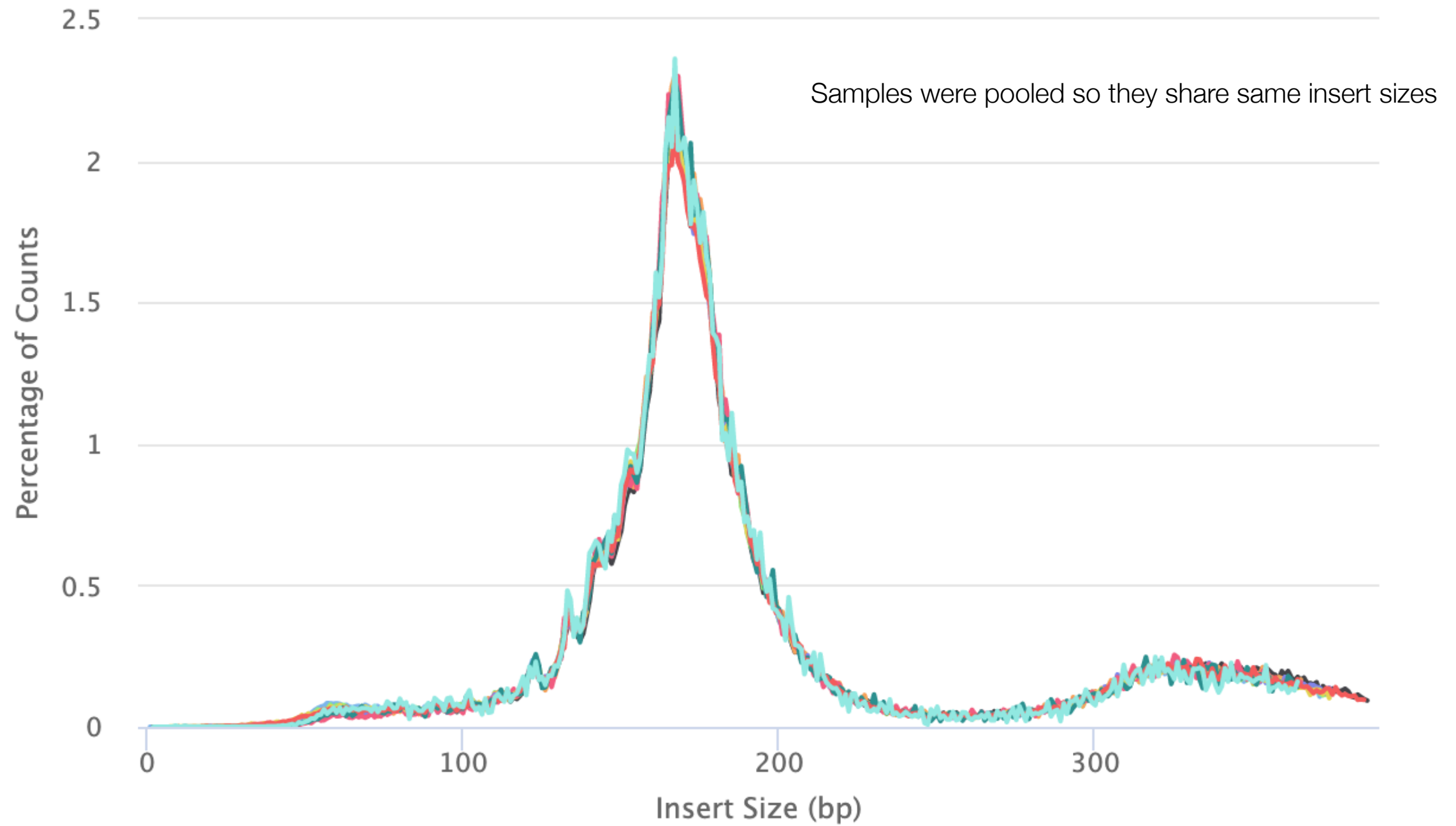
Sample Name		Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x		% UMI ≥ 2
		standard	final	standard	final	standard	final	standard	final	standard	final	
266R01	1:5	50,000,000	2,021,550	13,477	469	72.7%	62.5%	36.5%	30.9%	100.0%	99.4%	57%
266R02	1:10	50,000,000	1,291,048	13,597	251	73.5%	52.8%	36.7%	26.0%	100.0%	97.8%	73%
266R03	1:15	50,000,000	1,072,306	13,481	192	73.5%	48.9%	36.7%	24.2%	100.0%	96.4%	79%
266R04	1:20	50,000,000	720,616	13,537	98	74.3%	37.7%	37.1%	18.4%	100.0%	47.4%	71%
266R05	1:50	50,000,000	516,344	13,222	60	74.4%	32.7%	37.0%	16.0%	100.0%	3.0%	82%
263R Mean	1:1	50,000,000	8,076,796	12,282	2,448	78%	67%	40%	33%	100%	100%	27%

- ▶ All FASTQs downsampled to have 25m read pairs
- ▶ With increasing dilution:
 - ▶ Fewer UMI families (lower final total reads)
 - ▶ Lower median target coverage, lower % on bait/target bases
 - ▶ Above 1:20, significant loss of % bases with >100x coverage
- ▶ Our bait-to-target efficiency is about 50%
- ▶ 266R01-A01 has lower percentage of UMI families with ≥ 2 reads, however no clear trend for this value in other dilutions?

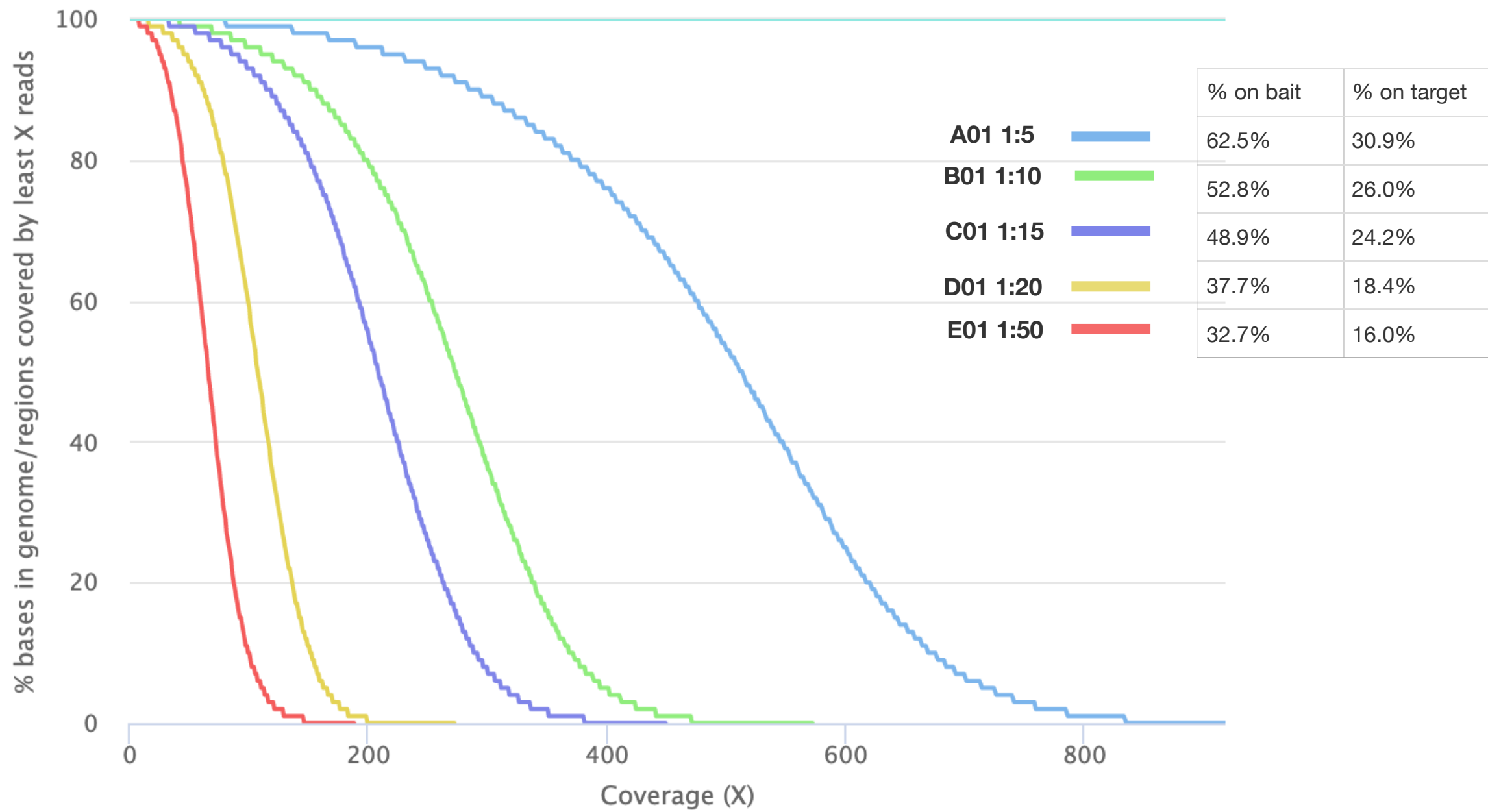
266R UMI family sizes



266R insert size



Coverage of final bam



280R

280R

Sample				Pre-PCR			Post-PCR		Pooling prior to hybridization	
Name	size	ng/uL	Fetal fraction	WellId	Input mass (ng)	Theoretical max coverage	Avg. Size [bp]	Conc. [ng/μl]	Mass added to pre-hyb pool (ng)	Moles added to pre-hyb pool
210P03	182	0.763	0.14 (XX)	A2	5ng	1667x	420	1.48	125	4.58E-13
210P03	182	0.763	0.14 (XX)	D2	1ng	333x	369	29.5	125	5.21E-13
211P12	198	0.725	0.056 (XY)	B2	5ng	1667x	377	26.7	125	5.10E-13
211P12	198	0.725	0.056 (XY)	E2	1ng	333x	335	22.1	125	5.74E-13
222P09	194	0.441	0.076 (XY)	C2	5ng	1667x	366	28.5	125	5.25E-13
222P09	194	0.441	0.076 (XY)	F2	1ng	333x	388	37.3	125	4.96E-13

- Repeat the best conditions from NGS266R (1/5 ligation into PCR)
- See whether approximately 1/5 of the input mass with the normal 20ul into PCR would produce similar results.
- Three remnant cell-free DNAs from the PDNAS assay were selected because they had enough leftover for two library preps (5ng, and 1ng). These were also normal samples with few flagged bins.

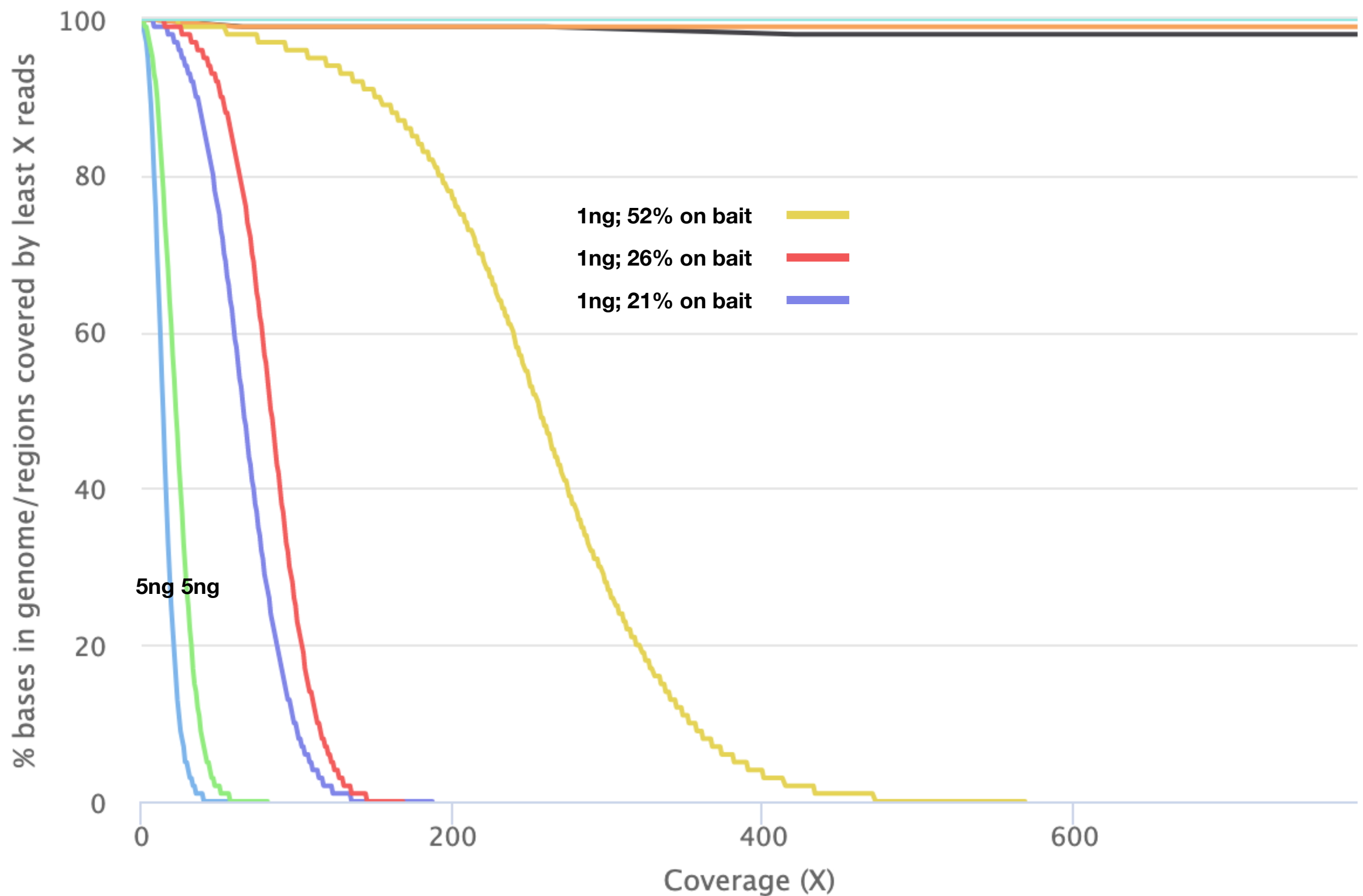
280R

Table 1

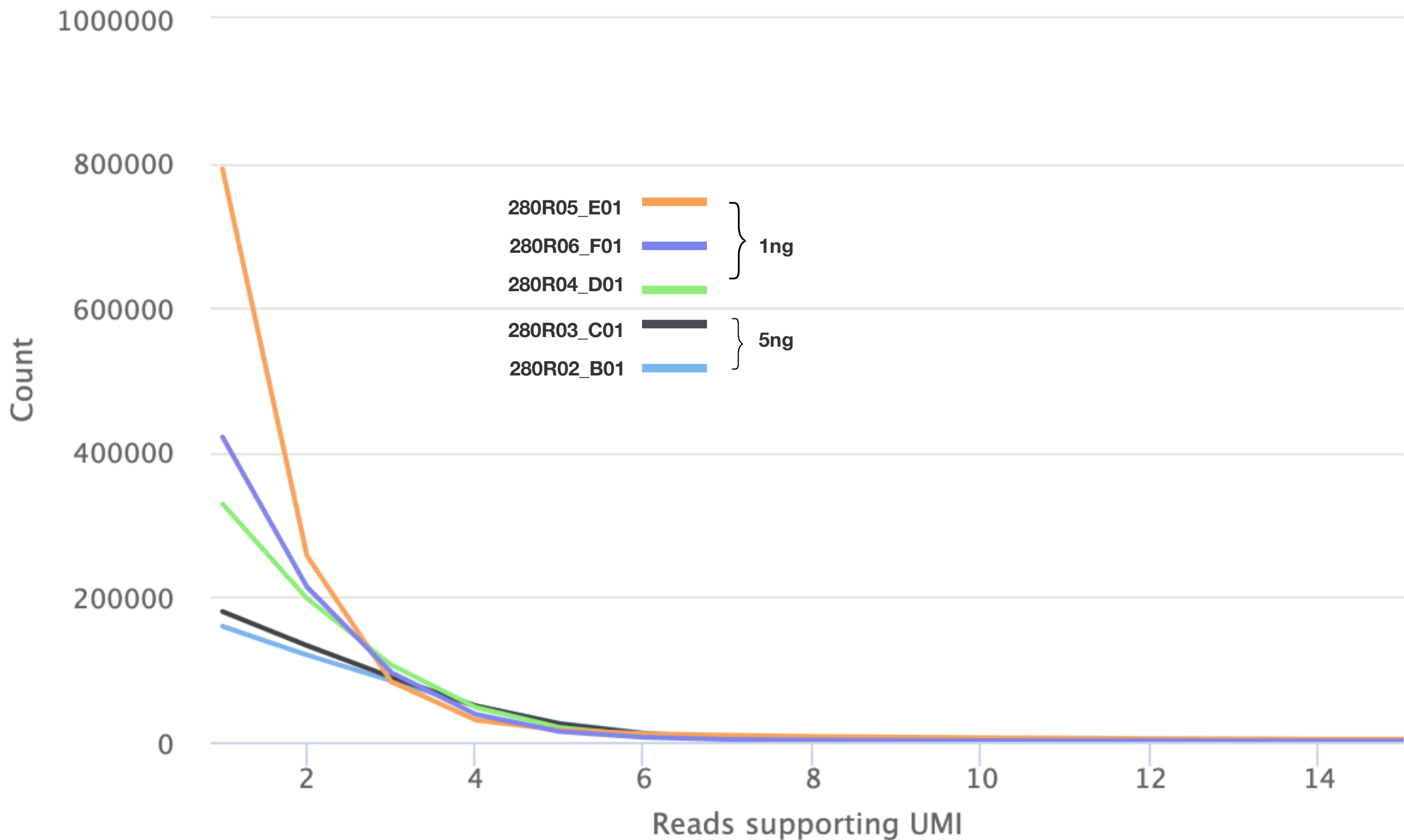
Sample Name			Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x	
			standard	final	standard	final	standard	final	standard	final	standard	final
280R01_A01	210P03	5ng	50,000,000	1,236,226	7	1	67.4%	1.7%	34.0%	0.9%	44.8%	0.0%
280R04_D01	210P03	1ng	50,000,000	593,438	12,643	60	70.7%	21.7%	35.0%	10.6%	99.9%	2.5%
280R02_B01	211P12	5ng	50,000,000	816,094	12,333	12	71.4%	6.5%	35.1%	3.1%	99.3%	0.0%
280R05_E01	211P12	1ng	50,000,000	547,794	12,789	219	74.6%	52.0%	34.9%	23.9%	99.9%	97.4%
280R03_C01	222P09	5ng	50,000,000	772,752	12,469	20	71.6%	9.6%	35.5%	4.7%	99.5%	0.0%
280R06_F01	222P09	1ng	50,000,000	343,036	13,085	76	70.6%	25.6%	35.7%	12.6%	100.0%	10.0%

- ▶ All FASTQs downsampled to have 25m read pairs
- ▶ A01 failed — disregard; note that it had normal *mean* coverage; only median target coverage is low.
- ▶ Metrics on standard bam do not predict final bam metrics.
- ▶ Comparing 5ng to 1ng:
 - ▶ 1ng has MORE on-bait and on-target bases and GREATER median target coverage.
 - ▶ BUT 1ng has FEWER reads in final/collapsed bam!
 - ▶ did 5ng and 1ng get switched?
- ▶ These results look like dilutions of 1:20 and 1:50, but don't follow the 263/266 trends

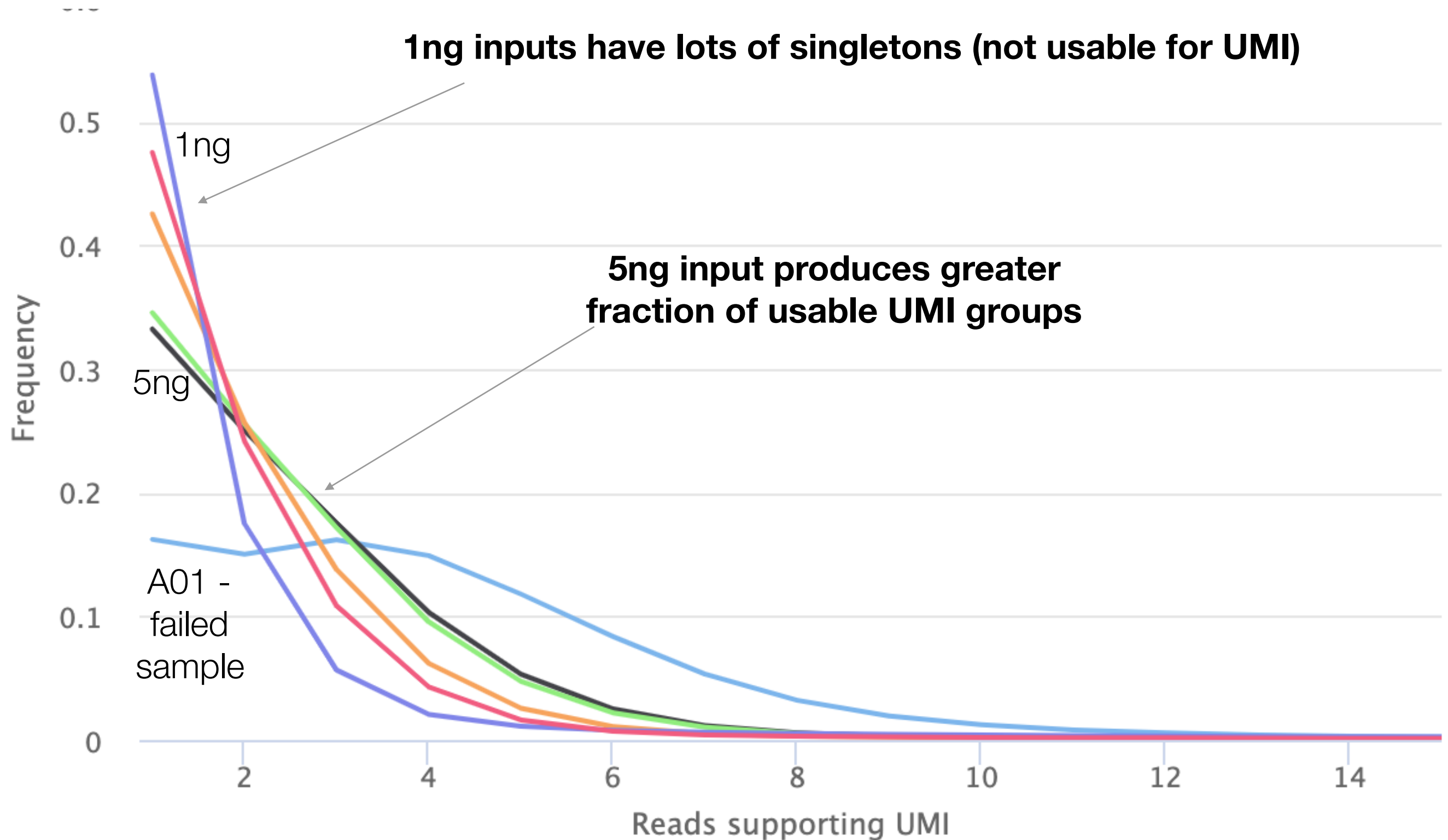
Coverage of final bam



fgbio: Family size count



UMI Histogram - percentages



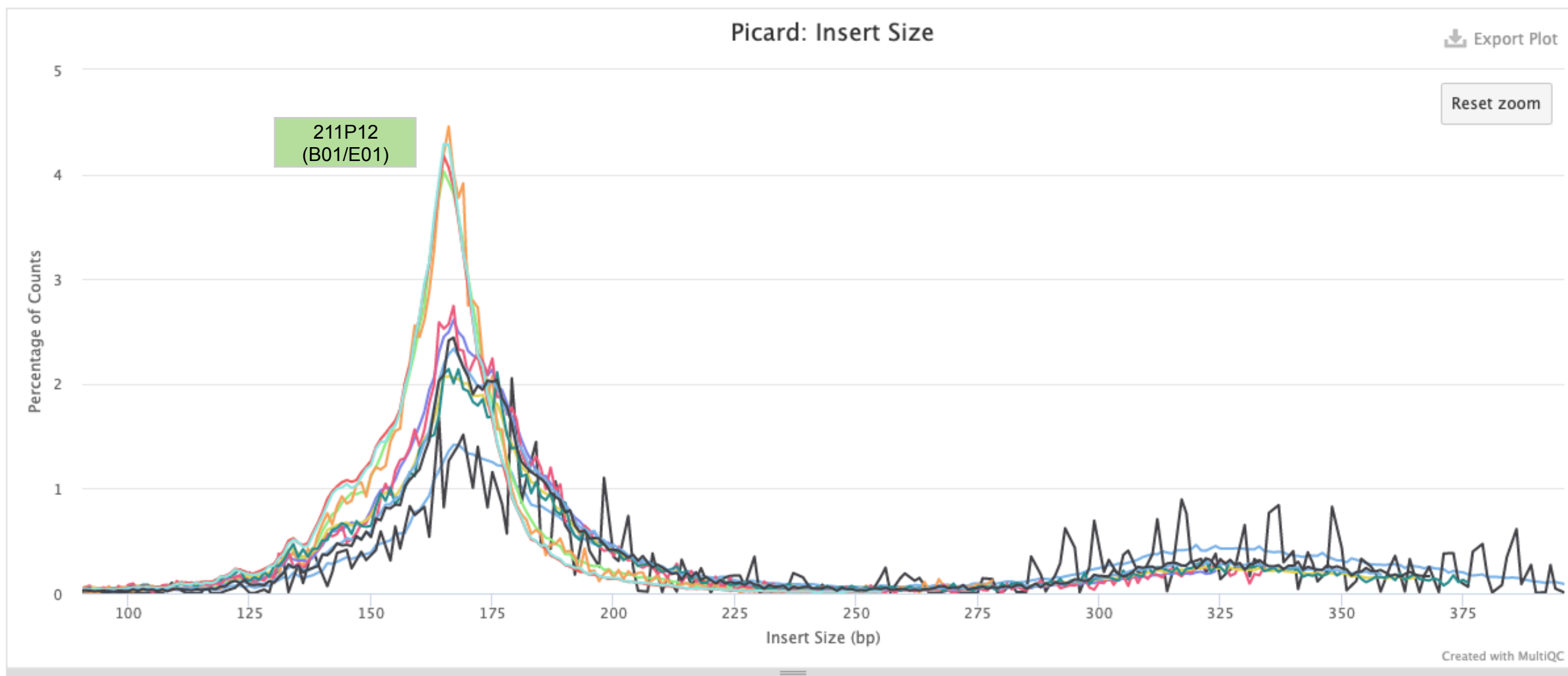
280R Insert Size

Insert Size

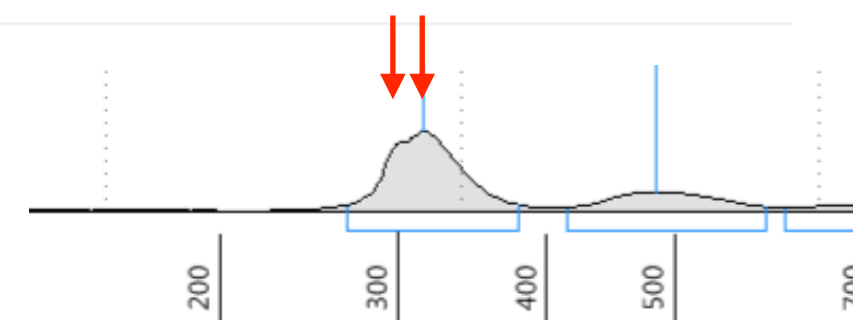
Plot shows the number of reads at a given insert size. Reads with different orientations are summed.

Y-Limits: ☒ on

Counts Percentages



211P12 (B01/E01) has slightly smaller insert size than other two samples.
(small bimodal peak seen on post-hyb TapeStation as well)



Comparing 266 vs 280

- 266R01 (1:5) should be comparable to 5ng condition of 280R? Very different numbers. What happened? **did 5ng and 1ng get switched??**

Table 1-1

Sample Name		Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x	
		standard	final	standard	final	standard	final	standard	final	standard	final
266R01	1:5	50,000,000	2,021,550	13,477	469	72.7%	62.5%	36.5%	30.9%	100.0%	99.4%
266R02	1:10	50,000,000	1,291,048	13,597	251	73.5%	52.8%	36.7%	26.0%	100.0%	97.8%
266R03	1:15	50,000,000	1,072,306	13,481	192	73.5%	48.9%	36.7%	24.2%	100.0%	96.4%
266R04	1:20	50,000,000	720,616	13,537	98	74.3%	37.7%	37.1%	18.4%	100.0%	47.4%
266R05	1:50	50,000,000	516,344	13,222	60	74.4%	32.7%	37.0%	16.0%	100.0%	3.0%

Table 1

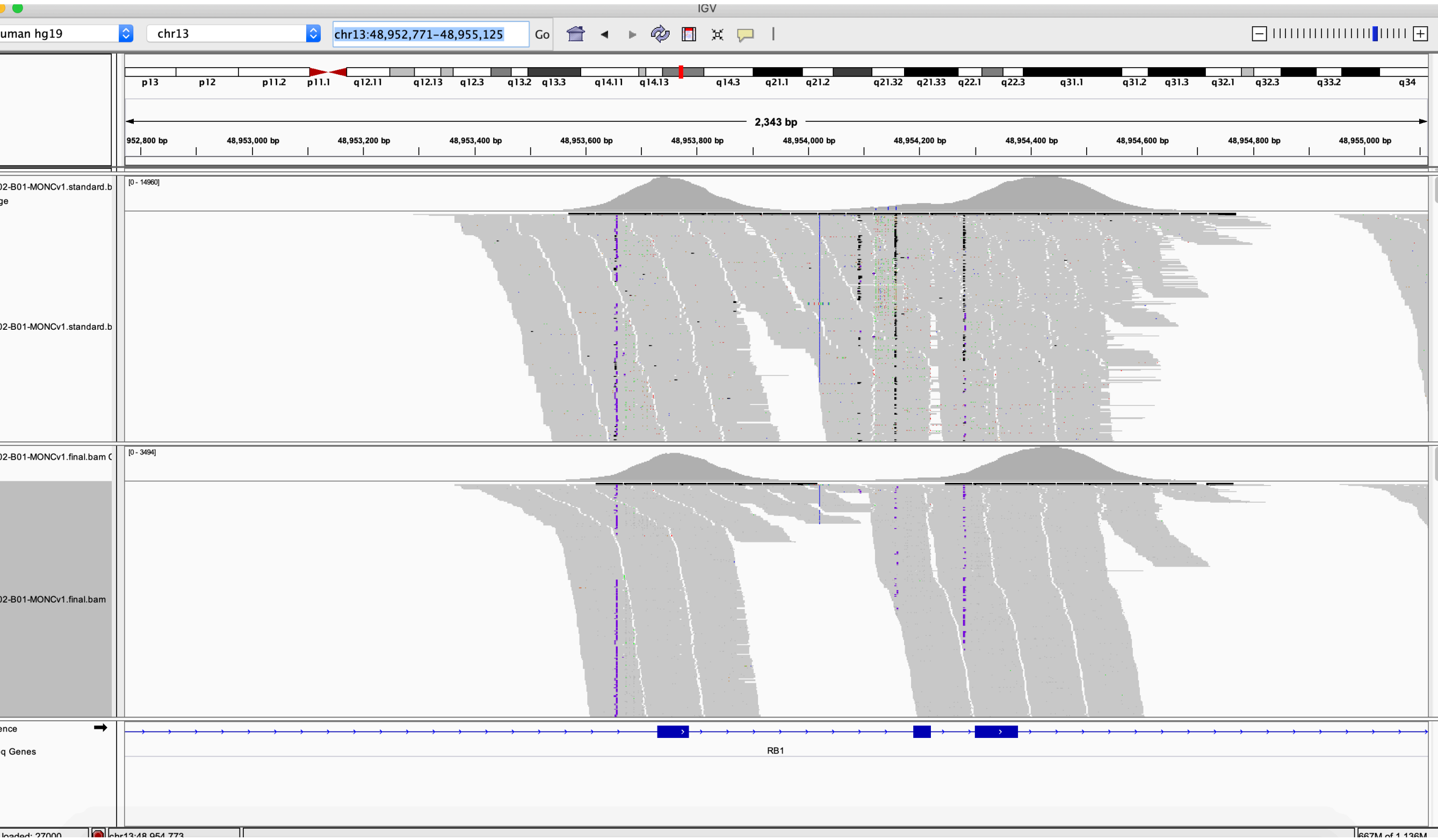
Sample Name			Total reads		Median Target Coverage		% on bait bases		% on target bases		% Bases >100x	
			standard	final	standard	final	standard	final	standard	final	standard	final
280R02_B01	211P12	5ng	50,000,000	816,094	12,333	12	71.4%	6.5%	35.1%	3.1%	99.3%	0.0%
280R03_C01	222P09	5ng	50,000,000	772,752	12,469	20	71.6%	9.6%	35.5%	4.7%	99.5%	0.0%

Table 1-2

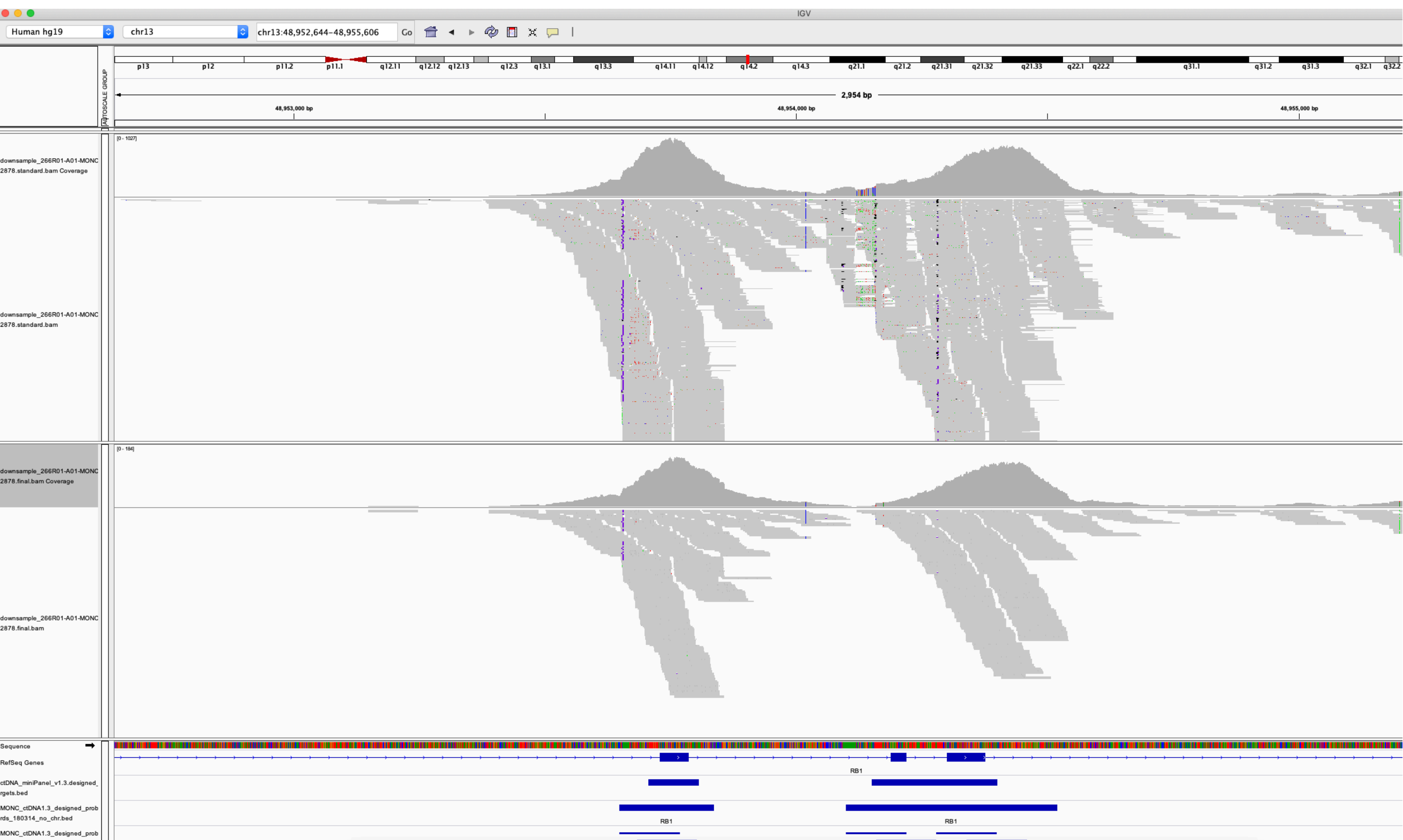
280R05_E01	211P12	1ng	50,000,000	547,794	12,789	219	74.6%	52.0%	34.9%	23.9%	99.9%	97.4%
280R06_F01	222P09	1ng	50,000,000	343,036	13,085	76	70.6%	25.6%	35.7%	12.6%	100.0%	10.0%

appendix

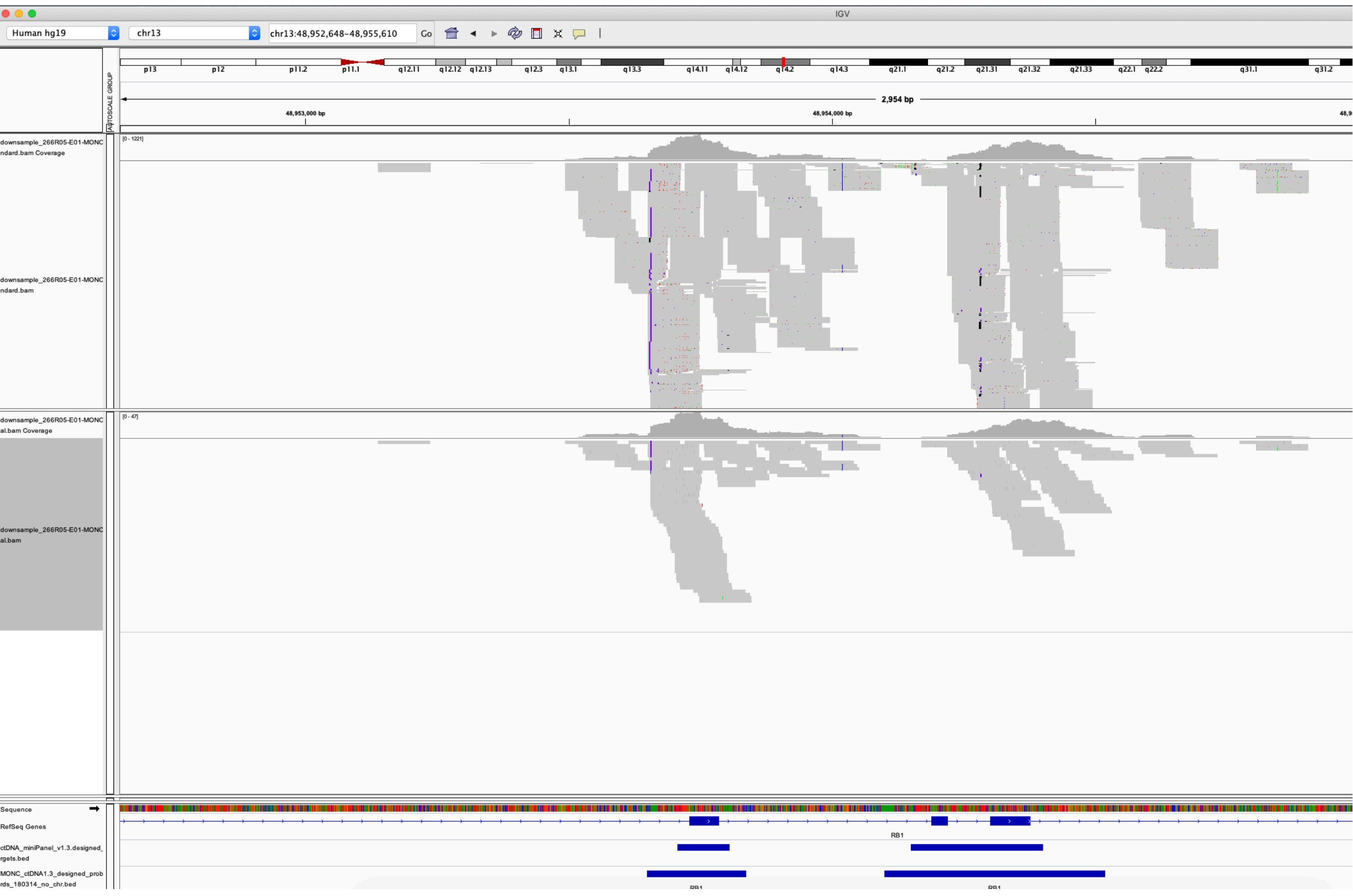
263R02



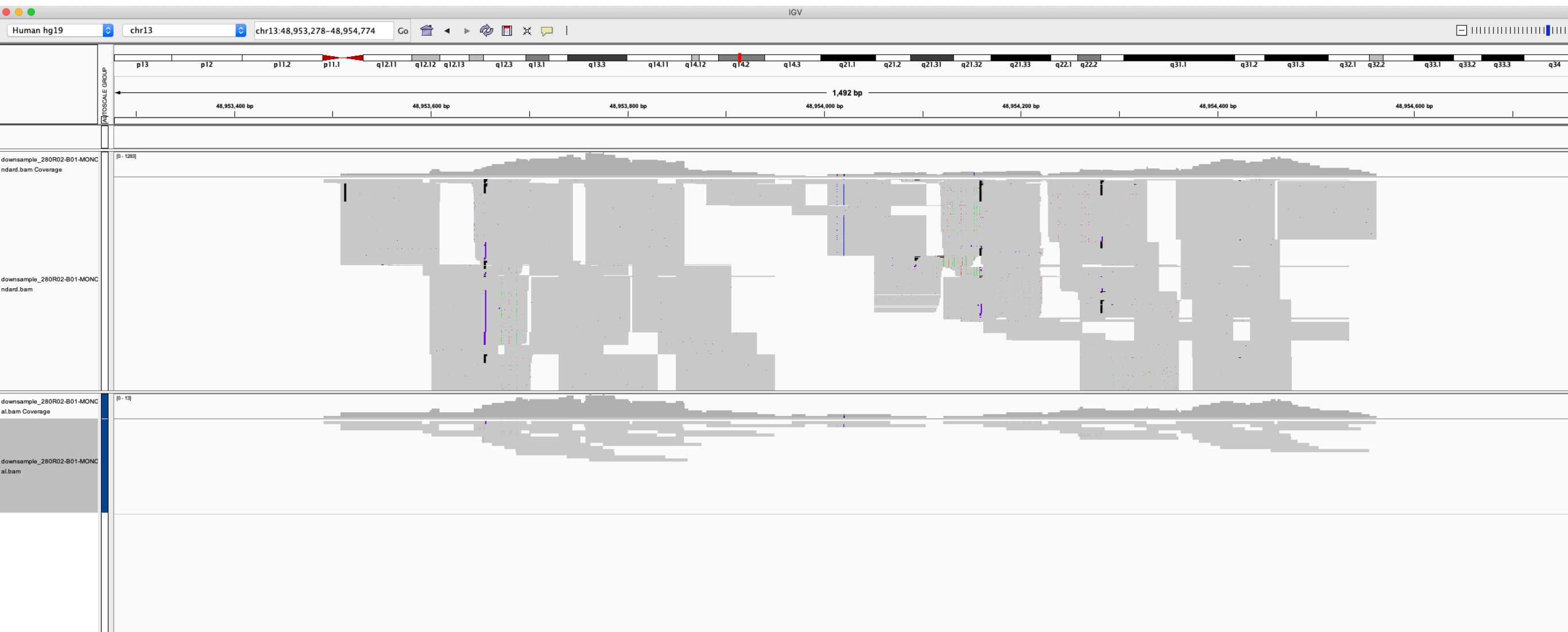
266R01



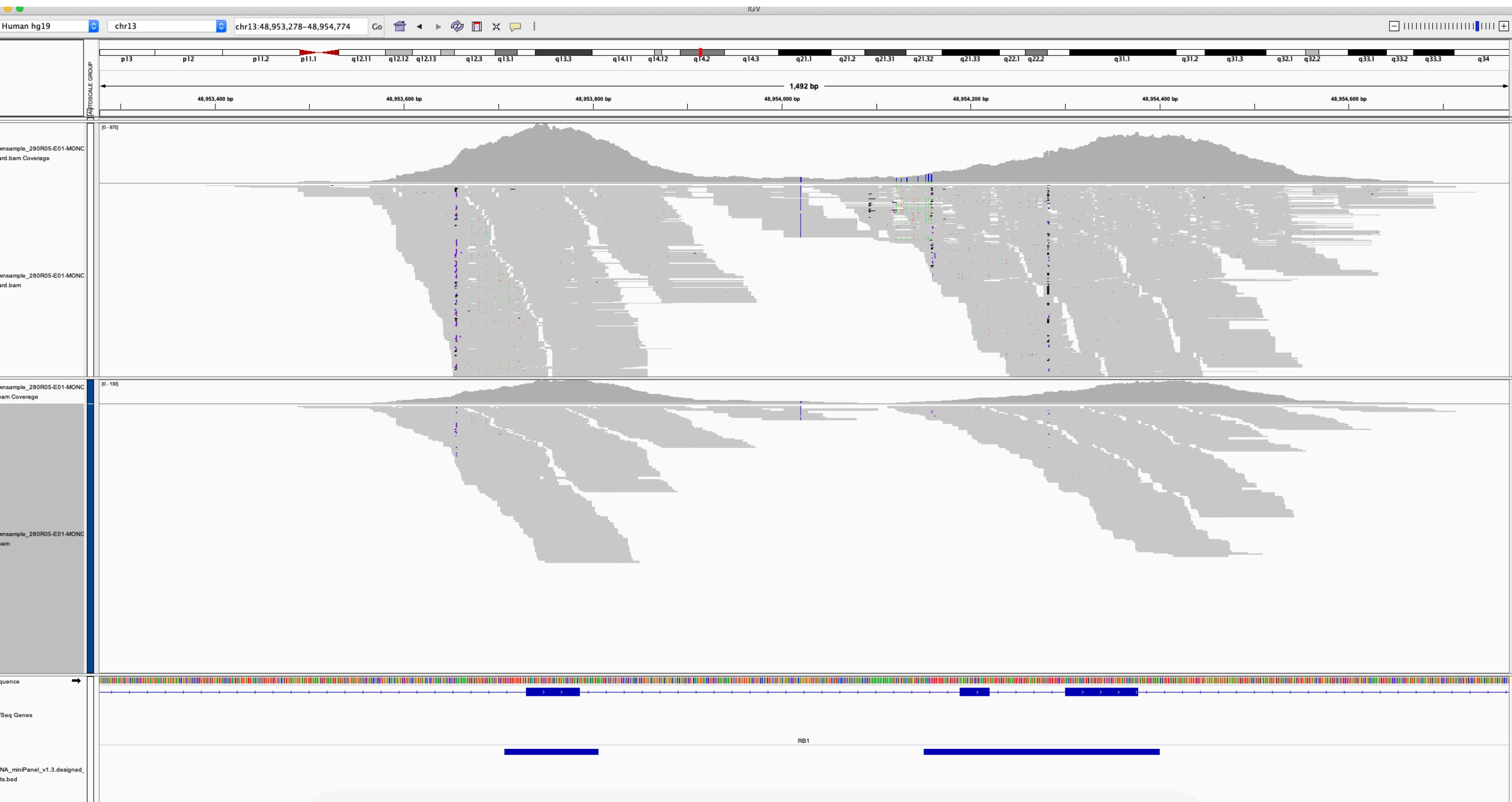
266R05



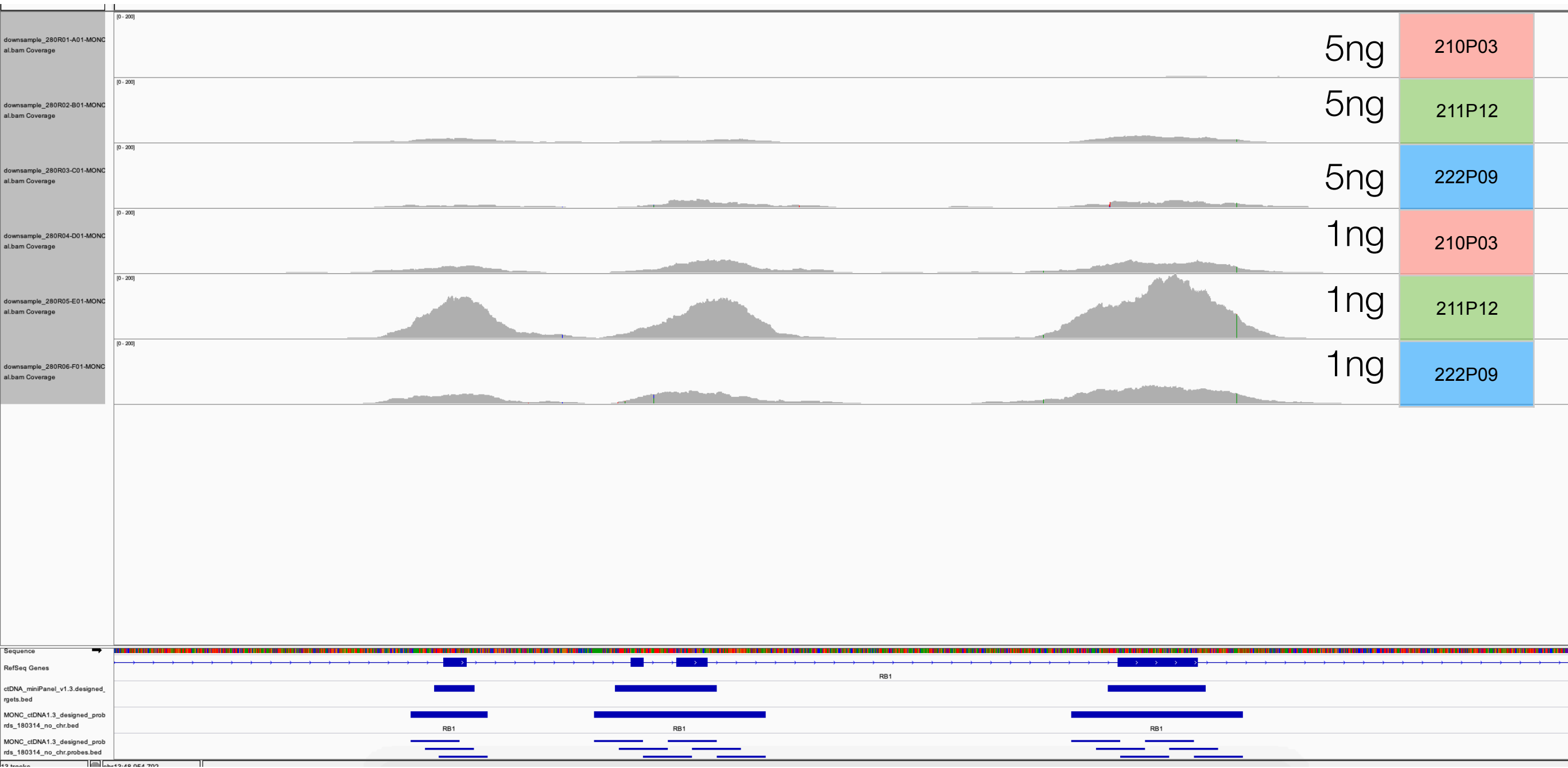
280R02 (5ng) standard vs final



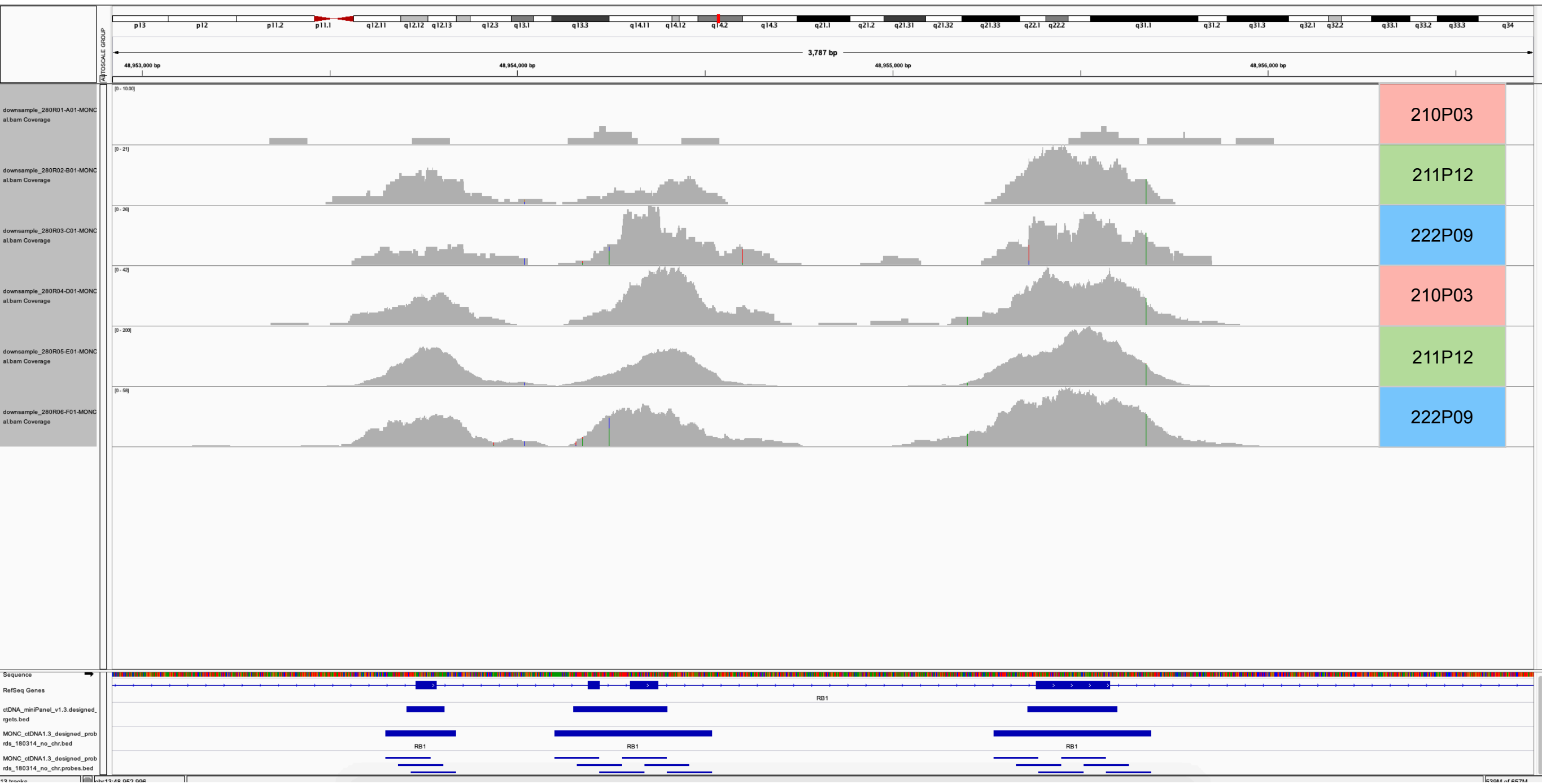
280R05 (1ng) standard vs final



280R coverage snapshot (scale 0-200)



280R coverage (autoscale)



280R01-A01 — failed (we knew this)

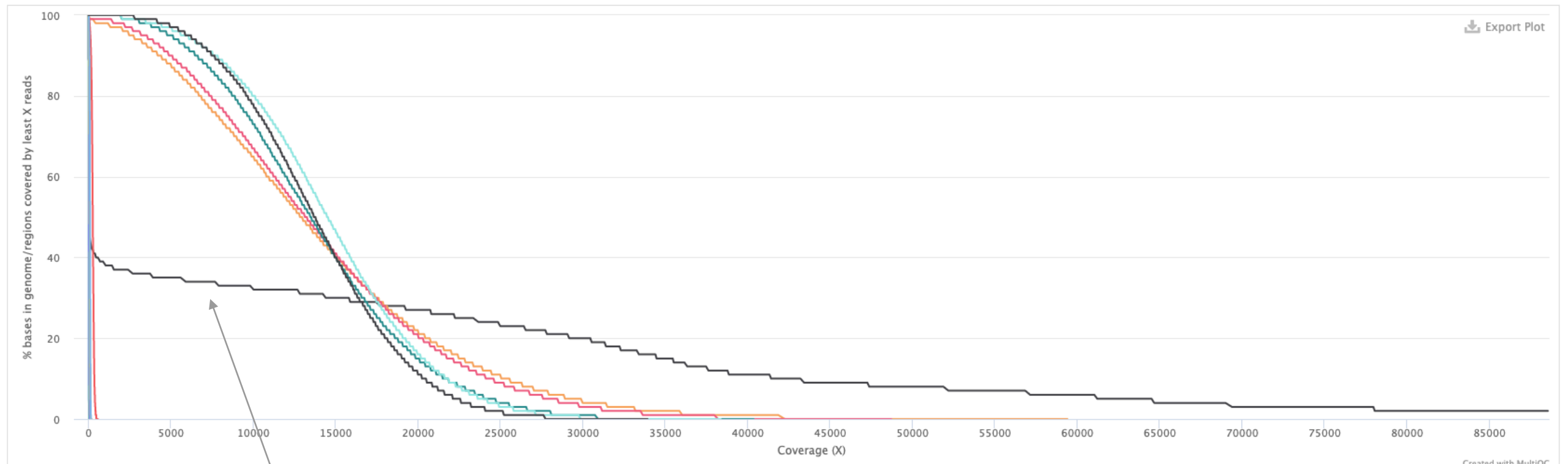
Coverage distribution

Distribution of the number of locations in the reference genome with a given depth of coverage

Help

Y-Limits: on

Export Plot



Indicates ~60% of targets have no coverage, remaining 40% have all reads

280R01-A01 failed but note that the MEAN coverage looks great, the median coverage is bad;
→ represents very strong jackpot effect (max target coverage = 150,000x vs 20-50k average)