# Package 'tugHall.3'

June 1, 2022
Title R-based script to simulate the cancer cell evolution
Version 3.0
<b>Description</b> tugHall (tumor gene-Hallmark) is a cancer-cell evolution model simulator, wherein gene mutations are linked to the hallmarks of cancer, which influence tumor cell behaviors.
License GPL (>= 3)
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# Description

Function to add deletion to gene map (chromosomal location data frame)

# Usage

```
add_deletion(gm, Ref_start, Ref_end, Chr)
```

# Arguments

gm Chromosomal location data frame
Ref\_start Starting position of deletion
Ref\_end Final position of deletion
Chr Chromosome name

### Value

Chromosomal location data frame with additional deletion info

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm_new = add_deletion( gm = gene_map, Ref_start = 112775658, Ref_end = 112775716, Chr = '5')
# to check add_del ... add_dupl, please, use diff library and:
# diff_data( gm_ref, gm, show_unchanged_columns = TRUE, always_show_order = TRUE)
```

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add_duplication	Function to add duplication to gene map (chromosomal location data frame)
-----------------	---

# Description

Function to add duplication to gene map (chromosomal location data frame)

# Usage

```
add_duplication(gm, Ref_start, Ref_end, Chr)
```

# Arguments

gm Chromosomal location data frame
Ref\_start Starting position of duplication
Ref\_end Final position of duplication
Chr Chromosome name

# Value

Chromosomal location data frame with additional duplication info

### **Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm_new = add_duplication( gm = gene_map, Ref_start = 112775658, Ref_end = 112775716, Chr = '5')
# to check add_del ... add_dupl, please, use diff library and:
# diff_data( gm_ref, gm, show_unchanged_columns = TRUE, always_show_order = TRUE )
```

add\_pnt\_mutation

Function to add point mutation to data.frame gene\_map (chromoso-mal location data frame)

### **Description**

Function to add point mutation to data.frame gene\_map (chromosomal location data frame)

# Usage

```
add_pnt_mutation(gm = gm, pos_pnt, Chr = Chr)
```

# Arguments

gm Chromosomal location data frame

pos\_pnt Position of point mutation

Chr Chromosome name

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### Value

Chromosomal location data frame with additional point mutation info

### **Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm2 = add_pnt_mutation( gm = gene_map, pos_pnt = 112775637 , Chr = '5' )
```

calc\_binom

Function to calculate binomial distribution including BIG NUMBERS like 10^12 and more using approximation with normal distribution

# **Description**

Function to calculate binomial distribution including BIG NUMBERS like 10^12 and more using approximation with normal distribution

### Usage

```
calc_binom(tr, n, p)
```

# Arguments

tr Length of vector with successes trials

n Number of independent Bernoulli trials

p Probability to get successes in trials

#### Value

Vector of integer numbers of successes trials

# **Examples**

```
calc_binom(tr = 3, n = 40, p = 0.9)
calc_binom(tr = 3, n = 4E20, p = 9E-9)
```

```
change_allele_A_by_cna
```

Function to change copy number of the allele A of the point mutation at the allele B due to CNA

# Description

Function to change copy number of the allele A of the point mutation at the allele B due to CNA

```
change_allele_A_by_cna(pnt1, start_end, t)
```

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#### **Arguments**

pnt1 Object of class 'Point\_Mutations'
start\_end Vector with initial and final positions of CNA
t 'dup' or 'del' for duplication or deletion respectively

#### Value

NULL, but data of pnt1 is updated due to CNA

### **Examples**

```
pnt1 = tugHall_dataset$pnt_clones[[ 2 ]]  # pnt of allele A
  start_end = c( pnt1$Phys_pos - 50 , pnt1$Phys_pos + 50 )
  message( pnt1$Copy_number )
  change_allele_A_by_cna( pnt1, start_end, t = 'dup' )  # View( pnt1 )
  message( pnt1$Copy_number )
  change_allele_A_by_cna( pnt1, start_end, t = 'del' )  # View( pnt1 )
  message( pnt1$Copy_number )
```

change\_pnt\_by\_cna

Function to change the point mutation due to CNA

# **Description**

Function to change the point mutation due to CNA

# Usage

```
change_pnt_by_cna(pnt1, start_end, t)
```

### **Arguments**

pnt1 Object of class 'Point\_Mutations'
start\_end Vector with initial and final positions of CNA
t 'dup' or 'del' for duplication or deletion respectively

# Value

NULL, but pnt1 data is updated due to CNA

```
pnt1 = tugHall_dataset$pnt_clones[[ 1 ]]
start_end = c( pnt1$Phys_pos - 50 , pnt1$Phys_pos + 50 )
message( pnt1$Copy_number )
change_pnt_by_cna( pnt1, start_end, t = 'dup' ) # View( pnt1 )
message( pnt1$Copy_number )
change_pnt_by_cna( pnt1, start_end, t = 'del' ) # View( pnt1 )
message( pnt1$Copy_number )
```

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check\_pkg

Check the installation of a package for some functions

# **Description**

Check the installation of a package for some functions

# Usage

```
check_pkg(pkg)
```

# **Arguments**

pkg

Package name

# Value

if the package is installed then it returns NULL else it returns error message

# **Examples**

```
check_pkg( pkg = 'grDevices' )
```

check\_pnts

Function to check what pnts do fall into the range?

# Description

Function to check what pnts do fall into the range?

### Usage

```
check_pnts(gm_w1)
```

# **Arguments**

gm\_w1

A row from data.frame gene\_map

# Value

Return the point mutations which fall into the range

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gene_map$pnts[6] = '112792451, 112792442'
gm_w1 = gene_map[6,]
check_pnts( gm_w1 )
```

8 Clone-class

<b>-</b> 1 <b>-</b>	n to check point mutations match or don't match into duplica- deletion
---------------------	---

# **Description**

Function to check point mutations match or don't match into duplication or deletion

# Usage

```
chk_pnt_mut(pnt1, Ref_start, Ref_end, Chr, prntl)
```

# **Arguments**

pnt1 Object of class 'Point\_Mutations'
Ref\_start Initial position of CNA
Ref\_end Final position of CNA
Chr Chromosome name
prnt1 Parental chromosome 1 or 2

#### Value

Logical: TRUE if point mutation matches CNA, FALSE if it doesn't match

# **Examples**

```
pnt1 = tugHall_dataset$pnt_clones[[ 5 ]]
pstn = pnt1$Phys_pos[1]
message( pstn )
prnt1 = pnt1$Parental_1or2
Chr = pnt1$Chr
chk_pnt_mut( pnt1 , Ref_start = pstn - 200, Ref_end = pstn + 200, Chr, prnt1 )
chk_pnt_mut( pnt1 , Ref_start = pstn - 200, Ref_end = pstn - 100, Chr, prnt1 )
```

Clone-class

Class 'Clone' for clones

# **Description**

Class 'Clone' for clones

#### Fields

```
id numeric. ID of a clone
parent numeric. Parent ID (for first - 0)

N_cells numeric. Number of cells in clone
c numeric. Split counter as average value for all cells in clone
d numeric. Probability of division
```

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```
i numeric. Probability of Hayflick limit
```

m numeric. Probability that gene normal function is destroyed due to epigenome abnormality / mutation rate

a numeric. Probability of apoptosis for a cell in the clone

s numeric. Coefficient in the sigmoid function of the mutation density

k numeric. Probability of cell death by environment

E numeric. Coefficient of friction term against to the split probability.

Nmax numeric. Coefficient for determination the max number of cells that can exist in the primary tumor (Nmax = 1/E)

im numeric. Probability of the invasion/ metastatic transformation

Ha numeric. Apoptosis hallmark value

Him numeric. Invasion/ metastasis hallmark

Hi numeric. Mitotic restriction hallmark (immortalization hallmark)

Hd numeric. Growth/antigrowth hallmark (division rate hallmark)

Hb numeric. Angiogenesis hallmark

gene numeric. Vector of flags for each genes if they have driver mutation

pasgene numeric. Vector of flags for each genes if they have passenger mutation

PointMut\_ID numeric. ID of point mutation in list of objects of class 'Point\_Mutations'

CNA\_ID numeric. ID of CNA mutation in list of objects of class 'CNA\_Mutations'

mutden numeric. Gene mutation density

invasion logical. Indicator that clone is metastatic (invasion/metastatic transformation occured or not)

primary logical. Logical variable is clone primary tumor or not (normal)

birthday numeric. Time step of birth of clone

# **Examples**

```
clone = tugHall_dataset$clones[[ 1 ]]
print(clone$Ha)
print(clone$N_cells)
clone$calcApoptosis()  # to calculate apoptosis death probability based on mutation density
```

clone\_copy

Function to make one copy for clone1 in clone\_init function

# Description

Function to make one copy for clone1 in clone\_init function

```
clone_copy(clone1)
```

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# **Arguments**

clone1 Object of class 'Clone'

### Value

New object of class 'Clone' with the same info and new ID

# **Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
env = tugHall_dataset$env
define_parameters()
clone_copy(clone1)
```

CNA\_Mutations-class

Class 'CNA\_Mutations'

# **Description**

Class 'CNA\_Mutations'

# **Fields**

```
CNA_ID numeric. ID of CNA mutation

Parental_1or2 numeric. Parental chromosome, could be 1 or 2

dupOrdel character. dup for duplication or del for deletion

Chr character. Chromosome name

Ref_start numeric. Reference start position

Ref_end numeric. Reference final position

Gene_names character. Names of genes involved in CNA

MalfunctionedByCNA logical.

mut_order numeric. True for driver mutation and False for passenger mutation
```

```
cna = tugHall_dataset$cna_clones[[ 1 ]]
cna$safe()  # to save as row of data.frame
cna$copy()
cna$initialize()
cna$show()  # After initialization
```

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copy\_CNA

Function to copy CNA info

# **Description**

Function to copy CNA info

# Usage

```
copy_CNA(CNA1)
```

# **Arguments**

CNA<sub>1</sub>

Object of class 'CNA\_Mutations'

#### Value

The same object of class 'CNA\_Mutations'

# **Examples**

```
cna = tugHall_dataset$cna_clones[[ 1 ]]
cna2 = copy_CNA( cna )
cna$safe()
cna2$safe()
```

copy\_files\_to\_Input

Function to copy the files by default from extdata folder in the library to Input folder in the working directory

# Description

Function to copy the files by default from extdata folder in the library to Input folder in the working directory

# Usage

### **Arguments**

```
Files to copy, vector of names of files by default:

files = c( 'CCDS.current.txt', 'CF.txt', 'cloneinit.txt', 'gene_hallmarks.txt', 'gene_map.txt', 'parameter
)

dir Folder to where files should be save, by default dir = 'Input'
```

#### Value

List of logic numbers for each copied file, TRUE - success, FALSE - not success

# **Examples**

```
\label{eq:files} files = \texttt{c('CF.txt', 'cloneinit.txt', 'gene\_hallmarks.txt', 'gene\_map.txt', 'parameters.txt')}
copy_files_to_Input( files, dir = 'Input' )
```

copy\_files\_to\_Output Function to copy the files of an example of simulation or from '/extdata/Output/' folder in the library to '/Output/' folder in the working directory

# **Description**

Function to copy the files of an example of simulation or from '/extdata/Output/' folder in the library to '/Output/' folder in the working directory

# Usage

```
copy_files_to_Output(
 files = c("cloneout.txt", "CNA_mutations.txt", "point_mutations.txt", "gene_MAP.txt",
   "geneout.txt", "log.txt", "order_genes_dysfunction.txt", "VAF_data.txt", "VAF.txt",
    "weights.txt"),
  dir = "Output"
```

### **Arguments**

dir

files Files to copy, vector of names of files by default: files = c('cloneout.txt', 'CNA\_mutations.txt', 'point\_mutations.txt', 'gene\_MAP.txt', 'geneout.txt', 'lo 'order\_genes\_dysfunction.txt', 'VAF\_data.txt', 'VAF.txt', 'weights.txt') Folder to where files should be save, by default dir = 'Output'

# Value

List of logic numbers for each copied file, TRUE - success, FALSE - not success

```
files = c( 'cloneout.txt', 'CNA_mutations.txt', 'point_mutations.txt', 'gene_MAP.txt')
copy_files_to_Output( files )
files = c('geneout.txt','log.txt', 'VAF_data.txt', 'VAF.txt', 'weights.txt')
copy_files_to_Output( files )
```

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copy\_pnt

Function to copy of point mutation info

# **Description**

Function to copy of point mutation info

# Usage

```
copy_pnt(pnt1)
```

# **Arguments**

pnt1

Object of class 'Point\_Mutations'

### Value

The same object of class 'Point\_Mutations' with different ID

# **Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
copy_pnt( pnt ) # View( pnt )
```

copy\_pnt\_no\_mutation

Function to copy of pnt1 without mutation info for allele A

# Description

Function to copy of pnt1 without mutation info for allele A

### Usage

```
copy_pnt_no_mutation(pnt1)
```

### **Arguments**

pnt1

Object of class 'Point\_Mutations'

# Value

Object of class 'Point\_Mutations' for another chromosome

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
copy_pnt_no_mutation( pnt ) # View( pnt )
```

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```
define_compaction_factor
```

Define compaction factor

# Description

Define compaction factor

### Usage

```
define_compaction_factor(
  cf = data.frame(Ha = 1, Hb = 1, Hd = 1, Hi = 1, Him = 1),
  read_fl = TRUE,
  file_name = "./Input/CF.txt"
)
```

### **Arguments**

```
cf Data frame with compaction factors for all the hallmarks, for example, data.frame(
Ha = 1, Hb = 1, Hd = 1, Him = 1)

read_fl Indicator to read file or not, logical type only

file_name File name to rad all the parameters, it is used only if read_fl == TRUE
```

#### Value

Data frame with with compaction factors for all the hallmarks

# Examples

```
copy_files_to_Input()
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
CF1 = CF
cf = data.frame( Ha = 0.1, Hb = 0.2, Hd = 0.7, Hi = 1, Him = 0.5 )
define_compaction_factor( cf = cf, read_fl = FALSE ) # View( c( CF, CF1 ) ) to compare
```

define\_files\_names

Function to define all the files names

# Description

Function to define all the files names

```
define_files_names(
  mainDir = getwd(),
  sbdr_Input = "/Input",
  sbdr_Output = "/Output"
)
```

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### **Arguments**

mainDir Working directory
sbdr\_Input Sub directory for input files
sbdr\_Output Sub directory for output files

# Value

NULL, but all file names are defined in GLOBAL environment

# **Examples**

```
define_files_names()
```

# **Description**

Define genes' location in chromosome

# Usage

```
define_gene_location(
  file_input = "Input/CCDS.current.txt",
  genes_list = c("CCDS4107.1", "CCDS8702.1", "CCDS43171.1", "CCDS11118.1")
)
```

### **Arguments**

file\_input is a name of file to input where the information about genes location is defined.

That is loaded from CCDS database https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current\_human/

genes\_list is a list of genes' names like CCDS4107.1 in the CCDS database.

#### Value

Function returns the table of genes' locations in DNA

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define\_parameters

Define all the parameters for a simulation

# **Description**

Define all the parameters for a simulation

# Usage

```
define_parameters(
  E0 = 1e-04,
 F0 = 10,
  m0 = 1e-07,
  uo = 0.9,
  us = 0.9,
  s0 = 10,
  k0 = 0.12,
  d0 = 0.4
  censore_n = 10^5,
  censore_t = 50,
  m_{dup} = 1e-08,
  m_{del} = 1e-08,
  lambda_dup = 5000,
  lambda_del = 7000,
  uo_dup = 0.8,
  us_dup = 0.5,
  uo_del = 0,
  us_del = 0.8,
  CF = TRUE,
 model = c("proportional_metastatic", "threshold_metastatic", "simplified")[1],
  time\_stop = 120,
  read_f1 = FALSE,
  file_name = "./Input/parameters.txt",
  n_repeat = 1000,
  monitor = TRUE
)
```

# Arguments

E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only
m0	Mutation probability for point mutation, numeric type only
uo	Oncogene mutation probability, numeric type only
us	Suppressor mutation probability, numeric type only
s0	Parameter in the sigmoid function, numeric type only
k0	Environmental death probability, numeric type only
d0	Initial probability to divide cells, numeric type only
censore_n	Max cell number where the program forcibly stops, integer type only

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censore_t	Max time where the program forcibly stops, integer type only
m_dup	Mutation probability for duplication, numeric type only
m_del	Mutation probability for deletion, numeric type only
lambda_dup	CNA duplication average length (of the geometrical distribution for the length), integer type only
lambda_del	CNA deletion average length (of the geometrical distribution for the length), integer type only
uo_dup	Gene malfunction probability by CNA duplication for oncogene, numeric type only
us_dup	Gene malfunction probability by CNA duplication for suppressor, numeric type only
uo_del	Gene malfunction probability by CNA deletion for oncogene, numeric type only
us_del	Gene malfunction probability by CNA deletion for suppressor, numeric type only
CF	Compaction factor, logical type only. True means 'to use', False means 'do not use' Compaction factor for hallmarks variables
model	Name of the model to use. Can be 'proportional_metastatic' or 'threshold_metastatic or 'simplified'
time_stop	Max time in seconds of running after that the program forcibly stops, integer type only
read_fl	Indicator to read file or not, logical type only
file_name	File name to rad all the parameters, it is used only if read_fl == TRUE
n_repeat	Max number of repetition of the program until the NON-ZERO output will be getting, integer type only
monitor	The indicator to make monitor file during a simulation or do not make, logical type only

# Value

Values of all the parameters

# **Examples**

```
copy_files_to_Input()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_parameters( read_fl = FALSE )
```

define\_par\_for\_plot Function to change par for plots and after plotting it returns par values

# Description

Function to change par for plots and after plotting it returns par values

```
define_par_for_plot(change_par_back)
```

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### **Arguments**

```
change_par_back
```

logical. If TRUE it changes par options back after finishing a function call

### Value

Change par() options and returns it (after finishing the function) to values before a function has started

# **Examples**

```
define_par_for_plot( change_par_back = TRUE )
```

Environ-class

Class 'Environ'

# **Description**

Class 'Environ'

#### **Fields**

T numeric. Time counter

N numeric. Number of normal cells

P numeric. Number of primary tumor cells

M numeric. Number of metastatic cells

F numeric. Coefficient that determines the maximal number of cells in pool of primary tumor cells

c numeric. Average number of divisions in pool of clones

d numeric. Mean value of splitting probability

i numeric. Average value of immortalization probability

a numeric. Average value of apoptosis probability

k numeric. Average probability of cell death via environment death

E numeric. Average value of coefficients of friction term

Nmax numeric. Maximal number of primary tumor cells that can exist in pool of clones

im numeric. Average value of invasion/metastasis probability

Ha numeric. Average value of apoptosis hallmark Ha

Him numeric. Average value of invasion/metastasis hallmark Him

Hi numeric. Average value of immortalization hallmark Hi

Hd numeric. Average value of growth/antigrowth hallmark Hd

Hb numeric. Average value of angiogenesis hallmark Hb

type numeric. Invasion / metastatic ratio

gene numeric. Cancer gene damage rate

mutden numeric. Average mutation rate

last\_id numeric. Maximal ID in the pool of clones.

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# **Examples**

```
env = tugHall_dataset$env
print( env )
env$initFields()
```

generate\_cna

Function to generate object of CNA mutation

# **Description**

Function to generate object of CNA mutation

# Usage

```
generate_cna(prntl, genes, start_end, onco1, dupOrdel)
```

# **Arguments**

prntl The 1st or 2nd parental chromosome

genes Genes names

start\_end vector with start and final positions of CNA

onco1 Object of class 'OncoGene'

dupOrdel It could be 'dup' or 'del' to denote duplication or deletion

# Value

Object of class 'CNA\_Mutations'

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
gene_map = tugHall_dataset$gene_map
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234
start_end = c(112775658, 112775716 )
generate_cna( prntl = 1, genes = 'APC', start_end = start_end, onco1, dupOrdel = 'dup' )
```

generate.	pnt	$F\iota$

Function to generate an object of class 'Point\_Mutations'

### **Description**

Function to generate an object of class 'Point\_Mutations'

### Usage

```
generate_pnt(prntl, gene, pos, onco1, Chr, mutation = NA)
```

#### **Arguments**

prnt1 Parental chromosome, could be 1 or 2

gene Gene name

pos Position of point mutation onco1 Object of class 'OncoGene'

Chr Chromosome name

mutation If mutation is NOT NA then MalfunctionedByPointMut = TRUE, else it is de-

fined by corresponding probabilities

### Value

Object of class 'Point\_Mutations'

### **Examples**

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
pnt_clones = tugHall_dataset$pnt_clones
copy_files_to_Input()
define_parameters()
mut_order = 234  # As an example
pnt1 = generate_pnt( prntl = 1, gene = 'APC', pos = 112767192, onco, Chr = '5', mutation = NA )
```

 ${\tt generate\_to\_copy\_pnt}$ 

Function to generate the same object of class 'Point\_Mutations' with coping all information from input object

# Description

Function to generate the same object of class 'Point\_Mutations' with coping all information from input object

```
generate_to_copy_pnt(pnt)
```

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# Arguments

pnt

Object of class 'Point\_Mutations'

### Value

The same object of class 'Point\_Mutations' with different ID

# **Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
pnt_clones = tugHall_dataset$pnt_clones
pnt2 = generate_to_copy_pnt( pnt )
```

gen\_colors

Function to make a large number of colors

# Description

Function to make a large number of colors

# Usage

```
gen\_colors(nm = 12)
```

# **Arguments**

nm

Number of colors

# Value

Vector of colors with length more than nm

# **Examples**

```
clrs = gen_colors( nm = 120 )
```

get\_cds\_rna

Function to get length of CDS and of genes from data.frame gene\_map and related probabilities

# Description

Function to get length of CDS and of genes from data.frame gene\_map and related probabilities

### Usage

```
get_cds_rna(gm)
```

### **Arguments**

gm

data.frame gene\_map with info about genes' location

22 get\_cna\_mutation

### Value

```
list( names, CDS, RNA, PROB, SUM, P0 )
```

# **Examples**

```
gene_map = tugHall_dataset$gene_map
define_parameters()
get_cds_rna( gm = gene_map )
```

get\_cna\_mutation

Generation CNA mutation info

# **Description**

Generation CNA mutation info

# Usage

```
get_cna_mutation(onco1, dupOrdel, gm_1_2)
```

### **Arguments**

onco1 Object of class 'OncoGene'

dupOrdel It could be 'dup' or 'del' to denote duplication or deletion

gm\_1\_2 List of two data frames (for 1st and 2nd parental chromosomes) with genes' location information

### Value

List of (prntl - 1 or 2 parental chromosome, Chr - name of chromosome, genes - genes names, start\_end - vector with start and end positions of CNA, w\_cna - rows of CNA in gene\_map data frame)

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
gm = tugHall_dataset$gene_map
get_cna_mutation( onco1 = onco, dupOrdel = 'dup', gm_1_2 = list(gm, gm) )
get_cna_mutation( onco1 = onco, dupOrdel = 'del', gm_1_2 = list(gm, gm) )
```

get\_flow\_data 23

get\_flow\_data Function to get data about last simulation from cloneoutfile

# **Description**

Function to get data about last simulation from cloneoutfile

# Usage

```
get_flow_data(
  cloneoutfile,
  genefile,
  mainDir = getwd(),
  sbdr_Output = "/Output"
)
```

# **Arguments**

cloneoutfile Name of file to read data about clone evolution

genefile Name of file with hallmarks values

mainDir Working directory, by default mainDir = getwd()
sbdr\_Output Directory for output data getting from mainDir

### Value

list of data.frames like onco, hall, data\_last (data of last time step), data\_avg (average data for all time steps), data\_flow (data without average rows), time\_max (max time step), pnt\_mut and pnt\_mut\_B (data.frame of point mutations for both alleles and for allele B only ) and cna\_mut (data.frame of CNA mutations)

```
copy_files_to_Input()
define_files_names()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
define_gene_location()
copy_files_to_Output()
dataset = get_flow_data(cloneoutfile, genefile, mainDir = getwd(), sbdr_Output = '/Output' )
# View(dataset)
```

Function to get length of CDS and whole gene from gene\_map get\_len\_cds\_rna data.frame

# **Description**

Function to get length of CDS and whole gene from gene\_map data.frame

# Usage

```
get_len_cds_rna(gene_map)
```

# **Arguments**

gene\_map data.frame with info about genes' locations

#### Value

list of (Name, CDS, LEN\_Genes) where Name is a vector of genes' names, CDS is a vector of CDS lengths, LEN\_Genes is a vector of length of whole genes including introns and exons

### **Examples**

```
gene_map = tugHall_dataset$gene_map
onco = tugHall_dataset$onco
get_len_cds_rna( gene_map)
```

```
get_order_of_genes_dysfunction
```

Function to get order of genes' dysfunction

### **Description**

Function to get order of genes' dysfunction

### Usage

```
get_order_of_genes_dysfunction(
  pnt_mut,
  data_last,
  cna_mut,
  file_name = "./Output/order_genes_dysfunction.txt"
```

# **Arguments**

pnt\_mut data.frame with info about all the point mutations data\_last data.frame with data of simulation at the last time step data.frame with info about all the CNA mutations cna\_mut

Name of file to save data file\_name

get\_point\_mutation 25

### Value

data.frame of genes' dysfunction and save it in a file

# **Examples**

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
cna_mut = tugHall_dataset$cna_mut
file_name = './Output/order_genes_dysfunction.txt'
rdr = get_order_of_genes_dysfunction( pnt_mut, data_last, cna_mut, file_name = file_name )
```

get\_point\_mutation

Generation point mutation info

# Description

Generation point mutation info

### Usage

```
get_point_mutation(onco1, gm_1_2)
```

# Arguments

onco1	Object of class 'OncoGene'
gm_1_2	List of two data frames (for 1st and 2nd parental chromosomes) with genes'
	location information

# Value

list of (prntl - 1 or 2 parental chromosome, gene - gene name, pos - position of point mutation, Chr - name of chromosome )

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
get_point_mutation( onco, gm_1_2 )
```

get\_rho\_VAF

```
get_point_mutation_for_gene
```

Generation point mutation info for the particular gene

# Description

Generation point mutation info for the particular gene

# Usage

```
get_point_mutation_for_gene(onco1, gm_1_2, gene)
```

# **Arguments**

onco1 Object of class 'OncoGene'

gm\_1\_2 List of two data frames (for 1st and 2nd parental chromosomes) with genes'

location information

gene Gene's name where point mutation should be occured

#### Value

list of (prntl - 1 or 2 parental chromosome, gene - gene name, pos - position of point mutation, Chr - name of chromosome )

### **Examples**

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
get_point_mutation_for_gene( onco, gm_1_2, gene = 'APC')
get_point_mutation_for_gene( onco, gm_1_2, gene = 'KRAS')
```

get\_rho\_VAF

Function to get Variant allele frequencies (VAF) based on rho input parameters

# Description

Function to get Variant allele frequencies (VAF) based on rho input parameters

# Usage

```
get_rho_VAF(vf = NULL, rho = c(0, 0.1, 0.5), file_name = "./Output/VAF.txt")
```

# Arguments

vf data.frame getting from get\_VAF() function rho Vector of rho parameter in the range (0,1)

file\_name Name of file to save VAF

get\_type 27

### Value

VAF for different rho with separation for metastatic cells and (primary tumor + speckled normal) cells

# **Examples**

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
if ( !dir.exists('./Output') ) dir.create('./Output')
vf = get_VAF( pnt_mut, data_last, file_name = 'Output/VAF_data.txt')
VAF = get_rho_VAF( vf = vf, rho = c( 0.0, 0.1, 0.5 ) , file_name = './Output/VAF.txt' )
```

get\_type

Function to get type of the clone: normal, primary or metastatic

# Description

Function to get type of the clone: normal, primary or metastatic

### Usage

```
get_type(clone1)
```

#### **Arguments**

clone1

Object of class 'Clone'

# Value

One of characters 'normal', 'primary' or 'metastatic'

# **Examples**

```
clone1 = tugHall_dataset$clones[[1]]
get_type( clone1 )
clone1 = tugHall_dataset$clones[[56]]
get_type( clone1 )
```

get\_u\_cna

Function to choose probability of CNA mutation for several genes

# Description

Function to choose probability of CNA mutation for several genes

```
get_u_cna(genes, dupOrdel)
```

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### **Arguments**

genes Names of genes, vector of names

dupOrdel It could be 'dup' or 'del' to denote duplication or deletion

### Value

Single value of maximal probability from probabilities for several genes

### **Examples**

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
get_u_cna( genes = 'APC', dupOrdel = 'dup' )
get_u_cna( genes = c('KRAS', 'APC'), dupOrdel = c('dup', 'del') )
```

get\_VAF

Function to get data about Variant allele frequencies (VAF)

# **Description**

Function to get data about Variant allele frequencies (VAF)

# Usage

```
get_VAF(pnt_mut, data_last, file_name = "Output/VAF_data.txt")
```

### **Arguments**

pnt\_mut data.frame with point mutation info

data\_last data.frame with data of simulation at the last time step

file\_name Name of file to save data

### Value

data.frame with info about Variant allele frequencies

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
vf = get_VAF( pnt_mut, data_last, file_name = 'Output/VAF_data.txt')
```

HallMark-class 29

HallMark-class

Class 'HallMark'

### **Description**

Class 'HallMark'

### **Fields**

Ha numeric. Apoptosis hallmark indexes of genes in onco\$name, where onco is object of class OncoGene

Hi numeric. Immortalization hallmark indexes of genes in onco\$name, where onco is object of class OncoGene

Hd numeric. Growth/antigrowth hallmark indexes of genes in onco\$name, where onco is object of class OncoGene

Hb numeric. Angiogenesis hallmark indexes of genes in onco\$name, where onco is object of class OncoGene

Him numeric. Invasion/metastatic transformation hallmark indexes of genes in onco\$name, where onco is object of class OncoGene

Ha\_w numeric. Apoptosis hallmark weights of genes

Hi\_w numeric. Immortalization hallmark weights of genes

Hd\_w numeric. Growth/antigrowth hallmark weights of genes

Hb\_w numeric. Angiogenesis hallmark weights of genes

Him\_w numeric. Invasion/metastatic transformation hallmark weights of genes

notHa numeric. Indexes of genes which are not in apoptosis hallmark

# Examples

```
hall = tugHall_dataset$hall
print( hall )
hall$copy()
hall$show()
```

init\_clones

Function to read file with initial clones

### **Description**

Function to read file with initial clones

# Usage

```
init_clones(clonefile, clone1)
```

# **Arguments**

clonefile File to read

clone1 Object of class 'Clone'

init\_onco\_clones

### Value

List of objects of class 'Clone

# **Examples**

```
copy_files_to_Input()
define_files_names()
env = tugHall_dataset$env
define_parameters()
onco = tugHall_dataset$onco
clone1 = tugHall_dataset$clones[[ 1 ]]
clones = init_clones(clonefile, clone1)  # View( clones )
```

init\_onco\_clones

Function to make list of objects of class 'OncoGene' and generate initial onco settings for all clones (onco\_clones)

# Description

Function to make list of objects of class 'OncoGene' and generate initial onco settings for all clones (onco\_clones)

# Usage

```
init_onco_clones(onco1, clones)
```

# **Arguments**

onco1 Object of class 'OncoGene'
clones List of objects of class 'Clone'

### Value

List of objects of class 'OncoGene'

```
copy_files_to_Input()
define_files_names()
env = tugHall_dataset$env
define_parameters()
onco = tugHall_dataset$onco
clone1 = tugHall_dataset$clones[[ 1 ]]
clones = init_clones(clonefile, clone1)  # View( clones )
onco_clones = init_onco_clones( onco1 = onco, clones )
```

init\_pnt\_clones 31

init\_pnt\_clones

Function to generate point mutations for initial clones

# **Description**

Function to generate point mutations for initial clones

# Usage

```
init_pnt_clones(clones, onco_clones)
```

# **Arguments**

clones List of objects of class 'Clone' onco\_clones List of objects of class 'OncoGene'

# **Examples**

```
clones = tugHall_dataset$clones
define_parameters()
onco = tugHall_dataset$onco
onco_clones = tugHall_dataset$onco_clones
copy_files_to_Input()
copy_files_to_Output()
define_gene_location()
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234
## Not run:
init_pnt_clones( clones, onco_clones ) # change pnt_clones for initialization
## End(Not run)
```

make\_map

Function to make a gene\_map data.frame with information of genes' locations

# Description

Function to make a gene\_map data.frame with information of genes' locations

```
make_map(
    f_out = "Input/map.txt",
    ls = c("CCDS4107.1", "CCDS8702.1", "CCDS43171.1", "CCDS11118.1"),
    f_in = "Input/CCDS.current.txt"
)
```

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#### **Arguments**

f_out	Name of file to save gene_map data.frame
ls	List of IDs of genes corresponding CCDS database https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_ht
f_in	Name of file to input downloaded from CCDS database

# Value

gene\_map data.frame with information of genes' locations for genes of interest

# **Examples**

```
url = 'https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_human/CCDS.current.txt'
download.file( url = url, destfile = 'CCDS.current.txt')
ls = c( 'CCDS4107.1', 'CCDS8702.1', 'CCDS43171.1', 'CCDS11118.1' )
gene_map = make_map(f_out = 'map.txt', ls = ls, f_in = 'CCDS.current.txt' )
```

mixed\_mut\_order

Function to get order of mutation for all possible types

# **Description**

Function to get order of mutation for all possible types

# Usage

```
mixed_mut_order(clone1)
```

# Arguments

clone1 Object of class 'Clone'

#### Value

data.frame with fields order, type, ID

```
clone = tugHall_dataset$clones[[ 46 ]]
clone$PointMut_ID
clone$CNA_ID
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mixed_mut_order( clone )
```

model 33

model Main function 'model' to simulate clones' evolution

# Description

Main function 'model' to simulate clones' evolution

# Usage

```
model(
  genefile,
  clonefile,
  geneoutfile,
  cloneoutfile,
  logoutfile,
  Ε0,
  F0,
  m0,
  uo,
  us,
  s0,
  k0,
  censore_n,
  censore_t,
  d0
)
```

# Arguments

genefile	Name of file with input data of hallmarks
clonefile	Name of file with initial clones data
geneoutfile	Name of file with hallmark data
cloneoutfile	Name of file with cloneout data
logoutfile	File name of log file
E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only
m0	Mutation probability for point mutation, numeric type only
uo	Oncogene mutation probability, numeric type only
us	Suppressor mutation probability, numeric type only
s0	Parameter in the sigmoid function, numeric type only
k0	Environmental death probability, numeric type only
censore_n	Max cell number where the program forcibly stops, integer type only
censore_t	Max time where the program forcibly stops, integer type only
d0	Initial probability to divide cells, numeric type only

34 modify\_gene\_map

#### Value

List of (clones, onco\_clones), where clones - list of objects of class 'Clone', and onco\_clones - list of objects of class 'OncoGene'. During a simulation it saves data to geneoutfile.

### **Examples**

```
copy_files_to_Input()
define_files_names()
define_gene_location()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
time_stop = 3  # Duration of simulation time is 3 sec
## Not run:
res = model( genefile, clonefile, geneoutfile, cloneoutfile, logoutfile,
E0, F0, m0, uo, us, s0, k0, censore_n, censore_t, d0 )
## End(Not run)
```

modify\_gene\_map

Function to add the mutations to the data.frame gene\_map

# **Description**

Function to add the mutations to the data.frame gene\_map

# Usage

```
modify_gene_map(clone1, onco1)
```

### **Arguments**

```
clone1 Object of class 'Clone'
onco1 Object of class 'OncoGene'
```

### Value

list(gm1, gm2), where gm1 and gm2 are data.frames gene\_maps with mutation information

```
clone = tugHall_dataset$clones[[ 46 ]]
onco = tugHall_dataset$onco_clones[[ 46 ]]
gene_map = tugHall_dataset$gene_map
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
gene_map$pnts = ''
## Not run:
gm_1_2 = modify_gene_map( clone , onco ) # View(gm_1_2)
## End(Not run)
```

number\_N\_P\_M 35

number\_N\_P\_M

Function to get number of cells of a clone with indicator (normal, primary tumor or metastatic)

### **Description**

Function to get number of cells of a clone with indicator (normal, primary tumor or metastatic)

### Usage

```
number_N_P_M(clone1)
```

# **Arguments**

clone1

Object of class 'Clone'

#### Value

```
Vector c( N_normal, N_primary, N_metastatic )
```

### **Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
number_N_P_M(clone1)
message( paste('Format is as follow: ', 'N_normal', 'N_primary', 'N_metastatic' ) )
```

OncoGene-class

Class 'OncoGene'

### **Description**

Class 'OncoGene'

# **Fields**

```
id numeric. ID is same as in clone (key for clones)

name character. Onco genes' names list

onsp character. Oncogene/suppressor indicator for each gene in list of names

len numeric. Lengths of onco genes

cds_1 numeric. Onco genes' CDS base lengths for parental chr 1

cds_2 numeric. Onco genes' CDS base lengths for parental chr 2

rna_1 numeric. Onco genes RNA base number length for parental chr 1 (exons+introns)

rna_2 numeric. Onco genes RNA base number length for parental chr 2 (exons+introns)

p0_1 numeric. Probability of absent of mutations for parental chr 1

p0_2 numeric. Probability of absent of mutations for parental chr 2

prob_1 numeric. Vector of relative probabilities for point mutation, deletion and duplication:

prob = c( m0 x sumCDS, m_del x sumRNA, m_dup x sumRNA ) / sum( m0 x sumCDS, m_del x sumRNA, m_dup x sumRNA)
```

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```
prob_2 numeric.
sum_prob_1 numeric.
sum_prob_2 numeric.
```

### **Examples**

```
onco = tugHall_dataset$onco
onco$copy()
```

onco\_copy

Function to make one copy for onco1 in init\_onco\_clones function

# **Description**

Function to make one copy for onco1 in init\_onco\_clones function

# Usage

```
onco_copy(onco1)
```

# **Arguments**

onco1

Object of class 'OncoGene'

# Value

New object of class 'OncoGene' with the same info

# **Examples**

```
onco1 = tugHall_dataset$onco_clones[[ 1 ]]
onco2 = onco_copy( onco1 ) # ID + 1
```

onco\_update

Function to update oncol after mutation (for usage in trial\_mutagenesis() function)

# **Description**

Function to update onco1 after mutation (for usage in trial\_mutagenesis() function)

# Usage

```
onco_update(onco1, gm)
```

# Arguments

onco1 Object of class 'OncoGene' gm data.frame gene\_map order\_gene\_map 37

#### Value

onco1 with updated info

## **Examples**

```
onco1 = tugHall_dataset$onco_clones[[ 1 ]]
copy_files_to_Input()
define_gene_location()
define_parameters()
onco_update( onco1, gm = list(gene_map, gene_map[1:42, ] ) ) # Check CDS length for TP53 gene
```

order\_gene\_map

Function to order info in gene\_map data.frame with information of genes' locations

# Description

Function to order info in gene\_map data.frame with information of genes' locations

## Usage

```
order_gene_map(gene_map)
```

#### **Arguments**

gene\_map

data.frame with information of genes' locations

#### Value

The same data.frame gene\_map with ordered positions for each gene and each chromosome

# Examples

```
gene_map = tugHall_dataset$gene_map
gene_map = order_gene_map( gene_map )
```

plot\_2D

Function to plot 2D figure of points y = y(x)

## **Description**

Function to plot 2D figure of points y = y(x)

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#### Usage

```
plot_2D(
    x,
    y,
    names = c("X", "Y"),
    pch = 18,
    col = "blue",
    cex = 1.2,
    xr = c(-10, 10),
    yr = c(-10, 10),
    safe_pdf = FALSE,
    filename = "./plot.pdf",
    change_par = TRUE
)
```

## **Arguments**

X	Input data for axes X
٧	Input data for axes Y

names Vector of two characters with names for X and Y axes

pch Parameter pch for plot function

col Colors of points

cex Parameter cex for plot function

xr Range for X yr Range for Y

safe\_pdf Indicator to save plot to a file or not

filename Name of file to save plot if safe\_pdf == TRUE

change\_par Indicator to change par() or not for a plot. By default change\_par = TRUE, after

plot it will be returned to initial values

## Value

NULL, making 2D plot using points

## **Examples**

```
plot_2D(x=-5:5, y=-3:7)
```

plot\_2D\_lines

Function to plot 2D figure of lines  $y_i = DF[, nl[i]]$ ), i - index

# Description

Function to plot 2D figure of lines  $y_i = DF[,nl[i]]$ ), i - index

plot\_2D\_lines 39

## Usage

```
plot_2D_lines(
  х,
  DF,
  n1 = 1:2,
  names = c("X", "Y"),
  legend_names = "",
  col = c("blue3", "darkmagenta", "red", "green4", "darkorange", "steelblue1"),
  cex = 1.2,
  1wd = 2,
  lt = c(1:6),
  xr = c(-10, 10),
  yr = c(-10, 10),
  safe_pdf = FALSE,
  filename = "./plot.pdf",
  type = "1",
  logscale = "",
  draw_key = TRUE,
  change_par = TRUE
)
```

## Arguments

x	Input data for axes X
DF	data.frame with data to plot
nl	indexes of columns in DF to plot
names	Vector of two characters with names for X and Y axes
legend_names	Name of legend
col	Vector of colors for lines
cex	Parameter cex for plot function
lwd	Vector of width of lines
lt	Vector of types of lines
xr	Range for X
yr	Range for Y
safe_pdf	Indicator to save plot to a file or not
filename	Name of file to save plot if safe_pdf == TRUE
type	Parameter type in plot function
logscale	Parameter logscale in plot function
draw_key	Indicator to draw key or not
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after plot it will be returned to initial values

## Value

NULL, making 2D plot using lines

40 plot\_clone\_evolution

#### **Examples**

```
DF = tugHall_dataset$data_avg
plot_2D_lines( x = DF[, 1 ], DF, nl = 8:12 , xr = c(1,max(DF$Time) ), yr = c(0,1) )
xr = c(1,max(DF$Time) )
yr = c(0,max(DF[,14],DF[,16],DF[,17] ))
plot_2D_lines( x = DF[, 1 ], DF, nl = c(14,16,17) , xr =xr, yr = yr )
plot_2D_lines( x = DF[, 1 ], DF, nl = 18:22 , xr = c(1,max(DF$Time) ), yr = c(0,1) )
```

```
plot_average_simulation_data
```

Function to plot main data from data.frame with average data

## **Description**

Function to plot main data from data.frame with average data

## Usage

```
plot_average_simulation_data(data_avg, time_max)
```

## **Arguments**

data\_avg data.frame with average values from cloneout.txt file

time\_max Maximal time step in a simulation

#### Value

NULL, draw many plot with average data

# **Examples**

```
data_avg = tugHall_dataset$data_avg
time_max = tugHall_dataset$time_max
plot_average_simulation_data( data_avg , time_max = time_max )
```

```
plot_clone_evolution Function to plot clone evolution
```

## **Description**

Function to plot clone evolution

plot\_order\_dysfunction

#### Usage

```
plot_clone_evolution(
   data_flow,
   threshold = c(0.05, 1),
   lwd = 2,
   hue = c(" ", "random", "red", "orange", "yellow", "green", "blue", "purple", "pink",
        "monochrome")[1],
   luminosity = c(" ", "random", "light", "bright", "dark")[5],
   yr = NA,
   add_initial = TRUE,
   log_scale = FALSE,
   change_par = TRUE
)
```

## **Arguments**

data_flow	data.frame with results of simulation at each time step	
threshold	Vector two numbers from 0 to 1 to show clones with relative final numbers of cells in the range of threshold	
lwd	Line width in the plot function	
hue	Parameter hue in the function randomColor from library randomcoloR	
luminosity	Parameter luminosity in the function randomColor from library randomcoloR	
yr	Range for Y axes	
add_initial	Indicator to add or do not add initial clones to plot	
log_scale	Indicator to use log_scale or not for Y axes	
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after	

plot it will be returned to initial values

## Value

NULL, making plot with clones evolution

# Examples

```
data_flow = tugHall_dataset$data_flow plot_clone_evolution( data_flow, threshold = c(0.01, 1), add_initial = TRUE, log_scale = FALSE) plot_clone_evolution( data_flow, threshold = c(0, 0.01), add_initial = FALSE, log_scale = TRUE)
```

```
plot_order_dysfunction
```

Function to plot order of genes dysfunction as a step function with number of cells related to each order

## **Description**

Function to plot order of genes dysfunction as a step function with number of cells related to each order

42 pnts\_add\_dlt

#### Usage

```
plot_order_dysfunction(
  rdr_dysf,
  pos = c(0, 100),
  logscale = "y",
  cex = 1,
  change_par = TRUE
)
```

## **Arguments**

rdr\_dysf Order of genes dysfunction as a data.frame

pos Coordinates of list of order of genes dysfunction

logscale Parameter logscale for plot function
cex Parameter cex for plot function

change\_par Indicator to change par() or not for a plot. By default change\_par = TRUE, after

plot it will be returned to initial values

#### Value

NULL, making plot with step function of order of genes' dysfunction

#### **Examples**

```
rdr_dysf = tugHall_dataset$rdr_dysf
plot_order_dysfunction( rdr_dysf , logscale = '', pos = c(8, 5000), cex = 1.4)
plot_order_dysfunction( rdr_dysf , logscale = 'y', pos = c(10, 100), cex = 1.2)
```

pnts\_add\_dlt

Function to subtract delta from position of point mutations

## **Description**

Function to subtract delta from position of point mutations

#### Usage

```
pnts_add_dlt(gm_w1, dlt)
```

#### **Arguments**

gm\_w1 A row from data.frame gene\_map

dlt Delta to subtract from positions of point mutations

## Value

Return the pnts - dlt for one row of data.frame gene\_map

Point\_Mutations-class 43

#### **Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gene_map$pnts[6] = '112792451'
gm_w1 = gene_map[6,]
pnts_add_dlt( gm_w1 , dlt = 1000 )
pnts_add_dlt( gm_w1 , dlt = -1001 )
```

Point\_Mutations-class Class 'Point\_Mutations'

# Description

Class 'Point\_Mutations'

## **Fields**

```
PointMut_ID numeric. ID of point mutation

Allele character. A or B allele

Parental_1or2 numeric. Parental chromosome, could be 1 or 2

Chr character. Chromosome name

Ref_pos numeric. Reference position

Phys_pos vector. Physical positions

Delta vector. Delta of positions

Copy_number numeric. Copy number of allele

Gene_name character. Gene's name

MalfunctionedByPointMut logical. True for driver mutation and False for passenger mutation

mut_order numeric. Number in order of mutation to reproduce the gene_map data.frame
```

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
print( pnt )
pnt$copy()
pnt$show()
pnt$initialize()
pnt$show()
pnt = tugHall_dataset$pnt_clones[[ 3 ]]
pnt$safe()  # save as row of data.frame
```

44 read\_file

print\_parameters

Function to print GLOBAL parameters

## **Description**

Function to print GLOBAL parameters

## Usage

```
print_parameters()
```

#### Value

Message with values of all the GLOBAL parameters

## **Examples**

```
copy_files_to_Input()
define_parameters( read_fl = FALSE )
define_compaction_factor()
print_parameters()
```

read\_file

Function to read file

## **Description**

Function to read file

# Usage

```
read_file(file_name = "", stringsAsFactors = FALSE, header = TRUE)
```

## **Arguments**

file\_name Name of file to read stringsAsFactors

Parameter for read.table function, by default stringsAsFactors = FALSE

header Logical type to read or do not read head of a file

#### Value

data.frame of data from a file

```
fl = system.file('extdata/Input', 'gene_map.txt',package = 'tugHall.3', mustWork = TRUE )
read_file(file_name = fl, stringsAsFactors = FALSE )
fl = system.file('extdata/Input', 'CF.txt',package = 'tugHall.3', mustWork = TRUE )
read_file(file_name = fl, stringsAsFactors = FALSE, header = FALSE )
```

safe\_pnt\_mut 45

safe\_pnt\_mut

Function to save 1 point mutation in a data frame

## Description

Function to save 1 point mutation in a data frame

## Usage

```
safe_pnt_mut(pnt)
```

#### **Arguments**

pnt

Object of class 'Point\_Mutations'

#### Value

data frame with 1 row of point mutation info

#### **Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
df = safe_pnt_mut( pnt ) # View( pnt )
```

simulation\_example

Example of simulation for lazy start

# Description

Example of simulation for lazy start

## Usage

```
simulation_example(verbose = TRUE, to_plot = TRUE, seed = NA)
```

## **Arguments**

verbose Logical type to show or do not show messages during execution to\_plot Logical type to plot or do not plot graphical results of a simulation

seed Numeric type to set seed for a simulation, if seed = NA (by default) then it will

be skipped

#### Value

List of results of simulation with default values for all the parameters

```
# it takes a time for a simulation and then it will demonstrates results, \cr
# so, please, wait for a while
simulation_example( verbose = FALSE , to_plot = FALSE )
```

46 sum\_mutation

sum\_cell

Aggregate data of a clone for environment object

## **Description**

Aggregate data of a clone for environment object

## Usage

```
sum_cell(env, clones)
```

## **Arguments**

env Object of class 'Environ'

clones List of all the objects of class 'Clone'

#### Value

NULL, but global variable env is updated

## **Examples**

```
clones = tugHall_dataset$clones
env = tugHall_dataset$env
sum_cell(env, clones)
message( paste0('Number of primary tumor cells in the pool of clones is ', env$P ) )
message( paste0('Number of normal cells in the pool of clones is ', env$N ) )
message( paste0('Number of metastatic cells in the pool of clones is ', env$M ) )
```

sum\_mutation

Serve function for sum\_cell() function

#### **Description**

Serve function for sum\_cell() function

## Usage

```
sum_mutation(clone1)
```

# Arguments

clone1

Object of class 'Clone'

## Value

vector of clone1 variables to aggregate in sum\_cell() function

```
clone1 = tugHall_dataset$clones[[ 1 ]]
sum_mutation(clone1)
```

 $sum_{-}N_{-}P_{-}M$ 

 $sum_N_P_M$ 

Function to calculate N and M numbers - normal and metastatic cells

## **Description**

Function to calculate N and M numbers - normal and metastatic cells

#### Usage

```
sum_N_P_M(env, clones)
```

## **Arguments**

env Object of class 'Environ'

clones List of all the objects of class 'Clone'

#### Value

Number of all the cells in a simulation (normal + primary tumor + metastatic)

## **Examples**

```
clones = tugHall_dataset$clones
env = tugHall_dataset$env
env$M = 0
env$P = 0
env$N = 0  # View( env )
sum_N_P_M(env, clones)  # View( env )
message( paste(env$N, env$P, env$M ) )
```

trial\_complex

Function trial for complex case of models

## Description

Function trial for complex case of models

## Usage

```
trial_complex(clone1, onco1)
```

#### **Arguments**

clone1 Object of class 'Clone' onco1 Object of class 'OncoGene'

## Value

Number of new clones originated by clone1

48 trial\_mutagenesis

#### **Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco
trial_complex( clone1, onco1 )
unlist( lapply( X = 1:20, FUN = function( x ) trial_complex( clone1, onco1 ) ) )
```

trial\_mutagenesis

Function for mutagenesis trial

#### **Description**

Function for mutagenesis trial

#### Usage

```
trial_mutagenesis(clone1, num_mut, onco1)
```

## **Arguments**

clone1 Object of class 'Clone'

num\_mut Number of mutations in this NEW clone1

oncol Object of class 'OncoGene' corresponding to clonel (with the same ID)

## Value

Changed object clone 1, add related mutations to the lists of point mutations and/or CNA mutations

```
copy_files_to_Input()
copy_files_to_Output()
define_parameters()
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco_clones[[ 1 ]]
onco = tugHall_dataset$onco
define_gene_location()
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234  # Just an example number
\label{local_message} $$ message( c('CNA mutation IDs ', paste(clone1$CNA_ID, collapse = ' ') ) ) $$ message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ') ) ) $$
## Not run:
trial_mutagenesis( clone1, num_mut = 1, onco1 ) # it adds info to clone1
message( c('CNA mutation IDs ', paste(clone1$CNA_ID, collapse = ' ') ) )
message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ') ) )
trial_mutagenesis( clone1, num_mut = 10, onco1 ) # it adds info to clone1
\label{local_collapse} message(\ c('CNA\ mutation\ IDs\ ',\ paste(clone1$CNA\_ID,\ collapse\ =\ '\ ')\ )\ )
message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ') ) )
## End(Not run)
```

trial\_simple 49

trial\_simple

Function trial for simplified case of model

## **Description**

Function trial for simplified case of model

#### Usage

```
trial_simple(clone1, onco1)
```

#### **Arguments**

clone1 Object of class 'Clone' onco1 Object of class 'OncoGene'

#### Value

Number of new clones originated by clone1

## **Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco
trial_simple( clone1, onco1 )
unlist( lapply( X = 1:20, FUN = function( x ) trial_simple( clone1, onco1 ) ) )
```

tugHall\_dataset

tugHall dataset named 'tugHall\_dataset'

#### **Description**

Dataset contains all the necessary data.frames and objects to check functions of tugHall. Description of each data.frame and object could be found in documentation to tugHall package.

## Usage

```
tugHall_dataset
```

#### **Format**

A data frame with 12 data.frames and 2 objects:

```
data_flow simulation data for all time steps, data from file cloneout.txt
data_last simulation data for the last time step, data from file cloneout.txt
data_avg simulation data averaged for the each time step, data from file cloneout.txt
pnt_mut data.frame with point mutation information
pnt_mut_B data.frame with point mutation information of mutated allele B
cna_mut data.frame with CNA mutation information
```

50 update\_Hallmarks

```
gene_map data.frame with genes' locations information
hall Object of class 'HallMark'
onco Object of class 'OncoGene'
time_max Value of maximal time step in an example simulation
CF Values of compaction factor
vf data.frame of preliminary data for VAF calculations
VAF_rho data.frame with VAF values for different rho
```

rdr\_dysf data.frame of order of genes dysfunction for each clone

update\_Hallmarks

Function to update Hallmark and variable after division or under initialization

# Description

Function to update Hallmark and variable after division or under initialization

## Usage

```
update_Hallmarks(clone1)
```

## **Arguments**

clone1

Object of class 'Clone'

## Value

The same object of class 'Clone' with updated fields

```
clone = tugHall_dataset$clones[[ 1 ]]
define_parameters()
hall = tugHall_dataset$hall
env = tugHall_dataset$env
update_Hallmarks( clone )
```

write\_cloneout 51

write_cloneout	Function to write data to cloneout file at a time step
----------------	--

## **Description**

Function to write data to cloneout file at a time step

## Usage

```
write_cloneout(outfile, env, clones, isFirst, onco_clones)
```

## **Arguments**

outfile File name for output info
env Object of class 'Environ'
clones List of objects of class 'Clone'
isFirst logical type = TRUE as default
onco\_clones List of objects of class 'OncoGene'

#### Value

NULL, but add rows to output file with clone evolution data

# **Examples**

```
env = tugHall_dataset$env
onco = tugHall_dataset$onco
if ( !dir.exists('./Output') ) dir.create('./Output')
clones = tugHall_dataset$clones
onco_clones = tugHall_dataset$onco_clones
write_header(outfile='./Output/exmpl.txt', env, onco)
write_cloneout( outfile = './Output/exmpl.txt', env, clones, isFirst = TRUE, onco_clones )
```

write\_geneout

Function to write info about HallMark data

## **Description**

Function to write info about HallMark data

## Usage

```
write_geneout(outfile, hall, Compaction_factor, CF)
```

52 write\_header

#### **Arguments**

outfile File name for output info
hall Object of class "HallMark"

Compaction\_factor

Compaction factor, logical type only. True means 'to use', False means 'do not

use' Compaction factor for hallmarks variables

CF Vector with values of compaction factor for each hallmark

#### Value

NULL, but data will save to a file

#### **Examples**

```
copy_files_to_Input()
define_files_names()
define_parameters()
if ( !dir.exists('./Output') ) dir.create('./Output')
hall = tugHall_dataset$hall
onco = tugHall_dataset$onco
define_compaction_factor()
write_geneout(outfile = geneoutfile, hall, Compaction_factor, CF)
```

write\_header

Function to write the header to a file

#### **Description**

Function to write the header to a file

#### Usage

```
write_header(outfile, env, onco)
```

# **Arguments**

outfile File name for output info env Object of class 'Environ' onco Object of class "OncoGene"

#### Value

NULL, but the header will save to a file and delete old info

```
env = tugHall_dataset$env
onco = tugHall_dataset$onco
if ( !dir.exists('./Output') ) dir.create('./Output')
write_header(outfile='./Output/exmpl.txt', env, onco)
```

write\_log 53

write\_log

Function to write log file

# Description

Function to write log file

# Usage

```
write_log(
  genefile,
  clonefile,
  geneoutfile,
  cloneoutfile,
  logoutfile,
  Ε0,
  F0,
  m0,
  uo,
  us,
  s0,
  k0,
  m_{dup},
  m_{del},
  lambda_dup,
  lambda_del,
  uo_dup,
  us_dup,
  uo_del,
  us_del,
  censore_n,
  censore_t,
  d0,
  Compaction_factor,
  model_name,
  time_stop,
  n_repeat,
  monitor
)
```

# Arguments

genefile	File name of initial OncoGene information
clonefile	File name of info about initial clones
geneoutfile	File name for output info about OncoGene information
cloneoutfile	File name for output info with clone evolution data
logoutfile	Name of log file with all the parameters
E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only

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m0	Mutation probability for point mutation, numeric type only	
uo	Oncogene mutation probability, numeric type only	
us	Suppressor mutation probability, numeric type only	
s0	Parameter in the sigmoid function, numeric type only	
k0	Environmental death probability, numeric type only	
m_dup	Mutation probability for duplication, numeric type only	
m_del	Mutation probability for deletion, numeric type only	
lambda_dup	CNA duplication average length (of the geometrical distribution for the length), integer type only	
lambda_del	CNA deletion average length (of the geometrical distribution for the length), integer type only	
uo_dup	Gene malfunction probability by CNA duplication for oncogene, numeric type only	
us_dup	Gene malfunction probability by CNA duplication for suppressor, numeric type only	
uo_del	Gene malfunction probability by CNA deletion for oncogene, numeric type only	
us_del	Gene malfunction probability by CNA deletion for suppressor, numeric type only	
censore_n	Max cell number where the program forcibly stops, integer type only	
censore_t	Max time where the program forcibly stops, integer type only	
d0	Initial probability to divide cells, numeric type only	
Compaction_fac		
	Compaction factor, logical type only. True means 'to use', False means 'do not use' Compaction factor for hallmarks variables	
model_name	Name of the model to use. Can be 'proportional_metastatic' or 'threshold_metastatic' or 'simplified'	
time_stop	Max time in seconds of running after that the program forcibly stops, integer type only	
n_repeat	Max number of repetition of the program until the NON-ZERO output will be getting, integer type only	
monitor	The indicator to make monitor file during a simulation or do not make, logical type only	

#### Value

NULL, write log file to Output folder

```
copy_files_to_Input()
define_files_names()
define_parameters()
if ( !dir.exists('./Output') ) dir.create('./Output')
## Not run:
write_log(genefile, clonefile, geneoutfile, cloneoutfile, logoutfile,
E0, F0, m0, uo, us, s0, k0, m_dup, m_del, lambda_dup, lambda_del,
uo_dup, us_dup, uo_del, us_del, censore_n, censore_t, d0,
Compaction_factor, model_name, time_stop, n_repeat, monitor )
## End(Not run)
```

write\_monitor 55

write\_monitor

Function to write a simulation monitoring data into the file\_monitor

#### **Description**

Function to write a simulation monitoring data into the file\_monitor

#### Usage

```
write_monitor(outfile = file_monitor, start = FALSE, env, clones)
```

## **Arguments**

outfile File name for output info

start Indicator to start from beginning (TRUE) or not (FALSE)

env Object of class 'Environ' clones List of objects of class 'Clone'

#### Value

NULL, but info about current state of simulation will write to a file

#### **Examples**

```
env = tugHall_dataset$env
if ( !dir.exists('./Output') ) dir.create('./Output')
clones = tugHall_dataset$clones
onco_clones = tugHall_dataset$onco_clones
cna_clones = tugHall_dataset$cna_clones
pnt_clones = tugHall_dataset$pnt_clones
write_monitor( outfile = './Sim_monitoring.txt', start = TRUE , env, clones )
write_monitor( outfile = './Sim_monitoring.txt', start = FALSE , env, clones )
```

write\_pnt\_clones

Function to write the point mutation info for all clones for all time steps, used at the last time step or after simulation

## **Description**

Function to write the point mutation info for all clones for all time steps, used at the last time step or after simulation

## Usage

```
write_pnt_clones(pnt_clones, file_out = "Output/point_mutations.txt")
```

## **Arguments**

pnt\_clones List of objects of class 'Point\_Mutations'

file\_out File name to write

56 write\_weights

#### Value

NULL, but info will write to a file

## **Examples**

```
pnt_clones = tugHall_dataset$pnt_clones
if ( !dir.exists('./Output') ) dir.create('./Output')
write_pnt_clones(pnt_clones, file_out = 'Output/point_mutations.txt')
```

write\_weights

Function to write info about relationship between genes and hallmarks

## **Description**

Function to write info about relationship between genes and hallmarks

# Usage

```
write_weights(outfile, hall)
```

# Arguments

outfile File name for output info hall Object of class 'HallMark'

## Value

NULL, but info about relationship between genes and hallmarks will write to a file

```
if ( !dir.exists('./Output') ) dir.create('./Output')
hall = tugHall_dataset$hall
onco = tugHall_dataset$onco
write_weights(outfile = './Output/weights.txt', hall)
```

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