

# Package ‘tugHall.3’

May 31, 2022

**Title** R-based script to simulate the cancer cell evolution

**Version** 3.0

**Description** tugHall (tumor gene-Hallmark) is a cancer-cell evolution model simulator, wherein gene mutations are linked to the hallmarks of cancer, which influence tumor cell behaviors.

**License** GPL (>= 3)

**Depends** R (>= 3.6.0)

**Imports** actuar,  
graphics,  
grDevices,  
methods,  
randomcoloR,  
stats,  
stringr,  
utils

**Suggests** rmarkdown,  
knitr,  
testthat,  
DiagrammeR

**Encoding** UTF-8

**Roxygen** list(markdown = TRUE)

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**LazyData** true

**VignetteBuilder** knitr

## R topics documented:

add_deletion . . . . .	3
add_duplication . . . . .	4
add_pnt_mutation . . . . .	4
calc_binom . . . . .	5
change_allele_A_by_cna . . . . .	5
change_pnt_by_cna . . . . .	6
check_pkg . . . . .	7
check_pnts . . . . .	7
chk_pnt_mut . . . . .	8

Clone-class . . . . .	8
clone_copy . . . . .	9
CNA_Mutations-class . . . . .	10
copy_CNA . . . . .	11
copy_files_to_Input . . . . .	11
copy_files_to_Output . . . . .	12
copy_pnt . . . . .	13
copy_pnt_no_mutation . . . . .	13
define_compaction_factor . . . . .	14
define_files_names . . . . .	14
define_gene_location . . . . .	15
define_parameters . . . . .	16
define_par_for_plot . . . . .	17
Environ-class . . . . .	18
generate_cna . . . . .	19
generate_pnt . . . . .	20
generate_to_copy_pnt . . . . .	20
gen_colors . . . . .	21
get_cds_rna . . . . .	21
get_cna_mutation . . . . .	22
get_flow_data . . . . .	23
get_len_cds_rna . . . . .	24
get_order_of_genes_dysfunction . . . . .	24
get_point_mutation . . . . .	25
get_point_mutation_for_gene . . . . .	26
get_rho_VAF . . . . .	26
get_type . . . . .	27
get_u_cna . . . . .	27
get_VAF . . . . .	28
HallMark-class . . . . .	29
init_clones . . . . .	29
init_onco_clones . . . . .	30
init_pnt_clones . . . . .	31
make_map . . . . .	31
mixed_mut_order . . . . .	32
model . . . . .	33
modify_gene_map . . . . .	34
number_N_P_M . . . . .	35
OncoGene-class . . . . .	35
onco_copy . . . . .	36
onco_update . . . . .	36
order_gene_map . . . . .	37
plot_2D . . . . .	37
plot_2D_lines . . . . .	38
plot_average_simulation_data . . . . .	40
plot_clone_evolution . . . . .	40
plot_order_dysfunction . . . . .	41
pnts_add_dlt . . . . .	42
Point_Mutations-class . . . . .	43
print_parameters . . . . .	44
read_file . . . . .	44
safe_pnt_mut . . . . .	45

simulation_example . . . . .	45
sum_cell . . . . .	46
sum_mutation . . . . .	46
sum_N_P_M . . . . .	47
trial_complex . . . . .	47
trial_mutagenesis . . . . .	48
trial_simple . . . . .	49
tugHall_dataset . . . . .	49
update_Hallmarks . . . . .	50
write_cloneout . . . . .	51
write_geneout . . . . .	51
write_header . . . . .	52
write_log . . . . .	53
write_monitor . . . . .	55
write_pnt_clones . . . . .	55
write_weights . . . . .	56

<b>Index</b>	<b>57</b>
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add_deletion	<i>Function to add deletion to gene map (chromosomal location data frame)</i>
--------------	---

## Description

Function to add deletion to gene map (chromosomal location data frame)

## Usage

```
add_deletion(gm, Ref_start, Ref_end, Chr)
```

## Arguments

gm	Chromosomal location data frame
Ref_start	Starting position of deletion
Ref_end	Final position of deletion
Chr	Chromosome name

## Value

Chromosomal location data frame with additional deletion info

## Examples

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm_new = add_deletion( gm = gene_map, Ref_start = 112775658, Ref_end = 112775716, Chr = '5')
# to check add_del ... add_dupl, please, use diff library and:
# diff_data( gm_ref, gm, show_unchanged_columns = TRUE, always_show_order = TRUE )
```

---

add_duplication	<i>Function to add duplication to gene map (chromosomal location data frame)</i>
-----------------	--

---

### Description

Function to add duplication to gene map (chromosomal location data frame)

### Usage

```
add_duplication(gm, Ref_start, Ref_end, Chr)
```

### Arguments

gm	Chromosomal location data frame
Ref_start	Starting position of duplication
Ref_end	Final position of duplication
Chr	Chromosome name

### Value

Chromosomal location data frame with additional duplication info

### Examples

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm_new = add_duplication( gm = gene_map, Ref_start = 112775658, Ref_end = 112775716, Chr = '5')
# to check add_del ... add_dupl, please, use diff library and:
# diff_data( gm_ref, gm, show_unchanged_columns = TRUE, always_show_order = TRUE )
```

---

add_pnt_mutation	<i>Function to add point mutation to data.frame gene_map (chromosomal location data frame)</i>
------------------	--

---

### Description

Function to add point mutation to data.frame gene\_map (chromosomal location data frame)

### Usage

```
add_pnt_mutation(gm = gm, pos_pnt, Chr = Chr)
```

### Arguments

gm	Chromosomal location data frame
pos_pnt	Position of point mutation
Chr	Chromosome name

**Value**

Chromosomal location data frame with additional point mutation info

**Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gm2 = add_pnt_mutation( gm = gene_map, pos_pnt = 112775637 , Chr = '5' )
```

---

calc_binom	<i>Function to calculate binomial distribution including BIG NUMBERS like 10<sup>12</sup> and more using approximation with normal distribution</i>
------------	---

---

**Description**

Function to calculate binomial distribution including BIG NUMBERS like 10<sup>12</sup> and more using approximation with normal distribution

**Usage**

```
calc_binom(tr, n, p)
```

**Arguments**

tr	Length of vector with successes trials
n	Number of independent Bernoulli trials
p	Probability to get successes in trials

**Value**

Vector of integer numbers of successes trials

**Examples**

```
calc_binom(tr = 3, n = 40, p = 0.9)
calc_binom(tr = 3, n = 4E20, p = 9E-9)
```

---

change_allele_A_by_cna	<i>Function to change copy number of the allele A of the point mutation at the allele B due to CNA</i>
------------------------	--

---

**Description**

Function to change copy number of the allele A of the point mutation at the allele B due to CNA

**Usage**

```
change_allele_A_by_cna(pnt1, start_end, t)
```

**Arguments**

pnt1                Object of class 'Point\_Mutations'  
 start\_end          Vector with initial and final positions of CNA  
 t                   'dup' or 'del' for duplication or deletion respectively

**Value**

NULL, but data of pnt1 is updated due to CNA

**Examples**

```
pnt1 = tugHall_dataset$pnt_clones[[ 2 ]] # pnt of allele A
start_end = c( pnt1$Phys_pos - 50 , pnt1$Phys_pos + 50 )
message( pnt1$Copy_number )
change_allele_A_by_cna( pnt1, start_end, t = 'dup' ) # View( pnt1 )
message( pnt1$Copy_number )
change_allele_A_by_cna( pnt1, start_end, t = 'del' ) # View( pnt1 )
message( pnt1$Copy_number )
```

---

change_pnt_by_cna	<i>Function to change the point mutation due to CNA</i>
-------------------	---

---

**Description**

Function to change the point mutation due to CNA

**Usage**

```
change_pnt_by_cna(pnt1, start_end, t)
```

**Arguments**

pnt1                Object of class 'Point\_Mutations'  
 start\_end          Vector with initial and final positions of CNA  
 t                   'dup' or 'del' for duplication or deletion respectively

**Value**

NULL, but pnt1 data is updated due to CNA

**Examples**

```
pnt1 = tugHall_dataset$pnt_clones[[ 1 ]]
start_end = c( pnt1$Phys_pos - 50 , pnt1$Phys_pos + 50 )
message( pnt1$Copy_number )
change_pnt_by_cna( pnt1, start_end, t = 'dup' ) # View( pnt1 )
message( pnt1$Copy_number )
change_pnt_by_cna( pnt1, start_end, t = 'del' ) # View( pnt1 )
message( pnt1$Copy_number )
```

---

check\_pkg*Check the installation of a package for some functions*

---

**Description**

Check the installation of a package for some functions

**Usage**

```
check_pkg(pkg)
```

**Arguments**

pkg                      Package name

**Value**

if the package is installed then it returns NULL else it returns error message

**Examples**

```
check_pkg( pkg = 'grDevices' )
```

---

check\_pnts*Function to check what pnts do fall into the range?*

---

**Description**

Function to check what pnts do fall into the range?

**Usage**

```
check_pnts(gm_w1)
```

**Arguments**

gm\_w1                    A row from data.frame gene\_map

**Value**

Return the point mutations which fall into the range

**Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gene_map$pnts[6] = '112792451, 112792442'
gm_w1 = gene_map[6,]
check_pnts( gm_w1 )
```

---

chk_pnt_mut	<i>Function to check point mutations match or don't match into duplication or deletion</i>
-------------	--

---

### Description

Function to check point mutations match or don't match into duplication or deletion

### Usage

```
chk_pnt_mut(pnt1, Ref_start, Ref_end, Chr, prnt1)
```

### Arguments

pnt1	Object of class 'Point_Mutations'
Ref_start	Initial position of CNA
Ref_end	Final position of CNA
Chr	Chromosome name
prnt1	Parental chromosome 1 or 2

### Value

Logical: TRUE if point mutation matches CNA, FALSE if it doesn't match

### Examples

```
pnt1 = tugHall_dataset$pnt_clones[[ 5 ]]
pstn = pnt1$Phys_pos[1]
message( pstn )
prnt1 = pnt1$Parental_1or2
Chr = pnt1$Chr
chk_pnt_mut( pnt1 , Ref_start = pstn - 200, Ref_end = pstn + 200, Chr, prnt1 )
chk_pnt_mut( pnt1 , Ref_start = pstn - 200, Ref_end = pstn - 100, Chr, prnt1 )
```

---

Clone-class	<i>Class 'Clone' for clones</i>
-------------	---------------------------------

---

### Description

Class 'Clone' for clones

### Fields

id	numeric. ID of a clone
parent	numeric. Parent ID (for first - 0)
N_cells	numeric. Number of cells in clone
c	numeric. Split counter as average value for all cells in clone
d	numeric. Probability of division



i numeric. Probability of Hayflick limit  
 m numeric. Probability that gene normal function is destroyed due to epigenome abnormality / mutation rate  
 a numeric. Probability of apoptosis for a cell in the clone  
 s numeric. Coefficient in the sigmoid function of the mutation density  
 k numeric. Probability of cell death by environment  
 E numeric. Coefficient of friction term against to the split probability.  
 Nmax numeric. Coefficient for determination the max number of cells that can exist in the primary tumor ( $N_{max} = 1/E$ )  
  
 im numeric. Probability of the invasion/ metastatic transformation  
 Ha numeric. Apoptosis hallmark value  
 Him numeric. Invasion/ metastasis hallmark  
 Hi numeric. Mitotic restriction hallmark (immortalization hallmark)  
 Hd numeric. Growth/antigrowth hallmark (division rate hallmark)  
 Hb numeric. Angiogenesis hallmark  
 gene numeric. Vector of flags for each genes if they have driver mutation  
 pasgene numeric. Vector of flags for each genes if they have passenger mutation  
 PointMut\_ID numeric. ID of point mutation in list of objects of class 'Point\_Mutations'  
 CNA\_ID numeric. ID of CNA mutation in list of objects of class 'CNA\_Mutations'  
 mutden numeric. Gene mutation density  
 invasion logical. Indicator that clone is metastatic (invasion/metastatic transformation occurred or not)  
 primary logical. Logical variable is clone primary tumor or not (normal)  
 birthday numeric. Time step of birth of clone

### Examples

```

clone = tugHall_dataset$clones[[ 1 ]]
print(clone$Ha)
print(clone$N_cells)
clone$calcApoptosis() # to calculate apoptosis death probability based on mutation density

```

---

clone\_copy

---

*Function to make one copy for clone1 in clone\_init function*


---

### Description

Function to make one copy for clone1 in clone\_init function

### Usage

```
clone_copy(clone1)
```

**Arguments**

clone1                      Object of class 'Clone'

**Value**

New object of class 'Clone' with the same info and new ID

**Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
env = tugHall_dataset$env
define_parameters()
clone_copy(clone1)
```

---

CNA\_Mutations-class      *Class 'CNA\_Mutations'*

---

**Description**

Class 'CNA\_Mutations'

**Fields**

CNA\_ID    numeric. ID of CNA mutation

Parental\_1or2    numeric. Parental chromosome, could be 1 or 2

dupOrdel    character. dup for duplication or del for deletion

Chr    character. Chromosome name

Ref\_start    numeric. Reference start position

Ref\_end    numeric. Reference final position

Gene\_names    character. Names of genes involved in CNA

MalfunctionedByCNA    logical.

mut\_order    numeric. True for driver mutation and False for passenger mutation

**Examples**

```
cna = tugHall_dataset$cna_clones[[ 1 ]]
cna$save()    # to save as row of data.frame
cna$copy()
cna$initialize()
cna$show()    # After initialization
```

---

copy_CNA	<i>Function to copy CNA info</i>
----------	----------------------------------

---

**Description**

Function to copy CNA info

**Usage**

```
copy_CNA(CNA1)
```

**Arguments**

CNA1	Object of class 'CNA_Mutations'
------	---------------------------------

**Value**

The same object of class 'CNA\_Mutations'

**Examples**

```
cna = tugHall_dataset$cna_clones[[ 1 ]]
cna2 = copy_CNA( cna )
cna$safe()
cna2$safe()
```

---

copy_files_to_Input	<i>Function to copy the files by default from extdata folder in the library to Input folder in the working directory</i>
---------------------	--

---

**Description**

Function to copy the files by default from extdata folder in the library to Input folder in the working directory

**Usage**

```
copy_files_to_Input(
  files = c("CCDS.current.txt", "CF.txt", "cloneinit.txt", "gene_hallmarks.txt",
    "gene_map.txt", "parameters.txt"),
  dir = "Input"
)
```

**Arguments**

files	Files to copy, vector of names of files by default: files = c( 'CCDS.current.txt', 'CF.txt', 'cloneinit.txt', 'gene_hallmarks.txt', 'gene_map.txt', 'parameter' )
dir	Folder to where files should be save, by default dir = 'Input'

**Value**

List of logic numbers for each copied file, TRUE - success, FALSE - not success

**Examples**

```
files = c('CF.txt', 'cloneinit.txt', 'gene_hallmarks.txt', 'gene_map.txt', 'parameters.txt' )
copy_files_to_Input( files, dir = 'Input' )
```

---

copy_files_to_Output	<i>Function to copy the files of an example of simulation or from '/ext-data/Output/' folder in the library to '/Output/' folder in the working directory</i>
----------------------	---

---

**Description**

Function to copy the files of an example of simulation or from '/extdata/Output/' folder in the library to '/Output/' folder in the working directory

**Usage**

```
copy_files_to_Output(
  files = c("cloneout.txt", "CNA_mutations.txt", "point_mutations.txt", "gene_MAP.txt",
    "geneout.txt", "log.txt", "order_genes_dysfunction.txt", "VAF_data.txt", "VAF.txt",
    "weights.txt"),
  dir = "Output"
)
```

**Arguments**

files	Files to copy, vector of names of files by default: files = c('cloneout.txt', 'CNA_mutations.txt', 'point_mutations.txt', 'gene_MAP.txt', 'geneout.txt', 'log.txt', 'order_genes_dysfunction.txt', 'VAF_data.txt', 'VAF.txt', 'weights.txt' )
dir	Folder to where files should be save, by default dir = 'Output'

**Value**

List of logic numbers for each copied file, TRUE - success, FALSE - not success

**Examples**

```
files = c( 'cloneout.txt', 'CNA_mutations.txt', 'point_mutations.txt', 'gene_MAP.txt' )
copy_files_to_Output( files )
files = c('geneout.txt', 'log.txt', 'VAF_data.txt', 'VAF.txt', 'weights.txt' )
copy_files_to_Output( files )
```

---

copy_pnt	<i>Function to copy of point mutation info</i>
----------	--

---

**Description**

Function to copy of point mutation info

**Usage**

```
copy_pnt(pnt1)
```

**Arguments**

pnt1                      Object of class 'Point\_Mutations'

**Value**

The same object of class 'Point\_Mutations' with different ID

**Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
copy_pnt( pnt ) # View( pnt )
```

---

copy_pnt_no_mutation	<i>Function to copy of pnt1 without mutation info for allele A</i>
----------------------	--

---

**Description**

Function to copy of pnt1 without mutation info for allele A

**Usage**

```
copy_pnt_no_mutation(pnt1)
```

**Arguments**

pnt1                      Object of class 'Point\_Mutations'

**Value**

Object of class 'Point\_Mutations' for another chromosome

**Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
copy_pnt_no_mutation( pnt ) # View( pnt )
```

---

```
define_compaction_factor
```

*Define compaction factor*

---

**Description**

Define compaction factor

**Usage**

```
define_compaction_factor(
  cf = data.frame(Ha = 1, Hb = 1, Hd = 1, Hi = 1, Him = 1),
  read_fl = TRUE,
  file_name = "./Input/CF.txt"
)
```

**Arguments**

cf	Data frame with compaction factors for all the hallmarks, for example, data.frame(Ha = 1, Hb = 1, Hd = 1, Hi = 1, Him = 1 )
read_fl	Indicator to read file or not, logical type only
file_name	File name to rad all the parameters, it is used only if read_fl == TRUE

**Value**

Data frame with with compaction factors for all the hallmarks

**Examples**

```
copy_files_to_Input()
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
CF1 = CF
cf = data.frame( Ha = 0.1, Hb = 0.2, Hd = 0.7, Hi = 1, Him = 0.5 )
define_compaction_factor( cf = cf, read_fl = FALSE ) # View( c( CF, CF1 ) ) to compare
```

---

```
define_files_names
```

*Function to define all the files names*

---

**Description**

Function to define all the files names

**Usage**

```
define_files_names(
  mainDir = getwd(),
  sbdr_Input = "/Input",
  sbdr_Output = "/Output"
)
```

**Arguments**

mainDir	Working directory
sbdr_Input	Sub directory for input files
sbdr_Output	Sub directory for output files

**Value**

NULL, but all file names are defined in GLOBAL environment

**Examples**

```
define_files_names()
```

---

define\_gene\_location    *Define genes' location in chromosome*

---

**Description**

Define genes' location in chromosome

**Usage**

```
define_gene_location(
  file_input = "Input/CCDS.current.txt",
  genes_list = c("CCDS4107.1", "CCDS8702.1", "CCDS43171.1", "CCDS11118.1")
)
```

**Arguments**

file_input	is a name of file to input where the information about genes location is defined. That is loaded from CCDS database <a href="https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_human/">https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_human/</a>
genes_list	is a list of genes' names like CCDS4107.1 in the CCDS database.

**Value**

Function returns the table of genes' locations in DNA

**Examples**

```
copy_files_to_Input()
define_gene_location()
file_input = 'Input/CCDS.current.txt'
genes_list = c('CCDS4107.1', 'CCDS8702.1', 'CCDS43171.1', 'CCDS11118.1')
define_gene_location( file_input = file_input, genes_list = genes_list )
```

---

define_parameters	<i>Define all the parameters for a simulation</i>
-------------------	---

---

## Description

Define all the parameters for a simulation

## Usage

```
define_parameters(
  E0 = 1e-04,
  F0 = 10,
  m0 = 1e-07,
  uo = 0.9,
  us = 0.9,
  s0 = 10,
  k0 = 0.12,
  d0 = 0.4,
  censure_n = 10^5,
  censure_t = 50,
  m_dup = 1e-08,
  m_del = 1e-08,
  lambda_dup = 5000,
  lambda_del = 7000,
  uo_dup = 0.8,
  us_dup = 0.5,
  uo_del = 0,
  us_del = 0.8,
  CF = TRUE,
  model = c("proportional_metastatic", "threshold_metastatic", "simplified")[1],
  time_stop = 120,
  read_fl = FALSE,
  file_name = "../Input/parameters.txt",
  n_repeat = 1000,
  monitor = TRUE
)
```

## Arguments

E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only
m0	Mutation probability for point mutation, numeric type only
uo	Oncogene mutation probability, numeric type only
us	Suppressor mutation probability, numeric type only
s0	Parameter in the sigmoid function, numeric type only
k0	Environmental death probability, numeric type only
d0	Initial probability to divide cells, numeric type only
censure_n	Max cell number where the program forcibly stops, integer type only



censore_t	Max time where the program forcibly stops, integer type only
m_dup	Mutation probability for duplication, numeric type only
m_del	Mutation probability for deletion, numeric type only
lambda_dup	CNA duplication average length (of the geometrical distribution for the length), integer type only
lambda_del	CNA deletion average length (of the geometrical distribution for the length), integer type only
uo_dup	Gene malfunction probability by CNA duplication for oncogene, numeric type only
us_dup	Gene malfunction probability by CNA duplication for suppressor, numeric type only
uo_del	Gene malfunction probability by CNA deletion for oncogene, numeric type only
us_del	Gene malfunction probability by CNA deletion for suppressor, numeric type only
CF	Compaction factor, logical type only. True means 'to use', False means 'do not use' Compaction factor for hallmarks variables
model	Name of the model to use. Can be 'proportional_metastatic' or 'threshold_metastatic' or 'simplified'
time_stop	Max time in seconds of running after that the program forcibly stops, integer type only
read_fl	Indicator to read file or not, logical type only
file_name	File name to read all the parameters, it is used only if read_fl == TRUE
n_repeat	Max number of repetition of the program until the NON-ZERO output will be getting, integer type only
monitor	The indicator to make monitor file during a simulation or do not make, logical type only

**Value**

Values of all the parameters

**Examples**

```
copy_files_to_Input()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_parameters( read_fl = FALSE )
```

---

define\_par\_for\_plot      *Function to change par for plots and after plotting it returns par values*

---

**Description**

Function to change par for plots and after plotting it returns par values

**Usage**

```
define_par_for_plot(change_par_back)
```

**Arguments**

change\_par\_back

logical. If TRUE it changes par options back after finishing a function call

**Value**

Change par() options and returns it (after finishing the function) to values before a function has started

**Examples**

```
define_par_for_plot( change_par_back = TRUE )
```

---

Environ-class

*Class 'Environ'*

---

**Description**

Class 'Environ'

**Fields**

T numeric. Time counter

N numeric. Number of normal cells

P numeric. Number of primary tumor cells

M numeric. Number of metastatic cells

F numeric. Coefficient that determines the maximal number of cells in pool of primary tumor cells

c numeric. Average number of divisions in pool of clones

d numeric. Mean value of splitting probability

i numeric. Average value of immortalization probability

a numeric. Average value of apoptosis probability

k numeric. Average probability of cell death via environment death

E numeric. Average value of coefficients of friction term

Nmax numeric. Maximal number of primary tumor cells that can exist in pool of clones

im numeric. Average value of invasion/metastasis probability

Ha numeric. Average value of apoptosis hallmark Ha

Him numeric. Average value of invasion/metastasis hallmark Him

Hi numeric. Average value of immortalization hallmark Hi

Hd numeric. Average value of growth/antigrowth hallmark Hd

Hb numeric. Average value of angiogenesis hallmark Hb

type numeric. Invasion / metastatic ratio

gene numeric. Cancer gene damage rate

mutden numeric. Average mutation rate

last\_id numeric. Maximal ID in the pool of clones.

**Examples**

```
env = tugHall_dataset$env
print( env )
env$initFields()
```

generate\_cna

*Function to generate object of CNA mutation***Description**

Function to generate object of CNA mutation

**Usage**

```
generate_cna(prnt1, genes, start_end, onco1, dupOrdel)
```

**Arguments**

prnt1	The 1st or 2nd parental chromosome
genes	Genes names
start_end	vector with start and final positions of CNA
onco1	Object of class 'OncoGene'
dupOrdel	It could be 'dup' or 'del' to denote duplication or deletion

**Value**

Object of class 'CNA\_Mutations'

**Examples**

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
gene_map = tugHall_dataset$gene_map
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234
start_end = c(112775658, 112775716 )
generate_cna( prnt1 = 1, genes = 'APC', start_end = start_end, onco1, dupOrdel = 'dup' )
```

---

generate_pnt	<i>Function to generate an object of class 'Point_Mutations'</i>
--------------	--

---

**Description**

Function to generate an object of class 'Point\_Mutations'

**Usage**

```
generate_pnt(prntl, gene, pos, onco1, Chr, mutation = NA)
```

**Arguments**

prntl	Parental chromosome, could be 1 or 2
gene	Gene name
pos	Position of point mutation
onco1	Object of class 'OncoGene'
Chr	Chromosome name
mutation	If mutation is NOT NA then MalfunctionedByPointMut = TRUE, else it is defined by corresponding probabilities

**Value**

Object of class 'Point\_Mutations'

**Examples**

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
pnt_clones = tugHall_dataset$pnt_clones
copy_files_to_Input()
define_parameters()
mut_order = 234 # As an example
pnt1 = generate_pnt( prntl = 1, gene = 'APC', pos = 112767192, onco, Chr = '5', mutation = NA )
```

---

generate_to_copy_pnt	<i>Function to generate the same object of class 'Point_Mutations' with coping all information from input object</i>
----------------------	--

---

**Description**

Function to generate the same object of class 'Point\_Mutations' with coping all information from input object

**Usage**

```
generate_to_copy_pnt(pnt)
```

**Arguments**

pnt                      Object of class 'Point\_Mutations'

**Value**

The same object of class 'Point\_Mutations' with different ID

**Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
pnt_clones = tugHall_dataset$pnt_clones
pnt2 = generate_to_copy_pnt( pnt )
```

---

gen_colors	<i>Function to make a large number of colors</i>
------------	--

---

**Description**

Function to make a large number of colors

**Usage**

```
gen_colors(nm = 12)
```

**Arguments**

nm                      Number of colors

**Value**

Vector of colors with length more than nm

**Examples**

```
clrs = gen_colors( nm = 120 )
```

---

get_cds_rna	<i>Function to get length of CDS and of genes from data.frame gene_map and related probabilities</i>
-------------	--

---

**Description**

Function to get length of CDS and of genes from data.frame gene\_map and related probabilities

**Usage**

```
get_cds_rna(gm)
```

**Arguments**

gm                      data.frame gene\_map with info about genes' location

**Value**

list( names, CDS, RNA, PROB, SUM, P0 )

**Examples**

```
gene_map = tugHall_dataset$gene_map
define_parameters()
get_cds_rna( gm = gene_map )
```

---

get_cna_mutation	<i>Generation CNA mutation info</i>
------------------	-------------------------------------

---

**Description**

Generation CNA mutation info

**Usage**

```
get_cna_mutation(onco1, dupOrdel, gm_1_2)
```

**Arguments**

onco1	Object of class 'OncoGene'
dupOrdel	It could be 'dup' or 'del' to denote duplication or deletion
gm_1_2	List of two data frames (for 1st and 2nd parental chromosomes) with genes' location information

**Value**

List of ( prntl - 1 or 2 parental chromosome, Chr - name of chromosome, genes - genes names, start\_end - vector with start and end positions of CNA, w\_cna - rows of CNA in gene\_map data frame )

**Examples**

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
gm = tugHall_dataset$gene_map
get_cna_mutation( onco1 = onco, dupOrdel = 'dup', gm_1_2 = list(gm, gm) )
get_cna_mutation( onco1 = onco, dupOrdel = 'del', gm_1_2 = list(gm, gm) )
```

---

get_flow_data	<i>Function to get data about last simulation from cloneoutfile</i>
---------------	---

---

## Description

Function to get data about last simulation from cloneoutfile

## Usage

```
get_flow_data(
  cloneoutfile,
  genefile,
  mainDir = getwd(),
  sbdr_Output = "/Output"
)
```

## Arguments

cloneoutfile	Name of file to read data about clone evolution
genefile	Name of file with hallmarks values
mainDir	Working directory, by default mainDir = getwd()
sbdr_Output	Directory for output data getting from mainDir

## Value

list of data.frames like onco, hall, data\_last (data of last time step), data\_avg (average data for all time steps), data\_flow (data without average rows), time\_max (max time step), pnt\_mut and pnt\_mut\_B (data.frame of point mutations for both alleles and for allele B only ) and cna\_mut (data.frame of CNA mutations)

## Examples

```
copy_files_to_Input()
define_files_names()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
define_gene_location()
copy_files_to_Output()
dataset = get_flow_data(cloneoutfile, genefile, mainDir = getwd(), sbdr_Output = '/Output' )
# View(dataset)
```

---

get_len_cds_rna	<i>Function to get length of CDS and whole gene from gene_map data.frame</i>
-----------------	--

---

**Description**

Function to get length of CDS and whole gene from gene\_map data.frame

**Usage**

```
get_len_cds_rna(gene_map)
```

**Arguments**

gene_map	data.frame with info about genes' locations
----------	---

**Value**

list of ( Name, CDS, LEN\_Genes ) where Name is a vector of genes' names, CDS is a vector of CDS lengths, LEN\_Genes is a vector of length of whole genes including introns and exons

**Examples**

```
gene_map = tugHall_dataset$gene_map
onco = tugHall_dataset$onco
get_len_cds_rna( gene_map)
```

---

get_order_of_genes_dysfunction	<i>Function to get order of genes' dysfunction</i>
--------------------------------	--

---

**Description**

Function to get order of genes' dysfunction

**Usage**

```
get_order_of_genes_dysfunction(
  pnt_mut,
  data_last,
  cna_mut,
  file_name = "../Output/order_genes_dysfunction.txt"
)
```

**Arguments**

pnt_mut	data.frame with info about all the point mutations
data_last	data.frame with data of simulation at the last time step
cna_mut	data.frame with info about all the CNA mutations
file_name	Name of file to save data



**Value**

data.frame of genes' dysfunction and save it in a file

**Examples**

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
cna_mut = tugHall_dataset$cna_mut
file_name = './Output/order_genes_dysfunction.txt'
rdr = get_order_of_genes_dysfunction( pnt_mut, data_last, cna_mut, file_name = file_name )
```

---

get_point_mutation	<i>Generation point mutation info</i>
--------------------	---------------------------------------

---

**Description**

Generation point mutation info

**Usage**

```
get_point_mutation(onco1, gm_1_2)
```

**Arguments**

onco1	Object of class 'OncoGene'
gm_1_2	List of two data frames (for 1st and 2nd parental chromosomes) with genes' location information

**Value**

list of (prntl - 1 or 2 parental chromosome, gene - gene name, pos - position of point mutation, Chr - name of chromosome )

**Examples**

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
get_point_mutation( onco, gm_1_2 )
```

---

```
get_point_mutation_for_gene
```

*Generation point mutation info for the particular gene*

---

### Description

Generation point mutation info for the particular gene

### Usage

```
get_point_mutation_for_gene(onco1, gm_1_2, gene)
```

### Arguments

onco1	Object of class 'OncoGene'
gm_1_2	List of two data frames (for 1st and 2nd parental chromosomes) with genes' location information
gene	Gene's name where point mutation should be occurred

### Value

list of (prntl - 1 or 2 parental chromosome, gene - gene name, pos - position of point mutation, Chr - name of chromosome )

### Examples

```
gm = tugHall_dataset$gene_map
gm_1_2 = list( gm, gm )
onco = tugHall_dataset$onco
get_point_mutation_for_gene( onco, gm_1_2, gene = 'APC')
get_point_mutation_for_gene( onco, gm_1_2, gene = 'KRAS')
```

---

```
get_rho_VAF
```

*Function to get Variant allele frequencies (VAF) based on rho input parameters*

---

### Description

Function to get Variant allele frequencies (VAF) based on rho input parameters

### Usage

```
get_rho_VAF(vf = NULL, rho = c(0, 0.1, 0.5), file_name = "./Output/VAF.txt")
```

### Arguments

vf	data.frame getting from get_VAF() function
rho	Vector of rho parameter in the range (0,1)
file_name	Name of file to save VAF

**Value**

VAF for different rho with separation for metastatic cells and (primary tumor + speckled normal) cells

**Examples**

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
if ( !dir.exists('./Output') ) dir.create('./Output')
vf = get_VAF( pnt_mut, data_last, file_name = 'Output/VAF_data.txt')
VAF = get_rho_VAF( vf = vf, rho = c( 0.0, 0.1, 0.5 ) , file_name = './Output/VAF.txt' )
```

get\_type

*Function to get type of the clone: normal, primary or metastatic***Description**

Function to get type of the clone: normal, primary or metastatic

**Usage**

```
get_type(clone1)
```

**Arguments**

clone1                      Object of class 'Clone'

**Value**

One of characters 'normal', 'primary' or 'metastatic'

**Examples**

```
clone1 = tugHall_dataset$clones[[1]]
get_type( clone1 )
clone1 = tugHall_dataset$clones[[56]]
get_type( clone1 )
```

get\_u\_cna

*Function to choose probability of CNA mutation for several genes***Description**

Function to choose probability of CNA mutation for several genes

**Usage**

```
get_u_cna(genes, dupOrdel)
```

**Arguments**

genes	Names of genes, vector of names
dupOrdel	It could be 'dup' or 'del' to denote duplication or deletion

**Value**

Single value of maximal probability from probabilities for several genes

**Examples**

```
copy_files_to_Input()
define_parameters()
onco = tugHall_dataset$onco
get_u_cna( genes = 'APC', dupOrdel = 'dup' )
get_u_cna( genes = c('KRAS','APC'), dupOrdel = c('dup', 'del') )
```

---

get\_VAF

---

*Function to get data about Variant allele frequencies (VAF)*


---

**Description**

Function to get data about Variant allele frequencies (VAF)

**Usage**

```
get_VAF(pnt_mut, data_last, file_name = "Output/VAF_data.txt")
```

**Arguments**

pnt_mut	data.frame with point mutation info
data_last	data.frame with data of simulation at the last time step
file_name	Name of file to save data

**Value**

data.frame with info about Variant allele frequencies

**Examples**

```
pnt_mut = tugHall_dataset$pnt_mut
data_last = tugHall_dataset$data_last
vf = get_VAF( pnt_mut, data_last, file_name = 'Output/VAF_data.txt')
```

---

HallMark-class	<i>Class 'HallMark'</i>
----------------	-------------------------

---

**Description**

Class 'HallMark'

**Fields**

Ha numeric. Apoptosis hallmark indexes of genes in onco\$name,  
where onco is object of class OncoGene

Hi numeric. Immortalization hallmark indexes of genes in onco\$name,  
where onco is object of class OncoGene

Hd numeric. Growth/antigrowth hallmark indexes of genes in onco\$name,  
where onco is object of class OncoGene

Hb numeric. Angiogenesis hallmark indexes of genes in onco\$name,  
where onco is object of class OncoGene

Him numeric. Invasion/metastatic transformation hallmark indexes of genes in onco\$name,  
where onco is object of class OncoGene

Ha\_w numeric. Apoptosis hallmark weights of genes

Hi\_w numeric. Immortalization hallmark weights of genes

Hd\_w numeric. Growth/antigrowth hallmark weights of genes

Hb\_w numeric. Angiogenesis hallmark weights of genes

Him\_w numeric. Invasion/metastatic transformation hallmark weights of genes

notHa numeric. Indexes of genes which are not in apoptosis hallmark

**Examples**

```
hall = tugHall_dataset$hall
print( hall )
hall$copy()
hall$show()
```

---

init_clones	<i>Function to read file with initial clones</i>
-------------	--

---

**Description**

Function to read file with initial clones

**Usage**

```
init_clones(clonefile, clone1)
```

**Arguments**

clonefile	File to read
clone1	Object of class 'Clone'

**Value**

List of objects of class 'Clone'

**Examples**

```
copy_files_to_Input()
define_files_names()
env = tugHall_dataset$env
define_parameters()
onco = tugHall_dataset$onco
clone1 = tugHall_dataset$clones[[ 1 ]]
clones = init_clones(clonefile, clone1) # View( clones )
```

---

init_onco_clones	<i>Function to make list of objects of class 'OncoGene' and generate initial onco settings for all clones (onco_clones)</i>
------------------	---

---

**Description**

Function to make list of objects of class 'OncoGene' and generate initial onco settings for all clones (onco\_clones)

**Usage**

```
init_onco_clones(onco1, clones)
```

**Arguments**

onco1	Object of class 'OncoGene'
clones	List of objects of class 'Clone'

**Value**

List of objects of class 'OncoGene'

**Examples**

```
copy_files_to_Input()
define_files_names()
env = tugHall_dataset$env
define_parameters()
onco = tugHall_dataset$onco
clone1 = tugHall_dataset$clones[[ 1 ]]
clones = init_clones(clonefile, clone1) # View( clones )
onco_clones = init_onco_clones( onco1 = onco, clones )
```

---

init_pnt_clones	<i>Function to generate point mutations for initial clones</i>
-----------------	--

---

**Description**

Function to generate point mutations for initial clones

**Usage**

```
init_pnt_clones(clones, onco_clones)
```

**Arguments**

clones	List of objects of class 'Clone'
onco_clones	List of objects of class 'OncoGene'

**Examples**

```
clones = tugHall_dataset$clones
define_parameters()
onco = tugHall_dataset$onco
onco_clones = tugHall_dataset$onco_clones
copy_files_to_Input()
copy_files_to_Output()
define_gene_location()
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234
## Not run:
init_pnt_clones( clones, onco_clones ) # change pnt_clones for initialization

## End(Not run)
```

---

make_map	<i>Function to make a gene_map data.frame with information of genes' locations</i>
----------	--

---

**Description**

Function to make a gene\_map data.frame with information of genes' locations

**Usage**

```
make_map(
  f_out = "Input/map.txt",
  ls = c("CCDS4107.1", "CCDS8702.1", "CCDS43171.1", "CCDS11118.1"),
  f_in = "Input/CCDS.current.txt"
)
```

**Arguments**

f_out	Name of file to save gene_map data.frame
ls	List of IDs of genes corresponding CCDS database <a href="https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_h">https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_h</a>
f_in	Name of file to input downloaded from CCDS database

**Value**

gene\_map data.frame with information of genes' locations for genes of interest

**Examples**

```
url = 'https://ftp.ncbi.nlm.nih.gov/pub/CCDS/current_human/CCDS.current.txt'
download.file( url = url, destfile = 'CCDS.current.txt')
ls = c( 'CCDS4107.1', 'CCDS8702.1', 'CCDS43171.1', 'CCDS11118.1' )
gene_map = make_map(f_out = 'map.txt', ls = ls, f_in = 'CCDS.current.txt' )
```

---

mixed_mut_order	<i>Function to get order of mutation for all possible types</i>
-----------------	---

---

**Description**

Function to get order of mutation for all possible types

**Usage**

```
mixed_mut_order(clone1)
```

**Arguments**

clone1	Object of class 'Clone'
--------	-------------------------

**Value**

data.frame with fields order, type, ID

**Examples**

```
clone = tugHall_dataset$clones[[ 46 ]]
clone$PointMut_ID
clone$CNA_ID
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mixed_mut_order( clone )
```



---

model

Main function 'model' to simulate clones' evolution

---

## Description

Main function 'model' to simulate clones' evolution

## Usage

```
model(
  genefile,
  clonefile,
  geneoutfile,
  cloneoutfile,
  logoutfile,
  E0,
  F0,
  m0,
  uo,
  us,
  s0,
  k0,
  censure_n,
  censure_t,
  d0
)
```

## Arguments

genefile	Name of file with input data of hallmarks
clonefile	Name of file with initial clones data
geneoutfile	Name of file with hallmark data
cloneoutfile	Name of file with cloneout data
logoutfile	File name of log file
E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only
m0	Mutation probability for point mutation, numeric type only
uo	Oncogene mutation probability, numeric type only
us	Suppressor mutation probability, numeric type only
s0	Parameter in the sigmoid function, numeric type only
k0	Environmental death probability, numeric type only
censure_n	Max cell number where the program forcibly stops, integer type only
censure_t	Max time where the program forcibly stops, integer type only
d0	Initial probability to divide cells, numeric type only

**Value**

List of (clones, onco\_clones), where clones - list of objects of class 'Clone', and onco\_clones - list of objects of class 'OncoGene'. During a simulation it saves data to geneoutfile.

**Examples**

```
copy_files_to_Input()
define_files_names()
define_gene_location()
define_parameters( read_fl = TRUE , file_name = './Input/parameters.txt' )
define_compaction_factor( read_fl = TRUE , file_name = './Input/CF.txt' )
time_stop = 3 # Duration of simulation time is 3 sec
## Not run:
res = model( genefile, clonefile, geneoutfile, cloneoutfile, logoutfile,
E0, F0, m0, uo, us, s0, k0, censore_n, censore_t, d0 )

## End(Not run)
```

---

modify\_gene\_map

*Function to add the mutations to the data.frame gene\_map*

---

**Description**

Function to add the mutations to the data.frame gene\_map

**Usage**

```
modify_gene_map(clone1, onco1)
```

**Arguments**

clone1	Object of class 'Clone'
onco1	Object of class 'OncoGene'

**Value**

list( gm1, gm2 ), where gm1 and gm2 are data.frames gene\_maps with mutation information

**Examples**

```
clone = tugHall_dataset$clones[[ 46 ]]
onco = tugHall_dataset$onco_clones[[ 46 ]]
gene_map = tugHall_dataset$gene_map
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
gene_map$pnts = ''
## Not run:
gm_1_2 = modify_gene_map( clone , onco ) # View(gm_1_2)

## End(Not run)
```

---

number_N_P_M	<i>Function to get number of cells of a clone with indicator (normal, primary tumor or metastatic)</i>
--------------	--

---

**Description**

Function to get number of cells of a clone with indicator (normal, primary tumor or metastatic)

**Usage**

```
number_N_P_M(clone1)
```

**Arguments**

clone1                      Object of class 'Clone'

**Value**

Vector c( N\_normal, N\_primary, N\_metastatic )

**Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
number_N_P_M(clone1)
message( paste('Format is as follow: ', 'N_normal', 'N_primary', 'N_metastatic' ) )
```

---

OncoGene-class	<i>Class 'OncoGene'</i>
----------------	-------------------------

---

**Description**

Class 'OncoGene'

**Fields**

id    numeric. ID is same as in clone (key for clones)

name    character. Onco genes' names list

onsp    character. Oncogene/suppressor indicator for each gene in list of names

len    numeric. Lengths of onco genes

cds\_1    numeric. Onco genes' CDS base lengths for parental chr 1

cds\_2    numeric. Onco genes' CDS base lengths for parental chr 2

rna\_1    numeric. Onco genes RNA base number length for parental chr 1 (exons+introns)

rna\_2    numeric. Onco genes RNA base number length for parental chr 2 (exons+introns)

p0\_1    numeric. Probability of absent of mutations for parental chr 1

p0\_2    numeric. Probability of absent of mutations for parental chr 2

prob\_1    numeric. Vector of relative probabilities for point mutation, deletion and duplication:  

$$\text{prob} = c( m0 \times \text{sumCDS}, m\_del \times \text{sumRNA}, m\_dup \times \text{sumRNA} ) / \text{sum}( m0 \times \text{sumCDS}, m\_del \times \text{sumRNA}, m\_dup \times \text{sumRNA} )$$

```

prob_2 numeric.
sum_prob_1 numeric.
sum_prob_2 numeric.

```

### Examples

```

onco = tugHall_dataset$onco
onco$copy()

```

---

onco_copy	<i>Function to make one copy for onco1 in init_onco_clones function</i>
-----------	---

---

### Description

Function to make one copy for onco1 in init\_onco\_clones function

### Usage

```
onco_copy(onco1)
```

### Arguments

onco1                      Object of class 'OncoGene'

### Value

New object of class 'OncoGene' with the same info

### Examples

```

onco1 = tugHall_dataset$onco_clones[[ 1 ]]
onco2 = onco_copy( onco1 ) # ID + 1

```

---

onco_update	<i>Function to update onco1 after mutation (for usage in trial_mutagenesis() function)</i>
-------------	--

---

### Description

Function to update onco1 after mutation (for usage in trial\_mutagenesis() function)

### Usage

```
onco_update(onco1, gm)
```

### Arguments

onco1                      Object of class 'OncoGene'

gm                          data.frame gene\_map

**Value**

onco1 with updated info

**Examples**

```
onco1 = tugHall_dataset$onco_clones[[ 1 ]]
copy_files_to_Input()
define_gene_location()
define_parameters()
onco_update( onco1, gm = list(gene_map, gene_map[1:42, ] ) ) # Check CDS length for TP53 gene
```

---

order_gene_map	<i>Function to order info in gene_map data.frame with information of genes' locations</i>
----------------	---

---

**Description**

Function to order info in gene\_map data.frame with information of genes' locations

**Usage**

```
order_gene_map(gene_map)
```

**Arguments**

gene\_map            data.frame with information of genes' locations

**Value**

The same data.frame gene\_map with ordered positions for each gene and each chromosome

**Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map = order_gene_map( gene_map )
```

---

plot_2D	<i>Function to plot 2D figure of points <math>y = y(x)</math></i>
---------	---

---

**Description**

Function to plot 2D figure of points  $y = y(x)$

**Usage**

```
plot_2D(
  x,
  y,
  names = c("X", "Y"),
  pch = 18,
  col = "blue",
  cex = 1.2,
  xr = c(-10, 10),
  yr = c(-10, 10),
  safe_pdf = FALSE,
  filename = "./plot.pdf",
  change_par = TRUE
)
```

**Arguments**

x	Input data for axes X
y	Input data for axes Y
names	Vector of two characters with names for X and Y axes
pch	Parameter pch for plot function
col	Colors of points
cex	Parameter cex for plot function
xr	Range for X
yr	Range for Y
safe_pdf	Indicator to save plot to a file or not
filename	Name of file to save plot if safe_pdf == TRUE
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after plot it will be returned to initial values

**Value**

NULL, making 2D plot using points

**Examples**

```
plot_2D( x=-5:5, y=-3:7 )
```

---

plot_2D_lines	<i>Function to plot 2D figure of lines <math>y_i = DF[, nl[i]]</math> , <math>i</math> - index</i>
---------------	--

---

**Description**

Function to plot 2D figure of lines  $y_i = DF[, nl[i]]$  ,  $i$  - index

**Usage**

```

plot_2D_lines(
  x,
  DF,
  nl = 1:2,
  names = c("X", "Y"),
  legend_names = "",
  col = c("blue3", "darkmagenta", "red", "green4", "darkorange", "steelblue1"),
  cex = 1.2,
  lwd = 2,
  lt = c(1:6),
  xr = c(-10, 10),
  yr = c(-10, 10),
  safe_pdf = FALSE,
  filename = "./plot.pdf",
  type = "l",
  logscale = "",
  draw_key = TRUE,
  change_par = TRUE
)

```

**Arguments**

x	Input data for axes X
DF	data.frame with data to plot
nl	indexes of columns in DF to plot
names	Vector of two characters with names for X and Y axes
legend_names	Name of legend
col	Vector of colors for lines
cex	Parameter cex for plot function
lwd	Vector of width of lines
lt	Vector of types of lines
xr	Range for X
yr	Range for Y
safe_pdf	Indicator to save plot to a file or not
filename	Name of file to save plot if safe_pdf == TRUE
type	Parameter type in plot function
logscale	Parameter logscale in plot function
draw_key	Indicator to draw key or not
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after plot it will be returned to initial values

**Value**

NULL, making 2D plot using lines

**Examples**

```
DF = tugHall_dataset$data_avg
plot_2D_lines( x = DF[, 1 ], DF, nl = 8:12 , xr = c(1,max(DF$Time) ), yr = c(0,1) )
xr = c(1,max(DF$Time) )
yr = c(0,max(DF[,14],DF[,16],DF[,17] ))
plot_2D_lines( x = DF[, 1 ], DF, nl = c(14,16,17) , xr =xr, yr = yr )
plot_2D_lines( x = DF[, 1 ], DF, nl = 18:22 , xr = c(1,max(DF$Time) ), yr = c(0,1) )
```

---

plot\_average\_simulation\_data

*Function to plot main data from data.frame with average data*

---

**Description**

Function to plot main data from data.frame with average data

**Usage**

```
plot_average_simulation_data(data_avg, time_max)
```

**Arguments**

data_avg	data.frame with average values from cloneout.txt file
time_max	Maximal time step in a simulation

**Value**

NULL, draw many plot with average data

**Examples**

```
data_avg = tugHall_dataset$data_avg
time_max = tugHall_dataset$time_max
plot_average_simulation_data( data_avg , time_max = time_max )
```

---

plot\_clone\_evolution    *Function to plot clone evolution*

---

**Description**

Function to plot clone evolution



**Usage**

```
plot_clone_evolution(
  data_flow,
  threshold = c(0.05, 1),
  lwd = 2,
  hue = c(" ", "random", "red", "orange", "yellow", "green", "blue", "purple", "pink",
    "monochrome")[1],
  luminosity = c(" ", "random", "light", "bright", "dark")[5],
  yr = NA,
  add_initial = TRUE,
  log_scale = FALSE,
  change_par = TRUE
)
```

**Arguments**

data_flow	data.frame with results of simulation at each time step
threshold	Vector two numbers from 0 to 1 to show clones with relative final numbers of cells in the range of threshold
lwd	Line width in the plot function
hue	Parameter hue in the function randomColor from library randomcoloR
luminosity	Parameter luminosity in the function randomColor from library randomcoloR
yr	Range for Y axes
add_initial	Indicator to add or do not add initial clones to plot
log_scale	Indicator to use log_scale or not for Y axes
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after plot it will be returned to initial values

**Value**

NULL, making plot with clones evolution

**Examples**

```
data_flow = tugHall_dataset$data_flow
plot_clone_evolution( data_flow, threshold = c(0.01, 1 ), add_initial = TRUE, log_scale = FALSE )
plot_clone_evolution( data_flow, threshold = c(0, 0.01 ), add_initial = FALSE, log_scale = TRUE )
```

---

plot\_order\_dysfunction

*Function to plot order of genes dysfunction as a step function with number of cells related to each order*

---

**Description**

Function to plot order of genes dysfunction as a step function with number of cells related to each order

Usage

```
plot_order_dysfunction(  
  rdr_dysf,  
  pos = c(0, 100),  
  logscale = "y",  
  cex = 1,  
  change_par = TRUE  
)
```

Arguments

rdr_dysf	Order of genes dysfunction as a data.frame
pos	Coordinates of list of order of genes dysfunction
logscale	Parameter logscale for plot function
cex	Parameter cex for plot function
change_par	Indicator to change par() or not for a plot. By default change_par = TRUE, after plot it will be returned to initial values

Value

NULL, making plot with step function of order of genes' dysfunction

Examples

```
rdr_dysf = tugHall_dataset$rdr_dysf  
plot_order_dysfunction( rdr_dysf , logscale = '', pos = c(8, 5000), cex = 1.4)  
plot_order_dysfunction( rdr_dysf , logscale = 'y', pos = c(10, 100), cex = 1.2)
```

---

pnts_add_dlt	<i>Function to subtract delta from position of point mutations</i>
--------------	--

---

Description

Function to subtract delta from position of point mutations

Usage

```
pnts_add_dlt(gm_w1, dlt)
```

Arguments

gm_w1	A row from data.frame gene_map
dlt	Delta to subtract from positions of point mutations

Value

Return the pnts - dlt for one row of data.frame gene\_map

**Examples**

```
gene_map = tugHall_dataset$gene_map
gene_map$pnts = ''
gene_map$pnts[6] = '112792451'
gm_w1 = gene_map[6,]
pnts_add_dlt( gm_w1 , dlt = 1000 )
pnts_add_dlt( gm_w1 , dlt = -1001 )
```

---

Point\_Mutations-class    *Class 'Point\_Mutations'*

---

**Description**

Class 'Point\_Mutations'

**Fields**

PointMut\_ID    numeric. ID of point mutation

Allele    character. A or B allele

Parental\_1or2    numeric. Parental chromosome, could be 1 or 2

Chr    character. Chromosome name

Ref\_pos    numeric. Reference position

Phys\_pos    vector. Physical positions

Delta    vector. Delta of positions

Copy\_number    numeric. Copy number of allele

Gene\_name    character. Gene's name

MalfunctionedByPointMut    logical. True for driver mutation and False for passenger mutation

mut\_order    numeric. Number in order of mutation to reproduce the gene\_map data.frame

**Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
print( pnt )
pnt$copy()
pnt$show()
pnt$initialize()
pnt$show()
pnt = tugHall_dataset$pnt_clones[[ 3 ]]
pnt$save()    # save as row of data.frame
```

---

print_parameters	<i>Function to print GLOBAL parameters</i>
------------------	--

---

**Description**

Function to print GLOBAL parameters

**Usage**

```
print_parameters()
```

**Value**

Message with values of all the GLOBAL parameters

**Examples**

```
copy_files_to_Input()
define_parameters( read_fl = FALSE )
define_compaction_factor()
print_parameters()
```

---

read_file	<i>Function to read file</i>
-----------	------------------------------

---

**Description**

Function to read file

**Usage**

```
read_file(file_name = "", stringsAsFactors = FALSE, header = TRUE)
```

**Arguments**

file_name	Name of file to read
stringsAsFactors	Parameter for read.table function, by default stringsAsFactors = FALSE
header	Logical type to read or do not read head of a file

**Value**

data.frame of data from a file

**Examples**

```
f1 = system.file('extdata/Input', 'gene_map.txt', package = 'tugHall1.3', mustWork = TRUE )
read_file(file_name = f1, stringsAsFactors = FALSE )
f1 = system.file('extdata/Input', 'CF.txt', package = 'tugHall1.3', mustWork = TRUE )
read_file(file_name = f1, stringsAsFactors = FALSE, header = FALSE )
```

---

safe_pnt_mut	<i>Function to save 1 point mutation in a data frame</i>
--------------	--

---

**Description**

Function to save 1 point mutation in a data frame

**Usage**

```
safe_pnt_mut(pnt)
```

**Arguments**

pnt                      Object of class 'Point\_Mutations'

**Value**

data frame with 1 row of point mutation info

**Examples**

```
pnt = tugHall_dataset$pnt_clones[[ 1 ]]
df = safe_pnt_mut( pnt ) # View( pnt )
```

---

simulation_example	<i>Example of simulation for lazy start</i>
--------------------	---

---

**Description**

Example of simulation for lazy start

**Usage**

```
simulation_example(verbose = TRUE, to_plot = TRUE, seed = NA)
```

**Arguments**

verbose                  Logical type to show or do not show messages during execution  
 to\_plot                  Logical type to plot or do not plot graphical results of a simulation  
 seed                      Numeric type to set seed for a simulation, if seed = NA (by default) then it will be skipped

**Value**

List of results of simulation with default values for all the parameters

**Examples**

```
# it takes a time for a simulation and then it will demonstrates results, \cr
# so, please, wait for a while
simulation_example( verbose = FALSE , to_plot = FALSE )
```

---

sum_cell	<i>Aggregate data of a clone for environment object</i>
----------	---

---

**Description**

Aggregate data of a clone for environment object

**Usage**

```
sum_cell(env, clones)
```

**Arguments**

env	Object of class 'Environ'
clones	List of all the objects of class 'Clone'

**Value**

NULL, but global variable env is updated

**Examples**

```
clones = tugHall_dataset$clones
env = tugHall_dataset$env
sum_cell(env, clones)
message( paste0('Number of primary tumor cells in the pool of clones is ', env$P ) )
message( paste0('Number of normal cells in the pool of clones is ', env$N ) )
message( paste0('Number of metastatic cells in the pool of clones is ', env$M ) )
```

---

sum_mutation	<i>Serve function for sum_cell() function</i>
--------------	---

---

**Description**

Serve function for sum\_cell() function

**Usage**

```
sum_mutation(clone1)
```

**Arguments**

clone1	Object of class 'Clone'
--------	-------------------------

**Value**

vector of clone1 variables to aggregate in sum\_cell() function

**Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
sum_mutation(clone1)
```

---

sum\_N\_P\_M

*Function to calculate N and M numbers - normal and metastatic cells*


---

**Description**

Function to calculate N and M numbers - normal and metastatic cells

**Usage**

```
sum_N_P_M(env, clones)
```

**Arguments**

env	Object of class 'Environ'
clones	List of all the objects of class 'Clone'

**Value**

Number of all the cells in a simulation (normal + primary tumor + metastatic)

**Examples**

```
clones = tugHall_dataset$clones
env = tugHall_dataset$env
env$M = 0
env$P = 0
env$N = 0 # View( env )
sum_N_P_M(env, clones) # View( env )
message( paste(env$N, env$P, env$M ) )
```

---

trial\_complex

*Function trial for complex case of models*


---

**Description**

Function trial for complex case of models

**Usage**

```
trial_complex(clone1, onco1)
```

**Arguments**

clone1	Object of class 'Clone'
onco1	Object of class 'OncoGene'

**Value**

Number of new clones originated by clone1

**Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco
trial_complex( clone1, onco1 )
unlist( lapply( X = 1:20, FUN = function( x ) trial_complex( clone1, onco1 ) ) ) )
```

---

trial_mutagenesis	<i>Function for mutagenesis trial</i>
-------------------	---------------------------------------

---

**Description**

Function for mutagenesis trial

**Usage**

```
trial_mutagenesis(clone1, num_mut, onco1)
```

**Arguments**

clone1	Object of class 'Clone'
num_mut	Number of mutations in this NEW clone1
onco1	Object of class 'OncoGene' corresponding to clone1 (with the same ID)

**Value**

Changed object clone1, add related mutations to the lists of point mutations and/or CNA mutations

**Examples**

```
copy_files_to_Input()
copy_files_to_Output()
define_parameters()
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco_clones[[ 1 ]]
onco = tugHall_dataset$onco
define_gene_location()
pnt_clones = tugHall_dataset$pnt_clones
cna_clones = tugHall_dataset$cna_clones
mut_order = 234 # Just an example number
message( c('CNA mutation IDs ', paste(clone1$CNA_ID, collapse = ' ')) )
message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ')) )
## Not run:
trial_mutagenesis( clone1, num_mut = 1, onco1 ) # it adds info to clone1
message( c('CNA mutation IDs ', paste(clone1$CNA_ID, collapse = ' ')) )
message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ')) )

trial_mutagenesis( clone1, num_mut = 10, onco1 ) # it adds info to clone1
message( c('CNA mutation IDs ', paste(clone1$CNA_ID, collapse = ' ')) )
message( c('Point mutation IDs ', paste(clone1$PointMut_ID, collapse = ' ')) )

## End(Not run)
```



---

trial_simple	<i>Function trial for simplified case of model</i>
--------------	--

---

**Description**

Function trial for simplified case of model

**Usage**

```
trial_simple(clone1, onco1)
```

**Arguments**

clone1	Object of class 'Clone'
onco1	Object of class 'OncoGene'

**Value**

Number of new clones originated by clone1

**Examples**

```
clone1 = tugHall_dataset$clones[[ 1 ]]
onco1 = tugHall_dataset$onco
trial_simple( clone1, onco1 )
unlist( lapply( X = 1:20, FUN = function( x ) trial_simple( clone1, onco1 ) ) ) )
```

---

tugHall_dataset	<i>tugHall dataset named 'tugHall_dataset'</i>
-----------------	--

---

**Description**

Dataset contains all the necessary data.frames and objects to check functions of tugHall. Description of each data.frame and object could be found in documentation to tugHall package.

**Usage**

```
tugHall_dataset
```

**Format**

A data frame with 12 data.frames and 2 objects:

**data\_flow** simulation data for all time steps, data from file cloneout.txt  
**data\_last** simulation data for the last time step, data from file cloneout.txt  
**data\_avg** simulation data averaged for the each time step, data from file cloneout.txt  
**pnt\_mut** data.frame with point mutation information  
**pnt\_mut\_B** data.frame with point mutation information of mutated allele B  
**cna\_mut** data.frame with CNA mutation information

**gene\_map** data.frame with genes' locations information

**hall** Object of class 'HallMark'

**onco** Object of class 'OncoGene'

**time\_max** Value of maximal time step in an example simulation

**CF** Values of compaction factor

**vf** data.frame of preliminary data for VAF calculations

**VAF\_rho** data.frame with VAF values for different rho

**rdr\_dysf** data.frame of order of genes dysfunction for each clone

---

update_Hallmarks	<i>Function to update Hallmark and variable after division or under initialization</i>
------------------	--

---

## Description

Function to update Hallmark and variable after division or under initialization

## Usage

```
update_Hallmarks(clone1)
```

## Arguments

clone1            Object of class 'Clone'

## Value

The same object of class 'Clone' with updated fields

## Examples

```
clone = tugHall_dataset$clones[[ 1 ]]
define_parameters()
hall = tugHall_dataset$hall
env = tugHall_dataset$env
update_Hallmarks( clone )
```

---

write_cloneout	<i>Function to write data to cloneout file at a time step</i>
----------------	---

---

**Description**

Function to write data to cloneout file at a time step

**Usage**

```
write_cloneout(outfile, env, clones, isFirst, onco_clones)
```

**Arguments**

outfile	File name for output info
env	Object of class 'Environ'
clones	List of objects of class 'Clone'
isFirst	logical type = TRUE as default
onco_clones	List of objects of class 'OncoGene'

**Value**

NULL, but add rows to output file with clone evolution data

**Examples**

```
env = tugHall_dataset$env
onco = tugHall_dataset$onco
if ( !dir.exists('./Output') ) dir.create('./Output')
clones = tugHall_dataset$clones
onco_clones = tugHall_dataset$onco_clones
write_header(outfile='./Output/exmpl.txt', env, onco)
write_cloneout( outfile = './Output/exmpl.txt', env, clones, isFirst = TRUE, onco_clones )
```

---

write_geneout	<i>Function to write info about HallMark data</i>
---------------	---

---

**Description**

Function to write info about HallMark data

**Usage**

```
write_geneout(outfile, hall, Compaction_factor, CF)
```

**Arguments**

outfile	File name for output info
hall	Object of class "HallMark"
Compaction_factor	Compaction factor, logical type only. True means 'to use', False means 'do not use' Compaction factor for hallmarks variables
CF	Vector with values of compaction factor for each hallmark

**Value**

NULL, but data will save to a file

**Examples**

```
copy_files_to_Input()
define_files_names()
define_parameters()
if ( !dir.exists('./Output') ) dir.create('./Output')
hall = tugHall_dataset$hall
onco = tugHall_dataset$onco
define_compaction_factor()
write_geneout(outfile = geneoutfile, hall, Compaction_factor, CF)
```

---

write\_header

*Function to write the header to a file*

---

**Description**

Function to write the header to a file

**Usage**

```
write_header(outfile, env, onco)
```

**Arguments**

outfile	File name for output info
env	Object of class 'Environ'
onco	Object of class "OncoGene"

**Value**

NULL, but the header will save to a file and delete old info

**Examples**

```
env = tugHall_dataset$env
onco = tugHall_dataset$onco
if ( !dir.exists('./Output') ) dir.create('./Output')
write_header(outfile='./Output/exmpl.txt', env, onco)
```

---

write_log	<i>Function to write log file</i>
-----------	-----------------------------------

---

## Description

Function to write log file

## Usage

```
write_log(
  genefile,
  clonefile,
  geneoutfile,
  cloneoutfile,
  logoutfile,
  E0,
  F0,
  m0,
  uo,
  us,
  s0,
  k0,
  m_dup,
  m_del,
  lambda_dup,
  lambda_del,
  uo_dup,
  us_dup,
  uo_del,
  us_del,
  censore_n,
  censore_t,
  d0,
  Compaction_factor,
  model_name,
  time_stop,
  n_repeat,
  monitor
)
```

## Arguments

genefile	File name of initial OncoGene information
clonefile	File name of info about initial clones
geneoutfile	File name for output info about OncoGene information
cloneoutfile	File name for output info with clone evolution data
logoutfile	Name of log file with all the parameters
E0	Parameter in the division probability, numeric type only
F0	Parameter in the division probability, numeric type only

m0	Mutation probability for point mutation, numeric type only
uo	Oncogene mutation probability, numeric type only
us	Suppressor mutation probability, numeric type only
s0	Parameter in the sigmoid function, numeric type only
k0	Environmental death probability, numeric type only
m_dup	Mutation probability for duplication, numeric type only
m_del	Mutation probability for deletion, numeric type only
lambda_dup	CNA duplication average length (of the geometrical distribution for the length), integer type only
lambda_del	CNA deletion average length (of the geometrical distribution for the length), integer type only
uo_dup	Gene malfunction probability by CNA duplication for oncogene, numeric type only
us_dup	Gene malfunction probability by CNA duplication for suppressor, numeric type only
uo_del	Gene malfunction probability by CNA deletion for oncogene, numeric type only
us_del	Gene malfunction probability by CNA deletion for suppressor, numeric type only
censore_n	Max cell number where the program forcibly stops, integer type only
censore_t	Max time where the program forcibly stops, integer type only
d0	Initial probability to divide cells, numeric type only
Compaction_factor	Compaction factor, logical type only. True means 'to use', False means 'do not use' Compaction factor for hallmarks variables
model_name	Name of the model to use. Can be 'proportional_metastatic' or 'threshold_metastatic' or 'simplified'
time_stop	Max time in seconds of running after that the program forcibly stops, integer type only
n_repeat	Max number of repetition of the program until the NON-ZERO output will be getting, integer type only
monitor	The indicator to make monitor file during a simulation or do not make, logical type only

### Value

NULL, write log file to Output folder

### Examples

```

copy_files_to_Input()
define_files_names()
define_parameters()
if ( !dir.exists('./Output') ) dir.create('./Output')
## Not run:
write_log(genefile, clonefile, geneoutfile, cloneoutfile, logoutfile,
E0, F0, m0, uo, us, s0, k0, m_dup, m_del, lambda_dup, lambda_del,
uo_dup, us_dup, uo_del, us_del, censore_n, censore_t, d0,
Compaction_factor, model_name, time_stop, n_repeat, monitor )

## End(Not run)

```

---

write_monitor	<i>Function to write a simulation monitoring data into the file_monitor</i>
---------------	---

---

**Description**

Function to write a simulation monitoring data into the file\_monitor

**Usage**

```
write_monitor(outfile = file_monitor, start = FALSE, env, clones)
```

**Arguments**

outfile	File name for output info
start	Indicator to start from beginning (TRUE) or not (FALSE)
env	Object of class 'Environ'
clones	List of objects of class 'Clone'

**Value**

NULL, but info about current state of simulation will write to a file

**Examples**

```
env = tugHall_dataset$env
if ( !dir.exists('./Output') ) dir.create('./Output')
clones = tugHall_dataset$clones
onco_clones = tugHall_dataset$onco_clones
cna_clones = tugHall_dataset$cna_clones
pnt_clones = tugHall_dataset$pnt_clones
write_monitor( outfile = './Sim_monitoring.txt', start = TRUE , env, clones )
write_monitor( outfile = './Sim_monitoring.txt', start = FALSE , env, clones )
```

---

write_pnt_clones	<i>Function to write the point mutation info for all clones for all time steps, used at the last time step or after simulation</i>
------------------	--

---

**Description**

Function to write the point mutation info for all clones for all time steps, used at the last time step or after simulation

**Usage**

```
write_pnt_clones(pnt_clones, file_out = "Output/point_mutations.txt")
```

**Arguments**

pnt_clones	List of objects of class 'Point_Mutations'
file_out	File name to write

**Value**

NULL, but info will write to a file

**Examples**

```
pnt_clones = tugHall_dataset$pnt_clones
if ( !dir.exists('./Output') ) dir.create('./Output')
write_pnt_clones(pnt_clones, file_out = 'Output/point_mutations.txt')
```

---

write_weights	<i>Function to write info about relationship between genes and hallmarks</i>
---------------	--

---

**Description**

Function to write info about relationship between genes and hallmarks

**Usage**

```
write_weights(outfile, hall)
```

**Arguments**

outfile	File name for output info
hall	Object of class 'HallMark'

**Value**

NULL, but info about relationship between genes and hallmarks will write to a file

**Examples**

```
if ( !dir.exists('./Output') ) dir.create('./Output')
hall = tugHall_dataset$hall
onco = tugHall_dataset$onco
write_weights(outfile = './Output/weights.txt', hall)
```



# Index

## \*Topic **datasets**

tugHall\_dataset, [49](#)

add\_deletion, [3](#)  
add\_duplication, [4](#)  
add\_pnt\_mutation, [4](#)

calc\_binom, [5](#)  
change\_allele\_A\_by\_cna, [5](#)  
change\_pnt\_by\_cna, [6](#)  
check\_pkg, [7](#)  
check\_pnts, [7](#)  
chk\_pnt\_mut, [8](#)  
clone (Clone-class), [8](#)  
Clone-class, [8](#)  
clone\_copy, [9](#)  
CNA\_Mutations (CNA\_Mutations-class), [10](#)  
CNA\_Mutations-class, [10](#)  
copy\_CNA, [11](#)  
copy\_files\_to\_Input, [11](#)  
copy\_files\_to\_Output, [12](#)  
copy\_pnt, [13](#)  
copy\_pnt\_no\_mutation, [13](#)

define\_compaction\_factor, [14](#)  
define\_files\_names, [14](#)  
define\_gene\_location, [15](#)  
define\_par\_for\_plot, [17](#)  
define\_parameters, [16](#)

environ (Environ-class), [18](#)  
Environ-class, [18](#)

gen\_colors, [21](#)  
generate\_cna, [19](#)  
generate\_pnt, [20](#)  
generate\_to\_copy\_pnt, [20](#)  
get\_cds\_rna, [21](#)  
get\_cna\_mutation, [22](#)  
get\_flow\_data, [23](#)  
get\_len\_cds\_rna, [24](#)  
get\_order\_of\_genes\_dysfunction, [24](#)  
get\_point\_mutation, [25](#)  
get\_point\_mutation\_for\_gene, [26](#)  
get\_rho\_VAF, [26](#)

get\_type, [27](#)  
get\_u\_cna, [27](#)  
get\_VAF, [28](#)

hallmark (HallMark-class), [29](#)  
HallMark-class, [29](#)

init\_clones, [29](#)  
init\_onco\_clones, [30](#)  
init\_pnt\_clones, [31](#)

make\_map, [31](#)  
mixed\_mut\_order, [32](#)  
model, [33](#)  
modify\_gene\_map, [34](#)

number\_N\_P\_M, [35](#)

onco\_copy, [36](#)  
onco\_update, [36](#)  
oncogene (OncoGene-class), [35](#)  
OncoGene-class, [35](#)  
order\_gene\_map, [37](#)

plot\_2D, [37](#)  
plot\_2D\_lines, [38](#)  
plot\_average\_simulation\_data, [40](#)  
plot\_clone\_evolution, [40](#)  
plot\_order\_dysfunction, [41](#)  
pnts\_add\_dlt, [42](#)  
Point\_Mutations  
(Point\_Mutations-class), [43](#)  
Point\_Mutations-class, [43](#)  
print\_parameters, [44](#)

read\_file, [44](#)

safe\_pnt\_mut, [45](#)  
simulation\_example, [45](#)  
sum\_cell, [46](#)  
sum\_mutation, [46](#)  
sum\_N\_P\_M, [47](#)

trial\_complex, [47](#)  
trial\_mutagenesis, [48](#)

trial\_simple, [49](#)  
tugHall\_dataset, [49](#)  
  
update\_Hallmarks, [50](#)  
  
write\_cloneout, [51](#)  
write\_geneout, [51](#)  
write\_header, [52](#)  
write\_log, [53](#)  
write\_monitor, [55](#)  
write\_pnt\_clones, [55](#)  
write\_weights, [56](#)