Supporting Information: Identifying signatures of proteolytic stability and monomeric propensity in *O*-glycosylated insulin using molecular simulation

Wei-Tse Hsu¹, Dominique A. Ramirez², Tarek Sammakia³, Zhongping Tan⁴, Michael R. Shirts¹

¹Department of Chemical & Biological Engineering, University of Colorado Boulder, Boulder, CO, USA 80309; ²Department of Biochemistry, University of Colorado Boulder, Boulder, CO, USA 80309; ³Department of Chemistry, University of Colorado Boulder, Boulder, CO, USA 80309; ⁴Institute of Materia Medica, Chinese Academy of Medical Sciences, Peking Union Medical College, Beijing, 100050, China

*For correspondence:

michael.shirts@colorado.edu (MRS); zhongping.tan@imm.pumc.edu.cn (ZT)

Submitted in the Journal of Computer-Aided Molecular Design.

1 Supplemental Tables

| | 4EYD | 4EY1 | 3I3Z | 4EY9 | 2MVC |
|---------------|----------|----------|----------|----------|----------|
| Total charges | -1 | -1 | -1 | -1 | -1 |
| pH value | 8.0 | 8.0 | 7.9 | 6.9 | 7.3 |
| HisB5 | HIP (+1) | HIP (+1) | HIP (+1) | HIP (+1) | HIE (+0) |
| HisB10 | HIE (+0) | HIE (+0) | HIE (+0) | HID (+0) | HIP (+1) |

Table S1. The comparison of histidine protonation states of wild-type structures.

| | GF 2 | GF 3 | GF 4 | GF 5 | GF 6 | GF 7 | GF 8 | GF 9 | GF 10 | GF 11 | GF 12 | GF 13 |
|------|------|------|-------|--------|--------|------|-------|--------|--------|-------|--------|--------|
| 4EYD | NA | NA | 3.45 | 63.56 | 69.11 | NA | NA | 45.79 | 121.70 | 21.65 | 140.40 | 24.77 |
| 4EY1 | NA | NA | 6.89 | 371.83 | 71.04 | NA | 19.37 | 205.24 | 159.14 | 58.17 | 360.45 | 637.20 |
| 4EY9 | NA | NA | 18.53 | 274.66 | 97.34 | NA | NA | 12.55 | 496.98 | 0.39 | 89.13 | 57.77 |
| 3I3Z | NA | NA | 93.49 | 62.86 | 199.65 | NA | NA | 45.27 | 62.80 | 42.78 | 97.81 | 281.52 |
| 2MVC | 3.98 | NA | 21.31 | 54.78 | 105.28 | NA | NA | 114.61 | 59.62 | 58.75 | 66.00 | 81.57 |

Table S2. Dimer occlusion autocorrelation lag times for each of the listed models. All numbers are listed in units of nanoseconds.

| # | 4EYD-based GFs | 4EY1-based GFs | 4EY9-based GFs | 3I3Z-based GFs | 2MVC-based GFs |
|----|--|---|---|--|--|
| 1 | N/A | N/A | N/A | N/A | N/A |
| 2 | None | ThrA8(OG1)-GalNAc[1](O2N):10% | None | None | GlnA5(NE2)-GalNAc[1](O6): 14% GlnA15(NE2)-GalNAc[1](O6): 13% |
| 3 | None | None | None | None | GlnA15(NE2)-52(O2N):11% |
| 4 | None | None | None | None | None |
| 5 | None | PheB24(N)-GalNAc[1](O3): 48% ThrB30(N)-GalNAc[1](O2N): 16% | PheB24(N)-GalNAc[1](O3): 11% | ThrB27(N)-GalNAc[1](O3):15% Glu4(N)-GalNAc[1](O2N): 14% | PheB24(N)-GalNAc[1](O3): 10% |
| 6 | ThrB27(N)-GalNAc[1](O3):26% | ThrB27(N)-GalNAc(1)(O3):26% | None | None | CysA7(N)-GalNAc[1](O2N): 18% ThrB27(N)-GalNAc(1)(O3): 12% |
| 7 | None | None | None | None | GlnA15(NE2)-Man[1](O4): 18% |
| 8 | GlnA5(NE2)-Man[2](O2): 12% | GlnA5(NE2)-Man[2](O2): 11% | None | GlnA5(NE2)-Man[1](O5): 12% | None |
| 9 | PheB24(N)-Man[1](O3): 22% TyrB16(OH)-Man[1](O4): 13% | None | PheB24(N)-Man[1](O3): 27% TyrB16(OH)-Man[1](O4): 19% | None | None |
| 10 | PheB24(N)-Man[1](O3): 52% ThrB30(N)-Man[2](O2): 35% ThrB27(N)-Man[2](O6): 23% | ThrB27(N)-Man[2](O6): 46% PheB24(N)-Man[1](O3): 30% TyrB16(OH)-Man[1](O4): 15% | ThrB27(N)-Man[2](O6): 45% PheB24(N)-Man[1](O3): 28% TyrB16(OH)-Man[1](O4): 11% | ThrB27(N)-Man[2](O6): 47% PheB24(N)-Man[1](O3): 15% | ThrB27(N)-Man[2](O6): 47% PheB24(N)-Man[1](O3): 37% TyrB16(OH)-Man[1](O4): 14% |
| 11 | None | None | None | None | None |
| 12 | ThrB30(N)-Man[2](O6): 31% GlyB23(N)-Man[1](O3): 10% | TyrA19(OH)-Man[1](O4): 51% ThrB27(N)-Man[1](O3): 43% ThrB30(N)-Man[2](O6): 19% | ThrB30(N)-Man[2](O6): 24% ValA3(N)-Man[2](O2): 15% | ThrB30(N)–Man[2](O6): 38% | ThrB30(N)-Man[2](O6): 49% GlyB8(N)-Man[2](O4): 10% |
| 13 | ThrB27(N)-Man[2](O6): 61% PheB24(N)-Man[1](O3): 49% TyrB16(OH)-Man[1](O4): 16% | ThrB27(N)-Man[2](O6): 31% ThrB27(N)-Man[3](O6): 14% PheB24(N)-Man[1](O3): 12% TyrB16(OH)-Man[1](O4): 11% | ThrB27(N)-Man[2](O6): 27% PheB24(N)-Man[1](O3): 17% ThrB27(N)-Man[3](O6): 15% TyrB16(OH)-Man[1](O4): 10% | ThrB27(N)-Man[2](O6): 43% PheB24(N)-Man[1](O3): 25% TyrB16(OH)-Man[1](O4): 12% | ThrB27(N)-Man[2](O6): 44% PheB24(N)-Man[1](O3): 22% |

Table S3. The glycan-involved hydrogen bonds and their existence percentages of each glycoform.

| Atom type | Role | Description |
|-----------|----------|--|
| N | Donor | An sp ² nitrogen in amide group |
| NE2 | Donor | An epsilon nitrogen. |
| ОН | Donor | An alcohol oxygen in Tyr |
| OG1 | Donor | An alcohol oxygen in Thr |
| 02 | Acceptor | The oxygen atom connected to the second carbon atom of the sugar |
| O3 | Acceptor | The oxygen atom connected to the third carbon atom of the sugar |
| 04 | Acceptor | The oxygen atom connected to the fourth carbon atom of the sugar |
| O5 | Acceptor | The oxygen atom connected to the fifth carbon atom of the sugar |
| 06 | Acceptor | The oxygen atom connected to the sixth carbon atom of the sugar |
| O2N | Acceptor | The oxygen atom of the N-acetyl group |

Table S4. The atom types involved in the glycan-involved hydrogen bonds.

| | least occlusion | | | | | | | | most occlusion | | | |
|------|-----------------|---|---|--------------|----|----|------------|----|----------------|----|----|----|
| 4EYD | 2 | 3 | 7 | 8 | 4 | 11 | 12 | 6 | 9 | 10 | 5 | 13 |
| 4EY1 | 2 | 3 | 7 | 8 | 4 | 11 | 6 | 12 | 9 | 10 | 5 | 13 |
| 4EY9 | 2 | 3 | 7 | 8 | 11 | 4 | 6 | 12 | 5 | 9 | 10 | 13 |
| 313Z | 2 | 3 | 7 | 8 | 4 | 6 | 11 | 12 | 9 | 5 | 13 | 10 |
| 2MVC | 3 | 7 | 8 | 2 | 4 | 11 | 6 | 12 | 9 | 5 | 10 | 13 |
| | low batch | | | medium batch | | | high batch | | | | | |

Table S5. Glycoforms ordered from most to least proportion occlusion, based on proportion of simulation with measured occlusion.

2 Supplemental Figures

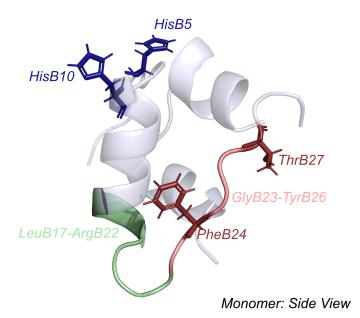


Figure S1. The histidine residues (HisB5, and HisB10, colored in blue) are shown with residues PheB24 and ThrB27 (colored red) which are important in enhancing proteolytic stability and residues LeuB17-ArgB22 (colored green) and GlyB23-TyrB26 (colored salmon), important in determining dimerization potential.

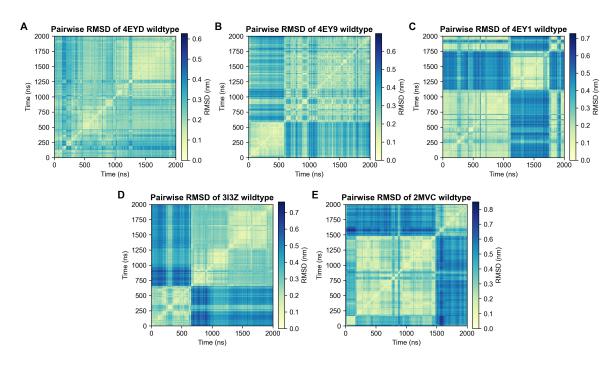


Figure S2. The pairwise RMSD of calculated from the 250ps-spaced MD trajectory of each wild-type model, including 4EYD (A), 4EY9 (B), 4EY1 (C), 3I3Z (D), and 2MVC (E). As shown in the figure, the major transitions occur around 500–1500 ns, we therefore concluded that at least 2000 ns was required to sample the configurational ensemble of insulin.

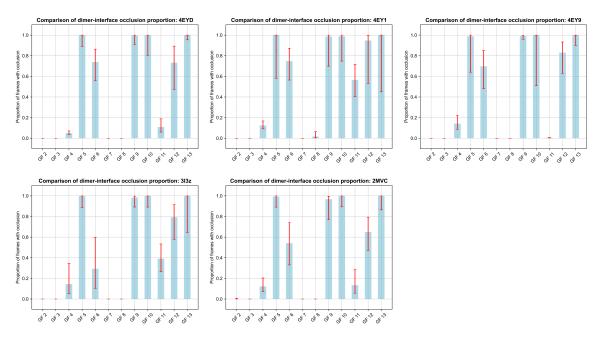


Figure S3. Proportion of frames with glycan-dimer occlusion for each glycoform. Red bars represent the asymmetric 95% Wilson score confidence interval.