Machine Learning 1

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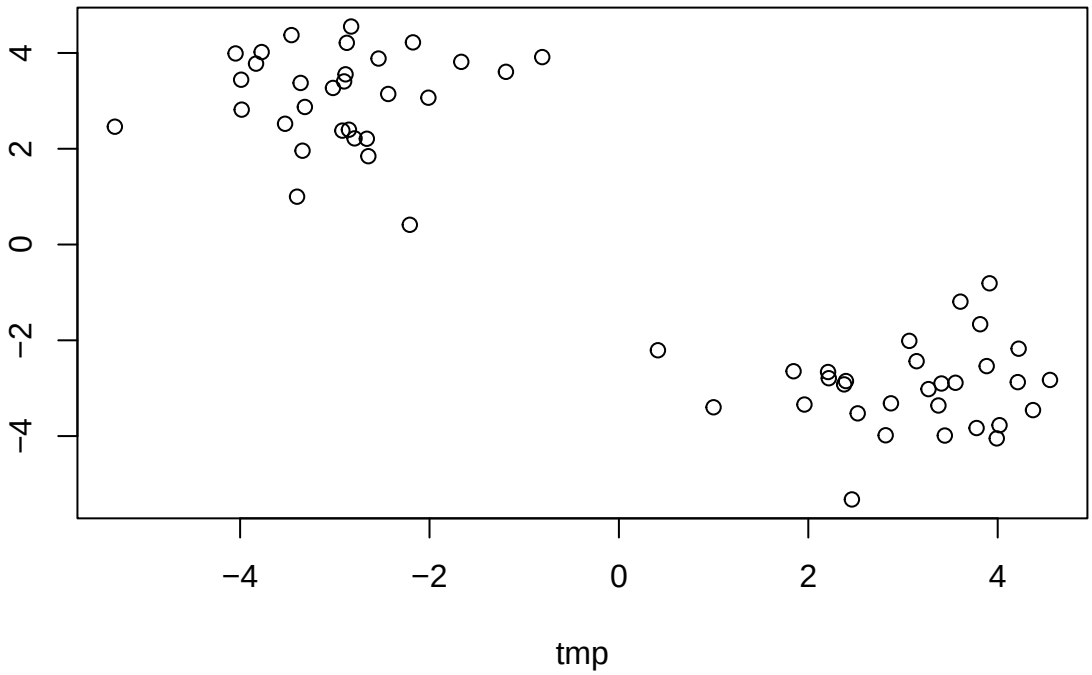
Clustering methods

Kmeansclustering in R is done with the `kmeans()` function.
Here we make up some data to test and learn with.

```
tmp <- c(rnorm(30, 3), rnorm(30, -3))
data <- cbind(tmp, rev(tmp))
plot(data)
```



Run `kmeans()` set k (# of centers) to 2 and nstart (iteration) to 20. The thing with `kmeans()` is that you need to tell it how many clusters you want.

```
km <- kmeans(data, centers=2, nstart=20)
km
```

```
## K-means clustering with 2 clusters of sizes 30, 30
##
## Cluster means:
```

```
##         tmp
## 1  3.090196 -2.958701
## 2 -2.958701  3.090196
##
## Clustering vector:
##  [1] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 2 2 2 2 2 2 2 2
## [39] 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
##
## Within cluster sum of squares by cluster:
## [1] 52.57245 52.57245
##  (between_SS / total_SS =  91.3 %)
##
## Available components:
##
## [1] "cluster"      "centers"      "totss"        "withinss"     "tot.withinss"
## [6] "betweenss"    "size"         "iter"         "ifault"
```

Q. How many points are in each cluster?

```
km$size
```

```
## [1] 30 30
```

Q. What 'component' of your result object details cluster alignment/membership?

```
km$cluster
```

```
##  [1] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 2 2 2 2 2 2 2 2
## [39] 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
```

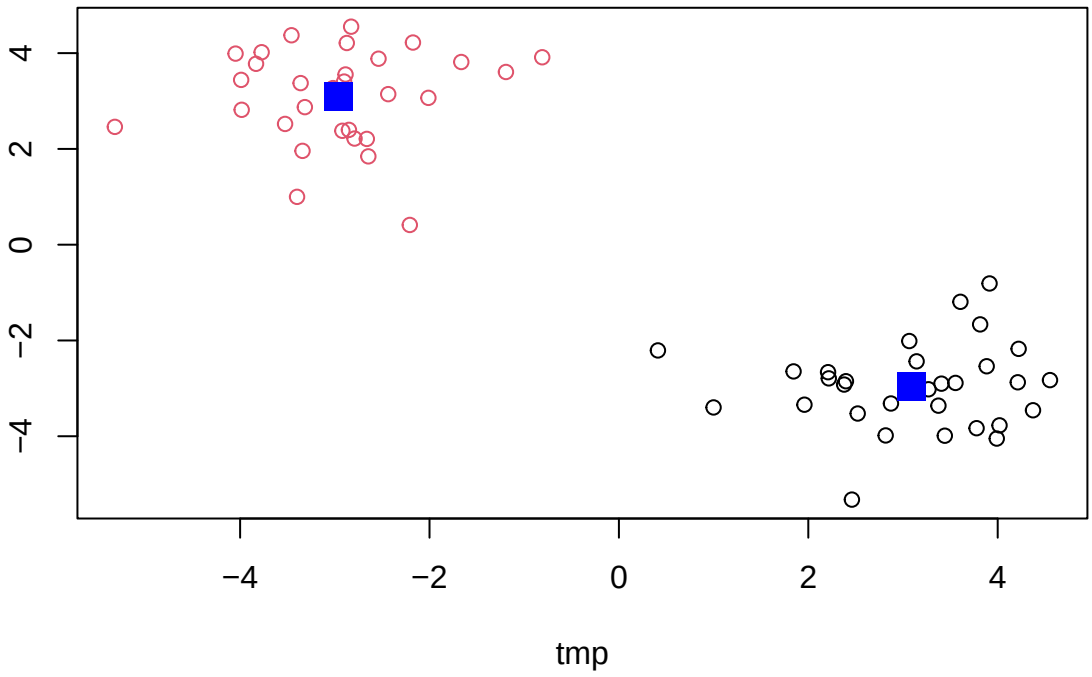
Q. What 'component' of your result object details cluster center?

```
km$centers
```

```
##         tmp
## 1  3.090196 -2.958701
## 2 -2.958701  3.090196
```

Q. Plot x colored by the kemans cluster assignment and add cluster centers as blue points.

```
plot(data, col=km$cluster)
points(km$centers, col="blue", pch=15, cex=2)
```



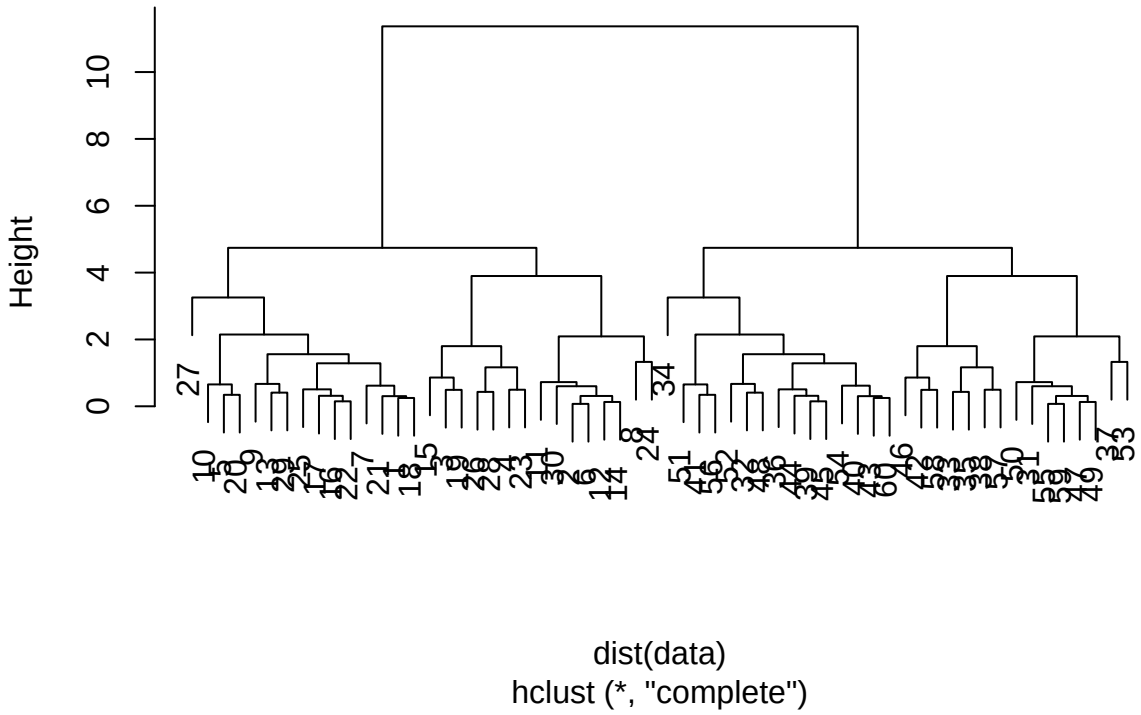
Hierarchical Clustering

We will use the `hclust()` function on the same data as before and see how this method works.

```
hc <- hclust(dist(data))
hc
```

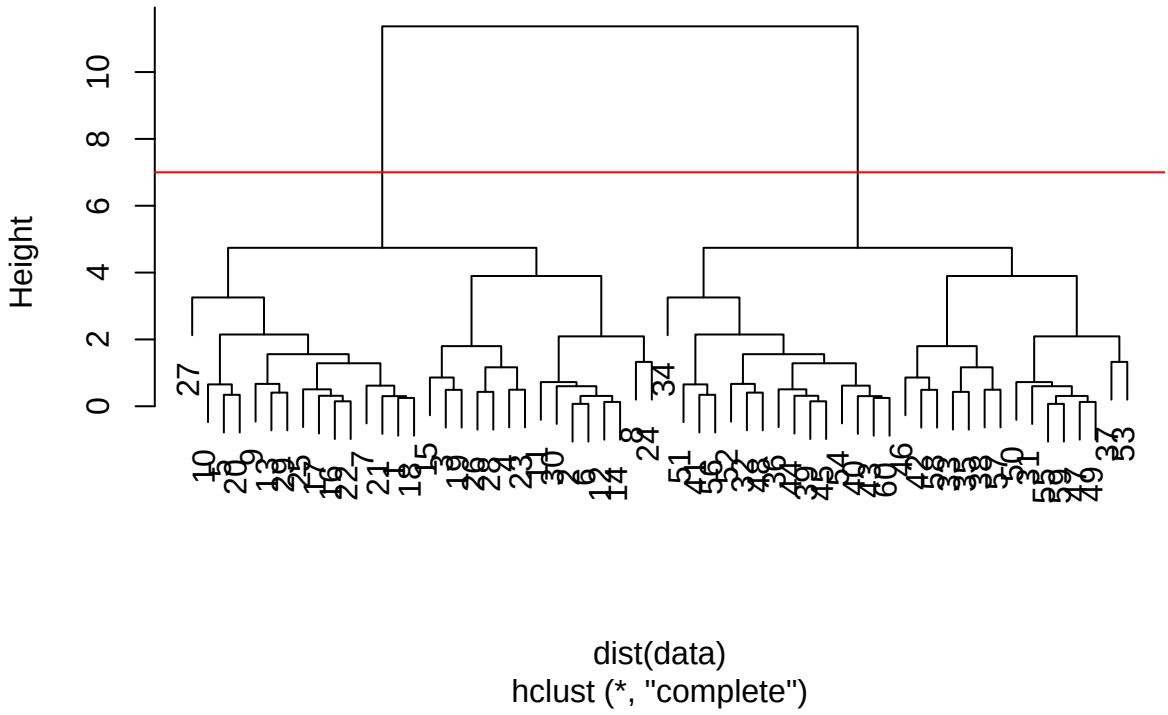
```
##
## Call:
## hclust(d = dist(data))
##
## Cluster method   : complete
## Distance         : euclidean
## Number of objects: 60
```

```
plot(hc)
```



To find our membership vector we need to "cut" the tree and for this we use the `cutree()` function and tell it the height to cut at.

```
plot(hc)
abline(h=7, col="red")
```

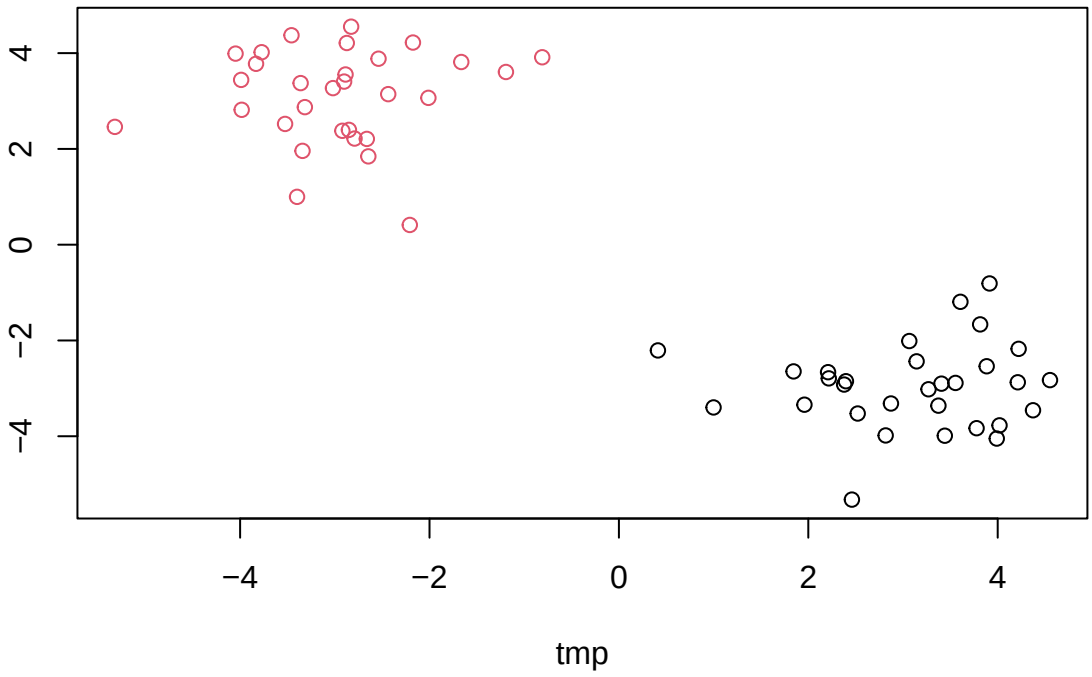



```
cutree(hc, h=7)
```

```
##  [1] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 2 2 2 2 2 2 2
## [39] 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
```

We can also use `cutree()` and state the number of k clusters we want:

```
grps <- cutree(hc, k=2)
plot(data, col=grps)
```



Principal Component Analysis (PCA)

Import the data from a CSV file

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url)
```

Q. How many rows and cols?

```
dim(x)
```

```
## [1] 17  5
```

Q. How do we inspect and clean up the data?

```
rownames(x) <- x[,1]
x <- x[, -1]
x
```

```
##                     England Wales Scotland N.Ireland
## Cheese                  105   103      103        66
## Carcass_meat            245   227      242       267
## Other_meat              685   803      750       586
## Fish                    147   160      122        93
## Fats_and_oils           193   235      184       209
## Sugars                  156   175      147       139
## Fresh_potatoes          720   874      566      1033
## Fresh_Veg               253   265      171       143
```

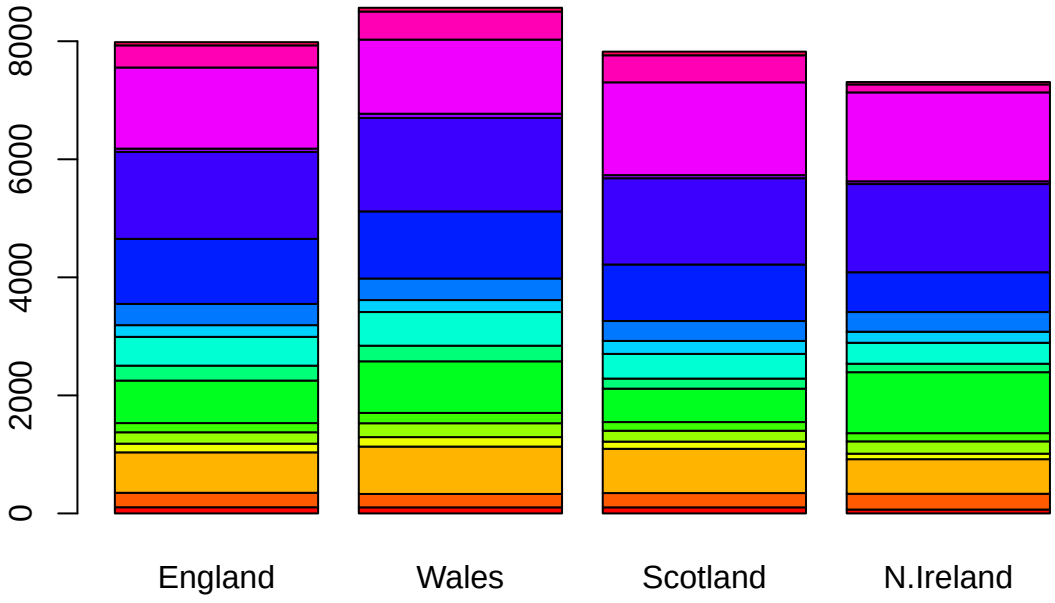
```
## Other_Veg               488   570      418       355
## Processed_potatoes      198   203      220       187
## Processed_Veg           360   365      337       334
## Fresh_fruit            1102  1137      957       674
## Cereals                1472  1582     1462      1494
## Beverages                57    73       53        47
## Soft_drinks            1374  1256     1572      1506
## Alcoholic_drinks        375   475      458       135
## Confectionery            54    64       62        41
```

Or we can read the data properly:

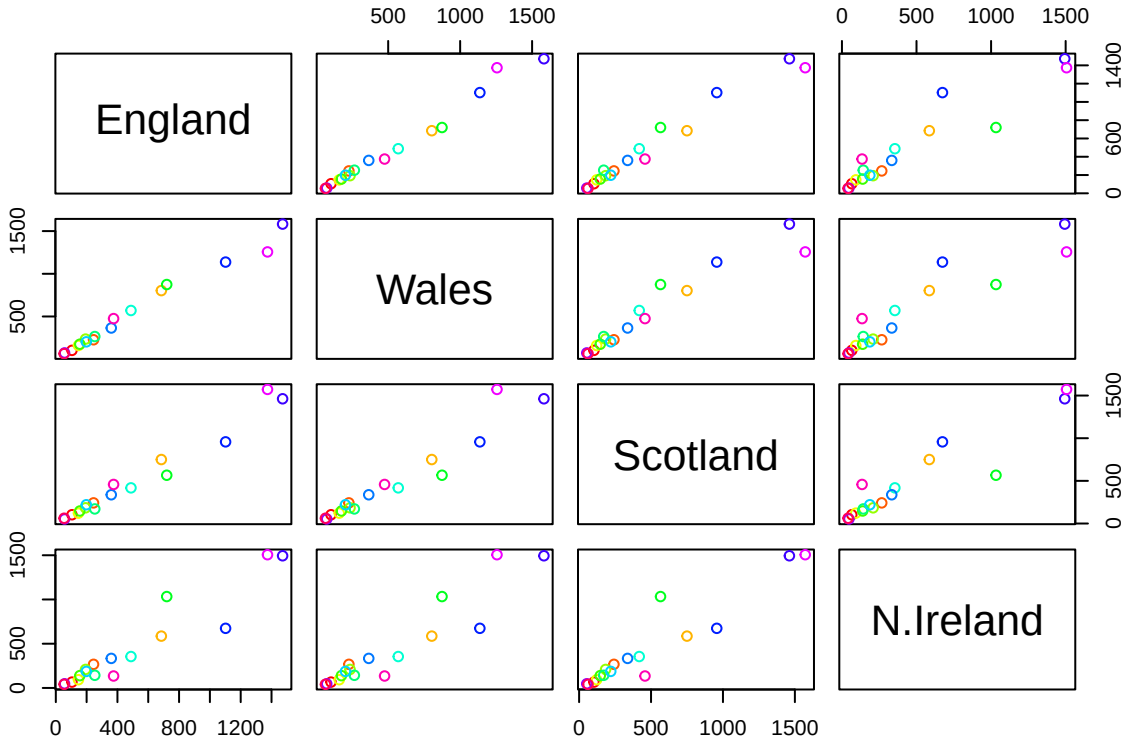
```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names=1)
x
```

```
##                     England Wales Scotland N.Ireland
## Cheese                  105   103      103        66
## Carcass_meat            245   227      242       267
## Other_meat              685   803      750       586
## Fish                    147   160      122        93
## Fats_and_oils           193   235      184       209
## Sugars                  156   175      147       139
## Fresh_potatoes          720   874      566      1033
## Fresh_Veg               253   265      171       143
## Other_Veg               488   570      418       355
## Processed_potatoes      198   203      220       187
## Processed_Veg           360   365      337       334
## Fresh_fruit            1102  1137      957       674
## Cereals                1472  1582     1462      1494
## Beverages                57    73       53        47
## Soft_drinks            1374  1256     1572      1506
## Alcoholic_drinks        375   475      458       135
## Confectionery            54    64       62        41
```

```
barplot(as.matrix(x), col=rainbow(17))
```




```
mycols <- rainbow(nrow(x))
pairs(x, col=mycols)
```



Anyways... None of these plots make sense.

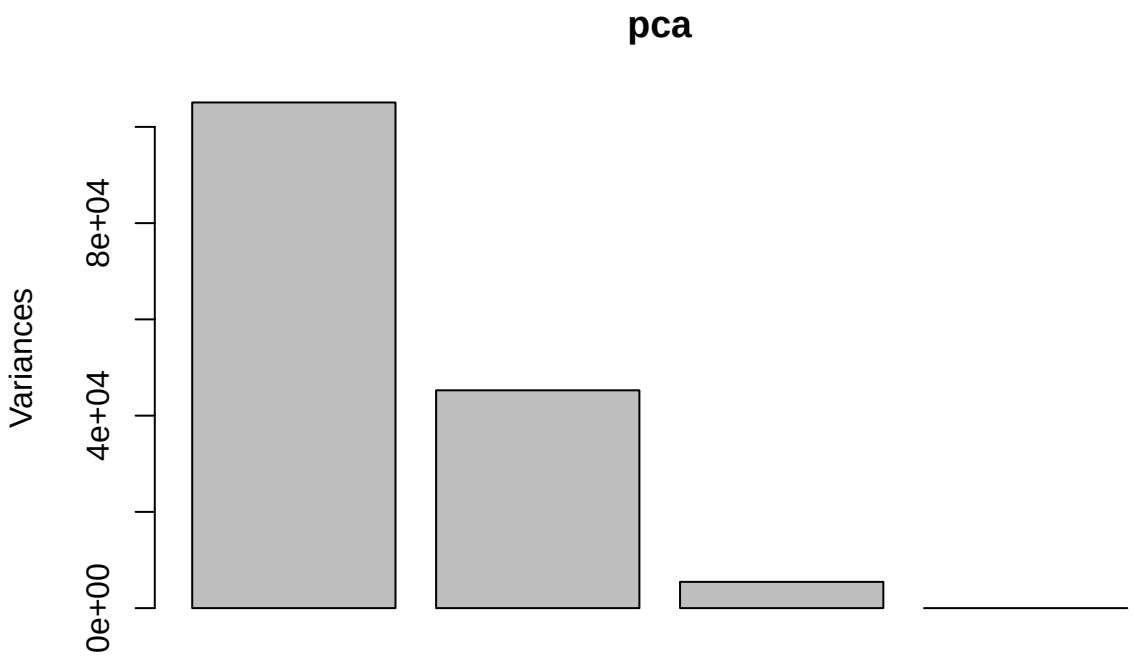
PCA to the rescue

Here we will use the base R function for PCA, which is called `prcomp()`. This function wants the transpose of our data.

```
pca <- prcomp(t(x))
summary(pca)
```

```
## Importance of components:
##                             PC1      PC2      PC3       PC4
## Standard deviation     324.1502 212.7478 73.87622 4.189e-14
## Proportion of Variance   0.6744   0.2905  0.03503 0.000e+00
## Cumulative Proportion    0.6744   0.9650  1.00000 1.000e+00
```

```
plot(pca)
```

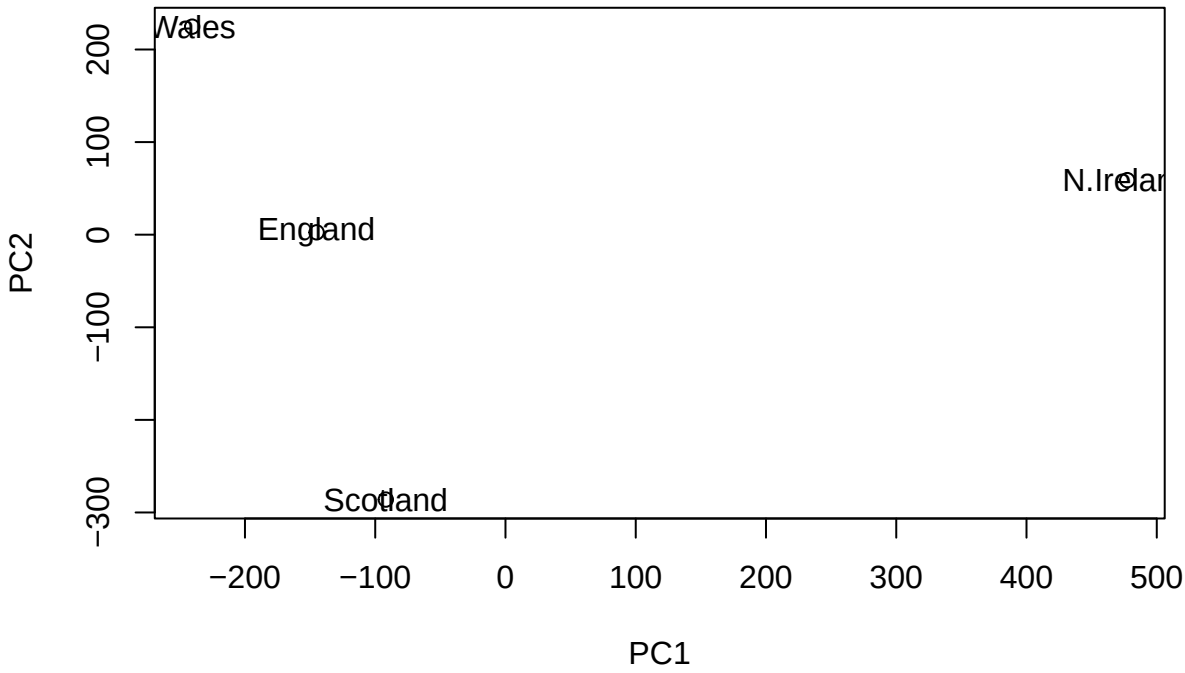


We want score plot(aka PCA plot). Basically, of PC1 vs PC2

```
attributes(pca)
```

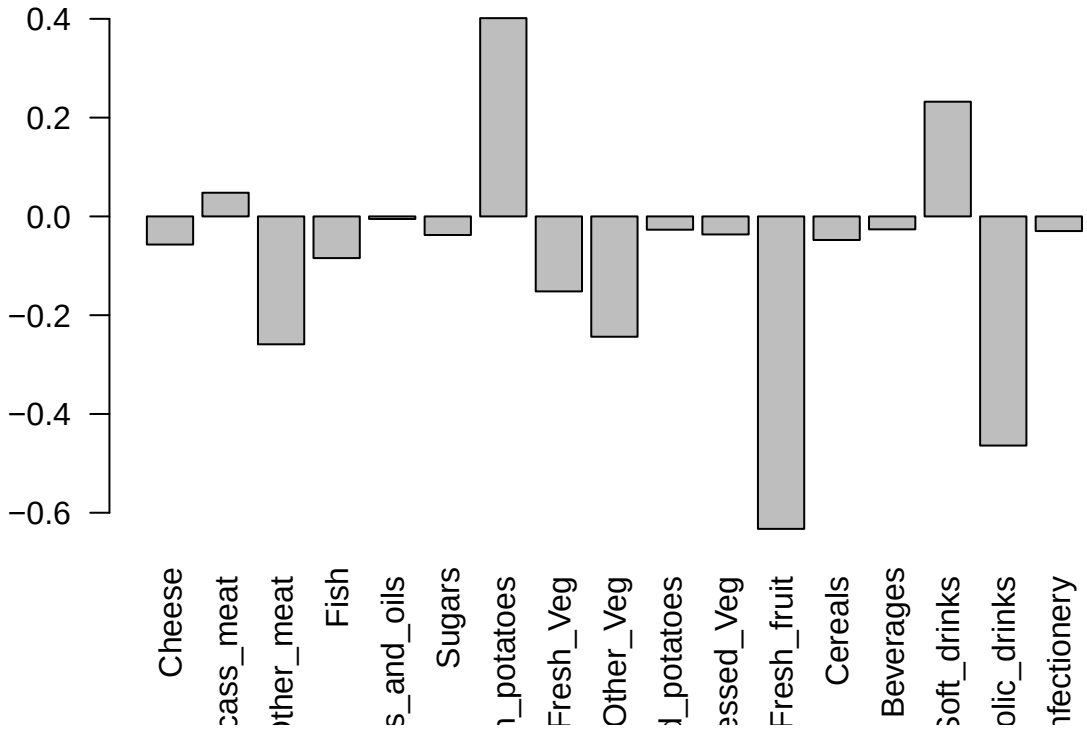
```
## $names
## [1] "sdev"     "rotation" "center"   "scale"    "x"
##
## $class
## [1] "prcomp"
```

```
plot(pca$x[, 1:2])
text(pca$x[, 1:2], labels=colnames(x))
```



We can also examine the PCA "loadings", which tell us how much the original variables contribute to each PC.

```
barplot(pca$rotation[,1], las=2)
```



PCA baby one more time!!!

```
url2 <- "https://tinyurl.com/expression-CSV"
rna.data <- read.csv(url2, row.names=1)
head(rna.data)
```

```
##        wt1 wt2  wt3  wt4 wt5 ko1 ko2 ko3 ko4 ko5
## gene1  439 458  408  429 420  90  88  86  90  93
## gene2  219 200  204  210 187 427 423 434 433 426
## gene3 1006 989 1030 1017 973 252 237 238 226 210
## gene4  783 792  829  856 760 849 856 835 885 894
## gene5  181 249  204  244 225 277 305 272 270 279
## gene6  460 502  491  491 493 612 594 577 618 638
```

Q. How many genes and samples?

```
nrow(rna.data)
```

```
## [1] 100
```

```
ncol(rna.data)
```

```
## [1] 10
```

```
colnames(rna.data)
```

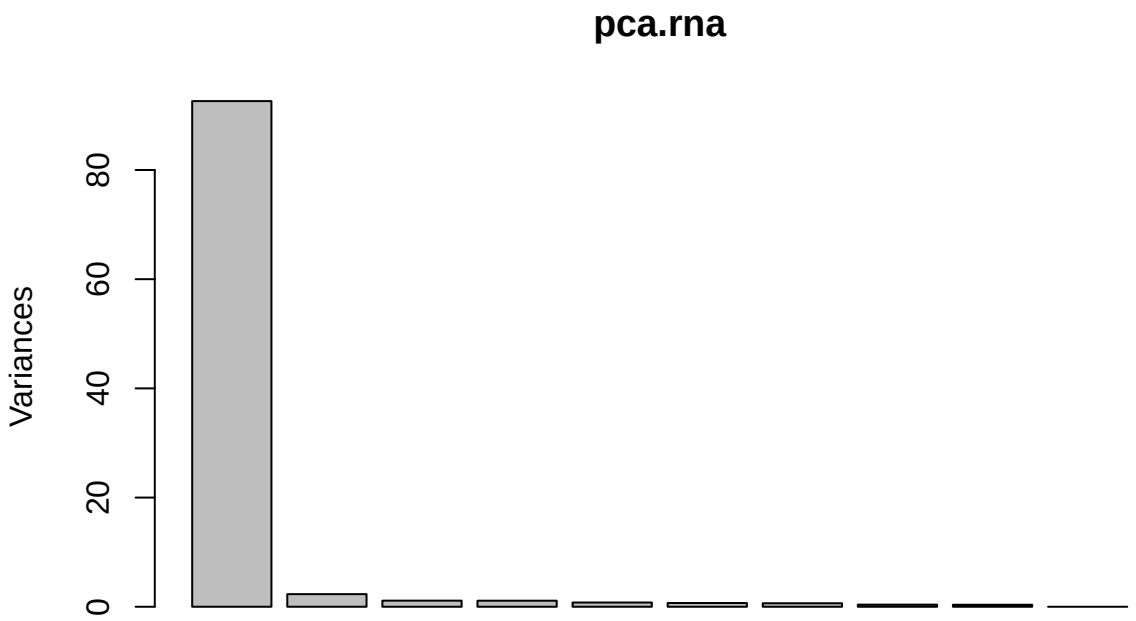
```
##  [1] "wt1" "wt2" "wt3" "wt4" "wt5" "ko1" "ko2" "ko3" "ko4" "ko5"
```

```
pca.rna <- prcomp(t(rna.data), scale=T)
summary(pca.rna)
```

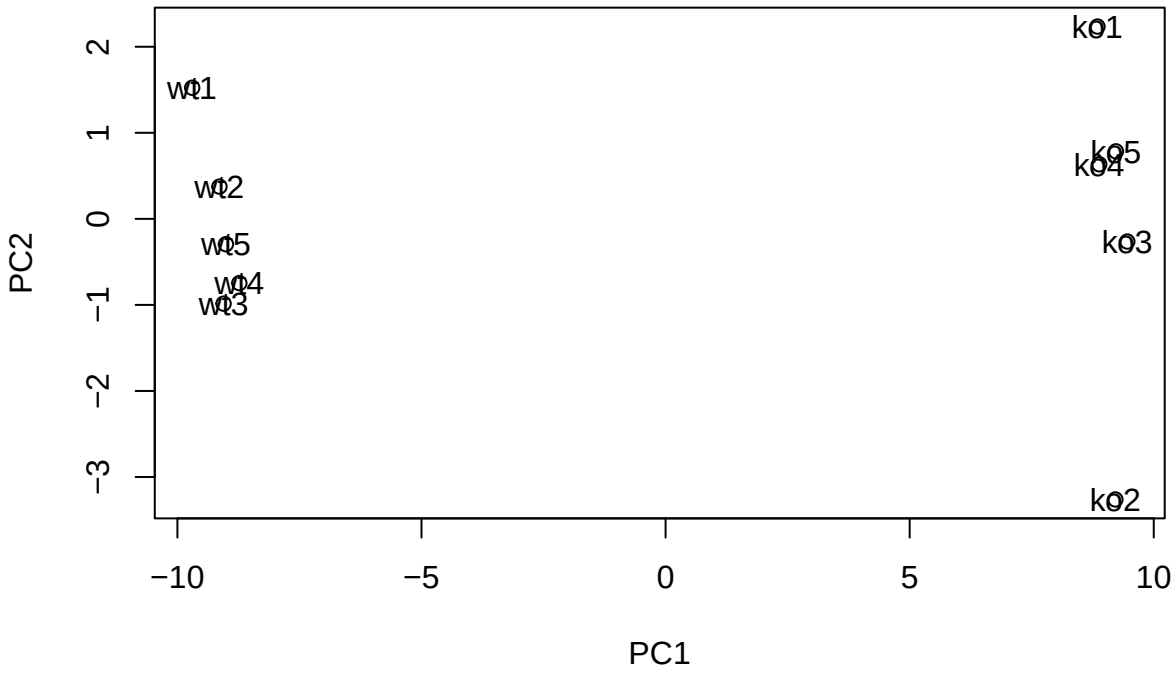
```
## Importance of components:
##                           PC1    PC2     PC3     PC4     PC5     PC6     PC7
## Standard deviation     9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
## Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
## Cumulative Proportion  0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
##                           PC8     PC9      PC10
## Standard deviation     0.62065 0.60342 3.348e-15
## Proportion of Variance 0.00385 0.00364 0.000e+00
## Cumulative Proportion  0.99636 1.00000 1.000e+00
```

Note here that nearly 93% of feature is captured by PC1.

```
plot(pca.rna)
```



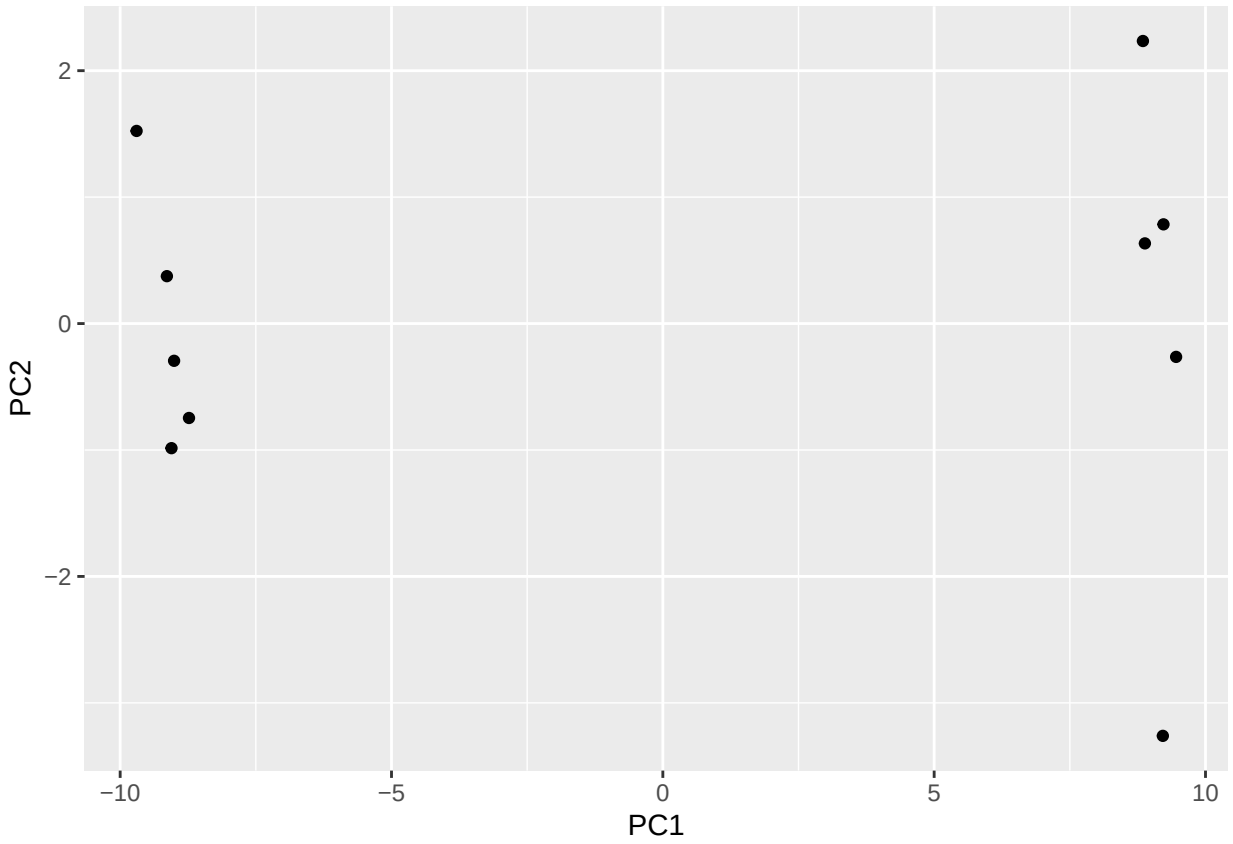

```
plot(pca.rna$x[, 1:2])
text(pca.rna$x[, 1:2], labels=colnames(rna.data))
```




```
library(ggplot2)

df <- as.data.frame(pca.rna$x)

# Our first basic plot
ggplot(df) +
  aes(PC1, PC2) +
  geom_point()
```

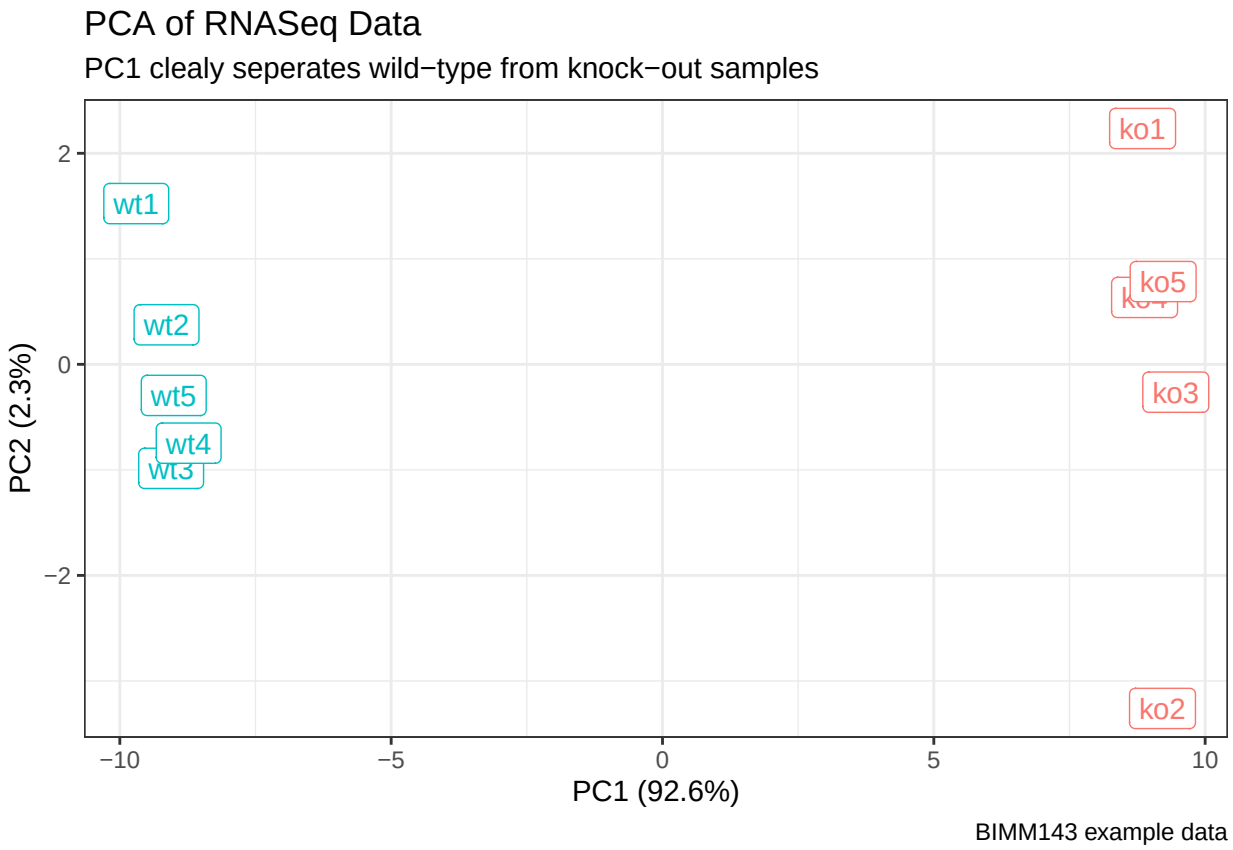



```
df$samples <- colnames(rna.data)
df$condition <- substr(colnames(rna.data),1,2)

pca.var <- pca.rna$sdev^2

pca.var.per <- round(pca.var/sum(pca.var)*100, 1)

p <- ggplot(df) +
        aes(PC1, PC2, label=samples, col=condition) +
        geom_label(show.legend = FALSE) +
labs(title="PCA of RNASeq Data",
       subtitle = "PC1 clealy seperates wild-type from knock-out samples",
            x=paste0("PC1 (", pca.var.per[1], "%)"),
       y=paste0("PC2 (", pca.var.per[2], "%)"),
       caption="BIMM143 example data") +
     theme_bw()
p
```


PCA of RNASeq Data
PC1 clealy seperates wild−type from knock−out samples
ko1
wt1
ko5
wt2
ko3
wt5
wt4
ko2
2
0
−2
PC2 (2.3%)
−10
−5
0
5
10
PC1 (92.6%)
BIMM143 example data