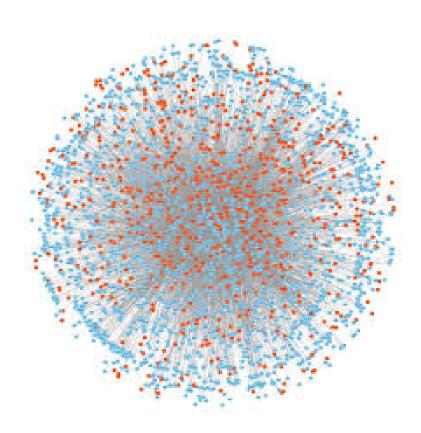
# **WS Networks and E. coli Transcription** Assignment 2



# **PS1: Watts-Strogatz network vs Random Network**

## **Methodology Overview**

#### 1. Network Generation

• Watts-Strogatz (WS):

3 networks created with N=100, K=5 (10 total neighbors symmetrically), and  $\beta$ = p  $\in$  {0.3, 0.7, 1.0}

- ullet Uses ring lattice rewiring with probability eta
- Edge count preserved at 500 during rewiring
- Erdős-Rényi (ER):

3 random networks were created with N=100 and exactly M=500 edges (same as WS networks)

#### 2. Key Metrics Calculated

- Degree distribution: Histogram of node degrees normalised to probabilities
- Average clustering coefficient: Mean of local clustering coefficients
- Characteristic path length: Average shortest path between all connected node pairs

#### 3. Implementation

- Built-in MATLAB functions for graph generation (WattsStrogatz, graph)
- Custom functions:
  - <u>clustering coefficients</u> for node-level clustering
  - <u>ErdosRenvi</u> for ER graph generation with exact edge control

# **Key Findings for Immediate Reporting**

#### 1. Degree Distributions

- WS networks retain near-regular structure at  $\beta$ =0.3 but develop Poisson-like distributions as  $\beta$  $\rightarrow$ 1
- ER networks show classic bell-shaped Poisson distribution

#### 2. Clustering Coefficients

- WS networks maintain *higher clustering* than ER at low  $\beta$  ( $\beta$ =0.3 preserves community structure)
- Clustering sharply decreases for WS as  $\beta \rightarrow 1$ , converging toward ER values

## 3. Path Length Dynamics

- WS networks achieve small-world properties (short paths, high clustering) at mid  $\beta$  (0.3-0.7)
- At  $\beta$ =1.0, WS path lengths match ER networks (random graph regime)

#### 4. Critical Transition

- At  $\beta$ =1.0, WS networks become structurally indistinguishable from ER graphs in:
  - Degree distribution shape
  - Global clustering magnitude
  - Average path length

# **Visualisation Strategy**

The code generates two key comparison plots along with all degree distributions of the graphs:

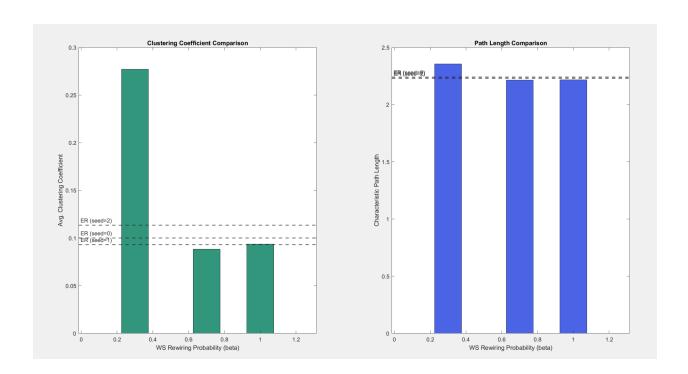
### 1. Clustering Coefficient vs β

- WS networks are shown as blue bars decreasing with β
- ER networks as horizontal red baselines

#### 2. Path Length vs β

- WS path lengths (green bars) decline rapidly with β
- ER values (red lines) mark the asymptotic lower-bound

These plots directly demonstrate the *small-world to random graph transition*. The comparison plot is shown below. All other plots of the degree distributions are attached in the zip file.



# E. Coli Transcription Network Centrality Analysis

#### **Network Construction**

#### **Data Processing**

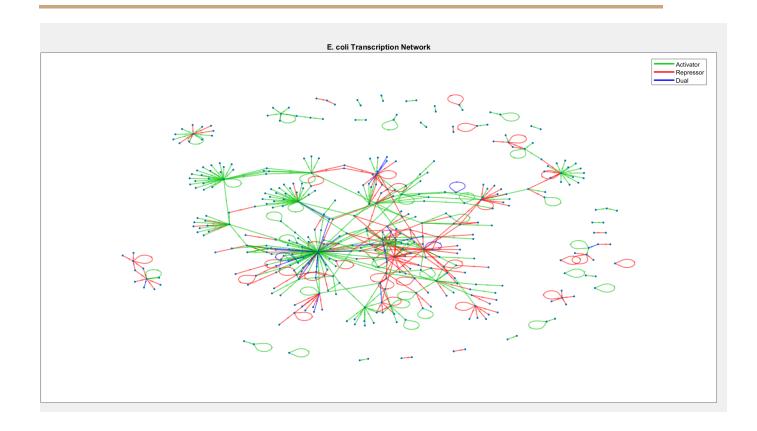
- Edge List Parsing:
  - Directed graph built from *E coli transcription network.txt* using MATLAB's *digraph* 
    - Nodes: 2 types (Transcription Factors [TFs] and Operons)
    - Edges: Directed interactions labelled as activator, repressor, or dual

#### <u>Visualisation Protocol</u>

- Layout: Force-directed algorithm ('force' with gravity)
- Edge Coloring:
  - Activator: Green ([0, 0.8, 0])
  - Repressor: Red ([1])
  - Dual: Blue ([1])

#### Key Implementation Note

Closeness centrality is calculated using outcloseness paths originating from TFs), aligning with transcriptional regulation dynamics.



## **A. Centrality Metrics**

Three measures were computed for all nodes:

- 1. <u>Degree Centrality</u>: Total edges (in + out)
- 2. <u>Closeness Centrality</u>: Reciprocal of average shortest path length from node to others
- 3. <u>Betweenness Centrality</u>: Fraction of shortest paths passing through a node

Filtering: Only TFs ranked (operons excluded).

#### **B. Subnetwork Analysis**

Activator Network: Activation Fraction Calculation:

$$Fraction_{activation} = \frac{TF's \ activator \ edges}{TF's \ total \ outgoing \ edges}$$

1. Top 5 Removed TFs:

CRP (16.1%), rpoE\_rseABC (7.5%), yhdG\_fis (7.5%), fnr (4.8%), nlpD\_rpoS (4.2%)

2. Edge Filtering: Retained only activator edges post-removal

Repressor/Dual Network: Repressor Fraction Calculation

$$Fraction_{repression} = \frac{TF's \; repressor \; edges}{TF's \; total \; outgoing \; edges}$$

1. Top 5 Removed TFs:

purR (7.9%), arcA (7.5%), lexA\_dinF (5.6%), fur (4.7%), himA (3.7%)

2. Edge Filtering: Retained repressor + dual edges post-removal

#### **Results**

Please find the detailed results in the zip folder in the file named **ecoli\_TF\_command\_terminal\_data**. Below are some key insights and a snapshot of the results.

#### A. Network Visualisation

- The force-directed layout reveals CRP as a central hub.
- Regulation type distribution:

Activator (43%) | Repressor (34%) | Dual (23%)

#### **B.** Centrality Rankings of the Original Network

Degree Centrality	Closeness Centrality	Betweenness Centrality
1. CRP (74)	1. CRP (0.00043481)	1. flhDC (49)
2. yhdG_fis (28)	2. rpoE_rseABC (0.00014753)	2. fliAZY (47)
3. rpoE_rseABC (26)	3. fnr (0.00014743)	3. rpoH (40)
4. fnr (24)	4. yhdG_fis (0.000146)	4. hns (22)
5. himA (23)	5. arcA (0.00011256)	5. ompR_envZ (18)

#### **C. Activator Network Post Removal**

1. <u>Degree Distribution</u>:

• Range: 0–12 (vs. original 0–74)

• Most frequent degree: 1 (41% nodes)

2. Top Degree Centrality:

hns (10), rpoH (9), cpxAR (9)

3. Closeness Centrality Range: 0-0.00022883

#### D. Repressor/Dual Network Post Removal

- 1. <u>Degree Distribution</u>:
  - Bimodal peaks at 1 (44 nodes) and 3 (18 nodes)
- 2. Top Degree Centrality:

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fliAZY (12), fnr (12), marRAB (7)
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3. Closeness Centrality Range: 0-0.00047959

## **Key Biological Insights**

- 1. CRP as 'Master Regulator':
  - The highest degree (74) and closeness centrality (0.00043481) confirm the role of the master regulator.
  - The absence of betweenness centrality in the top 5 suggests a specialised regulatory role rather than structural bridging.
  - CRP Regulator Modulates Multidrug Resistance of Escherichia coli by Repressing the mdtEF Multidrug Efflux Genes. This paper confirms the conclusion drawn about the role of CRP from network analysis alone.
- 2. Regulatory Bottlenecks:
  - <u>flhDC</u> (betweenness=49) and fliAZY (47)

#### Network Resilience:

1) Activator removal reduces max degree by 84% (74→12).

#### **Biological Implication:**

Activator-rich networks rely on a few "master regulators" – their removal collapses coordinated gene expression.

2) Repressor removal increases betweenness spread (marRAB=7 shows emerging control points), i.e. control in the original network is more centralised (CRP/flhDC dominate). Still, post-repressor removal, there is decentralised control (marRAB gains influence).

#### **Biological Implication**:

Repression-dominated networks develop backup control points under stress.