

Class 11

```
data <- read.csv('Data Export Summary.csv', row.names = 1)
data
```

```
##              X.ray   NMR   EM Multiple.methods Neutron Other  Total
## Protein (only)    142419 11807 6038             177     70    32 160543
## Protein/Oligosaccharide 8426   31  991             5      0     0  9453
## Protein/NA         7498   274 2000             3      0     0  9775
## Nucleic acid (only)   2368  1378  60             8      2     1  3817
## Other              149    31   3             0      0     0   183
## Oligosaccharide (only)  11     6   0             1      0     4    22
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
xray_percent <- 100 * sum(data$X.ray) / sum(data$Total)
xray_percent
```

```
## [1] 87.52836
```

```
em_perc <- 100 * sum(data$EM) / sum(data$Total)
em_perc
```

```
## [1] 4.94687
```

for every column

```
colSums(data) / sum(data$Total) * 100
```

```
##              X.ray              NMR              EM Multiple.methods
##      87.52836071      7.35991033      4.94686958      0.10555353
##              Neutron              Other              Total
##      0.03917451      0.02013134      100.00000000
```

Q2: What proportion of structures in the PDB are protein?

```
prot_perc <- 100 * sum(data$Total[1]) / sum(data$Total)
prot_perc
```

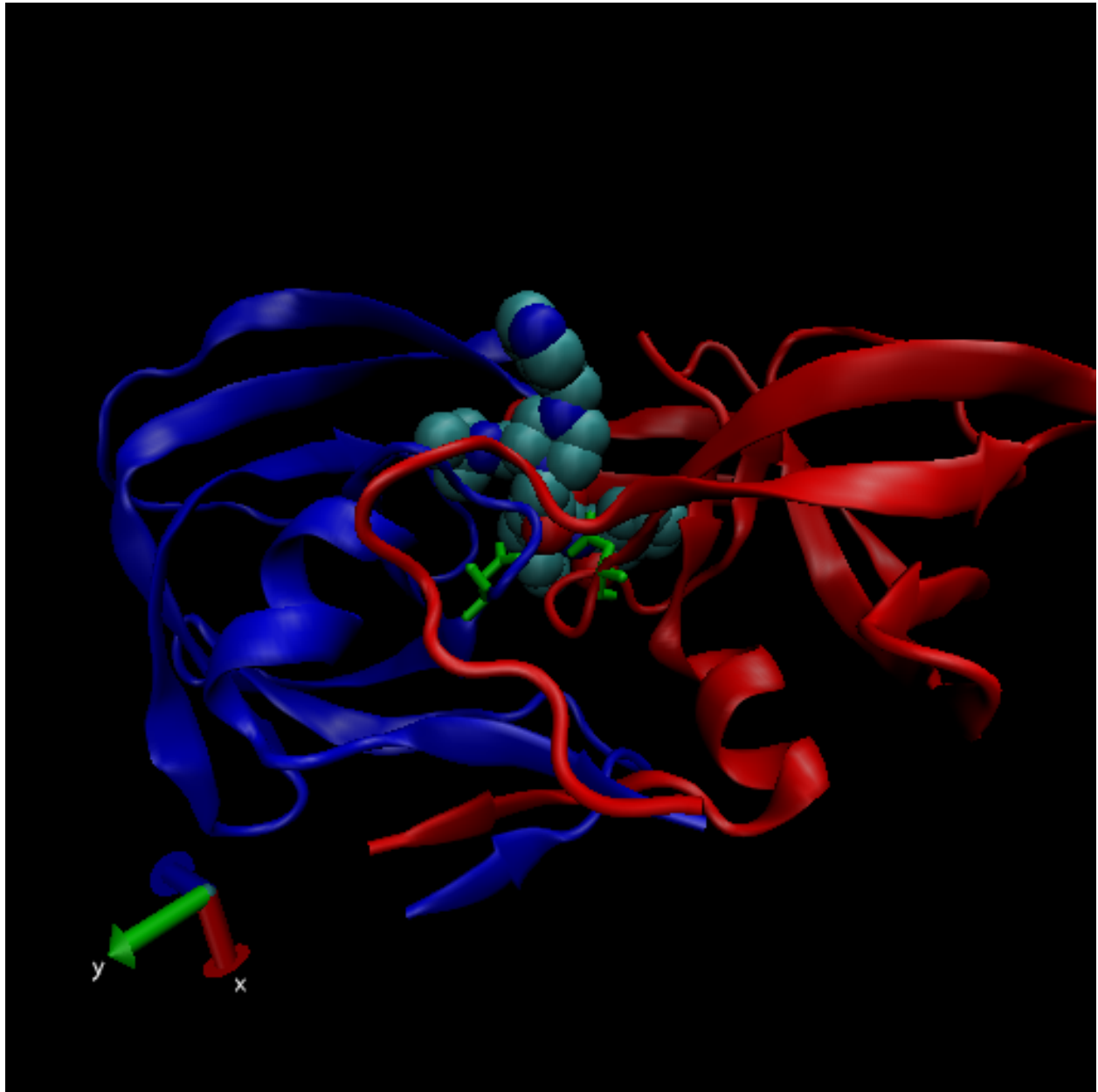
```
## [1] 87.3499
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

23409

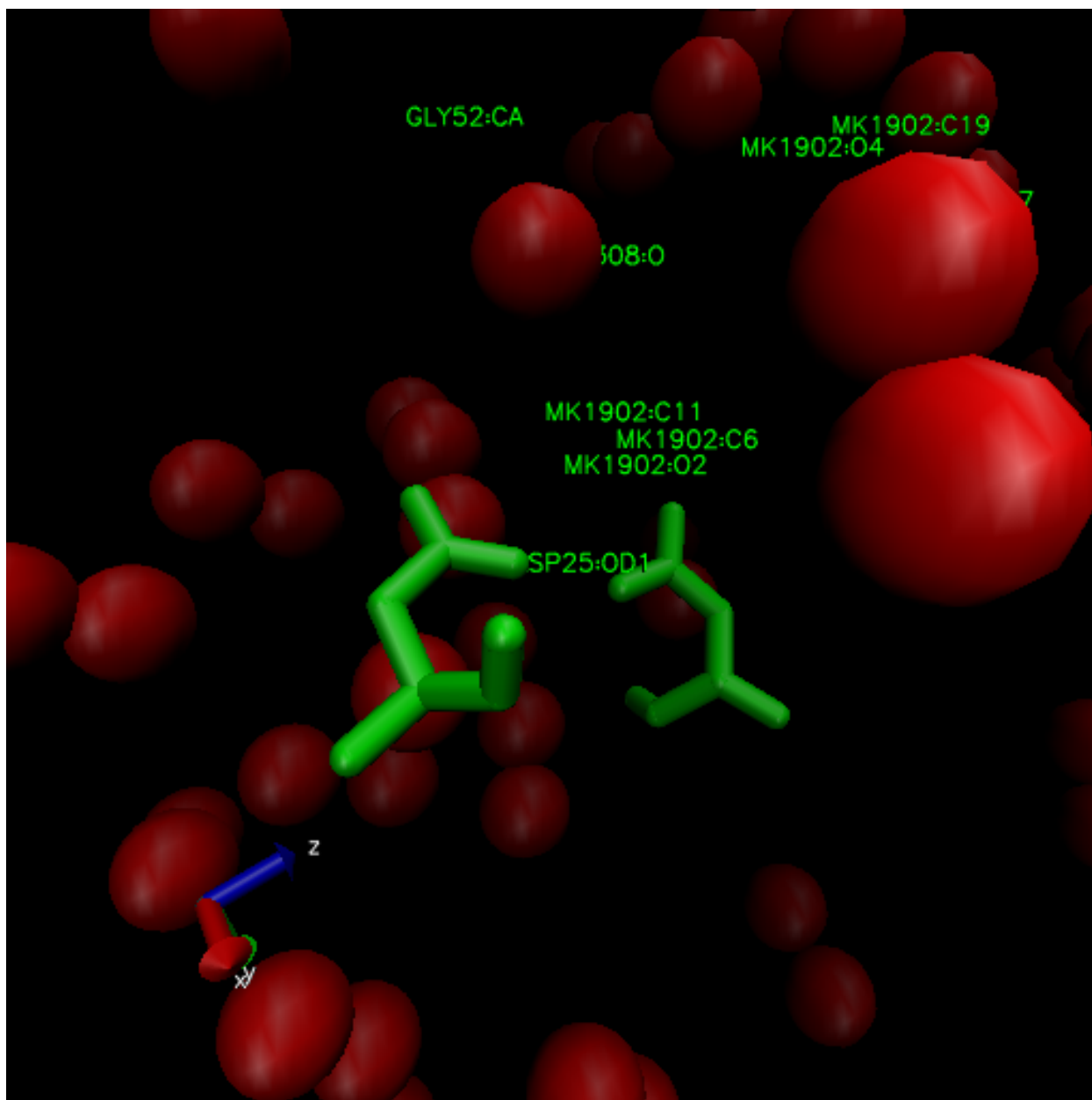
Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The resolution is higher than the size of hydrogen



Q5: There is a conserved water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have (see note below)?

HOH 308



```
library(bio3d)
pdb <- read.pdb('1hel')
```

```
## Note: Accessing on-line PDB file
```

```
pdb
```

```
##
## Call: read.pdb(file = "1hel")
##
## Total Models#: 1
## Total Atoms#: 1186, XYZs#: 3558 Chains#: 1 (values: A)
##
```

```
##      Protein Atoms#: 1001 (residues/Calpha atoms#: 129)
##      Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
##
##      Non-protein/nucleic Atoms#: 185 (residues: 185)
##      Non-protein/nucleic resid values: [ HOH (185) ]
##
##      Protein sequence:
##      KVFGRCELAAAMKRHGLDNYRGYSLGNWVCAAKFESNFNTQATNRNTDGSTDYGILQINS
##      RWWCNDGRTPGSRNLCNIPCSALLSSDITASVNC AKKIVSDGNGMNAWVAWRNRCKGTDV
##      QAWIRGCRL
##
## + attr: atom, xyz, seqres, helix, sheet,
##      calpha, remark, call
```

```
head(pdb$atom)
```

```
##      type eleno elety alt resid chain resno insert      x      y      z o      b
## 1 ATOM      1      N <NA>  LYS      A      1 <NA>  3.294 10.164 10.266 1 11.18
## 2 ATOM      2      CA <NA>  LYS      A      1 <NA>  2.388 10.533  9.168 1  9.68
## 3 ATOM      3      C <NA>  LYS      A      1 <NA>  2.438 12.049  8.889 1 14.00
## 4 ATOM      4      O <NA>  LYS      A      1 <NA>  2.406 12.898  9.815 1 14.00
## 5 ATOM      5      CB <NA>  LYS      A      1 <NA>  0.949 10.101  9.559 1 13.29
## 6 ATOM      6      CG <NA>  LYS      A      1 <NA> -0.050 10.621  8.573 1 13.52
##      segid elesy charge
## 1 <NA>      N <NA>
## 2 <NA>      C <NA>
## 3 <NA>      C <NA>
## 4 <NA>      O <NA>
## 5 <NA>      C <NA>
## 6 <NA>      C <NA>
```

Do a normal mode analysis (NMA), a prediction of the conformational variability and intrinsic dynamics of this protein

```
pdb <- read.pdb('1hel')
```

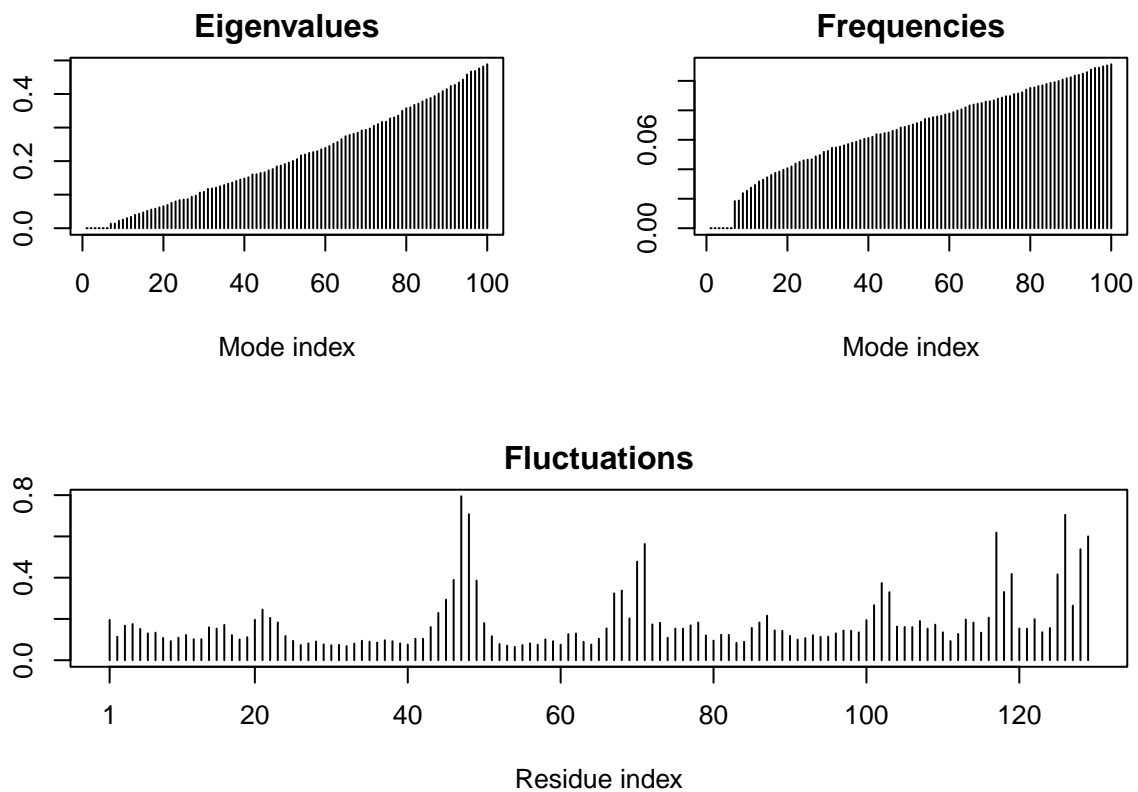
```
##      Note: Accessing on-line PDB file
```

```
## Warning in get.pdb(file, path = tempdir(), verbose = FALSE): /var/folders/5b/
## nx310gbn5pd_zqfnjpsgkt200000gn/T//Rtmpza3CdY/1hel.pdb exists. Skipping download
```

```
m <- nma(pdb)
```

```
## Building Hessian...      Done in 0.049 seconds.
## Diagonalizing Hessian... Done in 0.151 seconds.
```

```
plot(m)
```



Make a little movie (trajectory) for viewing in VMD.

```
mktrj(m, file = 'nma.pdb')
```