# Multiple Sequence Alignment Analysis Report: Saccharomyces cerevisiae: Cytochrome C subunit 1 (cox1)

1) Choose a gene from your favorite bacterium. What gene did you choose and from what bacterium? Remember to format the species name correctly (Genus name capitalized, specific epithet lower case, entire name italicized).

Gene: cox1 - cytochrome c oxidase sub unit 1

Organism: Saccharomyces cerevisiae S288C

2) Paste the gene sequence.

>NC\_027264.1:13017-21174 cox1 [organism=Saccharomyces cerevisiae] [GeneID=24573120] [chromosome=MT]

ATGGTACAAAGATGATTATATTCAACAAATGCAAAAGATATTGCAGTATTATATTTTATGTTAGCTA
TTT

TTAGTGGTATGGCAGGAACAGCAATGTCTTTAATCATTAGATTAGAATTAGCTGCACCTGGTTC ACAATA

TTTACATGGAAATTCACAATTATATAATGTTTTAGTAGTTGGTCATGCTGTATTAATGATTTTCTGTGCG

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ATG

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ATTAATACTAGAATTTTAAAATTAATGTCAGATATTAGAATGTTATTAATTGCTTATAAAAAATTAAA A

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AA

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ACATCTGGAGGATTTAGACCTTTAAGTGTTGGAAATCCTAGAGAAAAATTGTACAAGAAAGTA TGAGAA

TAATATTAGAAATTATCTATAATAATAATTATTCTCTTATTATTCTCATGGATTTAGACCTAACTTATCTT
G

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AT

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AC

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TAAATAAACATGGCTATTGTTCTCATGGTATTTTAGGAAAACCCAGAGGGGTTGGAAGATTAATT CATGA

AGAAATGAAAACCATTTTAATGCATTACTTAGCTGTTGGTAGAGGTATTATAAACTATTATAGATTA

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CAA

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TTC

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CATT

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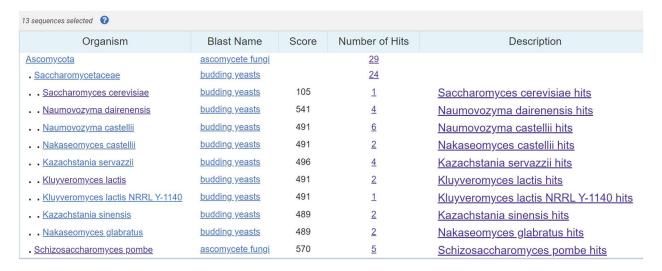
TCTAATCTTATCTTTAATTTAAATACAGTTAAATCTTCATCTATCGAATTCTTATTAACTTCTCCACC AG

#### **CTGTACACTCATTTAATACACCAGCTGTACAATCTTAA**

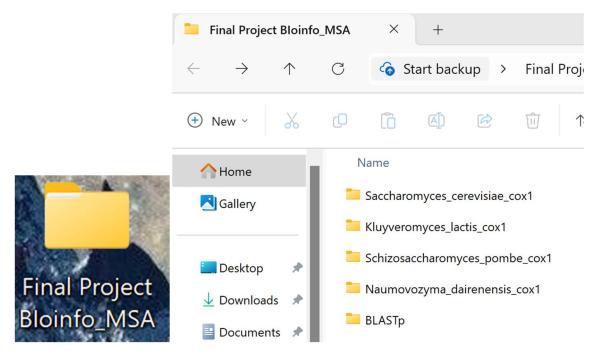
3) Using BLAST, find the 3 closest relatives from different genera (each homolog should be from a different genus as in the example AND a different genus from one another, meaning you should have 4 genera total) for your gene sequence. List the three species. Which type of BLAST search did you do for this step (e.g. BLASTn, BLASTx, etc)?

#### **Nucleotide BLASTx:**

- Saccharomyces cerevisiae S288C
- 1) Kluyveromyces lactis
- 2) Schizosaccharomyces pombe
- 3) Naumovozyma dairenensis



4) Create a file like the demo file for your gene and its closest homologs. Add a screenshot of the file. If you are having trouble creating a .fas file, you can download my demo file, replace my sequences with yours, then change the name.



5) Find the amino acid sequence for your gene of interest (the same gene you selected for number 1). Paste the amino acid sequence.

>YP\_009144706.1 cox1 [organism=Saccharomyces cerevisiae] [GeneID=24573120]

MVQRWLYSTNAKDIAVLYFMLAIFSGMAGTAMSLIIRLELAAPGSQYLHGNSQLYNVLVVGHAVLMI
FFL

VMPALIGGFGKYLLPLMIGATDTAFPRINNIAFWVLPMGLVCLVTSTLVESGAGTGWTVYPPLSSIQA HS

GPSVDLAIFALHLTSISSLLGAINFIVTTLNMRTNGMTMHKLPLFVWSIFITAFLLLLSLPVLSAGITML LLDRNFNTSFFEVAGGGDPILYEHLFWFFGHPEVYILIIPGFGIISHVVSTYSKKPVFGEISMVYAMASI GLLGFLVWSHHMYIVGLDADTRAYFTSATMIIAIPTGIKIFSWLATIYGGSIRLATPMLYAIAFLFLFTM GGLTGVALANASLDVAFHDTYYVVGHFHYVLSMGAIFSLFAGYYYWSPQILGLNYNEKLAQIQFWLI FIG

ANVIFFPMHFLGINGMPRRIPDYPDAFAGWNYVASIGSFIATLSLFLFIYILYDQLVNGLNNKVNNKS VI

YNKAPDFVESNTIFNLNTVKSSSIEFLLTSPPAVHSFNTPAVQS

6) Using BLAST, find the 3 closest relatives from different genera (each homolog should be from a different genus as in the example) for your amino acid sequence. List the three species. Which type of BLAST search did you do for this step (e.g. BLASTn, BLASTx, etc)?

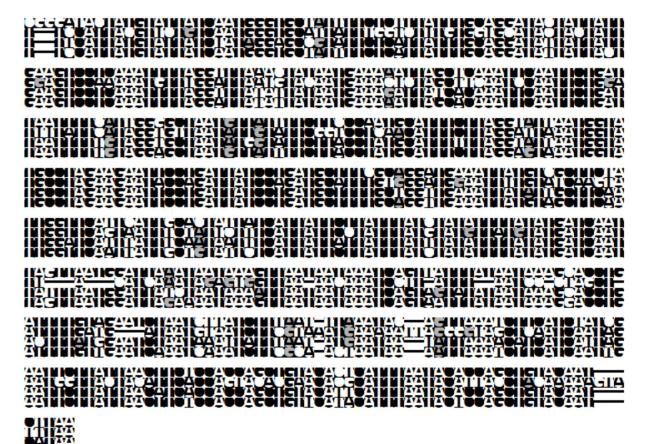
**BLASTp** 

- Saccharomyces cerevisiae S288C
- 1) Naumovozyma dairenensis
- 2) Nakaseomyces delphensis
- 3) Torulaspora quercuum

Organism	Blast Name	Score	Number of Hits	Description
Saccharomycetaceae	budding yeasts		31	
Saccharomyces	budding yeasts		8	
Saccharomyces cerevisiae	budding yeasts	1066	1	Saccharomyces cerevisiae hits
<ul> <li>Saccharomyces cerevisiae YJM993</li> </ul>	budding yeasts	1066	1	Saccharomyces cerevisiae YJM993 hits
Saccharomyces cerevisiae YJM978	budding yeasts	1066	1	Saccharomyces cerevisiae YJM978 hits
Saccharomyces cerevisiae YJM981	budding yeasts	1066	1	Saccharomyces cerevisiae YJM981 hits
Saccharomyces cerevisiae YJM984	budding yeasts	1066	1	Saccharomyces cerevisiae YJM984 hits
Saccharomyces cerevisiae YJM987	budding yeasts	1066	1	Saccharomyces cerevisiae YJM987 hits
Saccharomyces cerevisiae YJM990	budding yeasts	1066	1	Saccharomyces cerevisiae YJM990 hits
Saccharomyces cerevisiae YJM996	budding yeasts	1066	1	Saccharomyces cerevisiae YJM996 hits
<ul> <li>Naumovozyma dairenensis</li> </ul>	budding yeasts	993	2	Naumovozyma dairenensis hits
<ul> <li>Nakaseomyces delphensis</li> </ul>	budding yeasts	992	2	Nakaseomyces delphensis hits
Nakaseomyces nivariensis	budding yeasts	988	2	Nakaseomyces nivariensis hits
Nakaseomyces bracarensis	budding yeasts	988	2	Nakaseomyces bracarensis hits
Torulaspora quercuum	budding yeasts	977	4	Torulaspora quercuum hits
Torulaspora globosa	budding yeasts	976	2	Torulaspora globosa hits
<ul> <li>Nakaseomyces glabratus</li> </ul>	budding yeasts	974	2	Nakaseomyces glabratus hits
. Torulaspora microellipsoides	budding yeasts	974	2	Torulaspora microellipsoides hits
Kazachstania sinensis	budding yeasts	974	2	Kazachstania sinensis hits
<ul> <li>Kluyveromyces lactis</li> </ul>	budding yeasts	971	2	Kluyveromyces lactis hits
. Kluyveromyces lactis NRRL Y-1140	budding yeasts	971	1	Kluyveromyces lactis NRRL Y-1140 hits

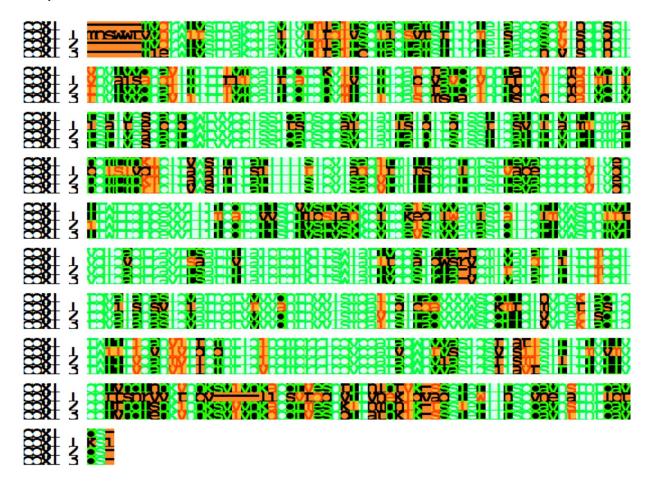
7) Perform a multiple sequence alignment of your gene AND protein sequences using T-COFFEE. Paste a screenshot of the results here. In a couple of sentences, use your own words to describe what a multiple sequencing alignment is and how it is

generated.	cox1 cox1_1 cox1_2 cox1_3	TTAAACTATAATGAAAAATTAGCTCAAATTCAATTCTGATTAATTTTCAT TTAATGTACAATGAAACTCTAGCTTCAATCCAATTCTGATTATTTCAT TTATATTATA
	cox1 cox1_1 cox1_2 cox1_3	TGGGGCTAATGTTATTTTCTTCCCAATGCATTTCTTAGGTATTAATGGTA AGGTGTTAATATTGTATTCGGTCCTCAACATTTCTTAGGATTAAATGGTA AGGTGCTAATATGGTATTCTTACCTATGCATTTTTTAGGTATTAATGGTA AGGAGCTAATGTTATTTCTTACCTATGCATTTCTTAGGAGTAAATGGTA ** * **** * * *** * * ***** * * *******
	cox1 cox1_1 cox1_2 cox1_3	TGCCTAGAAGAATTCCTGATTATCCTGATGCTTTCGCAGGATGAAATTAT TGCCTAGAAGAATACCAGATTATCCAGATGCATTTTGTGGGATGGAATTTT TGCCTAGAAGAATTCCTGATTATCCTGATGCTTTTGCTGGTTGAAATTAT TGCCTAGAAGAATTCCTGATTATCCTGATGCTTTTGCAGGTTGAAATTAT **************************
	cox1 cox1_1 cox1_2 cox1_3	GTCGCTTCTATTGGTTCATTCATTGCACTATTATCATTATTCTTATTTAT
	cox1 cox1_1 cox1_2 cox1_3	CTATATTTTATATGATCAATTAGTTAATGGATTAAATAATAAAGTTAATA GTATGTTATGTATG



A multiple sequence alignment is a method used to align two or more biological sequences, such as DNA, RNA, or proteins, to identify regions of similarity and differences. It is generated by algorithms that compare the sequences and insert gaps to maximize the alignment, highlighting conserved regions and differences among the sequences.

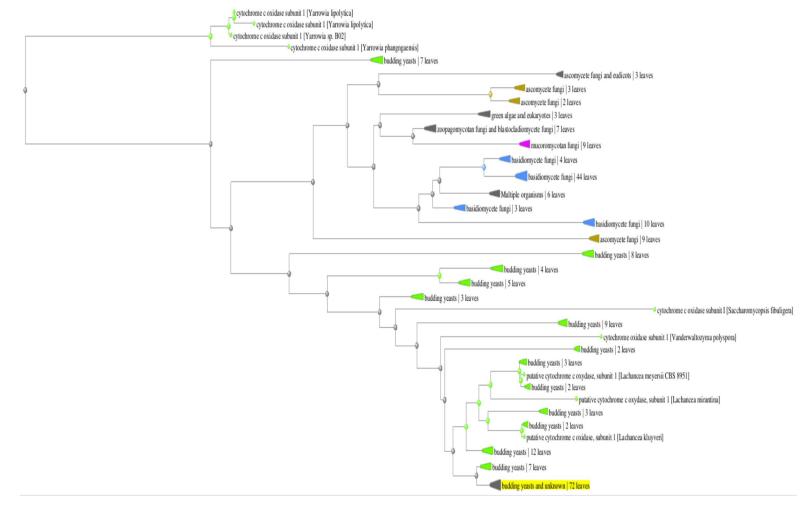
8) Generate a boxshade of your protein alignment (using whatever settings you would like!) and include a screenshot of the results here.



9) Using MEGA, please create a pairwise distance of your protein sequences. Paste a screenshot of the results here. Based on the results of your pairwise alignment, predict which two species are most closely related and which two species are most distantly related. How do you know?

In the analysis of phylogenetic trees, *Yarrowia lipolytica* and *Yarrowia phangngensis* are depicted as the most closely related species, sharing a branch early in the tree which points to a recent common ancestor and minimal evolutionary divergence. Conversely, *Saccharomyces cerevisiae* (S288C) and species from the basidiomycete fungi group, like

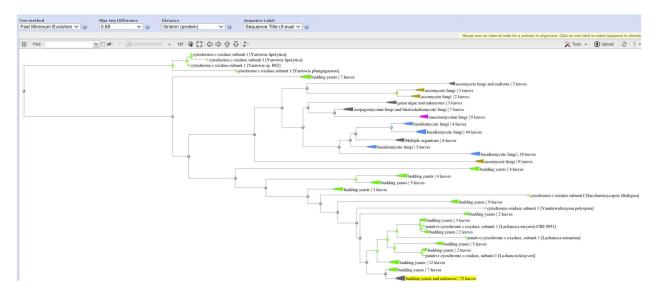
Tremella mesenterica, are found to be the most distantly related, positioned in entirely separate clades marked by significant distances and divergent evolutionary paths. In a separate query analysis, *Naumovozyma dairenensis* and *Nakaseomyces delphensis*, both members of the Saccharomycetaceae family, appear to be the closest genetically, potentially due to shared sequences within their family, whereas *Nakaseomyces delphensis* and *Torulaspora quercuum* represent the most distant relation within the same family, attributed to their belonging to different genera and evolutionary trajectories. These assessments rely on the assumption that genetic proximity and evolutionary closeness correlate with shorter connecting branches in phylogenetic trees, which can be further substantiated by detailed sequence alignments.



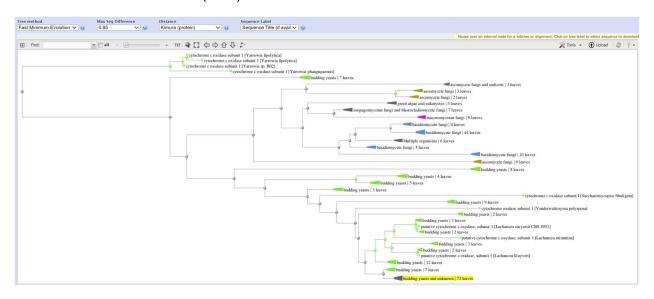
10) Generate two different trees from your protein alignment. For each tree, describe what type of tree you did, include a one sentence explanation of what this type of tree is, and how many bootstraps you did. You should also include a brief explanation (in your own words) of what bootstrapping is. Do your trees support your hypothesis about relatedness from your pairwise distances?

1. **Fast Minimum Evolution:** is a method for constructing phylogenetic trees by finding the topology that minimizes the total branch length, using bootstrapping to assess the stability of the tree topology by repeatedly resampling the data and constructing trees for each sample.

## Fast Minimum Evolution: Grishin (0.85)

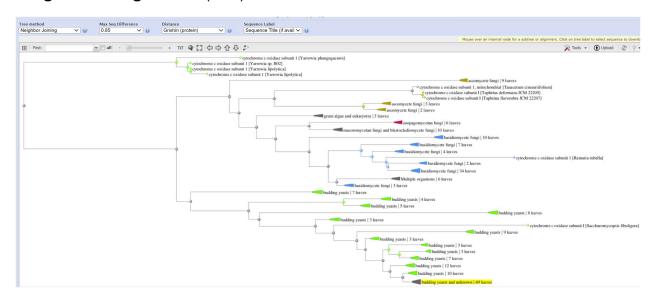


#### Fast Minimum Evolution: Kimura (0.85)

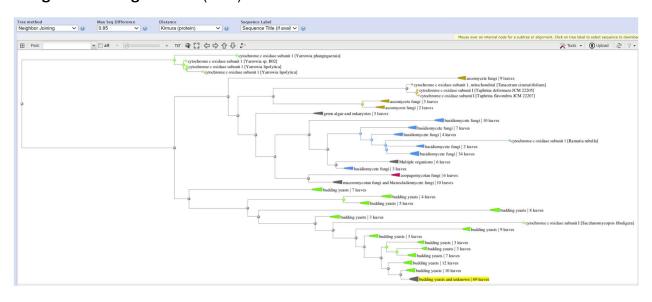


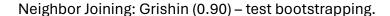
2. **Neighbor-Joining:** is a distance-based method for constructing phylogenetic trees that iteratively joins pairs of nodes with the smallest distance, using bootstrapping to estimate the confidence of the tree's branches by assessing the frequency of groupings across resampled datasets.

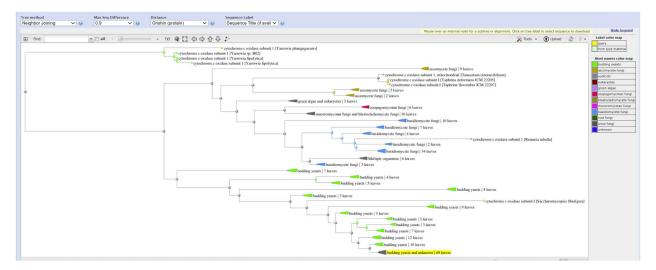
# Neighbor-Joining: Grishin (0.85)



## **Neighbor-Joining: Kimura** (0.85)







**Bootstrapping:** is a statistical method to estimate the reliability of phylogenetic trees. It involves repeatedly resampling the data and reconstructing the phylogenetic tree for each sample. The bootstrap values show how often a particular branch or grouping of species appears in the resampled trees, providing a measure of its support or reliability.

Yes, these data support the hypothesis that Naumovozyma dairenensis, Nakaseomyces delphensis, Torulaspora quercuum are closely related to Saccharomyces cerevisiae based on the analysis of these MSA and phylogenetic trees. This phylogenetic analysis, showed that Naumovozyma dairenensis and Nakaseomyces delphensis are both part of the Saccharomycetaceae family, suggesting a close phylogenetic relationship due to shared genetic markers typical for this group. These two species, belonging to different genera within the same family, show significant genetic similarities that suggest a common evolutionary path. In contrast, Torulaspora quercuum, also a member of the Saccharomycetaceae, is placed in a separate branch due to distinct genetic characteristics that differentiate it from Naumovozyma and Nakaseomyces. Torulaspora quercuum shares an evolutionary pathway that coincides with Candida and other fungi. This separation in the phylogenetic tree focuses on the genetic and evolutionary differences that define genus-level distinctions within Saccharomycetaceae.

### 11) Are these trees rooted or unrooted? How do you know?

**Rooted,** because rooted trees have a designated common ancestor, while unrooted trees do not.