

Meher Shrishti Nigam
20BRS1193

EDA LAB – 10
24 / 3 / 23

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20BRS1193

EDA Lab 10

options(prompt="MEHERSHRISHTI>", continue = " ")

options(prompt=">", continue = " ")

EDA-LAB-EXPERIMENT-10 (Date-25/3/2023)

library(dplyr)

library(ggplot2)

Q1) You have been given a gene expression data set (Ch10Ex11.csv) that consists of 40 tissue samples with

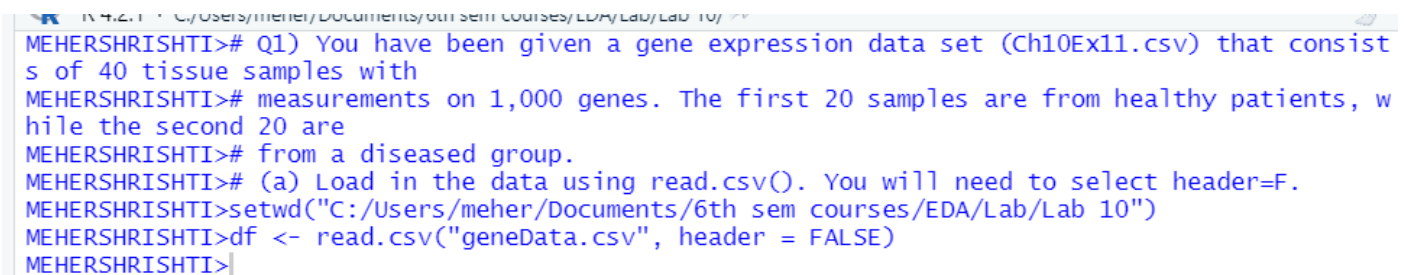
measurements on 1,000 genes. The first 20 samples are from healthy patients, while the second 20 are

from a diseased group.

(a) Load in the data using read.csv(). You will need to select header=F.

setwd("C:/Users/meher/Documents/6th sem courses/EDA/Lab/Lab 10")

df <- read.csv("geneData.csv", header = FALSE)

A screenshot of an R console window. The title bar shows the file path "C:/Users/meher/Documents/6th sem courses/EDA/Lab/Lab 10/". The console contains the following text: "MEHERSHRISHTI># Q1) You have been given a gene expression data set (Ch10Ex11.csv) that consists of 40 tissue samples with", "MEHERSHRISHTI># measurements on 1,000 genes. The first 20 samples are from healthy patients, while the second 20 are", "MEHERSHRISHTI># from a diseased group.", "MEHERSHRISHTI># (a) Load in the data using read.csv(). You will need to select header=F.", "MEHERSHRISHTI>setwd('C:/Users/meher/Documents/6th sem courses/EDA/Lab/Lab 10')", "MEHERSHRISHTI>df <- read.csv('geneData.csv', header = FALSE)", and "MEHERSHRISHTI>".

```
MEHERSHRISHTI># Q1) You have been given a gene expression data set (Ch10Ex11.csv) that consists of 40 tissue samples with
MEHERSHRISHTI># measurements on 1,000 genes. The first 20 samples are from healthy patients, while the second 20 are
MEHERSHRISHTI># from a diseased group.
MEHERSHRISHTI># (a) Load in the data using read.csv(). You will need to select header=F.
MEHERSHRISHTI>setwd("C:/Users/meher/Documents/6th sem courses/EDA/Lab/Lab 10")
MEHERSHRISHTI>df <- read.csv("geneData.csv", header = FALSE)
MEHERSHRISHTI>
```

(b) Apply hierarchical clustering to the samples using correlationbased distance, and plot the dendrogram.

Do the genes separate the samples into the two groups? Do your results depend on the type of linkage

used?

```
dists <- dist(cor(df))
```

```
methods <- c('centroid', 'average', 'single', 'complete')
```

```
par(mfrow = c(2,2))
```

```
for (method in methods) {
```

```
  clusts <- hclust(dists, method = method)
```

```
  plot(clusts,
```

```
    col = "#487AA1", col.main = "#45ADA8",
```

```
    col.lab = "#7C8071", col.axis = "#F38630",
```

```
    lwd = 3, lty = 1,
```

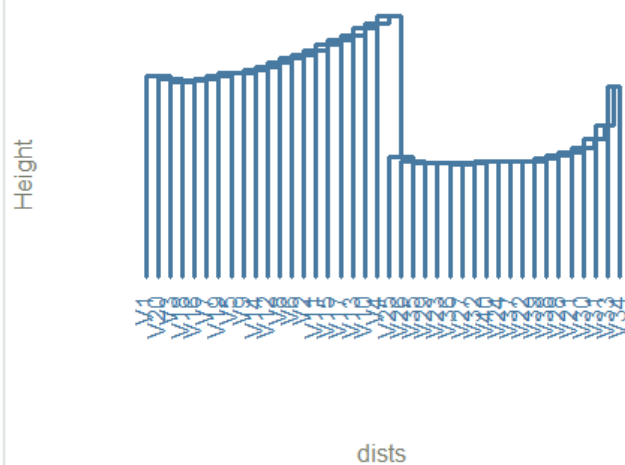
```
    sub = "", hang = -1,
```

```
    axes = FALSE,
```

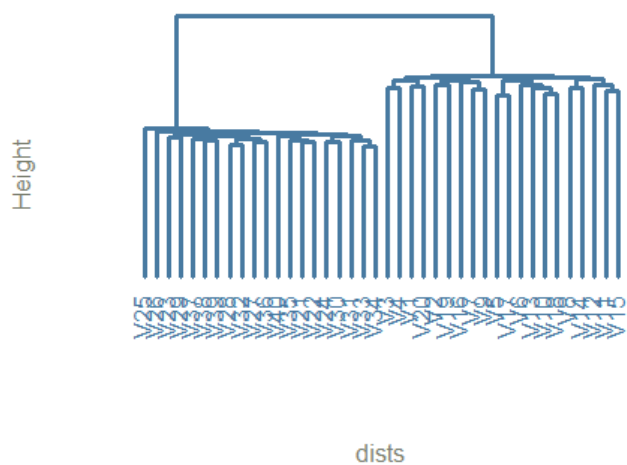
```
    main = paste0('Cluster Dendrogram using ', method, ' metric'))
```

```
}
```

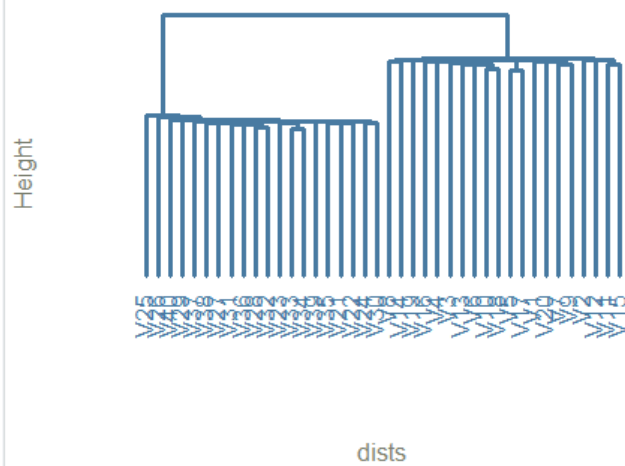
Cluster Dendrogram using centroid metric



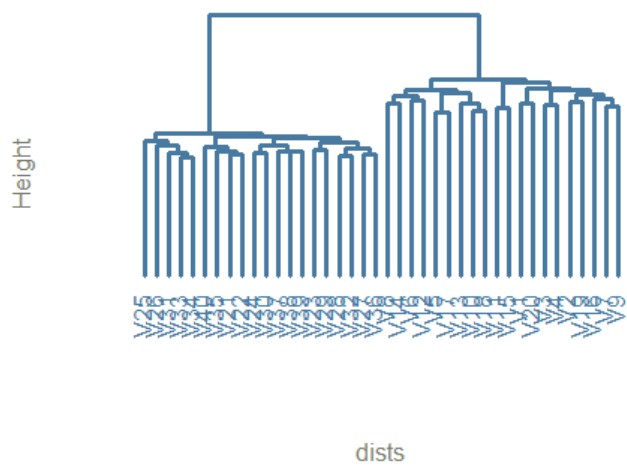
Cluster Dendrogram using average metric



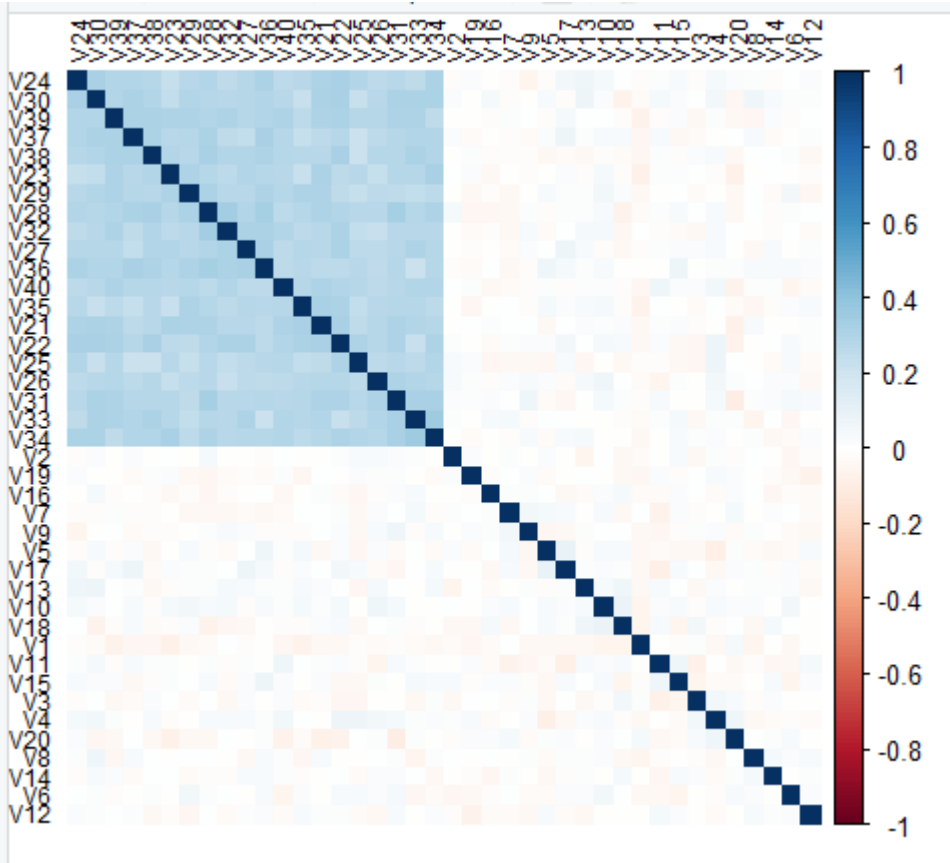
Cluster Dendrogram using single metric



Cluster Dendrogram using complete metric



```
require(corrplot)
corrplot(cor(df), method = 'color',
         order = 'hclust', hclust.method = 'complete',
         tl.col = 'black', tl.cex = 0.7)
```



```
patient_groups <- cutree(clusts, k = 2)
```

```
patient_groups
```

```
MEHERSHRISHTI># (b) Apply hierarchical clustering to the samples using correlationbased distance, and plot the dendrogram.
MEHERSHRISHTI># Do the genes separate the samples into the two groups? Do your results depend on the type of linkage
MEHERSHRISHTI># used?
MEHERSHRISHTI>dists <- dist(cor(df))
MEHERSHRISHTI>methods <- c('centroid', 'average', 'single', 'complete')
MEHERSHRISHTI>par(mfrow = c(2,2))
MEHERSHRISHTI>for (method in methods) {
  clusts <- hclust(dists, method = method)

  plot(clusts,
       col = "#487AA1", col.main = "#45ADA8",
       col.lab = "#7C8071", col.axis = "#F38630",
       lwd = 3, lty = 1,
       sub = "", hang = -1,
       axes = FALSE,
       main = paste0('Cluster Dendrogram using ', method, ' metric'))
}
MEHERSHRISHTI>require(corrplot)
MEHERSHRISHTI>corrplot(cor(df), method = 'color',
  order = 'hclust', hclust.method = 'complete',
  tl.col = 'black', tl.cex = 0.7)
MEHERSHRISHTI>patient_groups <- cutree(clusts, k = 2)
MEHERSHRISHTI>patient_groups
```

	V1	V2	V3	V4	V5	V6	V7	V8	V9	V10	V11	V12	V13	V14	V15	V16	V17	V18	V19	V20	V21	V22
V1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	2	2
V23	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2

```
MEHERSHRISHTI>
```

The basic dendrogram hints at the variables being separated into two groups. There seems to be strong evidence for the variables belonging to two distinct groups.

(c) Your collaborator wants to know which genes differ the most across the two groups. Suggest a way to

answer this question, and apply it here.

```
set.seed(702)
```

```
gene <- read.csv("geneData.csv", header = FALSE)
```

```
pr.gene <- prcomp(t(gene), scale=T)
```

```
plot(pr.gene)
```

```
summary(pr.gene)
```

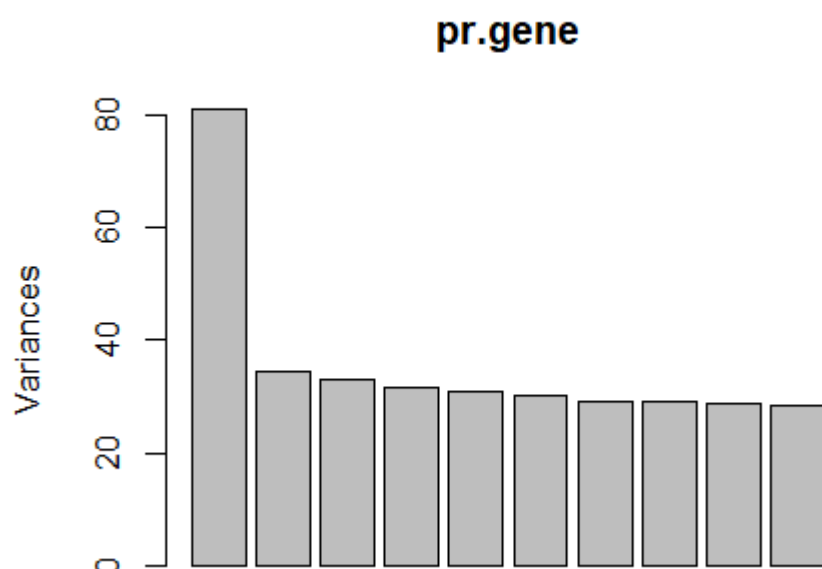
```
set.seed(702)
```

```
gl <- apply(pr.gene$rotation, 1, sum)
```

```
gl.dif <- order(abs(gl), decreasing=T)
```

```
top15 <- gl.dif[1:15]
```

```
top15
```



```
MEHERSHRISHTI>set.seed(702)
MEHERSHRISHTI>gene <- read.csv("geneData.csv", header = FALSE)
MEHERSHRISHTI>pr.gene <- prcomp(t(gene), scale=T)
MEHERSHRISHTI>plot(pr.gene)
MEHERSHRISHTI>summary(pr.gene)
Importance of components:
              PC1      PC2      PC3      PC4      PC5      PC6      PC7      PC8
Standard deviation  9.00460  5.87302  5.74347  5.61806  5.55344  5.50107  5.40069  5.38575
Proportion of Variance 0.08108 0.03449 0.03299 0.03156 0.03084 0.03026 0.02917 0.02901
Cumulative Proportion 0.08108 0.11558 0.14856 0.18013 0.21097 0.24123 0.27040 0.29940
              PC9      PC10     PC11     PC12     PC13     PC14     PC15     PC16
Standard deviation  5.3762  5.34146  5.31878  5.25016  5.18737  5.1667  5.10384  5.04667
Proportion of Variance 0.0289 0.02853 0.02829 0.02756 0.02691 0.0267 0.02605 0.02547
Cumulative Proportion 0.3283 0.35684 0.38513 0.41269 0.43960 0.4663 0.49234 0.51781
              PC17     PC18     PC19     PC20     PC21     PC22     PC23     PC24
Standard deviation  5.03288  4.98926  4.92635  4.90996  4.88803  4.85159  4.79974  4.78202
Proportion of Variance 0.02533 0.02489 0.02427 0.02411 0.02389 0.02354 0.02304 0.02287
Cumulative Proportion 0.54314 0.56803 0.59230 0.61641 0.64030 0.66384 0.68688 0.70975
              PC25     PC26     PC27     PC28     PC29     PC30     PC31     PC32
Standard deviation  4.70171  4.66105  4.64595  4.59194  4.53246  4.47381  4.4389  4.41670
Proportion of Variance 0.02211 0.02173 0.02158 0.02109 0.02054 0.02001 0.0197 0.01951
Cumulative Proportion 0.73185 0.75358 0.77516 0.79625 0.81679 0.83681 0.8565 0.87602
              PC33     PC34     PC35     PC36     PC37     PC38     PC39     PC40
Standard deviation  4.39404  4.3591  4.23504  4.2184  4.12936  4.0738  4.03658  4.64e-15
Proportion of Variance 0.01931 0.0190 0.01794 0.0178 0.01705 0.0166 0.01629 0.00e+00
Cumulative Proportion 0.89533 0.9143 0.93226 0.9501 0.96711 0.9837 1.00000 1.00e+00
MEHERSHRISHTI>set.seed(702)
MEHERSHRISHTI>gl <- apply(pr.gene$rotation, 1, sum)
MEHERSHRISHTI>gl.dif <- order(abs(gl), decreasing=T)
MEHERSHRISHTI>top15 <- gl.dif[1:15]
MEHERSHRISHTI>top15
[1] 889 676 755 960 907 19 475 673 374 174 716 878 327 567 840
MEHERSHRISHTI>
```

The first factor provides good separation between the two patient groups so variables that correlate highly with that factor are likely explaining the difference between diseased and healthy patients.

Q2. The Wage data set contains a number of other features, such as marital status (maritl), job class

#(jobclass),and others. Explore the relationships between some of these other predictors and wage, and

use non-linear fitting techniques in order to fit flexible models to the data. Create plots of the results

obtained, and write a summary of your findings.

```
library(ISLR)
```

```
library(boot)
```

```
set.seed(1)
```

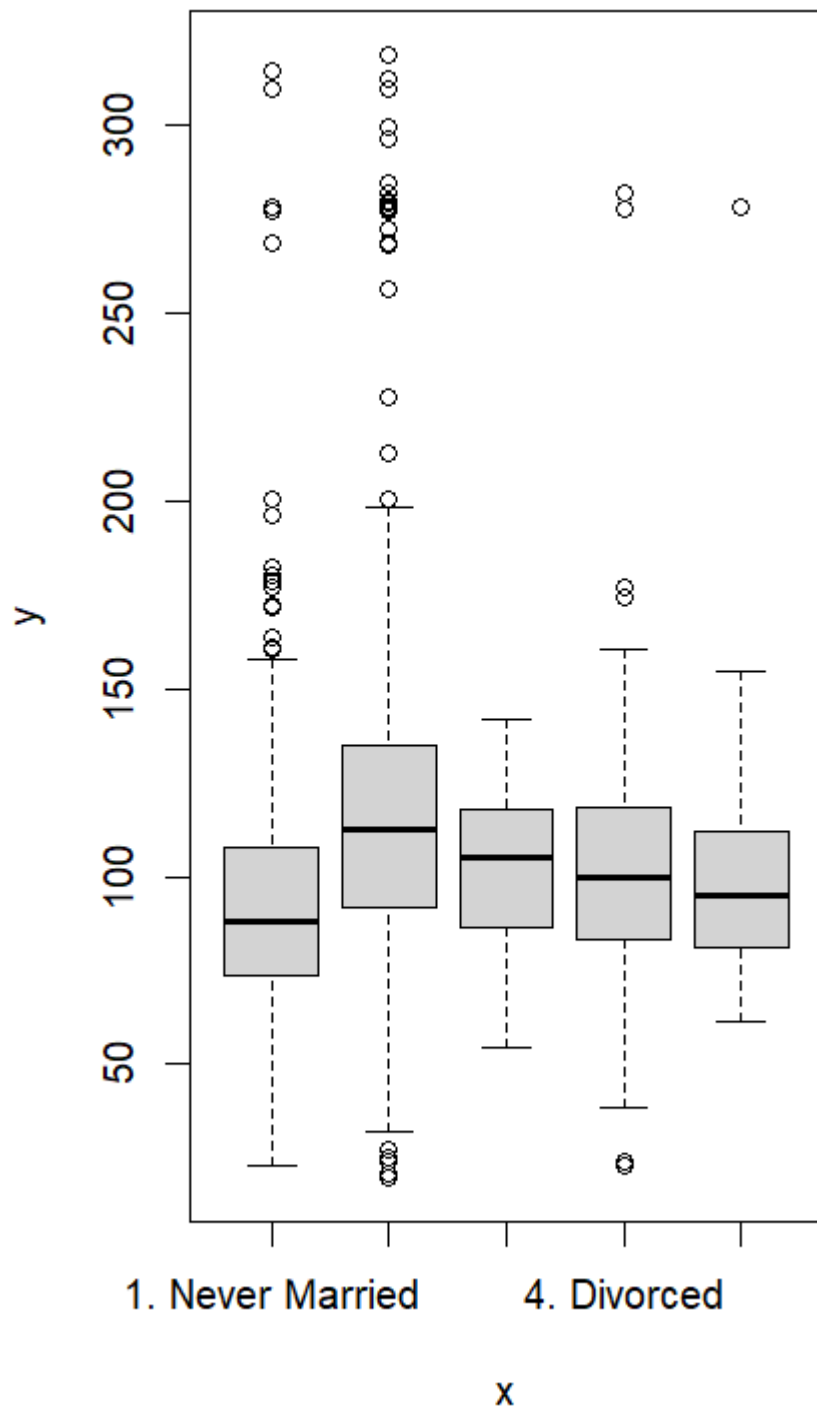
```
summary(Wage$maritl)
```

```
# table(Wage$maritl) the same with `summary`
```

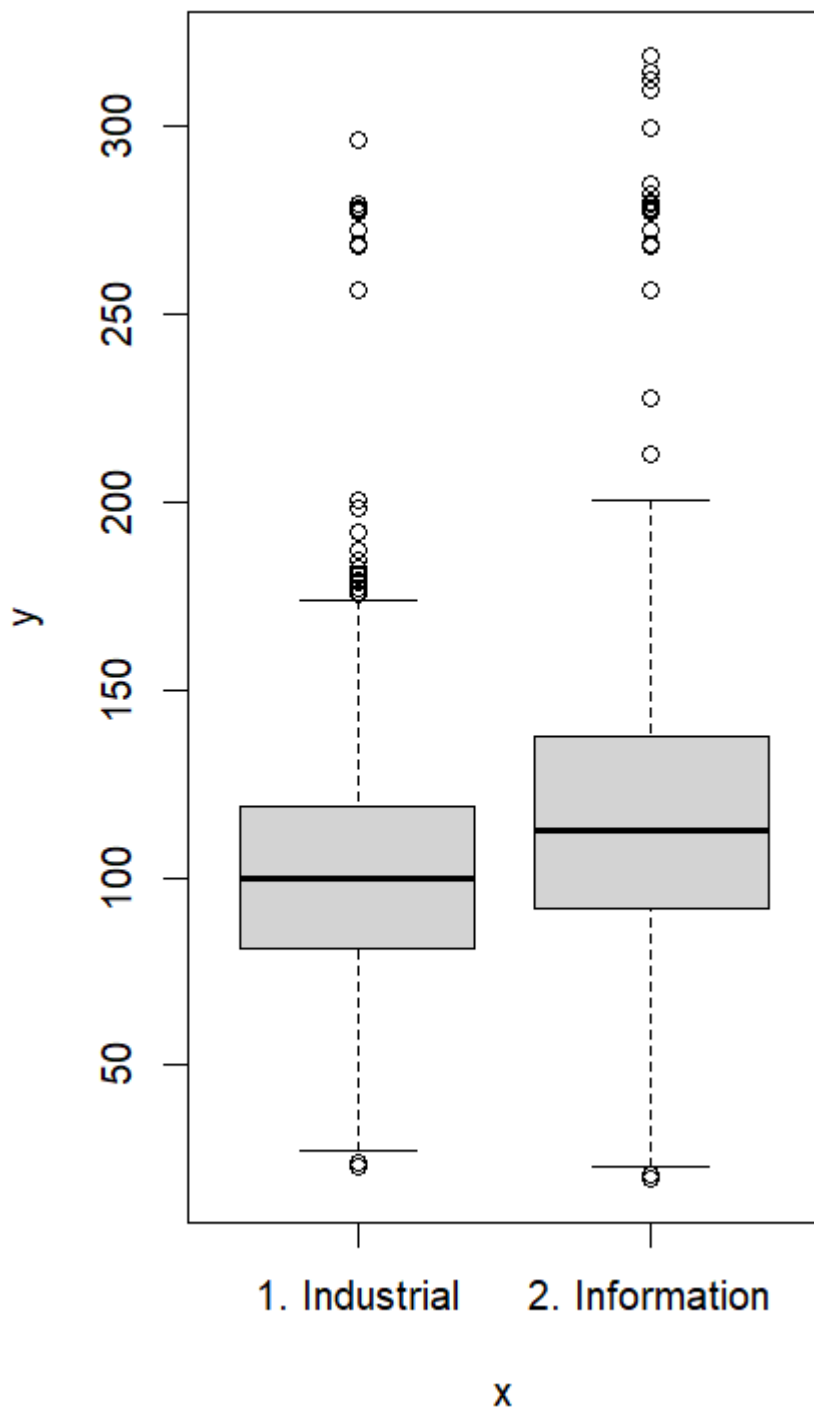
```
summary(Wage$jobclass)
```

```
par(mfrow = c(1, 2))
```

```
plot(Wage$maritl, Wage$wage)
```



```
plot(Wage$jobclass, Wage$wage)
```

```
install.packages("gam")
```

```
library(gam)
```

```
fit1 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education, data = Wage)
```

```
fit2 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass, data = Wage)
```

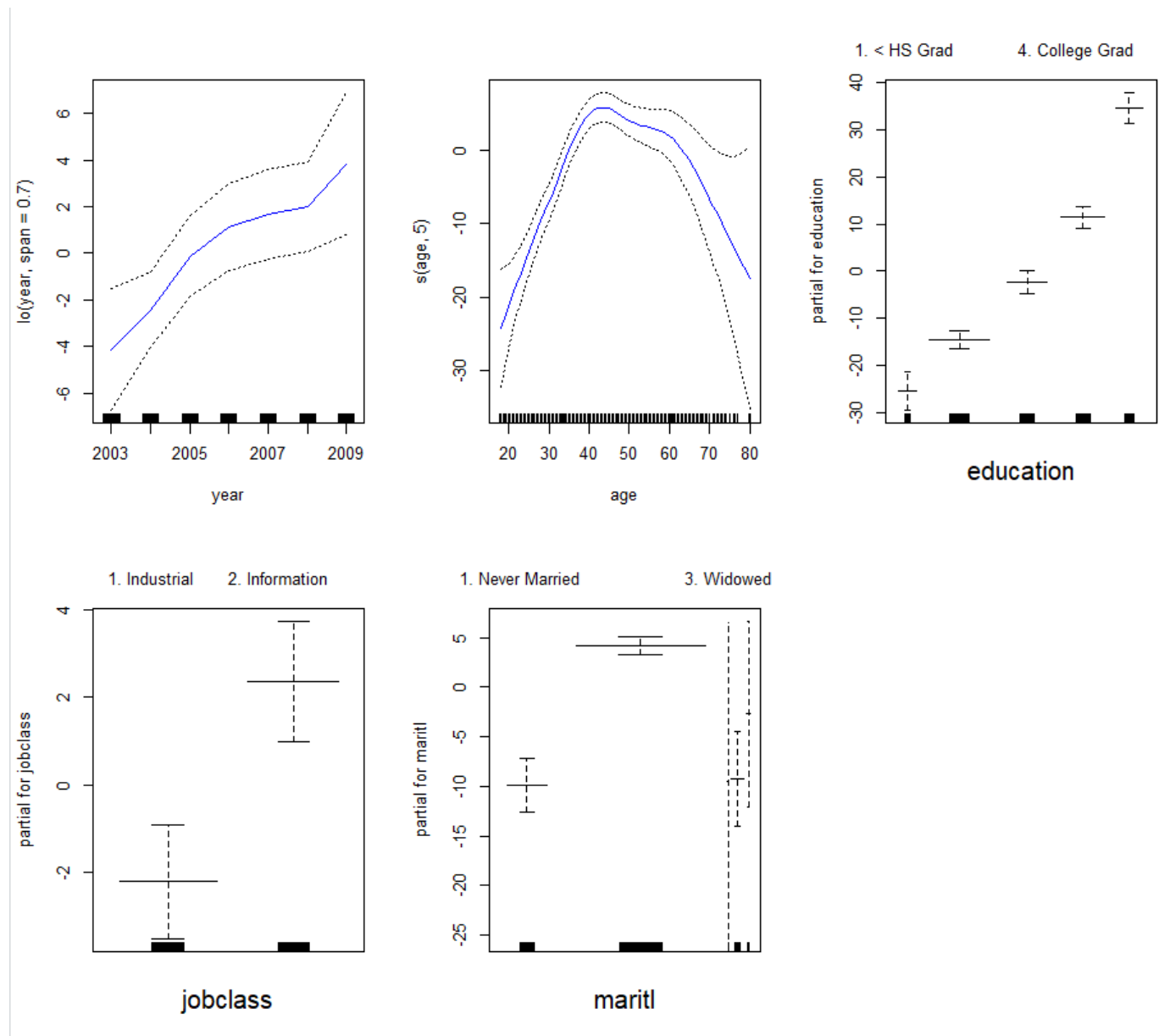
```
fit3 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + maritl, data = Wage)
```

```
fit4 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass +
maritl, data = Wage)
```

```
anova(fit1, fit2, fit3, fit4)
```

```
par(mfrow = c(2, 3))
```

```
plot(fit4, se = T, col = "blue")
```



```

MEHERSHRISHTI>library(ISLR)
MEHERSHRISHTI>library(boot)
MEHERSHRISHTI>set.seed(1)
MEHERSHRISHTI>summary(wage$maritl)
1. Never Married      2. Married      3. Widowed      4. Divorced      5. Separated
   648              2074              19              204              55
MEHERSHRISHTI># table(wage$maritl) the same with `summary`
MEHERSHRISHTI>summary(wage$jobclass)
1. Industrial 2. Information
   1544      1456
MEHERSHRISHTI>par(mfrow = c(1, 2))
MEHERSHRISHTI>plot(wage$maritl, Wage$wage)
MEHERSHRISHTI>plot(wage$jobclass, Wage$wage)
MEHERSHRISHTI>library(gam)
MEHERSHRISHTI>fit1 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education, data = Wage)
MEHERSHRISHTI>fit2 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass, data = Wage)
MEHERSHRISHTI>fit3 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + maritl, data = Wage)
MEHERSHRISHTI>fit4 <- gam(wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass + maritl, data = Wage)
MEHERSHRISHTI>anova(fit1, fit2, fit3, fit4)
Analysis of Deviance Table

Model 1: wage ~ lo(year, span = 0.7) + s(age, 5) + education
Model 2: wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass
Model 3: wage ~ lo(year, span = 0.7) + s(age, 5) + education + maritl
Model 4: wage ~ lo(year, span = 0.7) + s(age, 5) + education + jobclass + maritl
      Resid. Df Resid. Dev Df Deviance  Pr(>Chi)
1      2987.1      3691855
2      2986.1      3679689  1      12166 0.0014637 **
3      2983.1      3597526  3      82163 9.53e-15 ***
4      2982.1      3583675  1      13852 0.0006862 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
MEHERSHRISHTI>par(mfrow = c(2, 3))
MEHERSHRISHTI>plot(fit4, se = T, col = "blue")
MEHERSHRISHTI>

```

Q3. Fit some of the non-linear models to the Auto data set. Is there evidence for non-linear relationships in

this data set? Create some informative plots to justify your answer

```
set.seed(1)
```

```
pairs(Auto)
```

```
fit <- lm(mpg ~ poly(cylinders, 2) + poly(displacement, 5) + poly(horsepower, 5) + poly(weight, 5), data = Auto)
```

```
summary(fit)
```

```
anv1 <- gam(mpg ~ displacement + horsepower + weight, data = Auto)
```

```
anv2 <- gam(mpg ~ displacement + s(horsepower, 2) + weight, data = Auto)
```

```
anv3 <- gam(mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5), data = Auto)
```

```
anova(anv1, anv2, anv3, test = 'F')
```

```
summary(anv3)
```

```
par(mfrow=c(1,3))
```

```
plot.Gam(anv3, se=TRUE, col="red")
```

```
anv4 <- gam(mpg ~ s(displacement, 3) + s(horsepower, 3) + weight, data = Auto)
```

```
anova(anv4, anv3, test = 'F')
```

```
par(mfrow=c(1,3))
```

```
plot(anv4, se=TRUE, col="red")
```

```
lm1 <- glm(mpg ~ displacement + horsepower + weight, data = Auto)
```

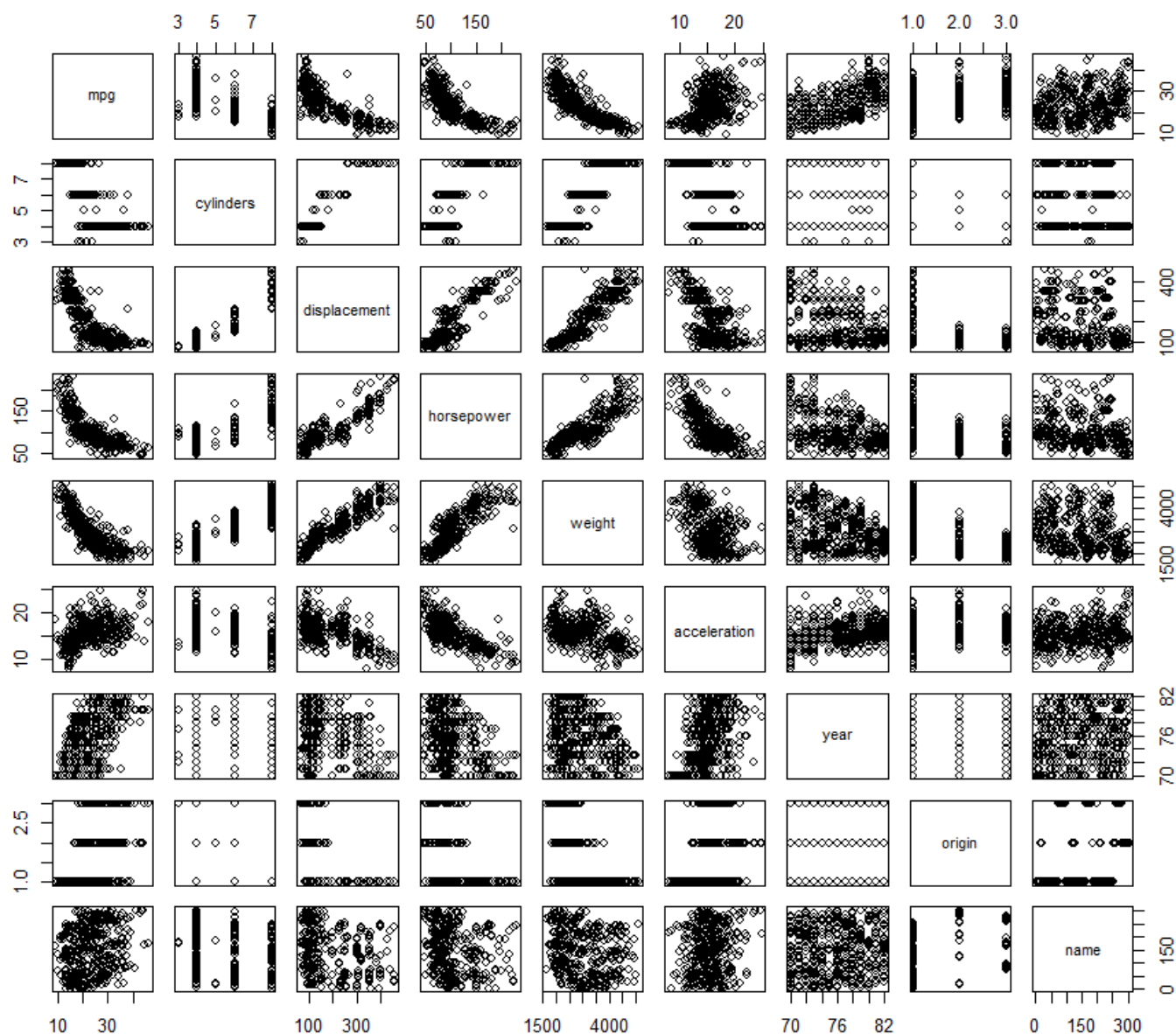
```
lm2 <- glm(mpg ~ poly(displacement, 3) + poly(horsepower, 3) + weight, data = Auto)
```

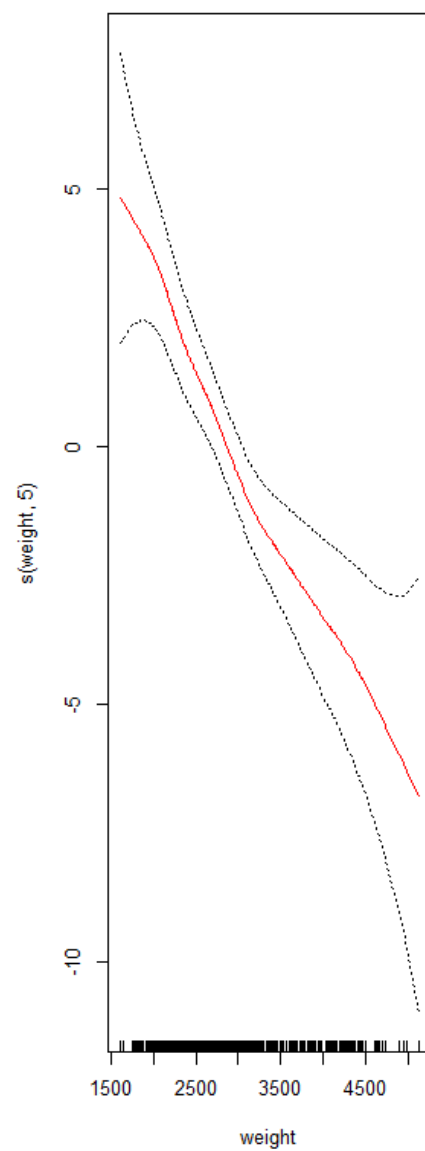
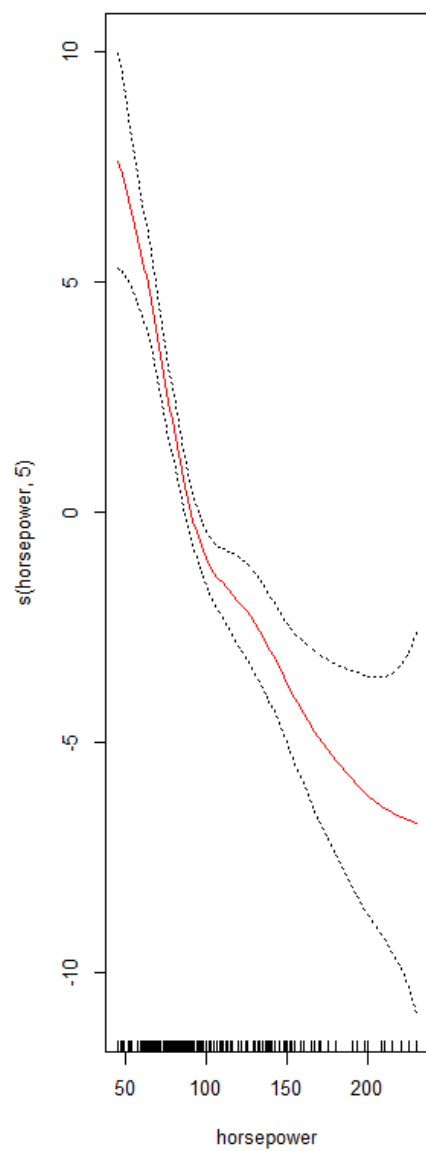
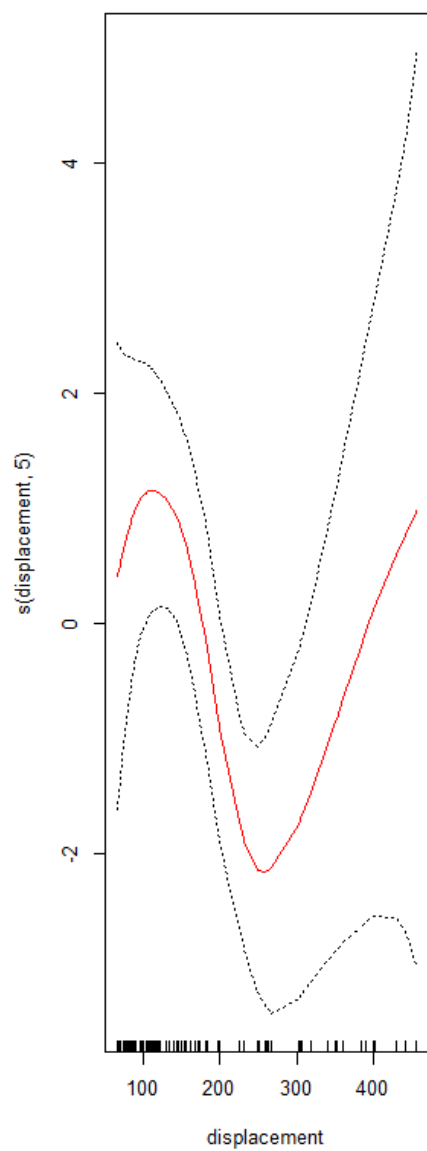
```
lm3 <- glm(mpg ~ poly(displacement, 5) + poly(horsepower, 5) + poly(weight, 5), data = Auto)
```

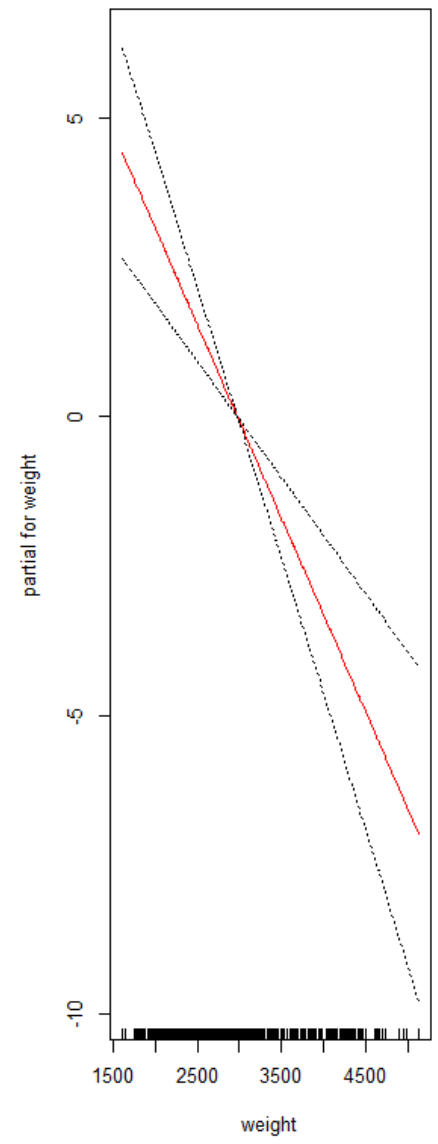
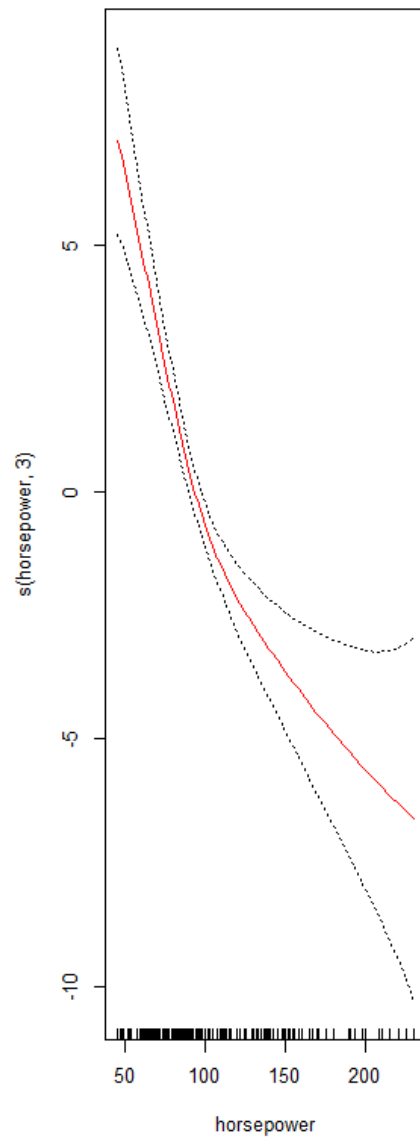
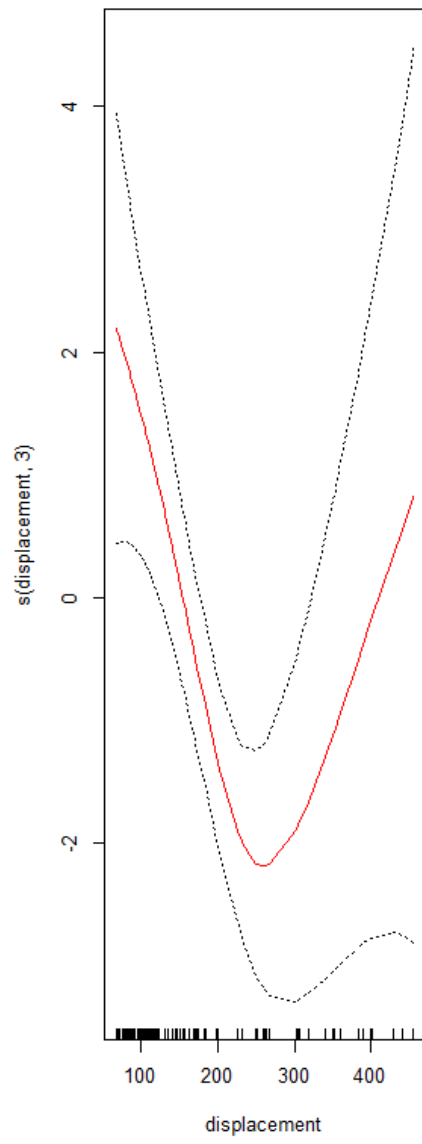
```
cv.glm(Auto, lm1, K = 10)$delta[1]
```

```
cv.glm(Auto, lm2, K = 10)$delta[1]
```

```
cv.glm(Auto, lm3, K = 10)$delta[1]
```







```
MEHERSHRISHTI>set.seed(1)
MEHERSHRISHTI>pairs(Auto)
MEHERSHRISHTI>fit <- lm(mpg ~ poly(cylinders, 2) + poly(displacement, 5) + poly(horsepower,
5) + poly(weight, 5), data = Auto)
MEHERSHRISHTI>summary(fit)
```

Call:

```
lm(formula = mpg ~ poly(cylinders, 2) + poly(displacement, 5) +
poly(horsepower, 5) + poly(weight, 5), data = Auto)
```

Residuals:

Min	1Q	Median	3Q	Max
-10.793	-2.219	-0.183	1.841	17.030

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	23.4459	0.1937	121.072	< 2e-16	***
poly(cylinders, 2)1	27.1932	18.4258	1.476	0.140834	
poly(cylinders, 2)2	-0.5902	7.4794	-0.079	0.937143	
poly(displacement, 5)1	-43.8830	19.8805	-2.207	0.027897	*
poly(displacement, 5)2	16.5805	9.8437	1.684	0.092942	.
poly(displacement, 5)3	12.7002	7.8850	1.611	0.108095	
poly(displacement, 5)4	-13.1163	5.7039	-2.300	0.022024	*
poly(displacement, 5)5	2.4590	4.9780	0.494	0.621607	
poly(horsepower, 5)1	-62.5295	12.1728	-5.137	4.51e-07	***
poly(horsepower, 5)2	21.5799	6.4347	3.354	0.000879	***
poly(horsepower, 5)3	-8.4355	6.7254	-1.254	0.210526	
poly(horsepower, 5)4	0.9338	4.4089	0.212	0.832378	
poly(horsepower, 5)5	8.5955	4.5355	1.895	0.058841	.
poly(weight, 5)1	-53.8275	12.9345	-4.162	3.93e-05	***
poly(weight, 5)2	6.3627	7.0359	0.904	0.366406	
poly(weight, 5)3	-3.1785	5.4532	-0.583	0.560333	
poly(weight, 5)4	-2.0484	4.6025	-0.445	0.656527	
poly(weight, 5)5	1.9338	4.1948	0.461	0.645062	

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 3.834 on 374 degrees of freedom
Multiple R-squared: 0.7692, Adjusted R-squared: 0.7587
F-statistic: 73.31 on 17 and 374 DF, p-value: < 2.2e-16

```
MEHERSHRISHTI>anv1 <- gam(mpg ~ displacement + horsepower + weight, data = Auto)
MEHERSHRISHTI>anv2 <- gam(mpg ~ displacement + s(horsepower, 2) + weight, data = Auto)
MEHERSHRISHTI>anv3 <- gam(mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5), data =
Auto)
MEHERSHRISHTI>anova(anv1, anv2, anv3, test = 'F')
```

Analysis of Deviance Table

Model 1: mpg ~ displacement + horsepower + weight

Model 2: mpg ~ displacement + s(horsepower, 2) + weight

Model 3: mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5)

	Resid. Df	Resid. Dev	Df	Deviance	F	Pr(>F)	
1	388	6980.0					
2	387	6145.6	0.99991	834.46	57.3356	2.879e-13	***
3	376	5472.8	10.99990	672.78	4.2021	6.952e-06	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1


```
MEHERSHRISHTI>anv1 <- gam(mpg ~ displacement + horsepower + weight, data = Auto)
MEHERSHRISHTI>anv2 <- gam(mpg ~ displacement + s(horsepower, 2) + weight, data = Auto)
MEHERSHRISHTI>anv3 <- gam(mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5), data =
Auto)
MEHERSHRISHTI>anova(anv1, anv2, anv3, test = 'F')
Analysis of Deviance Table
```

```
Model 1: mpg ~ displacement + horsepower + weight
Model 2: mpg ~ displacement + s(horsepower, 2) + weight
Model 3: mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5)
```

	Resid. Df	Resid. Dev	Df	Deviance	F	Pr(>F)
1	388	6980.0				
2	387	6145.6	0.99991	834.46	57.3356	2.879e-13 ***
3	376	5472.8	10.99990	672.78	4.2021	6.952e-06 ***

```
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
MEHERSHRISHTI>summary(anv3)
```

```
Call: gam(formula = mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight,
5), data = Auto)
```

```
Deviance Residuals:
```

Min	1Q	Median	3Q	Max
-10.5671	-2.0665	-0.2317	1.8421	16.3984

```
(Dispersion Parameter for gaussian family taken to be 14.5553)
```

```
Null Deviance: 23818.99 on 391 degrees of freedom
Residual Deviance: 5472.787 on 376.0002 degrees of freedom
AIC: 2179.87
```

```
Number of Local Scoring Iterations: NA
```

```
Anova for Parametric Effects
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
s(displacement, 5)	1	15397.9	15397.9	1057.889	< 2.2e-16 ***
s(horsepower, 5)	1	946.6	946.6	65.038	9.935e-15 ***
s(weight, 5)	1	400.7	400.7	27.528	2.592e-07 ***
Residuals	376	5472.8	14.6		

```
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
Anova for Nonparametric Effects
```

	Npar	Df	Npar F	Pr(F)
(Intercept)				
s(displacement, 5)	4	5.5978	0.000219	***
s(horsepower, 5)	4	9.8615	1.349e-07	***
s(weight, 5)	4	1.1977	0.311372	

```
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```

MEHERSHRISHTI>par(mfrow=c(1,3))
MEHERSHRISHTI>plot.Gam(anv3, se=TRUE, col="red")
MEHERSHRISHTI>anv4 <- gam(mpg ~ s(displacement, 3) + s(horsepower, 3) + weight, data = Auto)
MEHERSHRISHTI>anova(anv4, anv3, test = 'F')
Analysis of Deviance Table

Model 1: mpg ~ s(displacement, 3) + s(horsepower, 3) + weight
Model 2: mpg ~ s(displacement, 5) + s(horsepower, 5) + s(weight, 5)
  Resid. Df Resid. Dev    Df Deviance      F      Pr(>F)
1         384      5688.9
2         376      5472.8  7.9999    216.12  1.856  0.06572 .
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
MEHERSHRISHTI>par(mfrow=c(1,3))
MEHERSHRISHTI>plot(anv4, se=TRUE, col="red")
MEHERSHRISHTI>lm1 <- glm(mpg ~ displacement + horsepower + weight, data = Auto)
MEHERSHRISHTI>lm2 <- glm(mpg ~ poly(displacement, 3) + poly(horsepower, 3) + weight, data =
  Auto)
MEHERSHRISHTI>lm3 <- glm(mpg ~ poly(displacement, 5) + poly(horsepower, 5) + poly(weight,
  5), data = Auto)
MEHERSHRISHTI>cv.glm(Auto, lm1, K = 10)$delta[1]
[1] 18.21451
MEHERSHRISHTI>cv.glm(Auto, lm2, K = 10)$delta[1]
[1] 15.58109
MEHERSHRISHTI>cv.glm(Auto, lm3, K = 10)$delta[1]
[1] 15.51088
MEHERSHRISHTI>

```

The results also suggest model lm2 (same with an_v4) is good enough. So the conclusion of relationships with mpg: mpg ~ displacement: cubic; mpg ~ horsepower: cubic; mpg ~ weight: linear.