# Synthetic protein circuits for programmable control of mammalian cell death

Xia et al., 2024, Cell 187, 1-16

Reporter: 12212859 Sijie Li







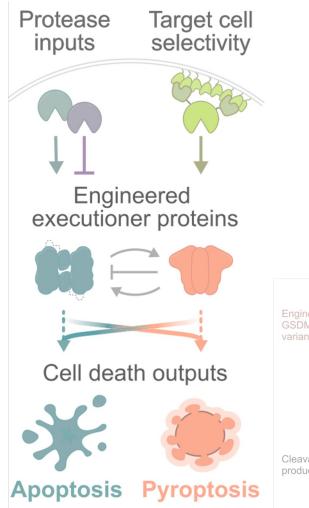


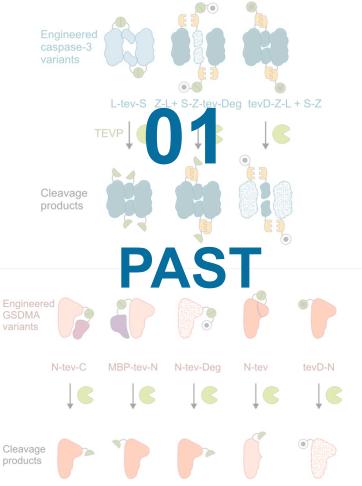


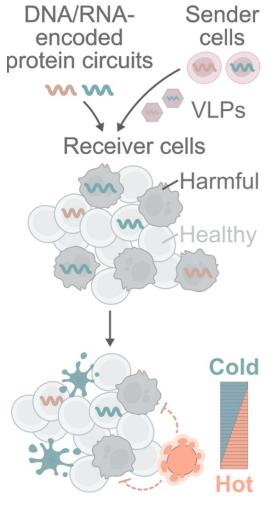












### **Cell Death?**

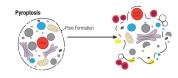
### **Apoptosis**

- > Triggered by caspase-3 and intrinsic signals
- > Cell shrinkage, membrane blebbing, nuclear fragmentation
- ➤ No leakage of intracellular contents
- Suitable for eliminating senescent or excess cells

# The cell draws Committee condenses Apoptosis Apoptosis Nuclear and experience colleges col

### **Pyroptosis**

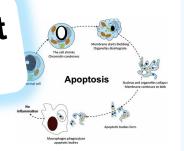
- Triggered by inflammatory caspases (e.g. caspase-1) or activated GSDMs
- > Causes membrane pore formation, cell swelling, lysis
- Releases IL-1β, IL-18, ATP, and DAMPs
- Useful for anti-tumor immunity or pathogen clearance
- > Needs careful control to avoid inflammatory storms



# Cell Death—main Pathways are uncovered

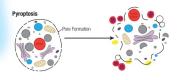
### **Apoptosis**

- > Cell shrinkage COLD: Immunologically silent
- → avoids inflammation
- Suitable for elim or excess cells



### **Pyroptosis**

- > Triggered by inflammatory caspass HOT: Immunologically active or activated GSDM
- ➤ Useful for a → inflammatory storms Causes me
- and the second s
- > Needs careful control to avoid inflammatory storms



### **Cell Death?**

### **Apoptosis**

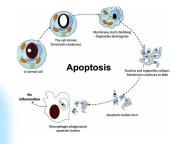
> Suitable for eliminating senescent or excess cells

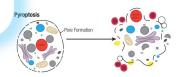
Expect to carry out the correct death modes under corresponding situation.
BUT, reality do not always work in this

way...SO?



Useful for anti-tumor immunity or pathogen clearance





### We want CELL DEATH

Before, To eliminate **senescent**, **cancerous**, or **infected** cells, we use...

# **✓** Cytotoxic Drugs

Induce apoptosis (mostly); poor specificity

# **V** CAR-T cells

Deliver granzymes → Induce either apoptosis or pyroptosis

# **✓** Chemogenetics / Optogenetics

Conditional expression or dimerization of apoptotic proteins

DVIV (DVIV 0---70

### We want SPECIFIC Cell Death

Before, To eliminate **senescent**, **cancerous**, or **infected** cells, we use...

# X Cytotoxic Drugs

Induce apoptosis (mostly); poor specificity

# **V** CAR-T cells

Deliver granzymes → Induce either apoptosis or pyroptosis

# X Chemogenetics / Optogenetics

Conditional expression or dimerization of apoptotic proteins → limit spatial targeting!

# We want SPECIFIC Cell Death in SPECIFIC MODE

Before, To eliminate **senescent**, **cancerous**, or **infected** cells, we use...

# X Cytotoxic Drugs

Induce apoptosis (mostly); poor specificity

# X CAR-T cells

Deliver granzymes → Induce

X Chemogenetics / Op Conditional expression or dime → limit spatial targeting! Synthetic Protein Circuits
Require New Method to
achieve Programmable
Control of Cell Death!!!

DAIA/DAIA Condar

# Why Synthetic Protein Circuits?

### gene circuits

operate at the transcriptional level, relying on promoter activity, mRNA synthesis, and protein translation.



### protein circuits

- function post-translationally, offering faster responses and finer temporal control
- more orthogonal to endogenous gene expression systems, reducing crosstalk and enhancing modularity

(Gao et al., 2018, Science)

DAIA /DAIA

# **Foundation of Synthetic Protein Circuits**

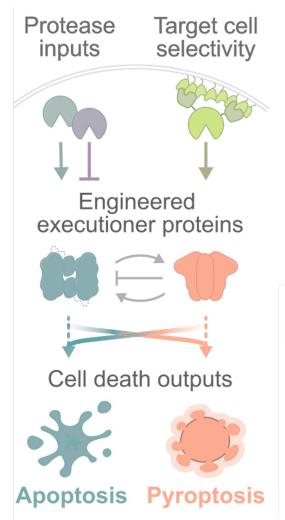
utilize engineered proteases (e.g., TEVP) to cleave specific sites on functional domains (e.g., caspase or gasdermin) → activating or inhibiting cellular processes (Gao et al., Science 2018)

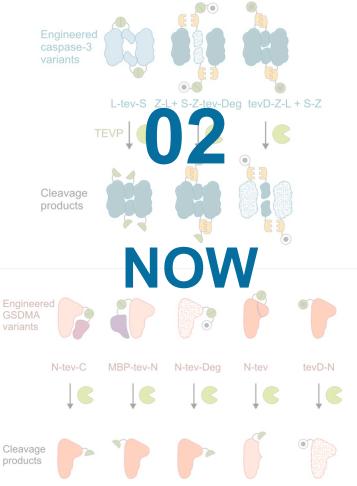


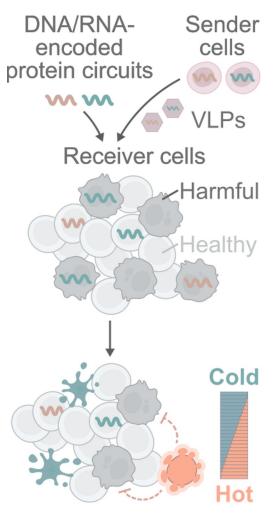
Protein degradation tags (degrons), caging domains, and heterodimerizing motifs (e.g., leucine zippers) are combined to control the stability, localization, or activity of key proteins (lwamoto et al., ChemBio 2010)

Degrons Masked (tevD)
Active (Deg) ●

✓ logical operations, signal integration, and programmable response behaviors within living cells

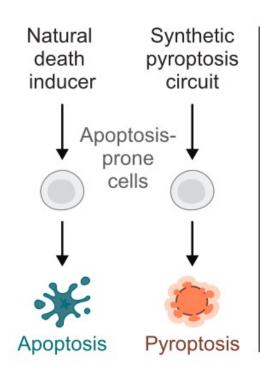


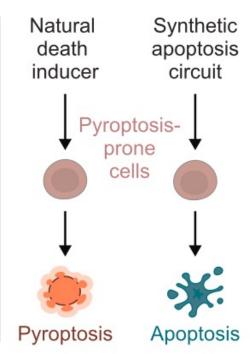




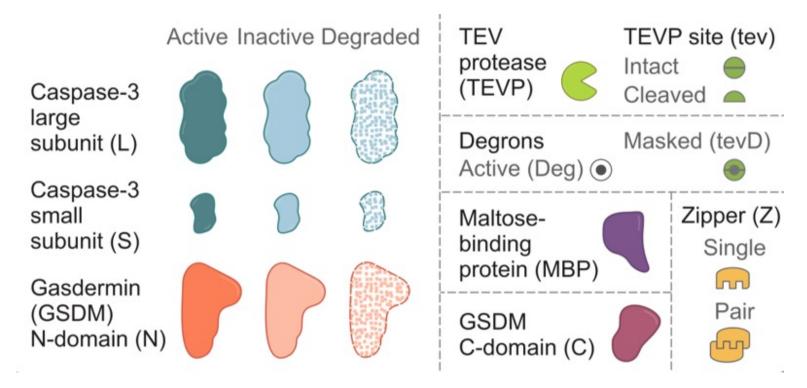
# **GOAL: Programmable Control of Cell Death**

- Control Over Cell Death Modes
- Selective Cell Elimination
- Adaptability to Delivery Methods
- Mimicry of Natural Processes





# **Building Blocks**



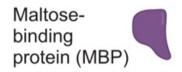
# **Building Blocks**

**TEV Protease (TEVP):** A viral protease used to cleave specific engineered sites (TEVP sites) in the circuit, enabling precise activation or deactivation of components like caspase-3 or GSDM.

**TEVP Site (tev):** The site (a range of sequence) that TEVP acted on.



**Maltose-Binding Protein (MBP):** Used as a bulky tag to sterically block activity (e.g., caging the N-domain of GSDM). Removal by TEVP cleavage restores function.



# **Building Blocks**

**Degrons (Deg):** Tags that promote protein degradation via proteasomal pathways, reducing the activity of the tagged protein. It can be masked by caging with TEVP sites, requiring cleavage to become functional.

Degrons Masked (tevD)
Active (Deg) •

**GSDM C-Terminal Domain (C):** Auto-inhibitory, it binds the N-terminal domain in intact gasdermin proteins, preventing pore formation and pyroptosis.

**Leucine Zippers (Z):** Designed as "single" or "paired" motifs for heterodimerization, these facilitate proximity interactions or artificial linkages in the circuits, such as stabilizing proteins or masking functional domains.

GSDM C-domain (C)

Zipper (Z)

Single

Pair

# **Apoptosis Modules**

### **Mechanism of Apoptosis**

- 1. Activation
- 2. caspase-3 proteolytic cleavage → separating its pro-domain, large subunit (L), and small subunit (S)
- 3. active caspase-3 forms a dimer (non-covalent interactions) between the large and small subunits → cleaves various downstream substrates, such as poly(ADP-ribose) polymerase (PARP) and lamin proteins
- 4. programmed cell death

Active Inactive Degraded Caspase-3 large subunit (L) Caspase-3 small subunit (S)

<sup>\*</sup> This pathway is conserved across the caspase family (so not only for caspase-3), with similar mechanisms observed in other caspase-driven apoptosis processes.

# **Apoptosis Modules**

### **Mechanism of Apoptosis**

- Regulate it! 1. Activation
- 2. caspase-3 proteolytic cleavage → separating its pro-domain, large subunit (L), and small subunit (S) Modulate its stability
- 3. active caspase-3 forms a dimer (noncovalent interactions) between the large and small subunits → cleaves various downstream substrates, such as poly(ADP-ribose) polymerase (PARP) and lamin proteins
- 4. programmed cell death

Caspase-3 large subunit (L)

small





Active Inactive Degraded









<sup>\*</sup> This pathway is conserved across the caspase family (so not only for caspase-3), with similar mechanisms observed in other caspase-driven apoptosis processes.

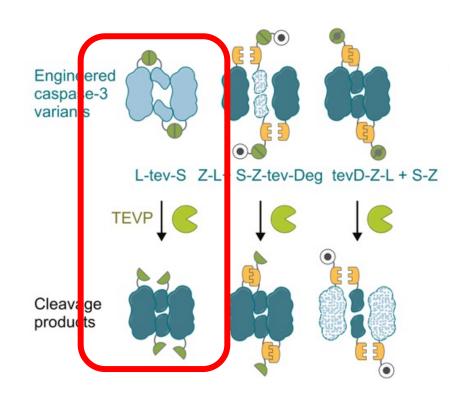
DAIA /DAIA

# **Apoptosis Modules**

### L-tev-S

### apoptosis induction

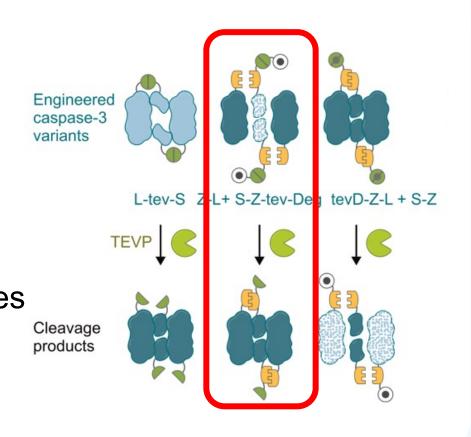
- caspase-3 engineered with a TEVP cleavage site between its large and small subunits
- ➤ TEVP cleavage →
  subunits separated →
  activate the dimer



# **Apoptosis Modules**

# **Z-L and S-tev-Deg** apoptosis induction

- ➤ active degron fused to the small subunit → suppresses caspase-3 activity
- ➤ TEVP cleavage removes the degron → leucine zippers facilitate dimerization

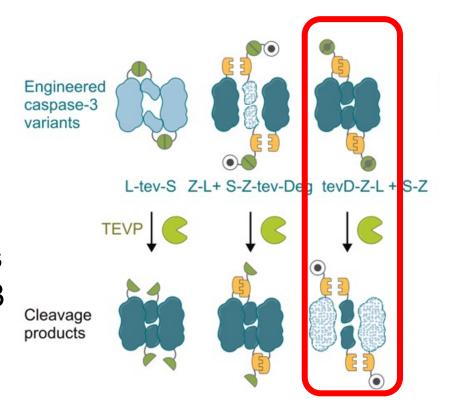


# **Apoptosis Modules**

### tevD-Z-L + S-Z

### apoptosis inhibition

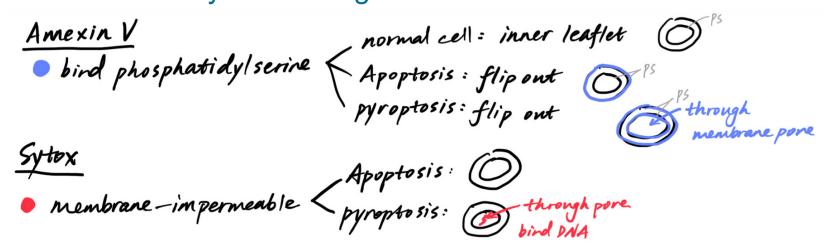
- ➤ caged degron fused to the large subunit → masking its activity
- ➤ TEVP cleavage exposes the degron → caspase-3 degrade



# **Apoptosis Modules Validation**

### **Death mode characterization**

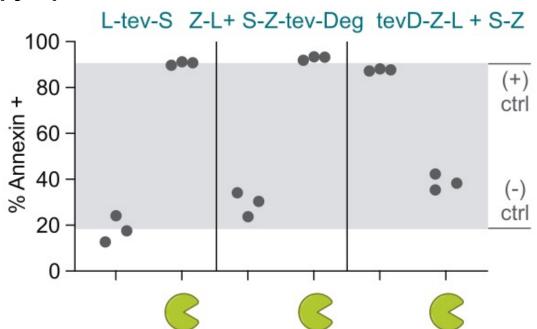
Annexin and Sytox staining



In brief, Annexin indicating apoptotic and pyroptotic states and Sytox specifically marking pyroptosis

# **Apoptosis Modules Validation**

- > Annexin: apoptosis and pyroptosis
- > Sytox: pyroptosis



### **Mechanism of Pyroptosis**

1. Activation

- Regulate it!
- 2. activated caspase-1/-4/-5 cleave GSDMA → generate: Modulate its stability
  - ➤ **N-terminal domain (active)** → inserts into membrane, forms pores
    - pores release IL-1β, IL-18, and DAMPs → trigger inflammation
    - ion imbalance → osmotic swelling → membrane rupture
  - C-terminal domain (inactive) → remains in cytosol, no pore formation

Active Inactive Degraded

Gasdermin (GSDM) N-domain (N)

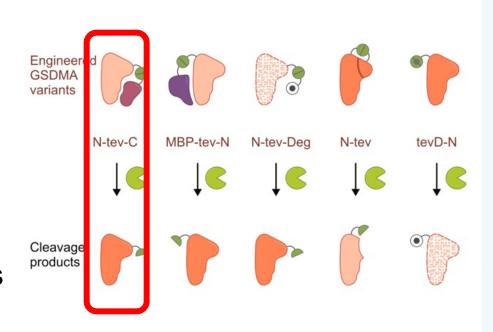
### 0----

# **Pyroptosis Modules**

### N-tev-C

### pyroptosis induction

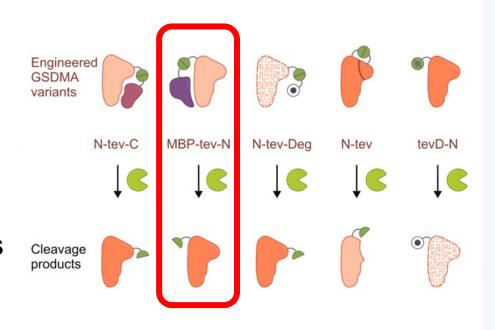
- gasdermin protein contains a TEVP cleavage site between its N-terminal (poreforming) and C-terminal (auto-inhibitory) domains
- > TEVP cleavage
  - → activates pyroptosis



### **MBP-tev-N**

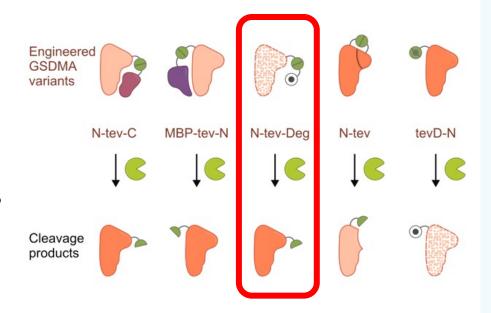
### pyroptosis induction

- ➤ maltose-binding protein fused to the N-terminal domain → block its pore-forming activity
- ➤ TEVP cleavage removesMBP → activatingpyroptosis



# N-tev-Deg pyroptosis induction

- N-domain fused to a degron → suppressing its activity
- ➤ TEVP cleavage removes the degron → activating the pyroptosis pathway





### **N-tev**

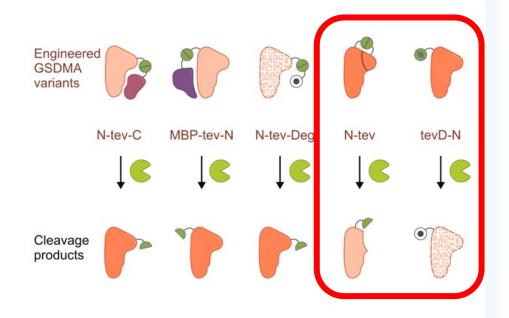
### pyroptosis inhibition

N-terminal domain with a TEVP cleavage site for direct activation

### tevD-N

### pyroptosis inhibition

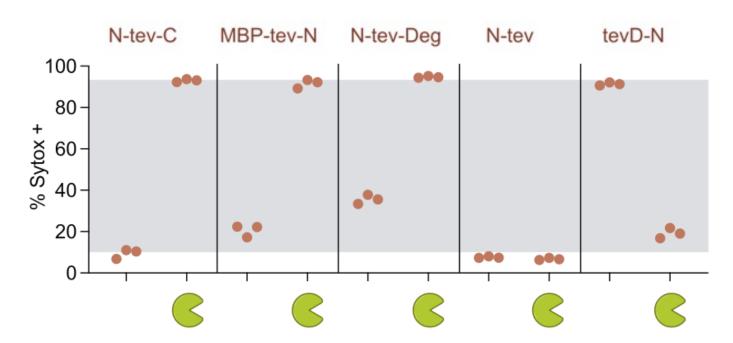
- caged degron masks the activity of the N-domain
- ➤ TEVP cleavage → degron exposed → marking the protein for degradation



# **Pyroptosis Modules Validation**

> Annexin: apoptosis and pyroptosis

> Sytox: pyroptosis

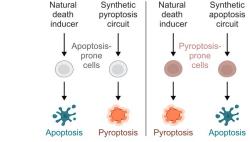


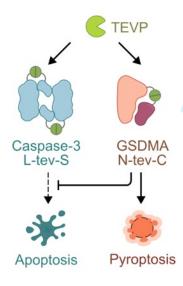
### What's Next?

- ✓ Apoptosis Module
- ✓ Pyroptosis Module

"Synostosis circuits"

Add A few elements to fullfill...

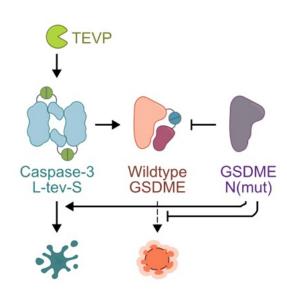




Driving pyroptosis in apoptosis-prone cells

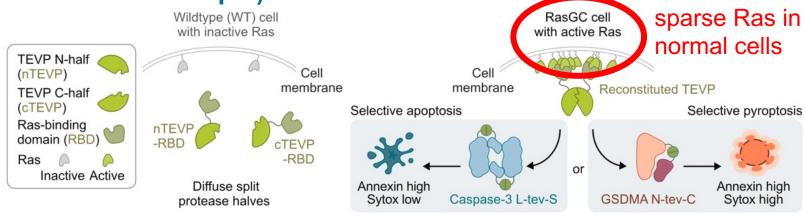
Driving apoptosis in pyroptosis-prone cells

**GSDMEN(mut)** is GSEME with its N domian mutant so that it can inhibit its WTcounterpart



# What's Next?--Specificity

Synostosis circuits selectively eliminate target cells (cancer cells as an example)



- Ras sensor with split TEVP fused to Ras-binding domains
- cancer cells have active Ras clustering at the membrane → brings the TEVP halves together → reconstituting active TEVP → activates downstream circuits → selective apoptosis or pyroptosis

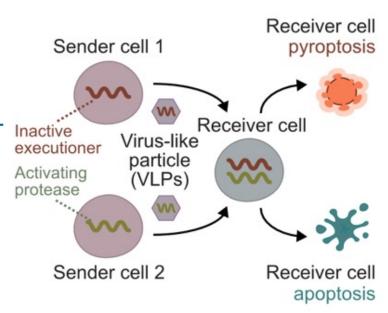
DAIA /DAIA

# What's Next?--Applicability

### Synostosis circuits support intercellular operations

# Split-Sender System

- ➤ Split pyroptosis circuit into an inactive executioner (e.g., GSDMA N-terminal domain) and an activating protease (e.g., TEVP) → Prevent self-destruction of sender cells
- Each component packaged into separate VLPs
- Receiver cells take up both components from the VLPs
  - → reconstitute the circuit → undergo pyroptosis or apoptosis

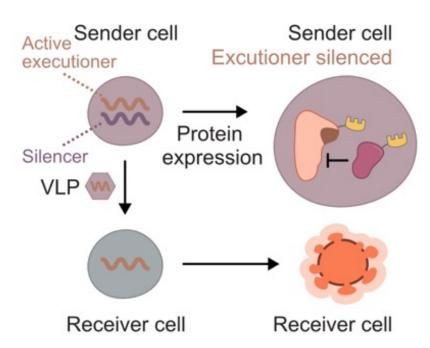


# What's Next?--Applicability

### Synostosis circuits support intercellular operations

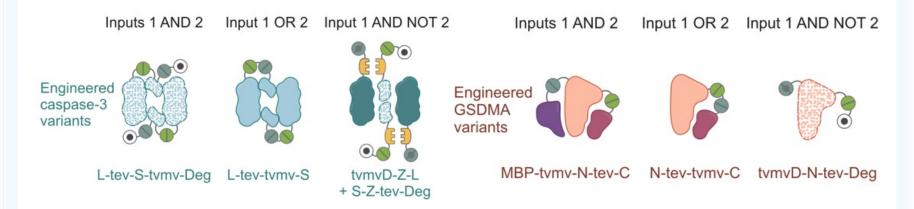
# Blocking Synpoptosis Activity in Senders

- a more compact system
- ➤ sender cells express silencer domains → inhibit executioner activity → protects the senders
- functional VLPs still trigger cell death in receivers

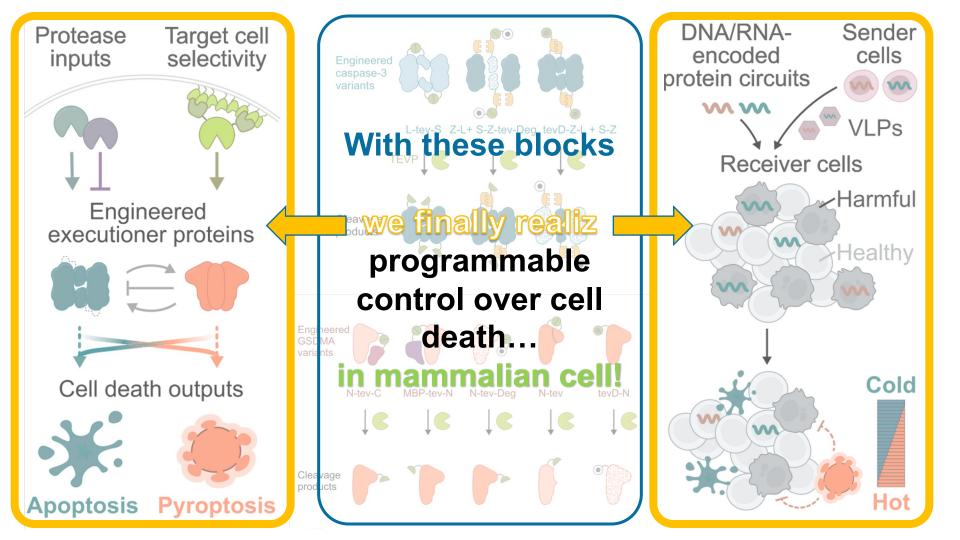


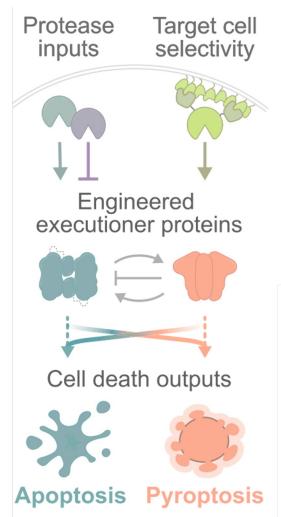
# What's Next?--Sophistication

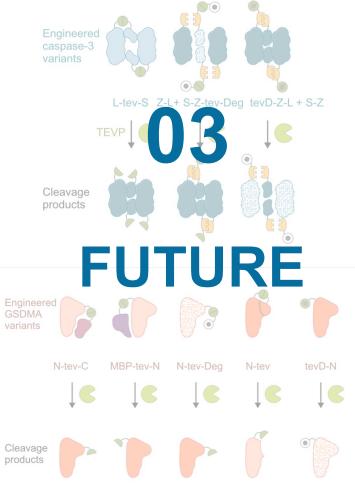
Synostosis circuits perform combinatorial computation!!

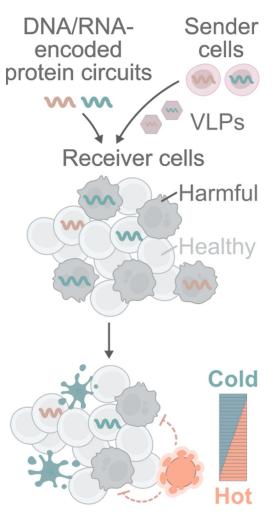


With these simple **boolean logic gates**, more complex implements can be done (just by combination), facilitating the death process to be more **adaptive**.



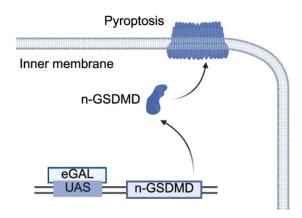






# Cited by...

- It is a relatively "fresh" article!
- So far, it has been cited by 12 articles (from scopus)
- And most of them are used as a background introduction / achievement (Qiu et al., 2025; Yu et al., 2025; Bai et al., 2025; Luca et al., 2025)



### A conceptual extension

"encoded PRCIS to express the N-terminal domain of gasdermin D (n-GSDMD), which is known to form membrane pores that facilitate pyroptosis" (Liu et al., 2025)

DAIA /DAIA

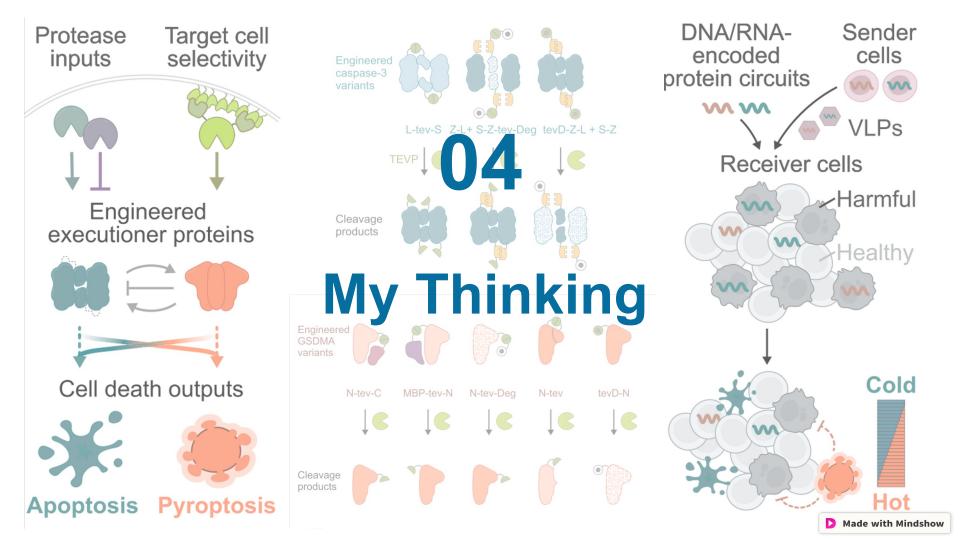
# Cited by...

- It is a relatively "fresh" article!
- So far, it has been cited by 12 articles (from scopus)
- ...and possible application (Orehek et al.,2025)

### **Design Inspirations for Inflammatory Skin Disorders**

"This technology can also serve as a targeted therapeutic approach to modulate the functions of pathogenic cells in **inflammatory skin diseases**" "a synthetic circuit tailored for **pathogenic KCs** can be designed to **respond to mechanical stress** signals mediated by piezo channels" (Yin et al., 2025)

A **different circumstance** aside from cell death.



DAIA/DAIA

# **Potential improvement**

From my perspective (some are noted in the Discussion too), this technique can further improve by overcome the following main points.

- ➤ Limited to Apoptosis and Pyroptosis: Other forms of cell death, such as necrosis or autophagy, are not directly addressed by the circuits, which limits their applicability to broader cellular contexts.
- ➤ Inability to Function in Non-Mammalian Systems: While the ultimate goal is to benefit human health, extending the circuit's applicability to common model organisms such as *Drosophila* or *C. elegans* would greatly enhance experimental flexibility and accelerate biological discovery.
- ➤ Unknown Single-Cell Dynamics of Synostosis Circuit: Exploring how individual cells respond to circuit activation could uncover cell-to-cell variability and reveal intermediate or hybrid death phenotypes. With the growing accessibility of single-cell technologies, I believe this direction holds great promise!

DAIA /DAIA

# **Potential improvement**

Of course, there are many points can be further improved (e.g. delivery optimization, advanced logic computation, orthogonality enhancement, etc.)

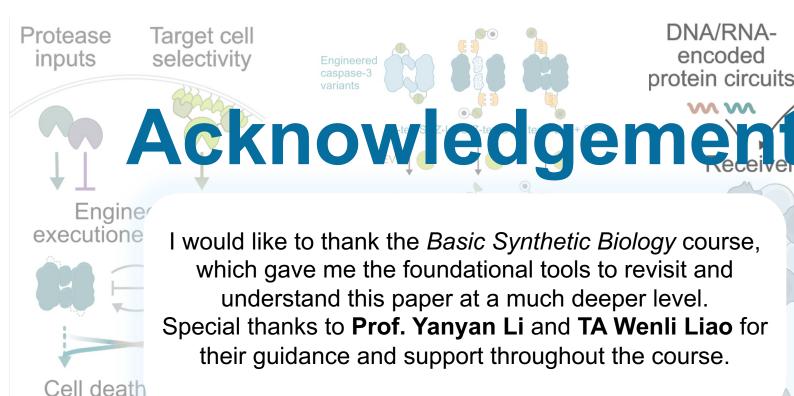
And I think, the following two directions can be starting points:

- ➤ Currently, the activity of synpoptosis circuits is **concentration-dependent** → **deepen understanding of cellular dynamics** to optimize concentration thresholds. This could involve **theoretical modeling.**
- Development of mechanisms to actively control intracellular concentration levels in real time. This could include engineering synthetic feedback loops, precise drug delivery systems, or leveraging molecular scaffolds to localize and stabilize circuit components. Such advancements would enable finer control over cell death activation, enhancing specificity and efficiency in therapeutic applications.

DNIA /DNIA

# **Inspirations**

- ➤ What really amazed me is how editable biological systems are!!!!
- ➤ At first it all looked super complex—but in the end, it's just basic biology, rearranged. Simple cleavage, degradation, dimerization... all wired together like logic blocks.
- ➤ The hard part isn't doing it—it's designing it, especially the fundamental block. Choosing the right tools, the right targets, and knowing how they'll behave inside a living cell... That takes deep understanding of both biological mechanisms and cellular dynamics, what I'll focus on as I learn more about synthetic biology.



DNA/RNAencoded protein circuits Sender cells



✓ Harmful

Healthy

Special thanks to Prof. Yanyan Li and TA Wenli Liao for



















Cold

### References

- Bai, Y., Pan, Y. & Liu, X. Mechanistic insights into gasdermin-mediated pyroptosis. Nat Rev Mol Cell Biol 26, 501–521 (2025). https://doi.org/10.1038/s41580-025-00837-0
- Chelur, D.S., and Chalfie, M. (2007). Targeted cell killing by reconstituted caspases. Proc. Natl. Acad. Sci. USA 104, 2283–2288. https://doi.org/ 10.1073/pnas.0610877104.
- Deng, W., Bai, Y., Deng, F., Pan, Y., Mei, S., Zheng, Z., Min, R., Wu, Z., Li, W., Miao, R., et al. (2022). Streptococcal pyrogenic exotoxin B cleaves GSDMA and triggers pyroptosis. Nature 602, 496–502. https://doi.org/ 10.1038/s41586-021-04384-4.
- Gao, X.J., Chong, L.S., Kim, M.S., and Elowitz, M.B. (2018). Programmable protein circuits in living cells. Science 361, 1252–1258. https://doi.org/10.1126/science.aat5062.
- He, K., Wan, T., Wang, D., Hu, J., Zhou, T., Tao, W., Wei, Z., Lu, Q., Zhou, R., Tian, Z., et al. (2023). Gasdermin D licenses MHCII induction to main- tain food tolerance in small intestine. Cell 186, 3033–3048.e20. https://doi.org/10.1016/j.cell.2023.05.027.
- Iwamoto, M., Bjo rklund, T., Lundberg, C., Kirik, D., and Wandless, T.J. (2010). A general chemical method to regulate protein stability in the mammalian central nervous system. Chem. Biol. 17, 981–988. https://doi.org/10.1016/j.chembiol.2010.07.009.
- Morsut, L., Roybal, K. T., Xiong, X., Gordley, R. M., Coyle, S. M., Thomson, M., & Lim, W. A. (2016). Engineering customized cell sensing and response behaviors using synthetic Notch receptors. Cell, 164(4), 780–791. https://doi.org/10.1016/j.cell.2016.01.012

### References

- Jorgensen, I., Rayamajhi, M., and Miao, E.A. (2017). Programmed cell death as a defence against infection. Nat. Rev. Immunol. 17, 151–164. <a href="https://doi.org/10.1038/nri.2016.147">https://doi.org/10.1038/nri.2016.147</a>.
- Liu, Y., Zhao, L., Long, J. et al. A generalizable approach for programming protease-responsive conformationally inhibited artificial transcriptional factors. Nat Commun 16, 4604 (2025). https://doi.org/10.1038/s41467-025-59828-6
- Orehek, S., Ramuta, T.Ž., Lainšček, D. et al. Cytokine-armed pyroptosis induces antitumor immunity against diverse types of tumors. Nat Commun15, 10801 (2024). https://doi.org/10.1038/s41467-024-55083-3
- Qiu, X., Zhu, L., Wang, H., & Xie, M. (2025). Biocomputing at the crossroad between emulating artificial intelligence and cellular supremacy. Current Opinion in Biotechnology, 92, 103264. https://doi.org/10.1016/j.copbio.2025.103264
- Xia et al., Synthetic protein circuits for programmable control of mammalian cell death, Cell (2024),https://doi.org/10.1016/j.cell.2024.03.031
- Yin, H., Chen, J., & Li, C. (2025). Immune memory: a new frontier in treating recurrent inflammatory skin diseases. Clinical Reviews in Allergy & Immunology, 68(1). https://doi.org/10.1007/s12016-025-09039-0
- Zhu, I., Liu, R., Garcia, J.M., Hyrenius-Wittsten, A., Piraner, D.I., Alavi, J., Israni, D.V., Liu, B., Khalil, A.S., and Roybal, K.T. (2022). Modular design of synthetic receptors for programmed gene regulation in cell therapies. Cell 185, 1431–1443.e16. https://doi.org/10.1016/j.cell.2022.03.023.