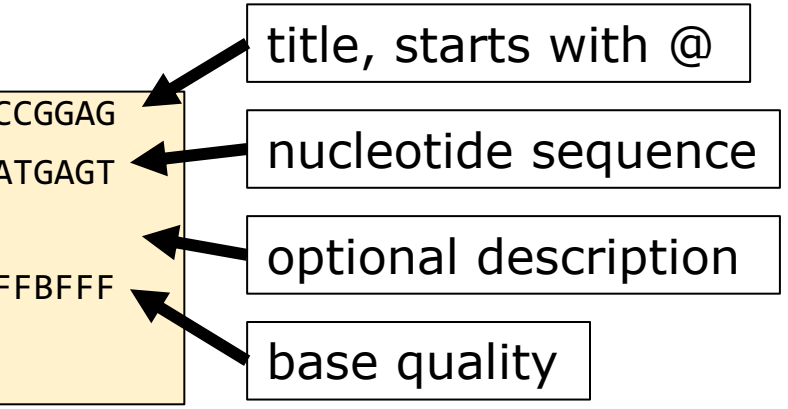
Long-read sequence analysis

QC & alignment

fastq

reads.fastq

```
@D00283R:66:CC611ANXX:4:2311:2596:2330 1:N:0:TCCGGAG
ACTCTACGCTCAATAAAGATTTCTGATACGGCTCCTGAAATGCAGAATGAGT
+
B/<<<B<FFFFFFFFFFBBFFFBFFFFBFFFF/FFFFFFFF/BFFFFFBFFF
```



fastq

fasta + basequality (fasta + q = fastq)

$$BASEQ = -10log_{10}\Pr\{base\ is\ wrong\}$$

$$-10log_{10}(0.01) = 20$$
$$-10log_{10}(0.1) = 10$$
$$-10log_{10}(0.5) = 3$$

Quiz question

What kind quality characteristics are important to long-read sequencing reads but less important for Illumina sequencing?

A. Base quality

B. Read length

C. GC content

D. Adapter content

Read quality control

- Number of reads
- Read length (mean and spread)
- Base quality
- Overrepresented sequences
- GC content
- Demultiplexing statistics
- Run duration/location dependency
- Others?

Read quality software

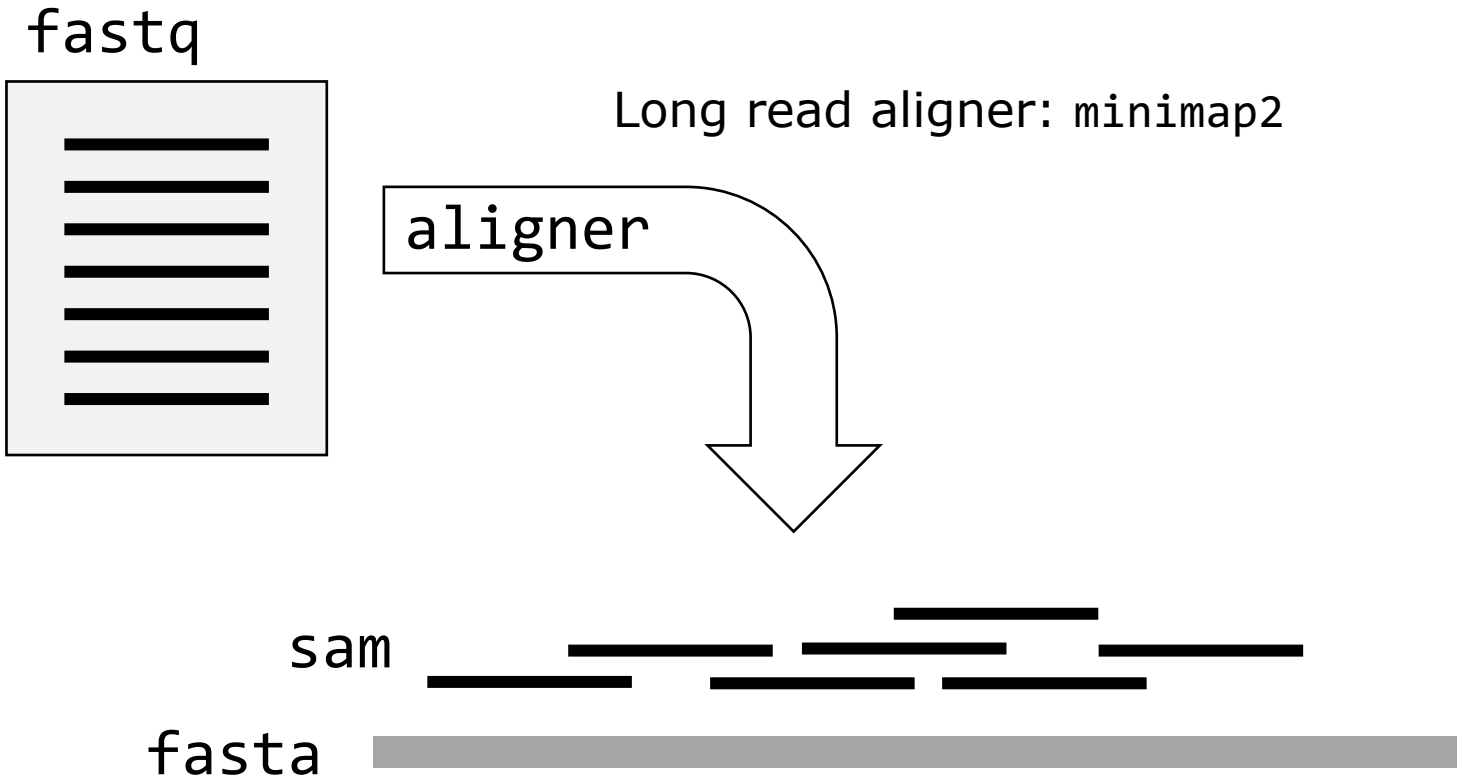
- Software of manufacturer
- NanoPlot (https://github.com/wdecoster/NanoPlot)
  - Takes many input formats
  - Basic statistics (fastq based)
- PycoQC (https://github.com/a-slide/pycoQC)
  - Specific for ONT
  - Requires so-called `sequencing_summary` file
- FastQC (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/)
  - Works also for long reads
  - Familiar output for many of us

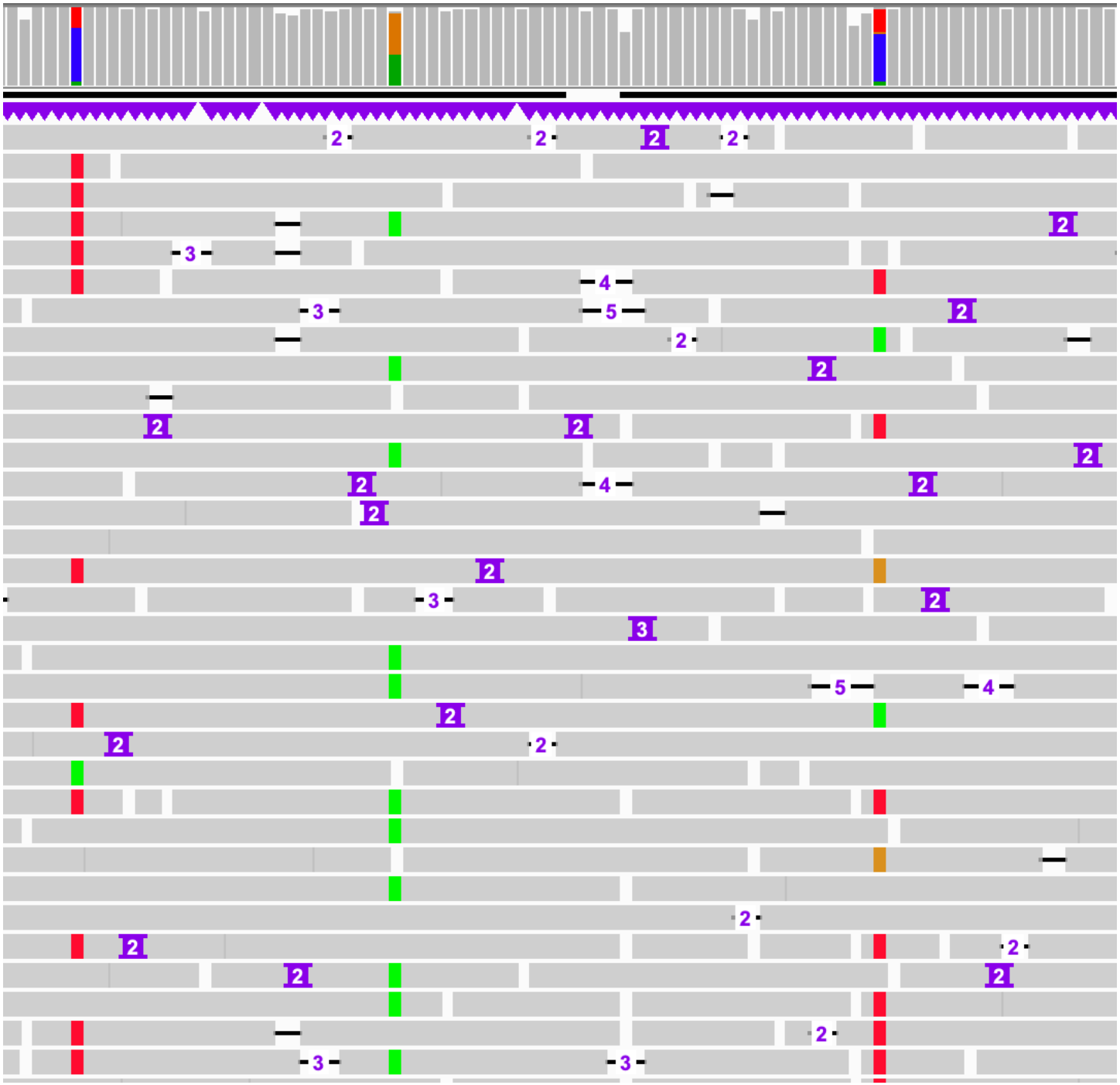
output of NanoPlot (https://github.com/wdecoster/NanoPlot)

Quality trimming

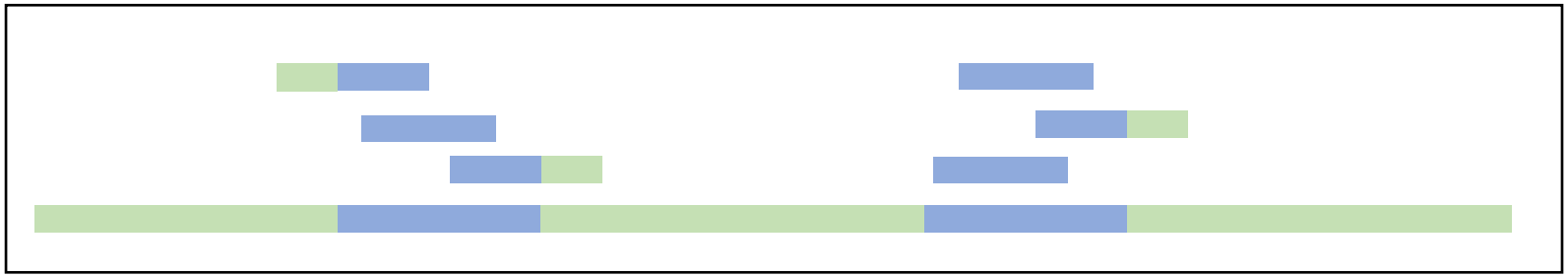
- Removal of:
  - Low quality sequences
  - Adapters/barcodes
- Oxford nanopore: On-instrument (`guppy`)
- PacBio:
  - On-instrument
  - During CCS generation (`pbccs`)

Read alignment (phred)





Mapping quality



$$MAPQ = -10log_{10}\Pr\{mapping\ position\ is\ wrong\}$$

$$-10log_{10}(0.01) = 20$$
$$-10log_{10}(0.5) = 3$$

sam header

```
@HD     VN:1.0  SO:coordinate
@SQ     SN:U00096.3     LN:4641652
@PG     ID:bowtie2      PN:bowtie2      VN:2.4.1        CL: bowtie2-
align-s --wrapper basic-0 -x ref.fasta -1 reads_1.fastq -2
reads_2.fastq"
```

SAM column	**example**
read name	SRR519926.5
flag	89
reference	chr20
start position	61
mapping quality	42
CIGAR string	150M
reference name mate is mapped	=
start position mate	476
fragment length	515
sequence	CATCACCATTCCCAC
base quality	@>4:4C@89+&9CC@
optional	AS:i:-2
optional	XN:i:0

samtools

- Convert `.sam` files into (a.o.)
  - `.fastq`
  - `.bam` (compressed `.sam`)
- Subset based on:
  - flag
  - region
- Ordering
- Mark alignment duplicates
- And many other things

Long-reads & fastq

- fastq format is limited to:
  - base
  - base-quality
- Long-read technologies -> need to store more information:
  - PacBio: (unaligned) bam
  - ONT: fast5