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Supervised Multiblock analyses

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Nutrimouse dataset

The data sets come from a nutrigenomic study in the mouse (Martin et al., 2007) in which the effects of five regimens with contrasted fatty acid compositions on liver lipids and hepatic gene expression in mice were considered.

Two sets of variables were acquired on forty mice: - genes: expressions of 120 genes measured in liver cells, selected (among about 30,000) as potentially relevant in the context of the nutrition study. These expressions come from a nylon macroarray with radioactive labelling - lipids: concentrations (in percentages) of 21 hepatic fatty acids measured by gas chromatography

Biological units (mice) were cross-classified according to two factors experimental design (4 replicates): - genotype: 2-levels factor, wild-type (WT) and PPARalpha -/- (PPAR) - diet: 5-levels factor. Oils used for experimental diets preparation were corn and colza oils (50/50) for a reference diet (REF), hydrogenated coconut oil for a saturated fatty acid diet (COC), sunflower oil for an Omega6 fatty acid-rich diet (SUN), linseed oil for an Omega3-rich diet (LIN) and corn/colza/enriched fish oils for the FISH diet (43/43/14)

```
data("nutrimouse")
genes <- nutrimouse$gene
lipids <- nutrimouse$lipid
metadata <- data.frame(genotype = nutrimouse$genotype, diet = nutrimouse$diet)
metadata$sample_name <- paste@(rownames(metadata), "_", metadata$genotype, "_", metadata$diet)
rownames(genes) <- metadata$sample_name
rownames(lipids) <- metadata$sample_name</pre>
```

Discriminant analysis of genotypes

Question 1: based on lipids and genes data, can we discriminate wt vs ppar samples?

Run block.plsda analysis with block.plsda() from mixomics package

```
# prepare data
blockPLS_data <- list(genes=genes, lipids=lipids)
genotype <- as.factor(metadata$genotype)

# run analysis
blockPLS_res <- block.plsda(X = blockPLS_data, Y = genotype, design = "full", all.outputs = T, ncomp = 10)</pre>
```

Question 2: Choose optimal number of latent variables?

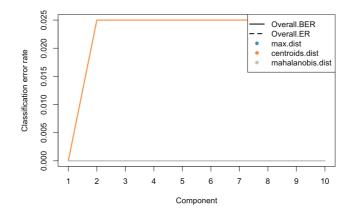
Run perf() plot the results with plot() Run the analysis with optimal number of latent variables

```
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blockPLS_perf <-- perf(blockPLS_res, validation = 'Mfold', folds = 7, nrepeat = 1, auc = TRUE, cpus=2, progressBar = FALS

E)

plot(blockPLS_perf)
```



```
blockPLS_res <- block.plsda(X = blockPLS_data, Y = genotype, design = "full", all.outputs = T, ncomp = 2)
```

Question 3: Is the model statistically significant?

Run a permutation test with DIABLO.test() from RVAideMemoire package

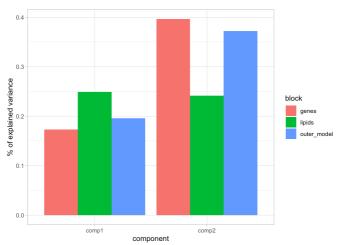
```
HIDE blockPLS_permtest <- DIABLO.test(blockPLS_res, progress = FALSE)
```

Question 4: what is the variance explained for each block by each latent variable and globally?

- for each block: AVE_X
- global: AVE[["AVE_outer"]]

```
blockPLS_expl <- do.call("rbind",blockPLS_res$AVE$AVE_X[1:2])
blockPLS_expl <- rbind(blockPLS_expl, blockPLS_res$AVE["AVE_outer"]])
rownames(blockPLS_expl)[3] <- "outer_model"
blockPLS_expl <- melt(blockPLS_expl)
colnames(blockPLS_expl) <- c("block", "comp", "value")

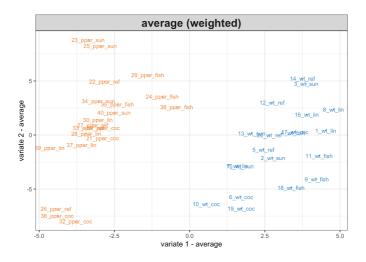
ggplot(blockPLS_expl, aes(x=comp, y=value, fill=block)) +
geom_bar(stat="identity", position=position_dodge()) +
labs(x="component",
    y="% of explained variance") +
theme_light()
```



Question 5: observe the samples distributions in the space of the latent variables.

plot scores with plotIndiv()

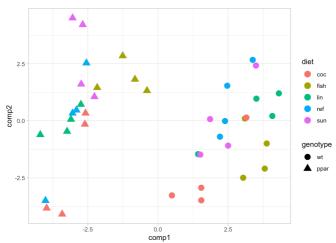
```
PlotIndiv(blockPLS_res, block = "weighted.average")
```



```
# ou:
blockPL5_variates.weighted <- blockPLS_res$variates[c("genes", "lipids")]
for(omic in c("genes", "lipids")){
    for(comp in c("comp1", "comp2")){
        blockPL5_variates.weighted[[omic]][,comp] <- blockPL5_variates.weighted[[omic]][,comp] * blockPLS_res$weights[omi c, comp]
    }
}
blockPL5_scores.weighted <- abind(blockPL5_variates.weighted[c("genes", "lipids")], along = 3)
blockPL5_scores.weighted <- apply(blockPL5_scores.weighted, c(1,2), mean)

blockPL5_scores.weighted <- data.frame(metadata, blockPL5_scores.weighted)

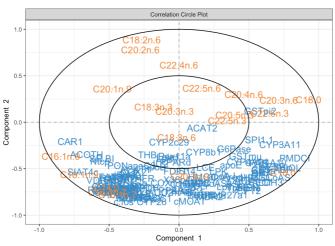
ggplot(blockPL5_scores.weighted, aes(x=comp1, y=comp2, col=diet, shape = genotype)) +
    geom_point(size=4) +
    theme_light()</pre>
```



Question 6: which genes and lipids are discriminant for genotype?

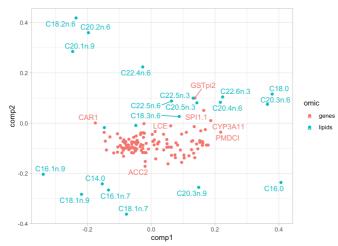
• plot loadings with plotVar()





```
# ou
blockPLS_loadings_genes <- blockPLS_res$loadings$genes
blockPLS_loadings_lipids <- blockPLS_res$loadings$lipids
blockPLS_loadings <- rbind.data.frame(blockPLS_loadings_genes, blockPLS_loadings_lipids)
blockPLS_loadings$mic <- c(rep("genes", dim(genes)[[2]]), rep("lipids", dim(lipids)[[2]]))
blockPLS_loadings$variable <- rownames(blockPLS_loadings)

ggplot(blockPLS_loadings, aes(x=comp1, y=comp2, col=omic, label=variable)) +
    geom_point() +
    geom_text_repel() +
    theme_light()</pre>
```



Consensus OPLS Discriminant analysis of genotypes

Question 1: based on lipids and genes data, can we discriminate wt vs ppar samples ?

Run ConsensusOPLS-DA analysis with ConsensusOPLS() from ConsensusOPLS package

```
COPLS_data <- list(genes=as.matrix(genes), lipids=as.matrix(lipids))

COPLS_data <- lapply(COPLS_data, scale)

genotype <- metadata$genotype

dummy_genotype <- as.matrix(data.frame(wt = ifelse(genotype == "wt", 1, 0),ppar = ifelse(genotype == "ppar", 1, 0)))

COPLS_res <- ConsensusOPLS(

data = COPLS_data,

Y = dummy_genotype,

maxPcomp = 1,

maxOcomp = 1,

modelType = "da",

cvType = "nfold",

nfold = 40,

nperm = 100,

verbose = T,

kernelParams = list(type='p', params = c(order=1.0))

}
```

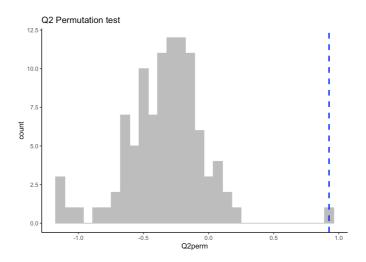
Question 2: Is the model statistically significant?

The results of permutations can be found in COPLS_res\$permStats . The results for the optimal model can be found in COPLS_res\$optimal\$modelCV and COPLS_res\$optimal\$modelCV\$cv

- plot Q2 permutations
- plot DQ2 permutations
- plot R2Y permutations

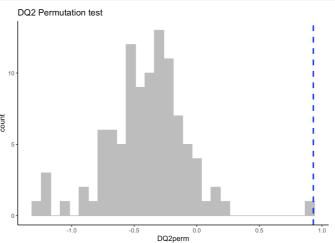
```
Q2perm <- data.frame(Q2perm = COPLS_res@permStats$Q2Y)

ggplot(data = Q2perm, aes(x = Q2perm)) +
geom_histogram(color="grey", fill="grey") +
geom_vline(aes(xintercept=COPLS_res@Q2["po1"]),color="blue", linetype="dashed", size=1) +
theme_classic() +
ggtitle("Q2 Permutation test")
```



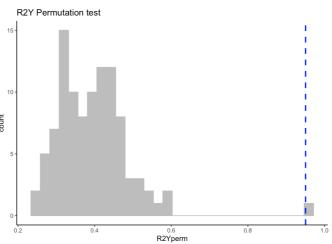
```
DQ2perm <- data.frame(DQ2perm = COPLS_res@permStats$DQ2Y)

ggplot(data = DQ2perm, aes(x = DQ2perm)) +
    geom_histogram(color="grey", fill="grey") +
    geom_vline(aes(xintercept=COPLS_res@DQ2["po1"]),color="blue", linetype="dashed", size=1) +
    theme_classic() +
    ggtitle("DQ2 Permutation test")
```



```
R2Yperm <- data.frame(R2Yperm = COPLS_res@permStats$R2Y)

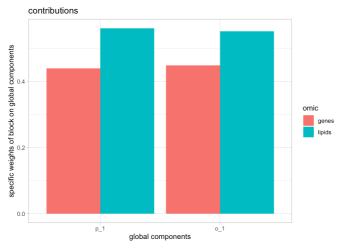
ggplot(data = R2Yperm, aes(x = R2Yperm)) +
    geom_histogram(color="grey", fill="grey") +
    geom_vline(aes(xintercept=COPLS_res@R2Y["po1"]),color="blue", linetype="dashed", size=1) +
    theme_classic() +
    ggtitle("R2Y Permutation test")
```



Question 3: What is the contribution of each data block?

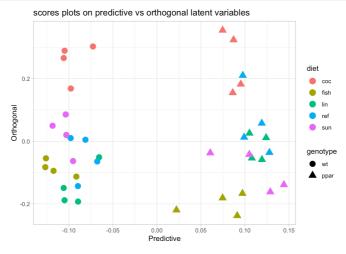
plot blockContribution of the optimal model

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Question 4: Show the distribution of samples in the space of the predictive and orthogonal latent variables?

plot scores of the optimal model



Question 5: Show the loadings of variables in the space of the predictive and orthogonal latent variables?

plot loadings of the optimal model

```
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loadings <- rbind.data.frame(COPLS_res@loadings$genes, COPLS_res@loadings$lipids)

loadings$dataset <- c(rep("genes", nrow(COPLS_res@loadings$genes)), rep("lipids", nrow(COPLS_res@loadings$lipids)))

loadings$variable <- rownames(loadings)

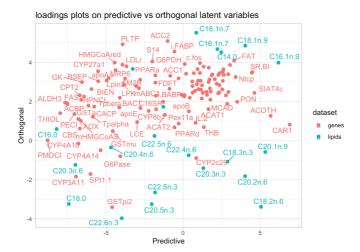
ggplot(loadings, aes(x=p_1, y=o_1, col=dataset, label = variable)) +

geom_point(size=2) +

labs(x="Predictive",
    y="Orthogonal",
    title = "loadings plots on predictive vs orthogonal latent variables") +

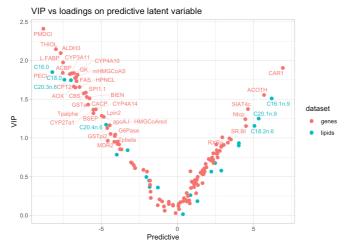
geom_text_repel() +

theme_light()
```



Question 6: Show the importance of variables in the model?

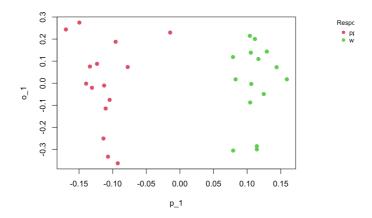
plot loadings and VIP of the optimal model



Question 7: train a model to discriminate between wt and ppar with 30 observations and test it with 10 observations?

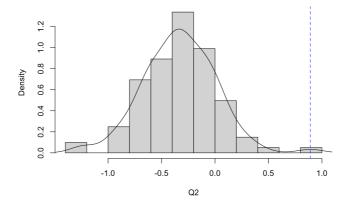
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```
test.observations <-c(sample(1:20, 5, replace = F),
                       sample(21:40, 5, replace = F))
{\tt train.data} \, \leftarrow \, {\tt list(genes=as.matrix(genes[-test.observations,])} \, ,
                    lipids=as.matrix(lipids[-test.observations,]))
train.data <- lapply(train.data, scale)</pre>
\texttt{test.data} \gets \texttt{list(genes=as.matrix(genes[test.observations,]),}
                   lipids=as.matrix(lipids[test.observations,]))
test.data <- lapply(test.data, scale)</pre>
train.genotype <- metadata$genotype[-test.observations]</pre>
train.COPLS_res <- ConsensusOPLS(
 data = train.data,
 Y = train.genotype,
 maxPcomp = 1,
 maxOcomp = 1,
 modelType = "da",
 cvType = "nfold",
 nfold = 30,
 nperm = 100,
  kernelParams = list(type='p', params = c(order=1.0))
plotScores(train.COPLS_res)
```



plotQ2(train.COPLS_res)

Q2 in models with permuted response

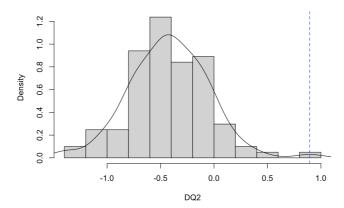


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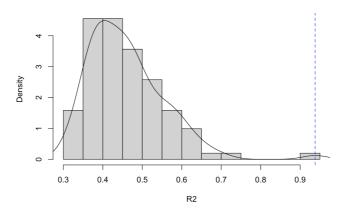
plotDQ2(train.COPLS_res)

DQ2 in models with permuted response

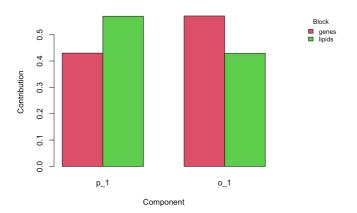


plotR2(train.COPLS_res)

R2 in models with permuted response



plotContribution(train.COPLS_res)

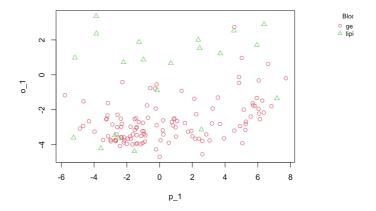


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 ${\tt ConsensusOPLS::plotLoadings(train.COPLS_res)} \ \textit{\#because mixOmics also has a function called plotLoadings()} \\$



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plotVIP(train.COPLS_res)

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```
class margin softmax.ppar softmax.wt True.genotype
## 20_wt_ref
               wt 1.494770
                                                             wt
## 8_wt_lin
                 wt 2.292263
                                                              wt
## 18_wt_fish
                 wt 2.598141
                                                              wt
## 13_wt_sun
                 wt 1.545651
                                                              wt
## 17_wt_coc
                wt 1.412837
                                                             wt
## 30_ppar_lin ppar 1.887535
                                                            ppar
## 24_ppar_fish ppar 1.279840
                                                            ppar
## 27_ppar_ref ppar 2.076832
                                                            ppar
## 22_ppar_ref ppar 2.533283
                                                            ppar
## 29_ppar_fish ppar 1.566171
```