



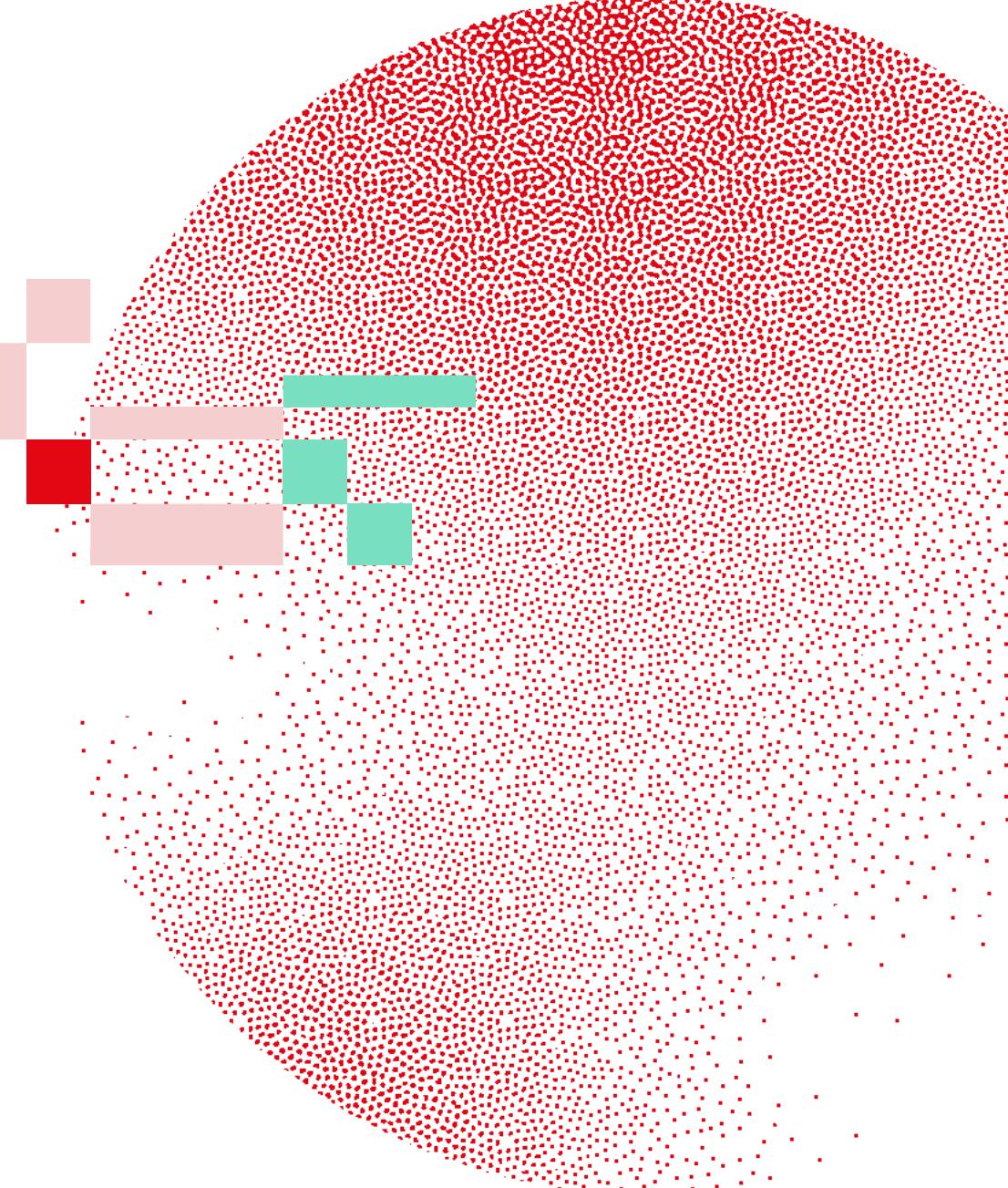
Swiss Institute of  
Bioinformatics

INTRODUCTION TO SEQUENCING-BASED SPATIAL  
TRANSCRIPTOMICS DATA ANALYSIS

# QC and Normalization

Deepak Tanwar

December 9-10, 2025



**Which QC do you use for single-cell data?**

QC for Visium HD is conceptually the same  
as single-cell QC – but computed *per High-  
Definition spot* rather than per cell.

Cell → Spot

# Why QC of spatial data?

High-density spots can capture technical artifacts (folds, bubbles, staining issues).

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Poor-quality regions can bias downstream analyses (clustering, spatial DE, trajectory analysis).

# QC in Visium HD

Same metrics as single-cell RNA-seq:

- Total UMIs per spot

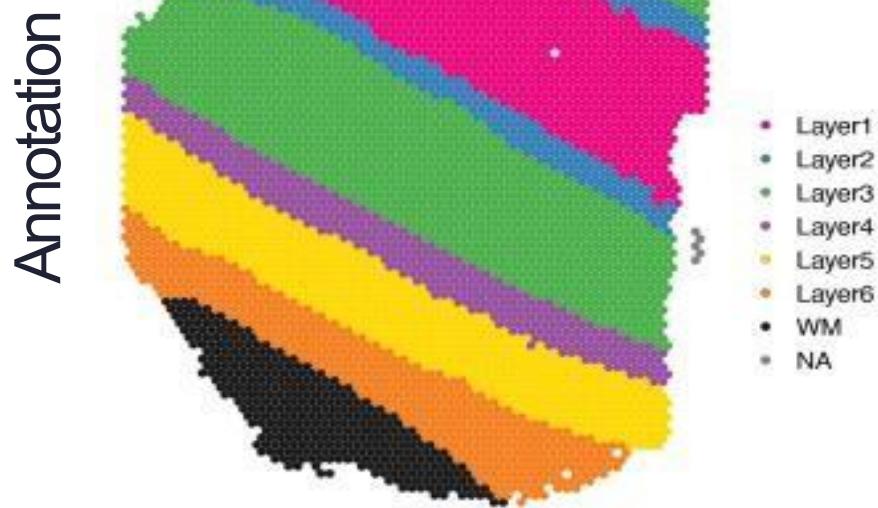
- Number of genes per spot

- Mitochondrial fraction

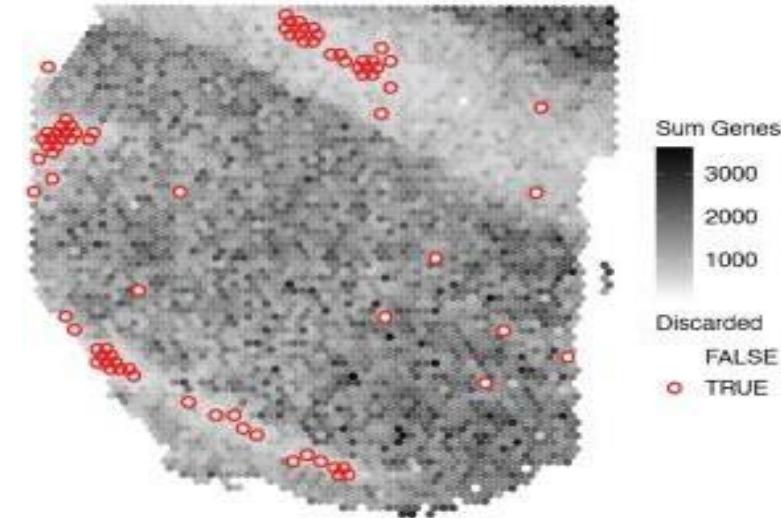
Key difference: unit is HD spot, not single cell.

Neighboring spots share microenvironment → spatial artifacts can emerge

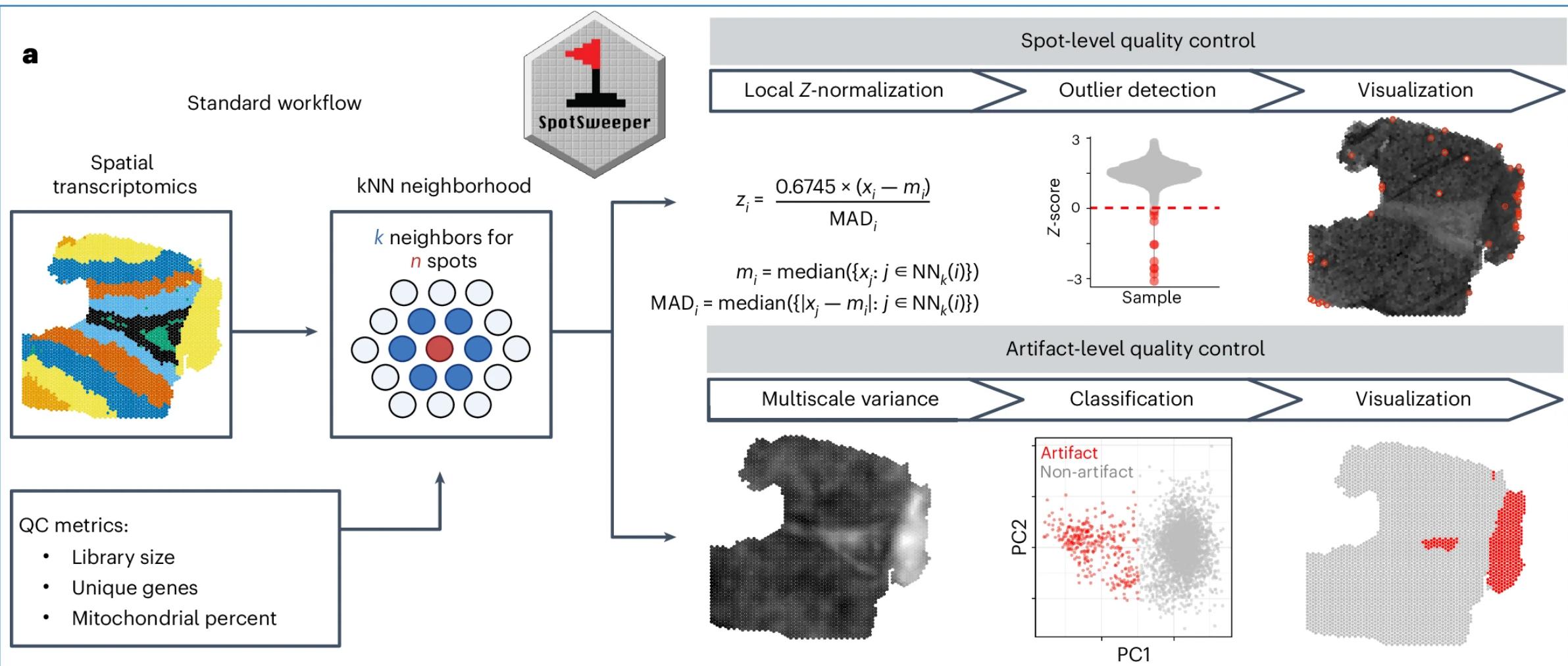
# Example of spots removal



Global

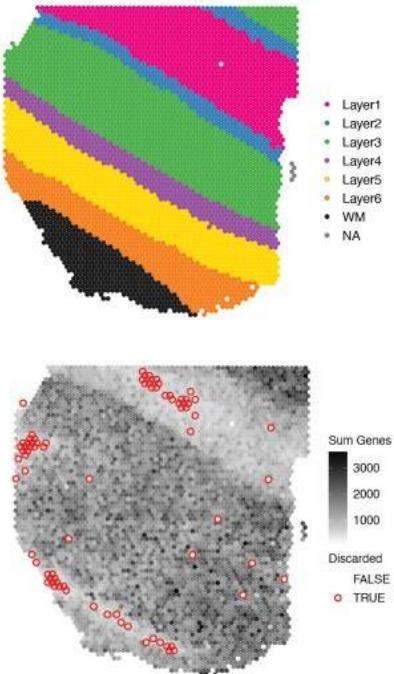


# SpotSweeper: spatially aware QC



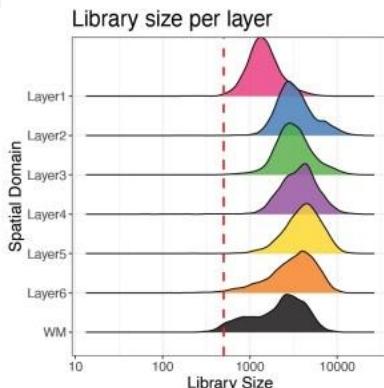
# SpotSweeper (spot-level artifacts)

Global Annotation

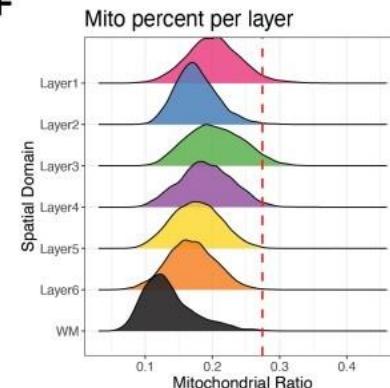


## Global outliers

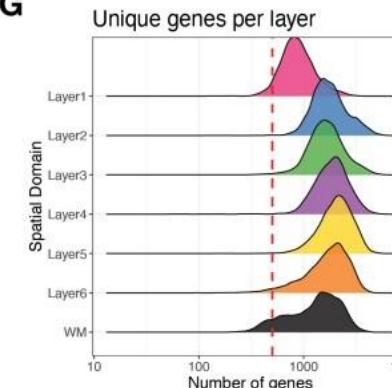
E



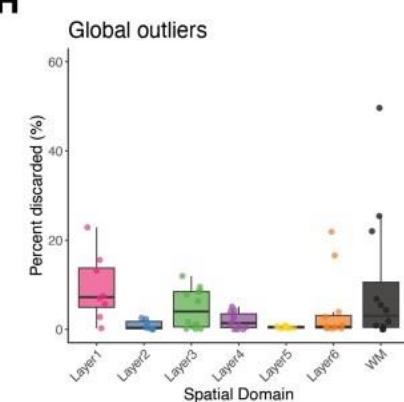
F



G

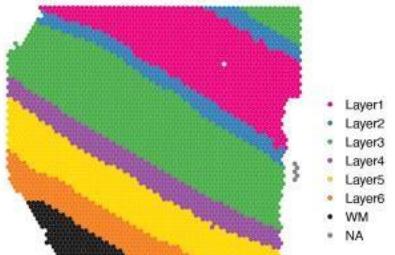


H

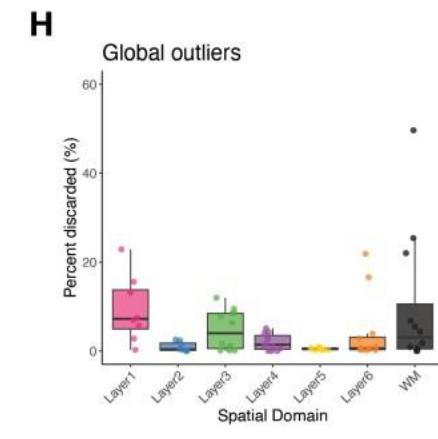
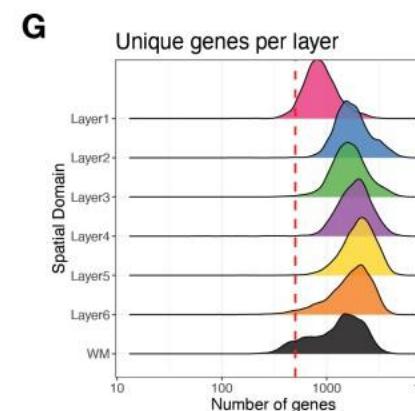
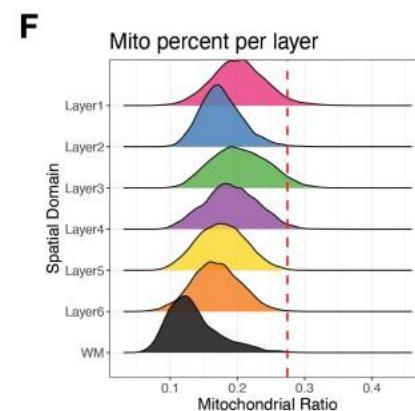
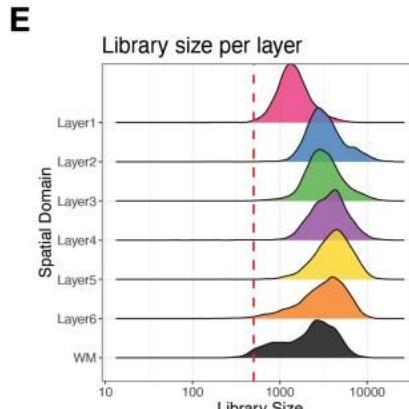


# SpotSweeper (spot-level artifacts)

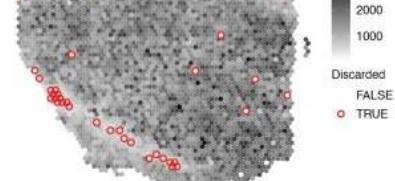
Global Annotation



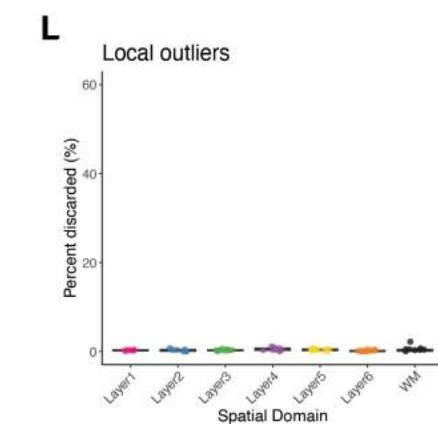
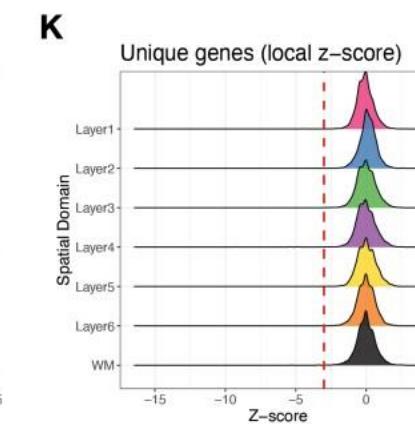
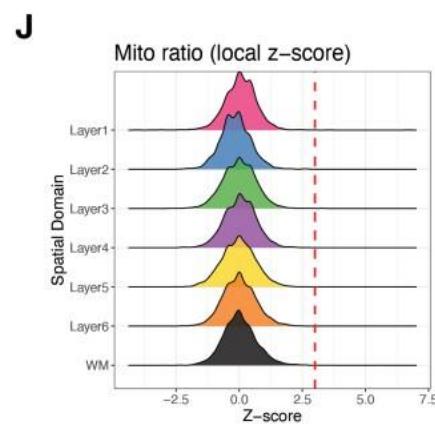
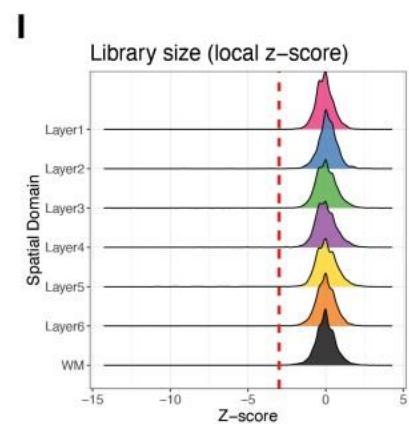
## Global outliers



Local Annotation



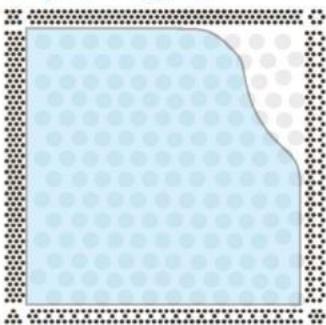
## Local outliers



# SpotSweeper (region-level artifacts)

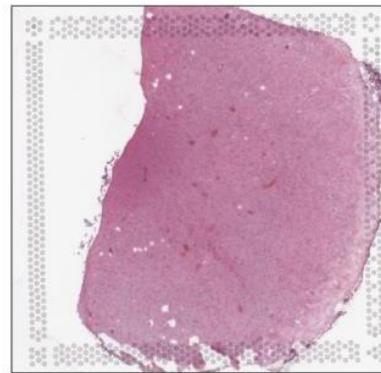
Incomplete coverage of Visium array

Liquid reagent

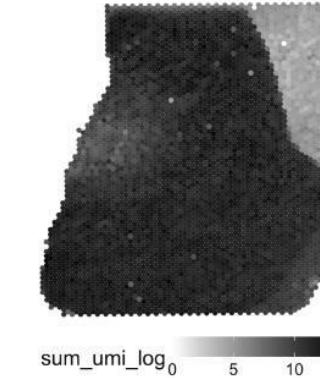


Dry spots result in  
smaller library size  
and fewer genes  
detected

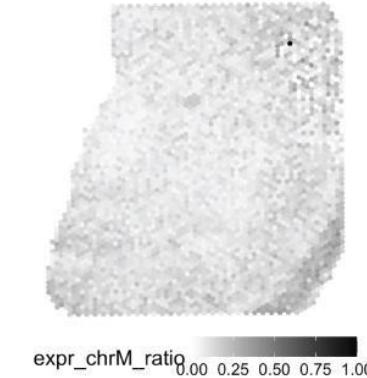
Br3942\_mid



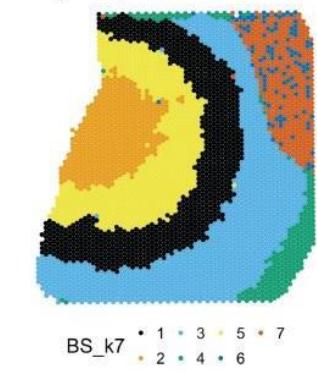
Library size



Mito Ratio



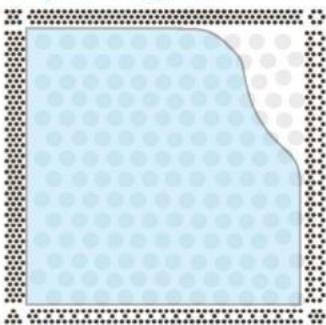
BayesSpace k=7



# SpotSweeper (region-level artifacts)

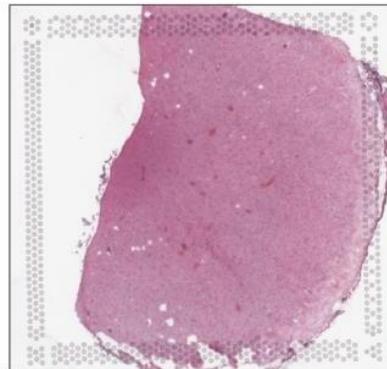
## Incomplete coverage of Visium array

Liquid reagent

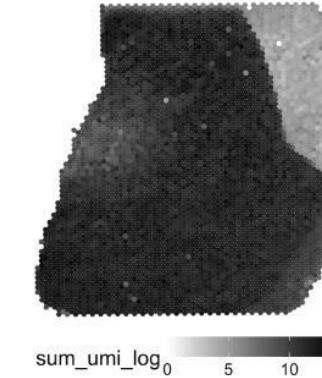


Dry spots result in smaller library size and fewer genes detected

Br3942\_mid



Library size



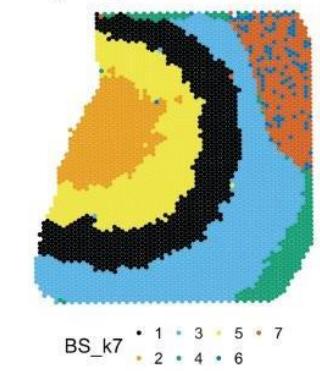
sum\_umi\_log<sub>0</sub> 5 10

Mito Ratio



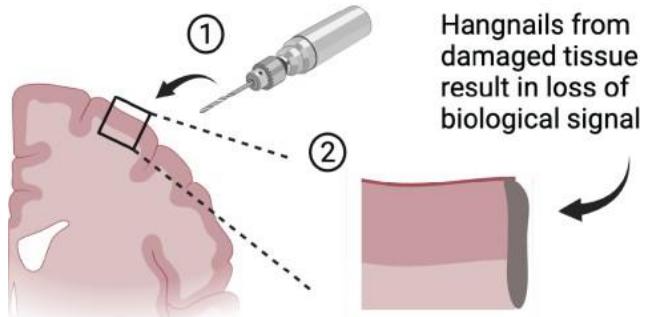
expr\_chrM\_ratio 0.00 0.25 0.50 0.75 1.00

BayesSpace k=7



BS\_k7 • 1 • 3 • 5 • 7  
• 2 • 4 • 6

## Tissue damage during dissection

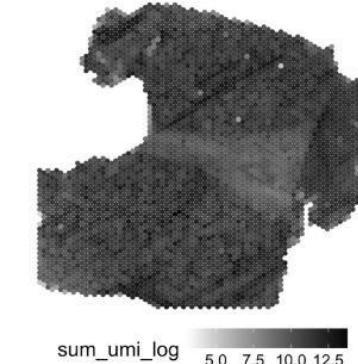


Hangnails from damaged tissue result in loss of biological signal

Br8325\_ant

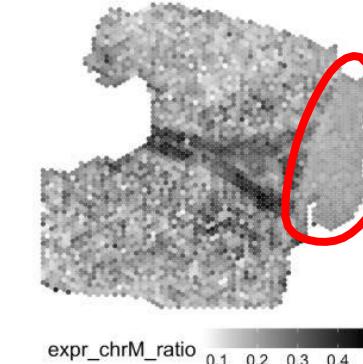


Library size



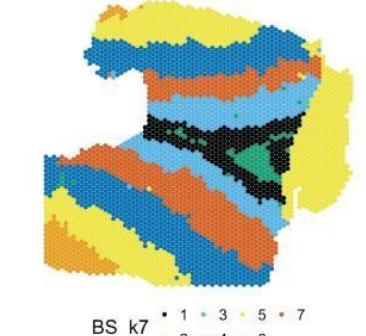
sum\_umi\_log 5.0 7.5 10.0 12.5

Mito Ratio



expr\_chrM\_ratio 0.1 0.2 0.3 0.4

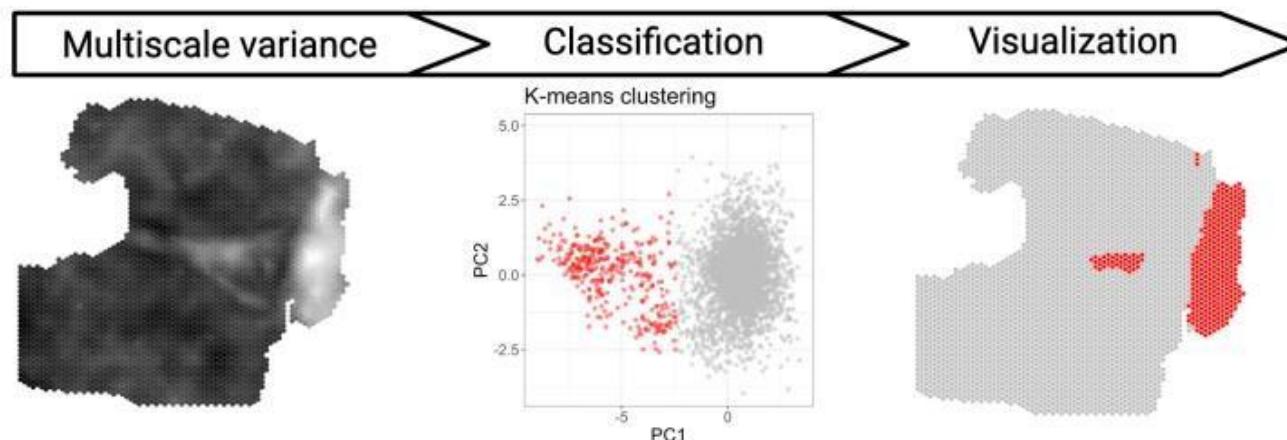
BayesSpace k=7



BS\_k7 • 1 • 3 • 5 • 7  
• 2 • 4 • 6

# SpotSweeper (region-level artifacts)

1. The k-NN for each spot are identified based on the spatial coordinates
2. For each neighborhood size (i.e., scale), local variance of the mitochondrial ratio is calculated and adjusted for a mean-variance relationship using linear regression
3. Mean-corrected local variance = the residuals of the linear regression
4. Perform PCA on the mean-corrected local variances of all neighborhood sizes
5. Apply k-means clustering ( $k=2$ ) in the first two PCs to identify regional artifacts compared to high-quality tissue



# Preserve biological variability while removing technical artifacts: [SpotSweeper](#)

Detects **spatially coherent low-quality regions** in Visium HD slides.

Uses **per-spot QC metrics** (mainly mitochondrial fraction) and **spatial neighborhood info**.

Flags artifacts for downstream masking/removal.

Works across **multiple neighborhood scales** to detect both small and large defects.

**How do you normalize your single-cell data?**

# Log-normalization

$$Y_{ij} = \log_e\left(\frac{X_{ij}}{\sigma_i X_{ij}} + 1\right)$$

- Simplest and most commonly-used normalization strategy
- Divide all counts for each spot by a spot-specific scaling factor (i.e. size factor)
- Assumes that any cell-specific bias (e.g., in capture or amplification efficiency) affects all genes equally via scaling of the expected mean count for that cell

**How about any other method?**

# Can we use scTransform?

Bhuva et al. *Genome Biology* (2024) 25:99  
<https://doi.org/10.1186/s13059-024-03241-7>

Genome Biology

SHORT REPORT

Open Access



## Library size confounds biology in spatial transcriptomics data

Dharmesh D. Bhuva<sup>1,2,3\*</sup> , Chin Wee Tan<sup>2,4</sup>, Agus Salim<sup>2,5</sup>, Claire Marceaux<sup>3,6</sup>, Marie A. Pickering<sup>7</sup>, Jinjin Chen<sup>2,3</sup>, Malvika Kharbanda<sup>1,2,3</sup>, Xinyi Jin<sup>2,3</sup>, Ning Liu<sup>1,2,3</sup>, Kristen Feher<sup>1,2,3</sup>, Giovanna Putri<sup>2,3</sup>, Wayne D. Tilley<sup>7</sup>, Theresa E. Hickey<sup>7</sup>, Marie-Liesse Asselin-Labat<sup>3,6</sup>, Belinda Phipson<sup>2,3†</sup> and Melissa J. Davis<sup>1,2,3,4,8†</sup>

- Tested the effects of normalization on spatial domain identification

Though sctransform removes library size effects effectively, their confounding with biology results in removal of biological effects as well.

See also for imaging-based SRT...

Atta et al. *Genome Biology* (2024) 25:153  
<https://doi.org/10.1186/s13059-024-03303-w>

Genome Biology

RESEARCH

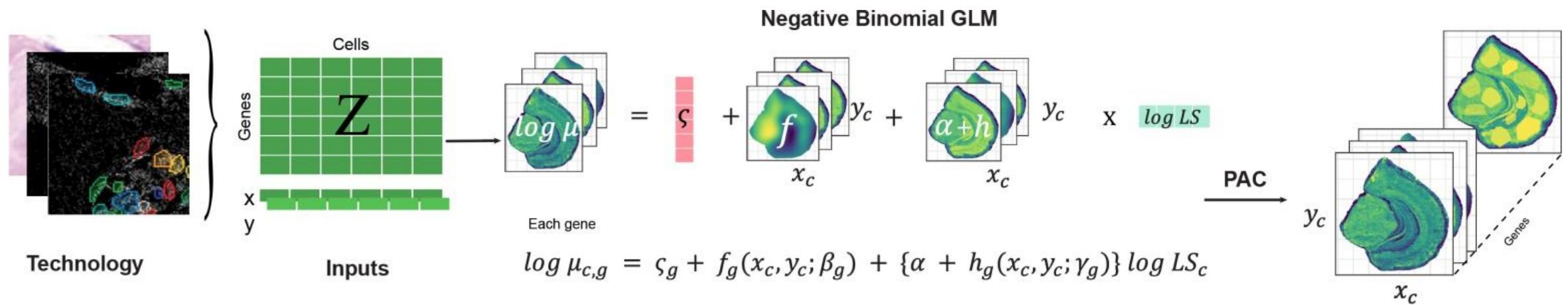
Open Access

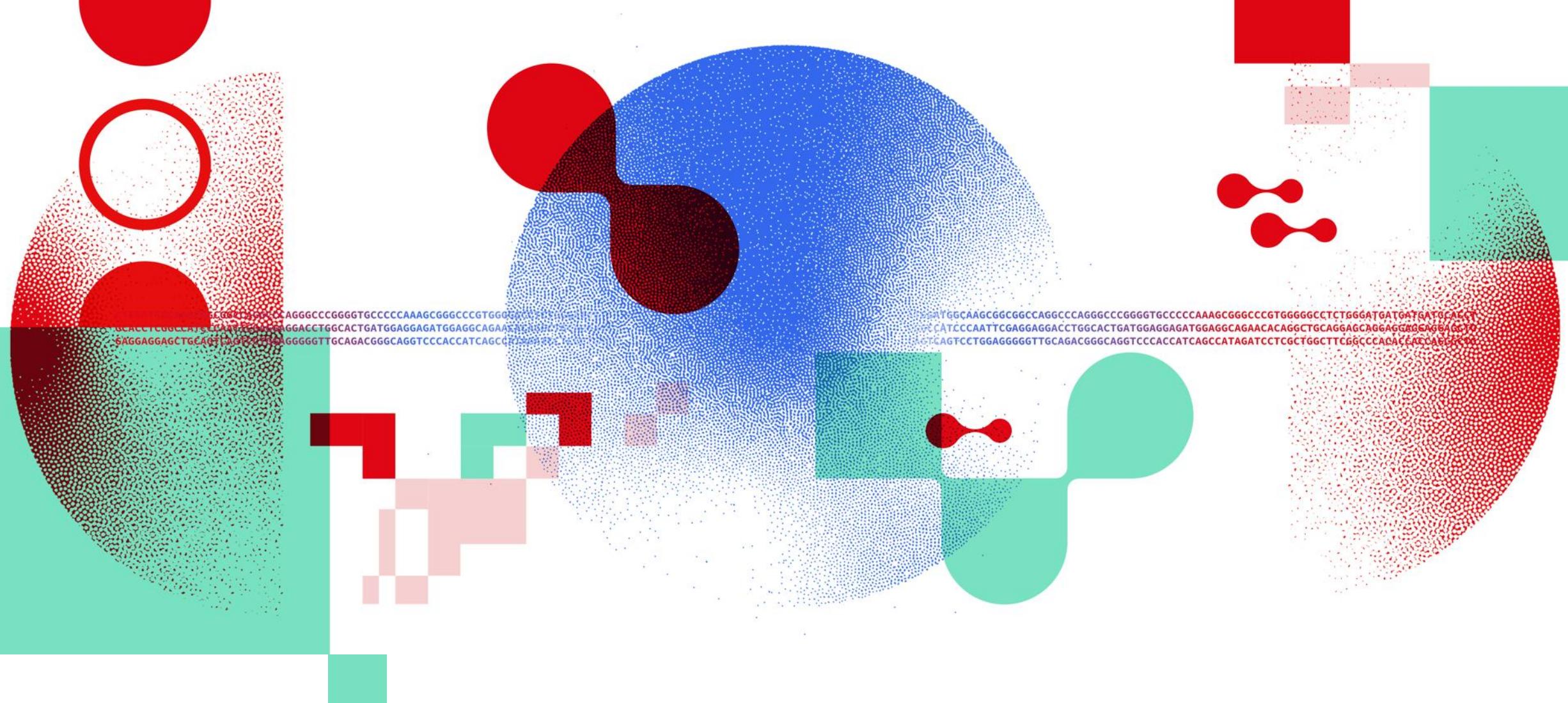


## Gene count normalization in single-cell imaging-based spatially resolved transcriptomics

Lyla Atta<sup>1,2</sup>, Kalen Clifton<sup>1,2</sup>, Manjari Anant<sup>2,3</sup>, Gohta Aihara<sup>1,2</sup> and Jean Fan<sup>1,2\*</sup>

# SpaNorm





# Thank you

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