



Swiss Institute of
Bioinformatics

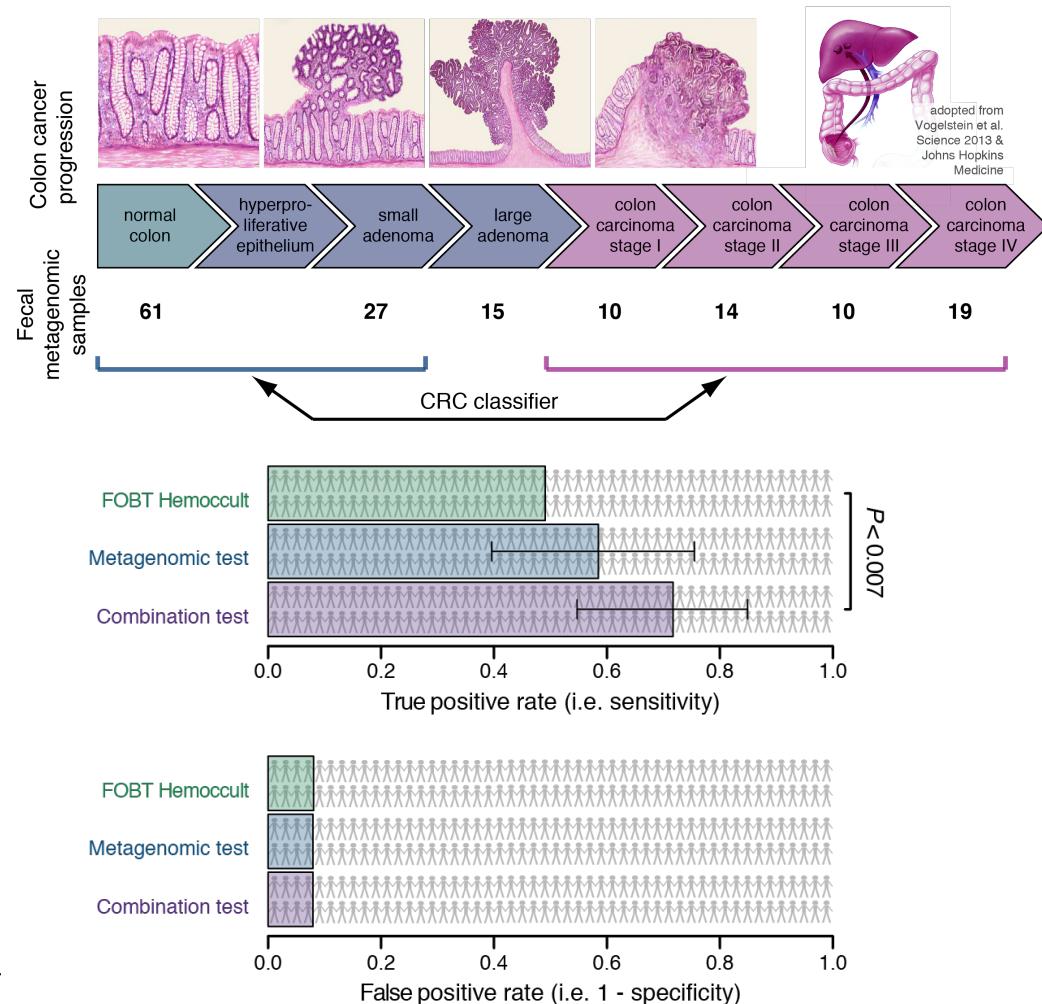
Machine learning / statistical modelling of metagenomic data

Project 3

Spring School Bioinformatics and
computational approaches in
Microbiology

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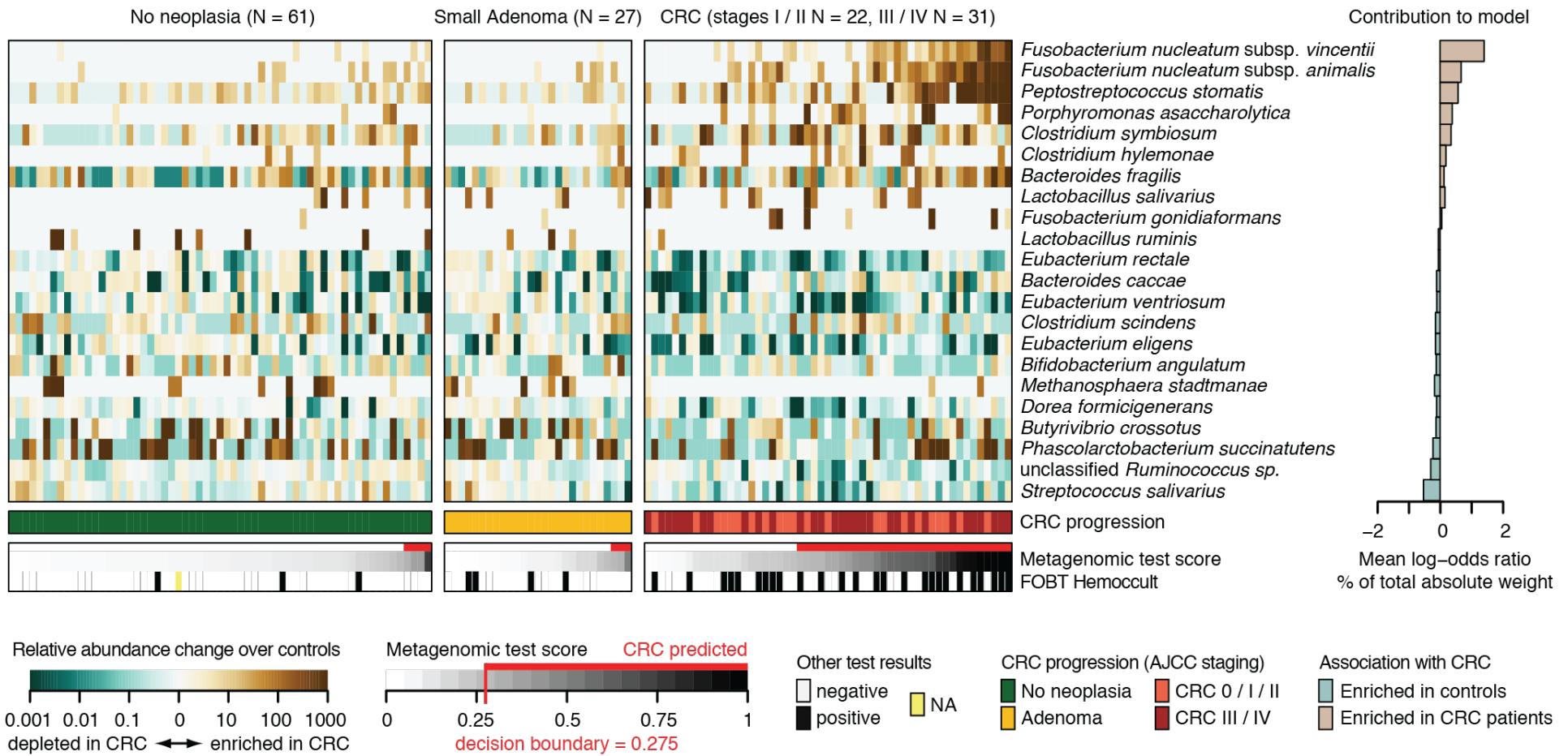
Colorectal cancer example (continued)



- Collected stool samples from 46 colorectal cancer (CRC) patients and 60 healthy controls
- Used metagenomic sequencing and profiled gut bacterial species
- Can microbiome differences be used for non-invasive detection of cancer?
- How does metagenomic detection compare to standard noninvasive diagnostic test (FOBT)?

[Zeller*, Tap*, Voigt* et al., *Mol. Syst. Biol.* 2014]

A microbiome “signature” of colorectal cancer



Descriptive statistics versus statistical modeling

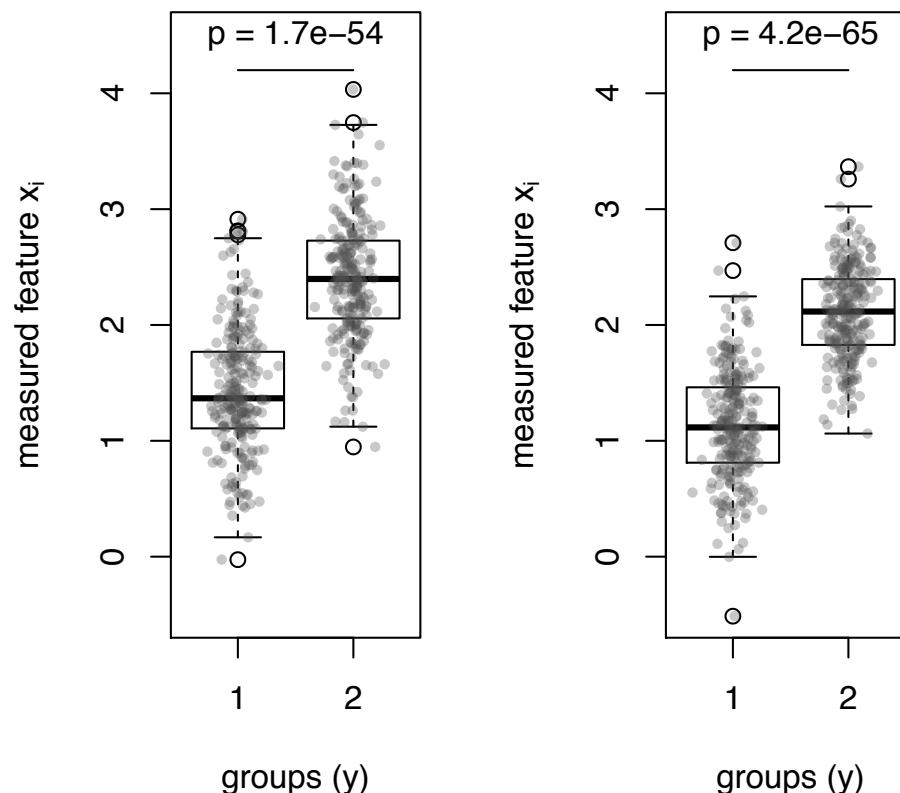
- **Hypothesis testing:**

Could the observed difference also be observed by chance?

- **Modeling:**

Given only the measurement, can we tell which group the measurement corresponds to?

- Recall that ***P*-values depend on both effect size and sample size!**



Why statistical modelling / machine learning?

- Modeling ideally **extracts the essence** of a biological phenomenon
- Model needed to **make predictions on new data**
(necessary e.g. for microbiome-based diagnostics)
- **Prediction accuracy** is often a more **meaningful measure of association** than statistical significance of differences
- Suitable methods can **select predictive taxa** (and ignore others)
- **Sparse statistical models** are based on only „few“ taxa,
therefore useful for microbiome **biomarker / signature extraction**

$$y_i = f(\mathbf{x}_i) + \epsilon$$

For i samples / patients

y_i – label (e.g. disease or control), always binary herein
 x_i – features (e.g. species abundance profile, a vector)

f – our model

ϵ – modeling error

Introduction to notation and input data format

- **Feature** data **X** (also observations, predictors):

$n \times p$ matrix x_{ij}

species/gene abundances in rows (i),

samples/patients in columns (j)

observations based on which we wish to make predictions

x_i denotes the feature vector, i.e. abundance profile, for the i-th sample

- **Label** data **y** (also dependent variable, response):

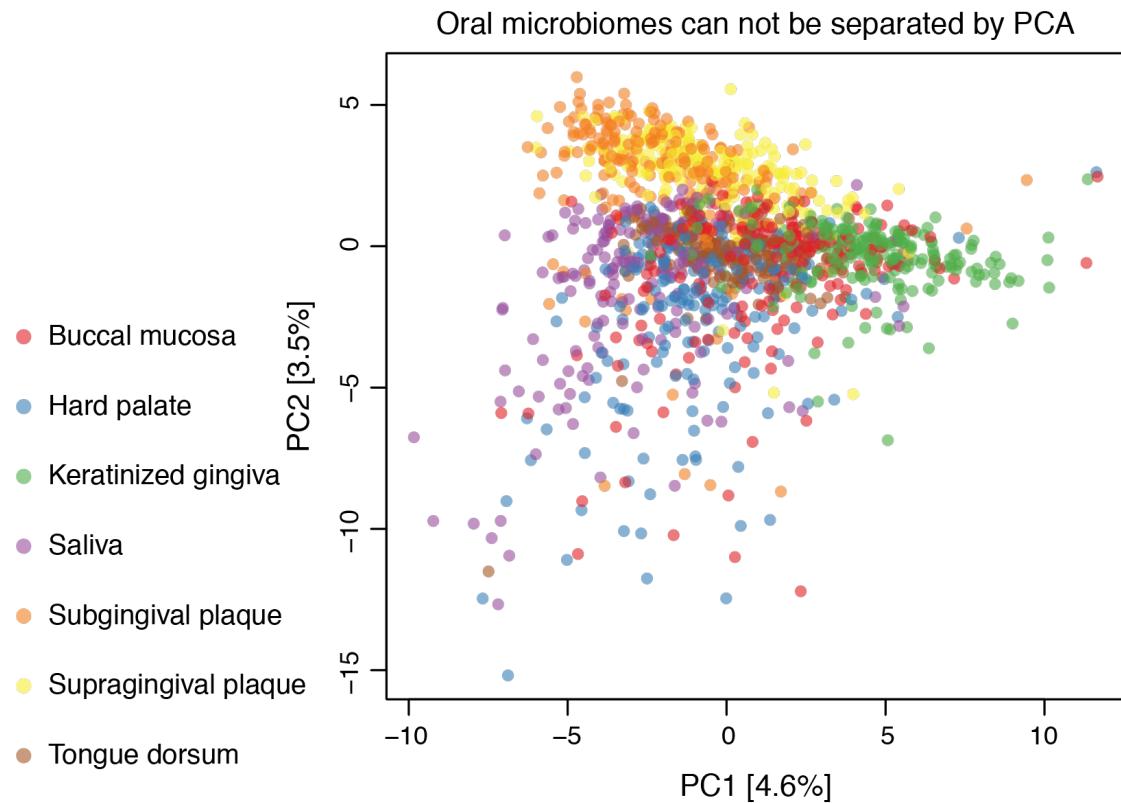
vector of length n, containing binary values in our cases

the phenomenon which we wish to predict:

disease vs. healthy, response vs. non-response etc.

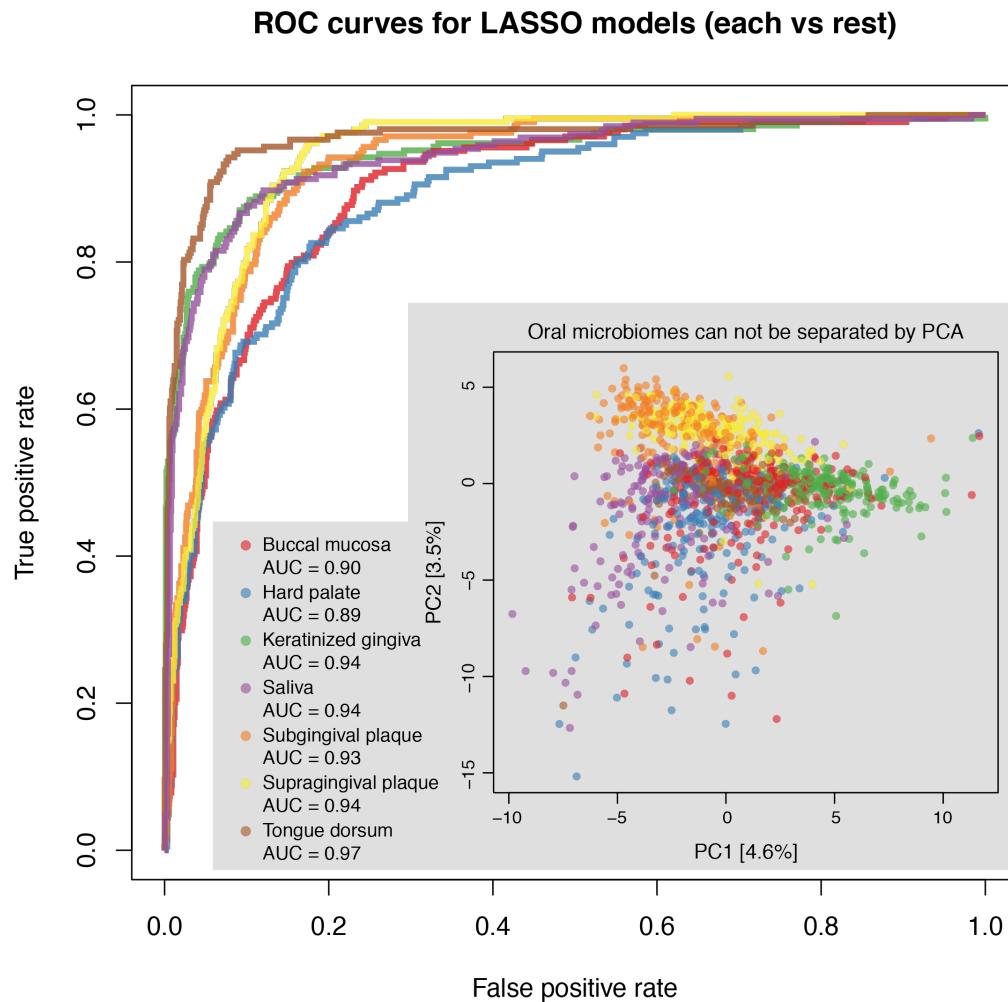
Ordination versus modelling (I)

- Using PCoA (with various dissimilarity measures), it is difficult to resolve for each oral microbiome sample the precise sampling site.



Ordination versus modelling (I)

- Using PCoA (with various dissimilarity measures), it is difficult to resolve for each oral microbiome sample the precise sampling site.
- Statistical models, in contrast, can very accurately recognize sample origin.



A typical machine learning workflow

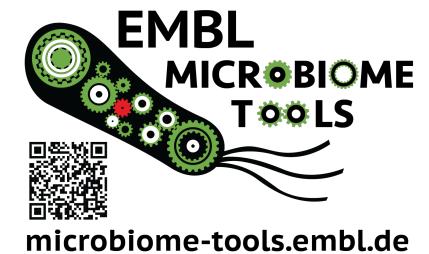


Starting with SIAMCAT

```
> source("https://bioconductor.org/biocLite.R")
> biocLite("SIAMCAT")
> browseVignettes("SIAMCAT")
```

File formats supported:

- phyloseq
- BIOM
- LEfSe
- MaAsLin
- metagenomeSeq



This workflow is implemented in the SIAMCAT Bioconductor package, which we will explore in detail in the practical.

What to use as input (features)?

- Use your **domain expertise** to engineer features that are likely predictive of the phenomenon of interest – microbiome examples:
 - Species abundances (or higher / lower resolution taxonomic profiles)
 - Metabolic pathway abundance (e.g. KEGG / CAZy maps)
 - Functional gene annotations (GO terms, domains, ...)
 - Orthologous gene families (COGs, eggNOG families, ...)
 - Toxins, virulence factors, ABX resistance genes, ...
- Consider **interpretability** –
predictive species/metabolic pathways may be preferred over k-mers or log-ratios
- Importantly, do **NOT use the label** information for selecting features for modeling
(more on this later)

Model evaluation (classification)

In many applications, classes aren't equal – neither are errors!

		True condition	
		positive ("cancer")	negative ("healthy")
Predicted condition	positive ("predicted to have cancer")	True positives TP	False positives FP (Type I errors)
	negative ("predicted not to have cancer")	False negatives FN (Type II errors)	True negatives TN

True positive rate (TPR, **sensitivity, recall**)

True negative rate (TNR, **specificity**)

False positive rate (FPR, $1 - \text{specificity}$)

- are all **independent of prevalence**
(fraction of positives in the population)

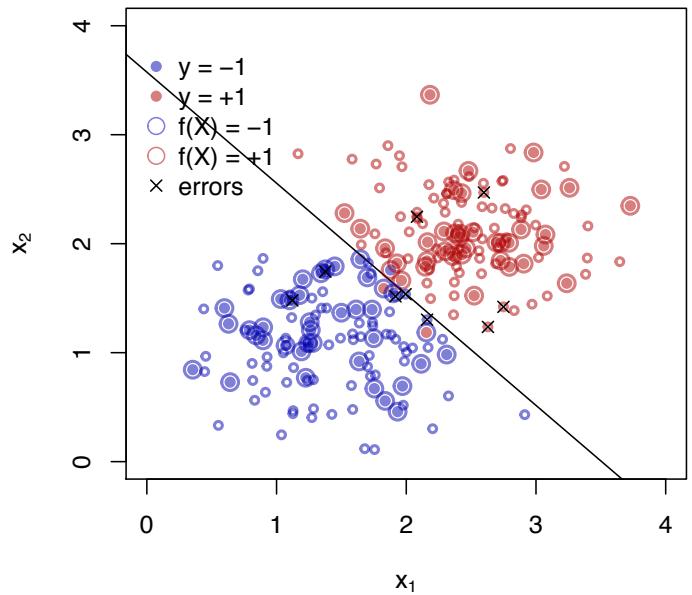
Precision (positive pred. value, PPV)

False discovery rate (FDR, $1 - \text{precision}$)

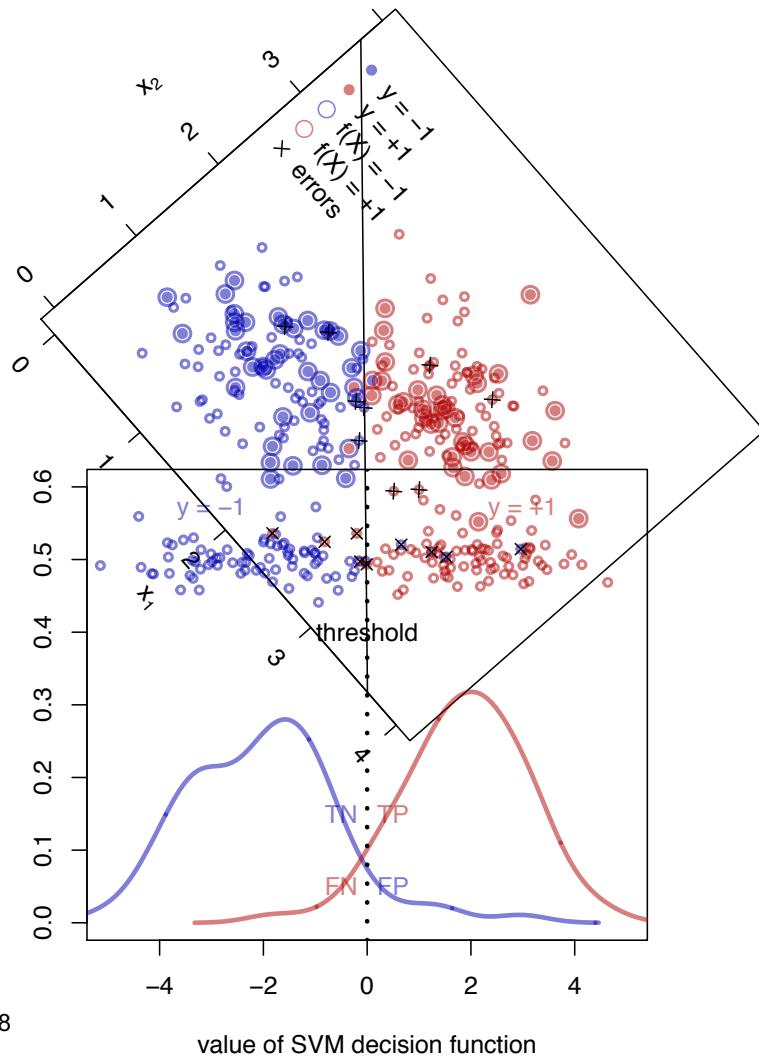
- are both **dependent on prevalence**
(fraction of positives in the population)

[these and more measures on en.wikipedia.org/wiki/Evaluation_of_binary_classifiers]

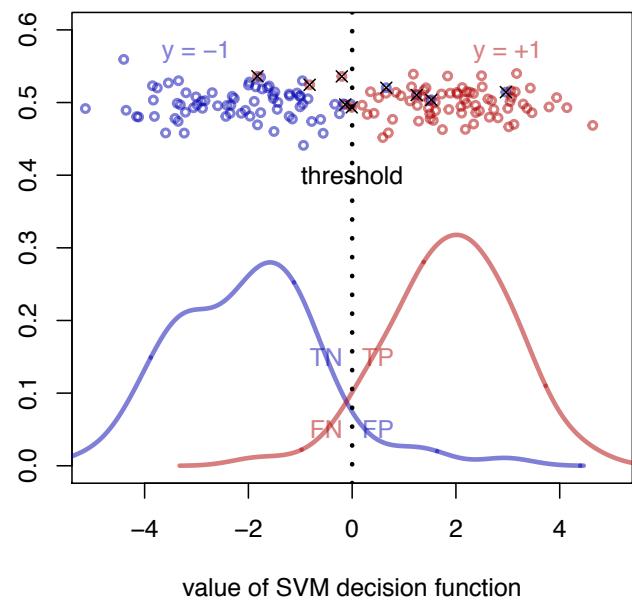
Model evaluation II – ROC curves



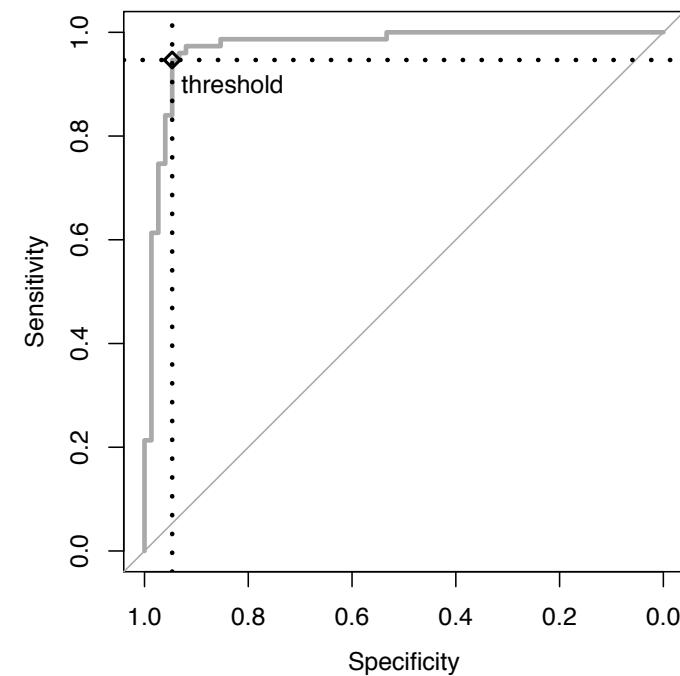
Model evaluation II – ROC curves



Model evaluation II – ROC curves



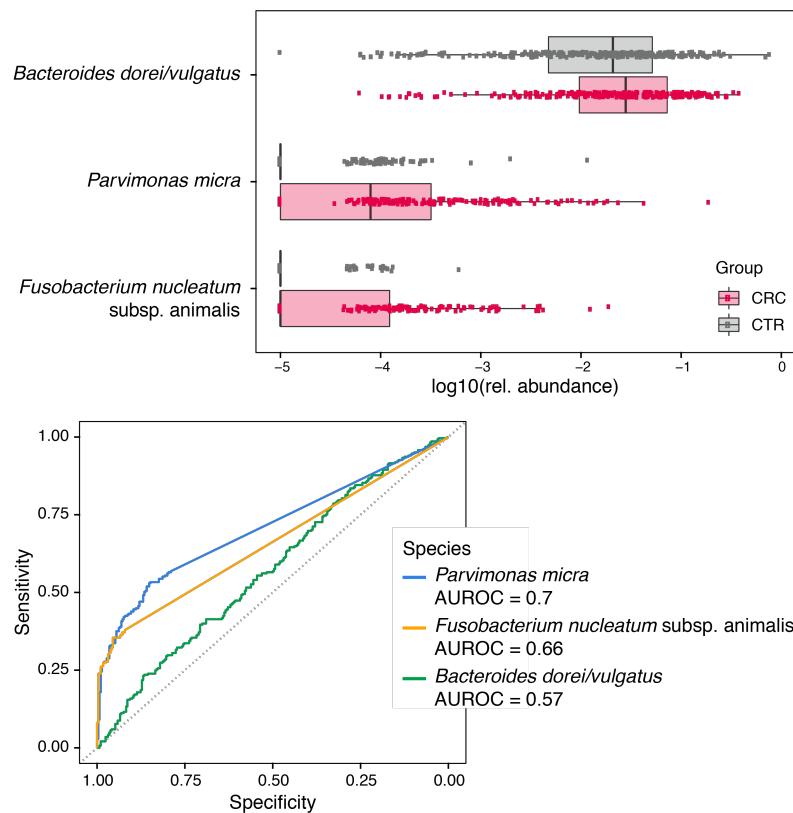
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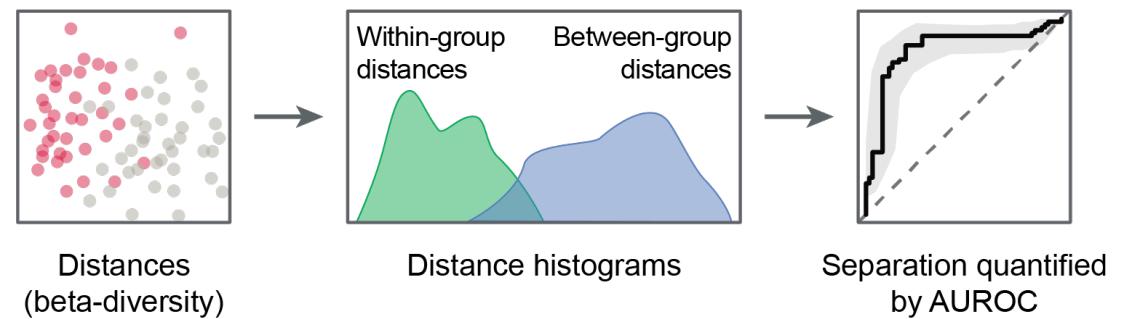
- Change decision threshold to obtain other **trade-offs between sensitivity and specificity**
- Receiver operating characteristic (ROC) curve plots all of them
- **Area under the ROC curve as a summary statistic**

ROC curves from single features / distances

- Enrichment of a species in disease group can be directly quantified using ROC curves (disease biomarker).

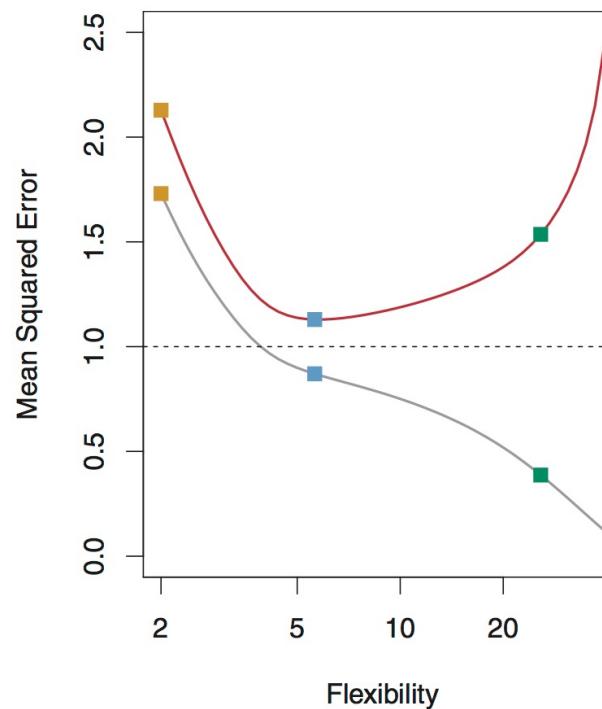
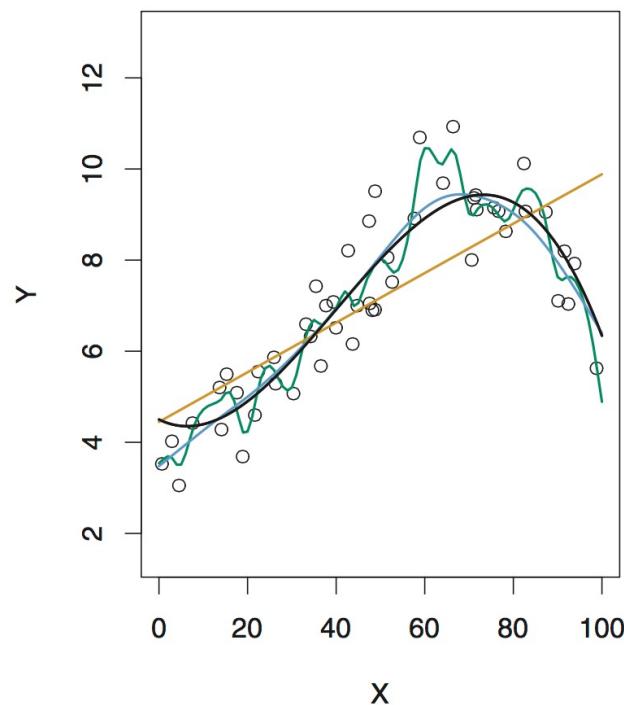


- Separation between groups in terms of pairwise dissimilarities can also be assessed using ROC curves.



Model evaluation III – assessing generalization

- What might seem a good idea at first: Minimizing the **training error**...
But with increasing flexibility, models will fit the training data better and better.
- Better: maximize **generalization** to new data sets...
Since **overfitting** the training data will result in poor generalization (i.e. large **test error**)



Here for illustration,
smoothing splines are
used where model
flexibility / complexity
increases with the
degree of the
polynomials.

[James, Witten, Hastie &
Tibshirani, Springer 2013]

Resampling data for external validation or cross validation

Some data needs to be reserved for model evaluation....

Resampling data for external validation or cross validation

Some data – **always!** – needs to be reserved for model evaluation....

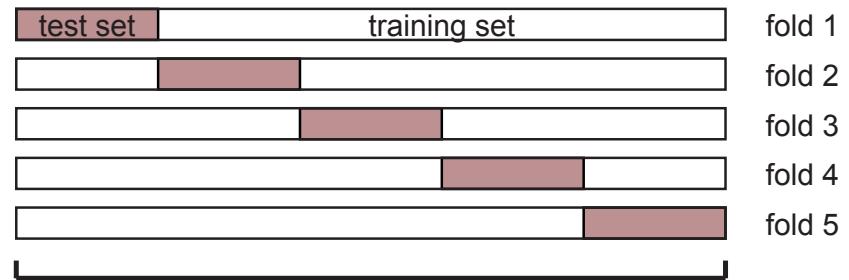
- Validation on external data



total number of samples (split into 2 subsets)

- Train model on training set
- Test on test set
- Assess error on test predictions

- Cross-validation (CV)



total number of samples (split into 5 subsets)

- For each CV fold:
 - Train a model on training set
 - Predict on the test set
- Either concatenate or average predictions from (all) test sets to estimate error
- More efficient use of (training) data

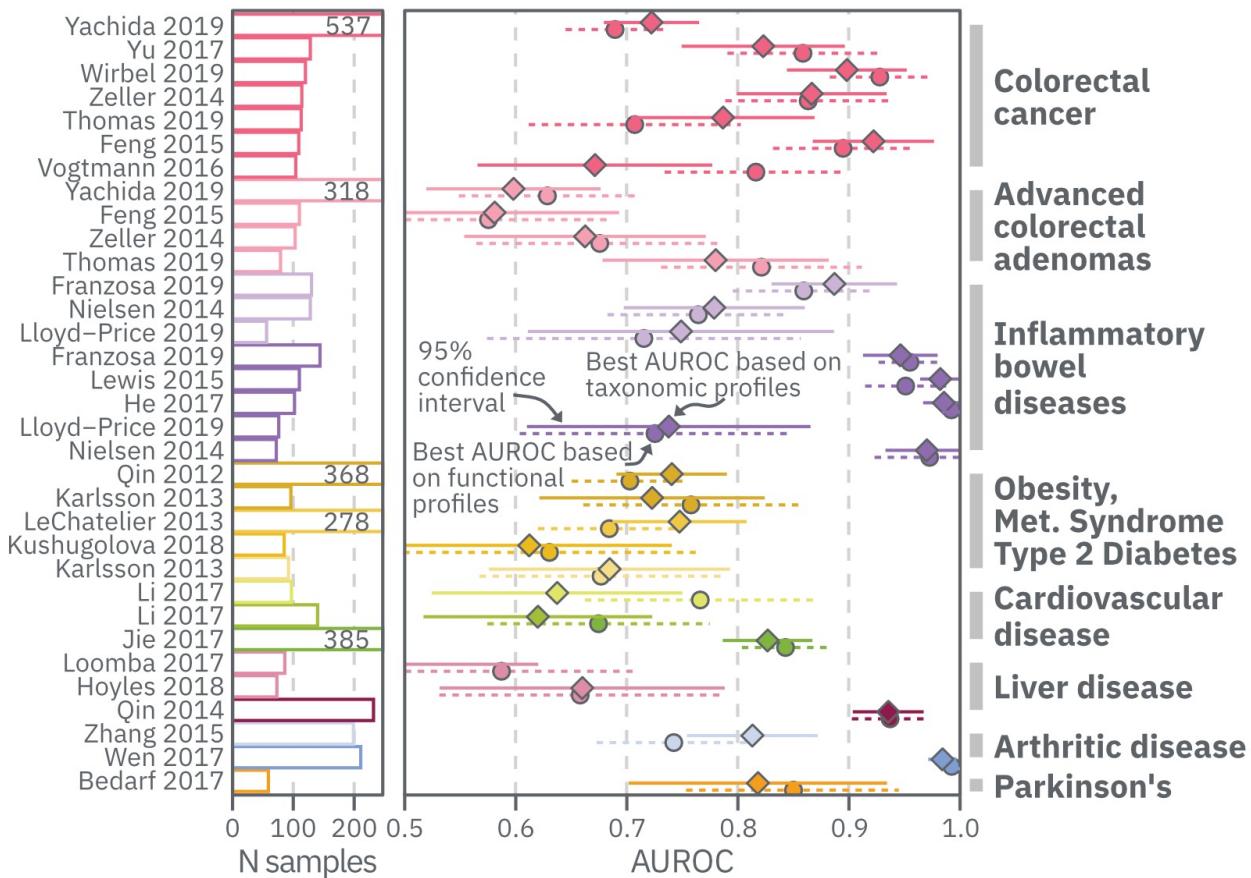
Cross-validation pitfalls II

- **Cross validation works under the i.i.d. assumption** (observations have the same probability distribution and are mutually independent)
 - E.g. a series of (fair or unfair) coin flips is i.i.d. as the next flip doesn't depend on the previous ones.
- However, biological samples are **rarely completely independent**:
 - Multiple time-point measurements from the same subject or related subjects
 - Spatial structure / dependencies between measurements
- Data (sets) are **not always identically distributed**
 - Batch effects: e.g. experiments or diagnostic tests performed in different labs (by different technicians, at different times, using different reagent lots, ...) may exhibit (subtle) distributional shifts

Take home messages

- **Model fitting is easy, model evaluation is not at all!**
Understand the generalization assessed – consult experts!
- Beware of **overfitting** – especially on small data sets, especially with complex algorithms!
Typically $N > 50$, better > 100 per group is a requirement; start with simple algorithms first
- **Trade off interpretability** (white-box models) **and maximal prediction accuracy** wisely!
- Diagnostic application is relatively straightforward, but underlying **mechanisms are generally difficult to glean** from models (predictability does NOT imply causality!)

Outlook – disease classification using SIAMCAT



www.siamcat.embl.de

[Wirbel et al., *Genome Biol.* 2020]