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Lecture 1.3

# **Phylogenetic Data**

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# Phylogenetic data

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## 1. **Data preparation**

- Taxon and gene sampling
- Sequence alignment (if needed)
- Data filtering

## 2. **Phylogenetic inference**

- Model selection
- Estimation of tree
- Further analysis and interpretation

# Phylogenetic data

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- **Select data to optimise signal:noise**
  - Slowly evolving markers for deep evolutionary events
  - Rapidly evolving markers for recent evolutionary events
- **Homoplasy**
  - Taxa share similarities that do not reflect evolutionary history
- **Take advantage of existing resources**



# Data types

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- **Sequence data**
  - Nucleotides
  - Amino acids
- **Binary data** (presence/absence of genomic features)
- **Microsatellites** (repeat numbers)
- **Single-nucleotide polymorphisms (SNPs)**
- **Reduced-representation sequences**

# Morphological data

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- Morphological characters from extant and extinct taxa

## Current Biology

Volume 25, Issue 19, 5 October 2015, Pages R922–R929

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Review

### Morphological Phylogenetics in the Genomic Age

Michael S.Y. Lee<sup>1, 2</sup>,  , Alessandro Palci<sup>1, 2</sup>

# Sequence data

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- **Coding sequences**
  - Ribosomal RNA
  - Protein-coding genes
- **Non-coding sequences**
  - Intergenic sites
  - Introns
- **Amino acid sequences**



# Sequence data

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non-coding region

bat	CGTTAG--CATGAGAGAACCCCTACTCTAGG
whale	CGATAG--TCATGAGAGAACCCCTACTCTAGG
rabbit	CGTTAG--TTATGAGGGAATCCTACCCTAGG
elephant	CA--GGTTTATGAGGCATTCC---TCTAGG
kangaroo	CA--GGT--ATGAGGCATTCC-----ATG

Diagram illustrating a DNA microarray layout with four rows of DNA sequences. Each row is associated with a header sequence: M R E P Y S R. The sequences are color-coded to match the header letters.

M	R	E	P	Y	S	R
ATGAGAGAGAGAACCCCTACTCTAGG						
ATGAGAGAGAGAACCCCTACTCTAGG						
ATGAGGGGAATCCTACCCCTAGG						
ATGAGGCGATTCC---TCTAGG						

CGTTAG--CATGAGAGAACCTACTCTAGG

CGATAG-TCATGAGAGAACCCCTACTCTAGG

CGTTAG-TTATGAGGGAATCCTACCCCTAGG

CA--GGTTTATGAGGCATTCC---TCTAGG
















CA--GGT--ATGAGGCATTCC-----ATG

nonsynonymous



# Data partitioning

- Sites evolve at different rates
- Separate substitution model for each gene and codon position?

	Gene A	Gene B	Gene C
bat			
whale			
rabbit			
elephant			
kangaroo			

- **Biological**

- Genome
- Genes
- Codon positions
- RNA stems vs loops
- Hydrophobic vs hydrophilic

- **Statistical**

# PartitionFinder

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- Too many possible partitioning schemes
  - 15 schemes for 4 genes
  - 52 schemes for 5 genes
  - 203 schemes for 6 genes

## **PartitionFinder 2: New Methods for Selecting Partitioned Models of Evolution for Molecular and Morphological Phylogenetic Analyses** □

[Robert Lanfear](#) □, [Paul B. Frandsen](#), [April M. Wright](#), [Tereza Senfeld](#), [Brett Calcott](#)

*Molecular Biology and Evolution*, Volume 34, Issue 3, March 2017, Pages 772–773,

# Gaps and missing data

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- **Delete sites with any missing data**
  - Potential loss of informative data
  - Problematic in analyses of data supermatrices
- **Treat gaps as unresolved data**
  - Gap is simultaneously A, C, G, and T
  - Most common approach
- **Code gaps as binary characters**

# Gaps and missing data

- Impact of missing data remains poorly understood
- Filter data according to chosen threshold of missing data

	Gene 1	Gene 2	Gene 3	Gene 4	Gene 5	
Taxon 1						Maximise gene sampling
Taxon 2						
Taxon 3						
Taxon 4						Maximise taxon sampling
Taxon 5						
Taxon 6						

# Mutational saturation

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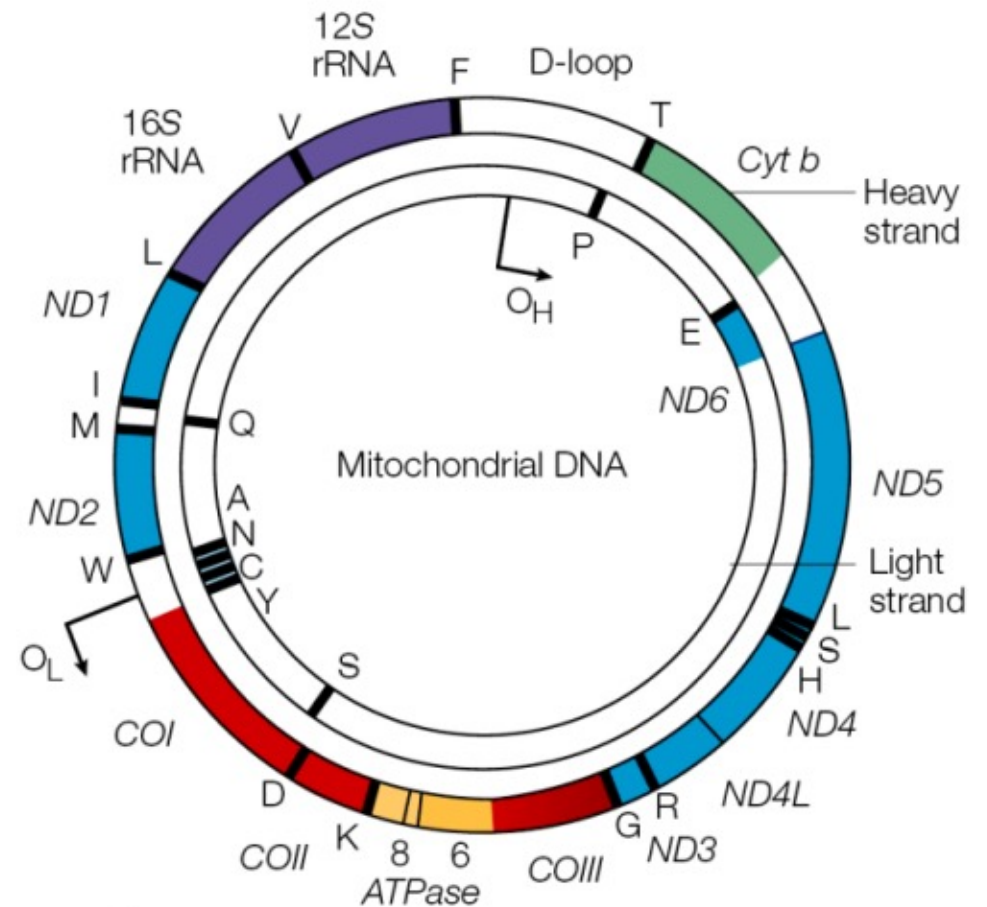
- Some sites can evolve very rapidly
  - 3rd codon positions
  - Loop regions in RNA
- Multiple hits can erode phylogenetic signal
- Various ways of testing for saturation (e.g., Xia's test in DAMBE, PhyloMAd)

Saturated sites can be removed to improve signal:noise

# High-Throughput Data

# Mitochondrial genomes

- Maternally inherited
- Protein-coding genes (e.g., *COI*)
- RNA genes (e.g., *12S*, *16S*)
- Control region



# Single-nucleotide polymorphisms

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- Single sites sampled from throughout the genome
- More common in intraspecific (population) studies
- Issues to consider:
  - **Recombination**  
SNPs are usually unlinked so they are likely to have different (gene) trees
  - **Ascertainment bias**  
SNPs are selected for variability and this can mislead estimates of population sizes, rates, and other parameters



# Reduced-representation sequences

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- Markers identified by cutting genome with restriction enzymes
- Process creates binary data and short sequences
- Examples include RADseq and DArTseq
- Issues to consider:
  - **Recombination**  
Markers are usually unlinked so they are likely to have different (gene) trees
  - **Missing data**  
Typically a large proportion of missing data



# Transcriptomes and exon capture

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- Large panels of protein-coding loci
- Sequences are easier to align
- Good for inferring deep relationships
- Issues to consider:
  - **Variability**  
Might not be much variation at the population level
  - **Selection**  
Differences in selection will lead to rate differences across exons

# Whole-genome sequencing

- Typically NOT (yet) the entire genome
- Many challenges: Jarvis et al *Science* 2014 >400 years of computing using a single processor
- **Issues to consider**
  - Single-copy genes
  - Selectively neutral
  - Unlinked loci



# Useful references

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