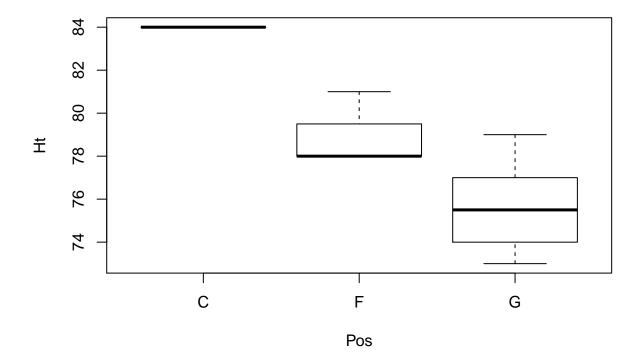
STAT 578 - Advanced Bayesian Modeling - Fall 2019 Assignment 6

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Solution for Problem 1

```
perf_data = read.csv("illinimensbb.csv", header=TRUE)
plot(Ht ~ Pos, data= perf_data)
```



By checking the plot, we do see height and position are highly correlated. *center* has highest mean of height, forward has shortest mean of height and forward has in between these two. Their value ranges also don't seem to cross each other significantly.

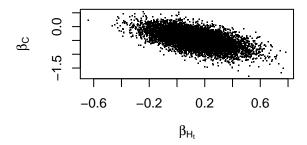
Solution for Problem 2

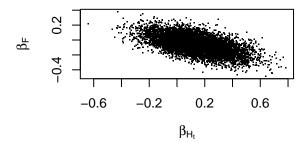
(a)

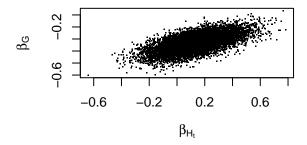
```
model {
   for (i in 1:length(FGM)) {
     FGM[i] ~ dbin(prob[i], FGA[i])
```

```
logit(prob[i]) <- beta_pos[Pos[i]] + beta_ht * Ht_Scaled[i]</pre>
        FGM_rep[i] ~ dbin(prob[i], FGA[i])
    }
    for (j in 1:max(Pos)) {
        beta_pos[j] ~ dt(0, 0.01, 1)
    beta ht \sim dt(0, 0.16, 1)
}
library(rjags)
df_jags_1 <- list( FGM = perf_data$FGM, FGA = perf_data$FGA,</pre>
                   Pos = unclass(perf_data$Pos),
                   Ht_Scaled = as.vector(scale(perf_data$Ht, scale=2*sd(perf_data$Ht))))
initial_vals_1 <- list(list(beta_pos = c(10,10,10), beta_ht=10),</pre>
                        list(beta pos = c(10, 10, -10), beta ht=-10),
                        list(beta_pos = c(10, -10, 10), beta_ht=-10),
                        list(beta pos = c(10,-10,-10), beta ht=10))
model_1 <- jags.model("perf_1.bug", df_jags_1, initial_vals_1, n.chains = 4,</pre>
                       n.adapt = 1000)
update(model_1, 1000)
#Need only check top-level parameters (in the DAG) for convergence.
x1 <- coda.samples(model_1, c("beta_pos", "beta_ht"), n.iter = 2000)</pre>
gelman.diag(x1, autoburnin=FALSE)
## Potential scale reduction factors:
##
##
               Point est. Upper C.I.
## beta ht
                        1
                                 1.01
## beta_pos[1]
                        1
                                 1.01
## beta_pos[2]
                        1
                                 1.01
## beta_pos[3]
                        1
                                 1.01
## Multivariate psrf
##
## 1
coef_sample_1 <- coda.samples(model_1, c("beta_pos","beta_ht","prob","FGM_rep"),</pre>
                               n.iter = 10000, thin = 5)
effectiveSize(coef_sample_1[,c("beta_pos[1]", "beta_pos[2]", "beta_pos[3]", "beta_ht")])
## beta_pos[1] beta_pos[2] beta_pos[3]
                                             beta_ht
##
      6051.941
                  6250.811
                               5725.131
                                           4706.478
(b)
summary(coef_sample_1[, c("beta_pos[1]", "beta_pos[2]", "beta_pos[3]", "beta_ht")])
##
```

```
## Iterations = 4005:14000
## Thinning interval = 5
## Number of chains = 4
## Sample size per chain = 2000
## 1. Empirical mean and standard deviation for each variable,
     plus standard error of the mean:
##
                            SD Naive SE Time-series SE
##
                   Mean
## beta_pos[1] -0.45744 0.2923 0.0032677
                                              0.0037629
## beta_pos[2] -0.06157 0.1115 0.0012462
                                              0.0014130
## beta_pos[3] -0.33432 0.0708 0.0007915
                                              0.0009416
## beta ht
               0.13828 0.1804 0.0020175
                                              0.0026321
##
## 2. Quantiles for each variable:
##
##
                  2.5%
                                    50%
                                             75%
                            25%
                                                   97.5%
## beta_pos[1] -1.0332 -0.64850 -0.4573 -0.25674 0.1085
## beta_pos[2] -0.2787 -0.13610 -0.0629 0.01292 0.1577
## beta_pos[3] -0.4750 -0.38122 -0.3334 -0.28628 -0.1981
## beta ht
              -0.2095 0.01519 0.1368 0.26105 0.4919
(c)
par(mfrow=c(2, 2))
plot(as.matrix(coef_sample_1)[,"beta_pos[1]"] ~ as.matrix(coef_sample_1)[,"beta_ht"],
     xlab = expression(paste(beta[H[t]])), ylab = expression(paste(beta[C])), pch='.')
plot(as.matrix(coef_sample_1)[,"beta_pos[2]"] ~ as.matrix(coef_sample_1)[,"beta_ht"],
     xlab = expression(paste(beta[H[t]])), ylab = expression(paste(beta[F])), pch='.')
plot(as.matrix(coef_sample_1)[,"beta_pos[3]"] ~ as.matrix(coef_sample_1)[,"beta_ht"],
     xlab = expression(paste(beta[H[t]])), ylab = expression(paste(beta[G])), pch='.')
```



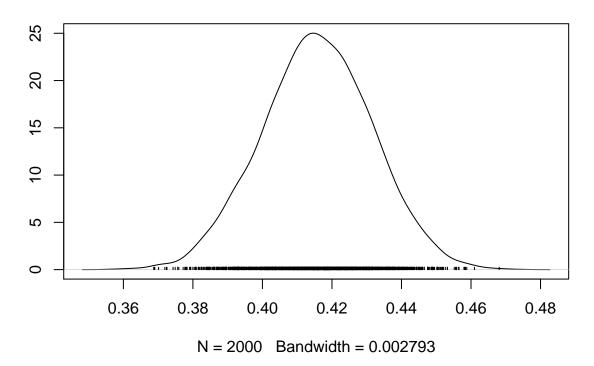




According to the plots, β_C , β_F , β_G are correlated with β_{H_t} .

(d)

Density of Probability for Ayo Dosunmu



(e)

Probability of $\beta_F > \beta_G$,

```
beta_F = as.matrix(coef_sample_1)[, "beta_pos[2]"]
beta_G = as.matrix(coef_sample_1)[, "beta_pos[3]"]
mean(beta_F > beta_G)
```

[1] 0.966125

Bayes factor favoring $\beta_F > \beta_G$ versus $\beta_F < \beta_G$,

```
mean(beta_F > beta_G) / mean(beta_F < beta_G)</pre>
```

[1] 28.5203

Given the Bayes factor is between 20 to 150, we can say that the data has **Strong** evidence that $\beta_F > \beta_G$.

(f)

```
probs <- as.matrix(coef_sample_1)[, paste("prob[",1:nrow(perf_data),"]", sep="")]
FGM_rep <- as.matrix(coef_sample_1)[, paste("FGM_rep[",1:nrow(perf_data),"]", sep="")]
Tchi <- numeric(nrow(FGM_rep))
Tchirep <- numeric(nrow(FGM_rep))</pre>
```

```
for(s in 1:nrow(FGM_rep)){
  Tchi[s] <- sum((perf_data$FGM - perf_data$FGA * probs[s,])^2 /</pre>
                   (perf_data$FGA * probs[s,] * (1 - probs[s,])))
  Tchirep[s] <- sum((FGM rep[s,] - perf data$FGA * probs[s,])^2 /
                      (perf_data$FGA * probs[s,] * (1 - probs[s,])))
}
mean(Tchirep >= Tchi)
```

[1] 0.045875

The posterior predictive p-value is small, although not exceedingly so. Given we don't find any outliers, we conclude that there is a problem of overdispersion.

(g)

##

beta ht ## beta_pos[1]

```
(i)
model {
    for (i in 1:length(FGM)) {
        FGM[i] ~ dbin(prob[i], FGA[i])
        logit(prob[i]) <- beta_pos[Pos[i]] + beta_ht * Ht_Scaled[i] + epsilon[i]</pre>
        epsilon[i] ~ dnorm(0, 1 / sigma_epsilon^2)
        FGM_rep[i] ~ dbin(prob[i], FGA[i])
    }
    for (j in 1:max(Pos)) {
        beta_pos[j] ~ dt(0, 0.01, 1)
    }
    beta_ht ~ dt(0, 0.16, 1)
    sigma_epsilon ~ dunif(0,10)
}
df_jags_2 <- list( FGM = perf_data$FGM, FGA = perf_data$FGA,</pre>
                   Pos = unclass(perf_data$Pos),
                   Ht_Scaled = as.vector(scale(perf_data$Ht, scale=2*sd(perf_data$Ht))))
initial vals 2 \leftarrow list(list(beta pos = c(10,10,10), beta ht=10, sigma epsilon = 0.01),
                        list(beta_pos = c(10,10,-10), beta_ht=-10, sigma_epsilon = 9),
                        list(beta_pos = c(10,-10,10), beta_ht=-10, sigma_epsilon = 0.01),
                       list(beta_pos = c(10,-10,-10), beta_ht=10, sigma_epsilon = 9))
model_2 <- jags.model("perf_2.bug", df_jags_2, initial_vals_2, n.chains = 4,</pre>
                      n.adapt = 1000)
update(model_2, 1000)
x2 <- coda.samples(model_2, c("beta_pos", "beta_ht", "sigma_epsilon"), n.iter = 10000)
gelman.diag(x2, autoburnin=FALSE)
## Potential scale reduction factors:
```

Point est. Upper C.I. 1.02

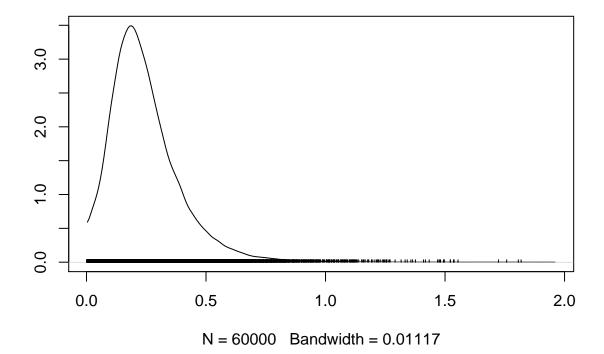
1.01

1.04

1.02

```
## beta_pos[2]
                        1.01
                                   1.02
## beta_pos[3]
                        1.01
                                   1.01
                        1.01
                                   1.03
## sigma_epsilon
## Multivariate psrf
##
## 1.01
coef_sample_2 <- coda.samples(model_2, c("beta_pos", "beta_ht", "prob", "FGM_rep",</pre>
                                          "sigma_epsilon"), n.iter = 60000)
effectiveSize(coef_sample_2[,c("beta_pos[1]", "beta_pos[2]", "beta_pos[3]", "beta_ht",
                                "sigma_epsilon")])
##
     beta_pos[1]
                                  beta_pos[3]
                   beta_pos[2]
                                                     beta_ht sigma_epsilon
        8335.420
                       6418.475
                                     7133.235
                                                    5479.946
                                                                  5458.723
##
(ii)
densplot(coef_sample_2[, "sigma_epsilon"],
         main = expression(paste("Desity of ", sigma[epsilon])))
```

Desity of σ_{ϵ}



```
(iii)
beta_F = as.matrix(coef_sample_2)[, "beta_pos[2]"]
beta_G = as.matrix(coef_sample_2)[, "beta_pos[3]"]
mean(beta_F > beta_G)
```

```
## [1] 0.78665
```

This posterior probability is smaller than previous model.

```
mean(beta_F > beta_G) / mean(beta_F < beta_G)</pre>
```

```
## [1] 3.687134
```

This Bayes factor favoring $\beta_F > \beta_G$ versus $\beta_F < \beta_G$ is much smaller than previous model, and we can only say the data has **Positive** (between 3 to 30) evidence that $\beta_F > \beta_G$.

Also Chi-square discrepancy,

```
## [1] 0.3826792
```

Thus we says no overdispersion problems for this model.

Solution for Problem 3

(a)

```
model {
    for (i in 1:length(BLK)) {
        BLK[i] ~ dpois(lambda[i])
        log(lambda[i]) <- log_MIN[i] + beta_pos[Pos[i]] + beta_ht * Ht_Scaled[i]</pre>
        BLK_rep[i] ~ dpois(lambda[i])
    }
    for (j in 1:max(Pos)) {
        beta_pos[j] ~ dnorm(0, 0.0001)
    beta_ht ~ dnorm(0, 0.0001)
}
df_jags_3 <- list( BLK = perf_data$BLK,</pre>
                   Pos = unclass(perf data$Pos),
                    log_MIN = log(perf_data$MIN),
                   Ht_Scaled = as.vector(scale(perf_data$Ht, scale=sd(perf_data$Ht))))
initial_vals_3 \leftarrow list(list(beta_pos = c(100,100,100), beta_ht=100),
                        list(beta_pos = c(100, 100, -100), beta_ht=-100),
                        list(beta_pos = c(100, -100, 100), beta_ht=-100),
                        list(beta_pos = c(100,-100,-100), beta_ht=100))
model_3 <- jags.model("perf_3.bug", df_jags_3, initial_vals_3, n.chains = 4,</pre>
                       n.adapt = 1000)
update(model_3, 1000)
x3 <- coda.samples(model_3, c("beta_pos","beta_ht"), n.iter = 2000)
gelman.diag(x3, autoburnin=FALSE)
## Potential scale reduction factors:
##
##
               Point est. Upper C.I.
## beta_ht
                         1
                                    1
## beta_pos[1]
                         1
                                    1
```

```
## beta_pos[2]
                        1
## beta_pos[3]
##
## Multivariate psrf
##
## 1
coef_sample_3 <- coda.samples(model_3, c("beta_pos", "beta_ht", "lambda", "BLK_rep"),</pre>
                              n.iter = 20000, thin = 5)
effectiveSize(coef_sample_3[,c("beta_pos[1]", "beta_pos[2]", "beta_pos[3]", "beta_ht")])
## beta_pos[1] beta_pos[2] beta_pos[3]
                                            beta_ht
##
      4881.227
                  5760.599
                              9429.966
                                           4620.907
(b)
summary(coef_sample_3[, c("beta_pos[1]", "beta_pos[2]", "beta_pos[3]", "beta_ht")])
##
## Iterations = 4005:24000
## Thinning interval = 5
## Number of chains = 4
## Sample size per chain = 4000
## 1. Empirical mean and standard deviation for each variable,
##
      plus standard error of the mean:
##
##
                           SD Naive SE Time-series SE
                  Mean
## beta_pos[1] -5.2725 0.6009 0.004750
                                              0.008628
## beta_pos[2] -4.5005 0.2844 0.002248
                                              0.003795
## beta_pos[3] -4.4522 0.1774 0.001402
                                              0.001831
## beta ht
                0.9969 0.2727 0.002156
                                              0.004027
##
## 2. Quantiles for each variable:
##
##
                  2.5%
                           25%
                                    50%
                                           75% 97.5%
## beta_pos[1] -6.4756 -5.6724 -5.2571 -4.857 -4.138
## beta pos[2] -5.0832 -4.6864 -4.4929 -4.304 -3.972
## beta_pos[3] -4.8080 -4.5709 -4.4499 -4.329 -4.116
                0.4797 0.8097 0.9896 1.179 1.543
## beta_ht
(c)
beta_ht = as.matrix(coef_sample_3)[, "beta_ht"]
quantile(exp(beta_ht), c(0.025, 0.975))
       2.5%
               97.5%
## 1.615524 4.679455
```

The values within 95% central posterior credible interval are all greater than 1 and thus we can conclude that greater height is associated with a higher rate of blocking shots.

(d)

##

name

p_value

```
lambdas <- as.matrix(coef_sample_3)[, paste("lambda[",1:nrow(perf_data),"]", sep="")]</pre>
BLK_rep <- as.matrix(coef_sample_3)[, paste("BLK_rep[",1:nrow(perf_data),"]", sep="")]
Tchi <- numeric(nrow(BLK_rep))</pre>
Tchirep <- numeric(nrow(BLK_rep))</pre>
for(s in 1:nrow(BLK_rep)){
  Tchi[s] <- sum((perf_data$BLK - lambdas[s,])^2 / lambdas[s,])</pre>
  Tchirep[s] <- sum((BLK_rep[s,] - lambdas[s,])^2 / lambdas[s,])</pre>
 }
mean(Tchirep >= Tchi)
## [1] 0.0069375
The posterior predictive p-value is extremely small. Thus this could indicate a problem of overdispersion.
(e)
(i)
p_sample <- matrix(FALSE, nrow = nrow(BLK_rep), ncol = nrow(perf_data))</pre>
for(s in 1:nrow(BLK_rep)){
  p_sample[s,] <- BLK_rep[s,] > perf_data$BLK
}
p = apply(p_sample, 2, mean)
p_df = data.frame(name=perf_data$Player, p_value=p)
p_df
##
                        name
                                p_value
## 1
      Bezhanishvili, Giorgi 0.5311250
## 2
                 Cayce, Drew 0.0592500
## 3
         De La Rosa, Adonis 0.9865625
## 4
               Dosunmu, Ayo 0.7095000
## 5
               Feliz, Andres 0.8339375
## 6
             Frazier, Trent 0.8236250
               Griffin, Alan 0.0075000
## 7
## 8
             Griffith, Zach 0.1917500
## 9
               Jones, Tevian 0.8935625
## 10
               Jordan, Aaron 0.1326875
                 Kane, Samba 0.0022500
## 11
## 12
            Nichols, Kipper 0.2376250
## 13
          Oladimeji, Samson 0.1830625
## 14
           Underwood, Tyler 0.2636250
## 15
         Williams, Da'Monte 0.0436875
(ii)
p_df[p_df^p_value < 0.05,]
```

By looking at the original data, **Adonis** played in center position and was 84 height. He played 225 minutes but blocked only 1 shot. Samba in another hand, also played in center position and was also 84 height. For 86 minutes he played, blocked 10 shots. This makes the model always overestimate the blocks by Adonis. Thus the p-value is very high.