

Università degli Studi di Padova

Random subsampling techniques for sea bass mortality prediction

Giovanni Gaio, Simone Moretti July 24, 2025

Overview

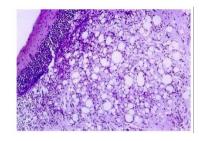


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VNN and Sea Basses



Viral nervous necrosis (**VNN**) is a highly spread disease among sealife.

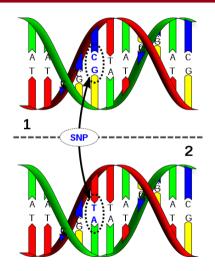




We concentrate our efforts on predicting the mortality of a population of **sea basses** affected by VNN.

SNPs for predicting mortality





The **genome** might be useful to predict mortality.

SNPs: Single nucleotide polymorphisms.

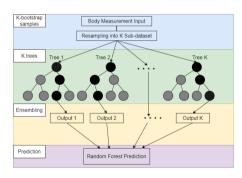
Machine Learning Approach



Predict if a sea bass will die by watching its genome: **Machine Learning**.

In particular, we use the XGBoost classifier.

dmlc **XGBoost**



Challenges with Genomic Data

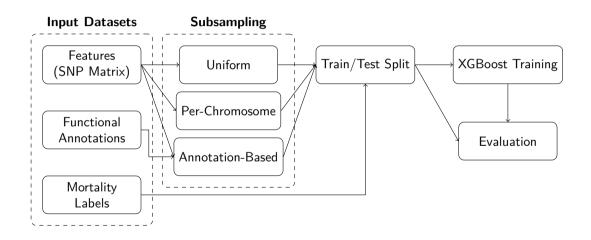


- Each fish: over 6 million SNP positions.
- Sample size: only 990 sea bass individuals.
- Traditional models may overfit due to high dimensionality.
- We mitigate through subsampling.



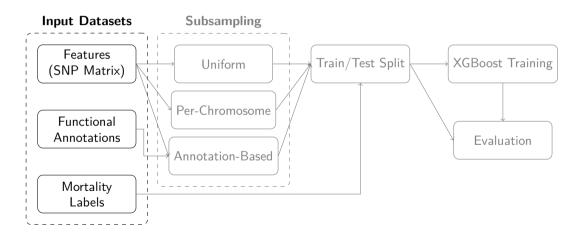
Pipeline Overview





Input Datasets





SNP Dataset Structure



- 990 rows (fish), each with 6,072,853 SNP features.
- SNP values: 0 (no mutation), 1 (heterozygous), 2 (homozygous alt).
- Each fish is paired with a mortality label.

| id | mortality |
|-----------|-----------|
| PL06-B12 | 1 |
| PL06-B06 | 1 |
| PL06-E06 | 1 |
| PL08n-B05 | 0 |
| PL08n-G09 | 0 |
| : | : |

| | CAJNNU010000001.1:299 | CAJNNU010000001.1:903 | CAJNNU010000001.1:986 | |
|----------|-----------------------|-----------------------|-----------------------|---|
| PL04-A06 | 1 | 0 | 0 | |
| PL04-A08 | 0 | 0 | 1 | |
| PL04-A09 | 0 | 1 | 1 | |
| PL04-A10 | 0 | 0 | 0 | |
| PL04-A11 | 2 | 2 | 1 | |
| : | : | : | : | |
| • | • | • | • | 1 |

Annotation Metadata

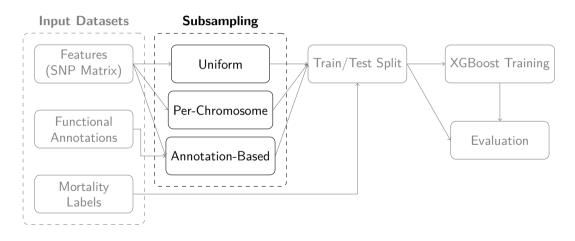


- Annotations include function: Promoter, Enhancer, Open Chromatin.
- Tissue number (0–25) indicates location-specific relevance.

| snp_id | funct | n_tissue |
|------------------------|----------------|----------|
| CAJNNU010000001.1:7825 | Open_chromatin | 10 |
| CAJNNU010000001.1:7865 | Open_chromatin | 4 |
| CAJNNU010000001.1:8046 | Enhancer | 21 |
| CAJNNU010000001.1:8084 | Open_chromatin | 5 |
| CAJNNU010000001.1:8116 | Promoter | 12 |
| : | i i | : |

Subsampling techniques

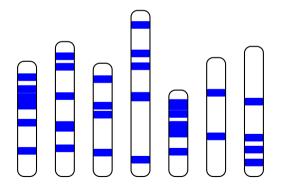




Uniform Subsampling



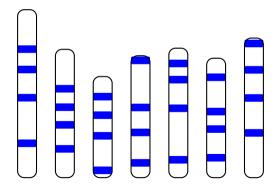
- Randomly sample a fixed proportion p of all SNPs.
- Simple but may cause imbalance across chromosomes.



Per-Chromosome Subsampling



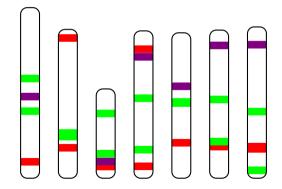
- Ensures balanced representation from each chromosome.
- Randomly sample same number of SNPs per chromosome.



Annotation-Based Subsampling

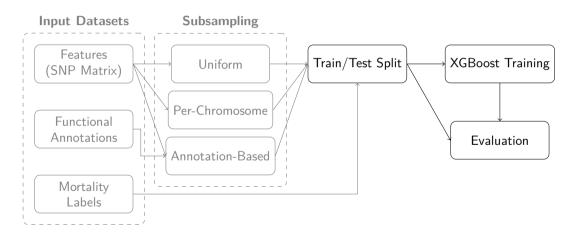


- Filter SNPs by biological annotation.
- Then apply uniform subsampling to relevant regions.



Parameter Evaluation





Let's get to the substance

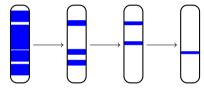




Subsampling Ratios and baseline results



• Subsampled with multiple subasmpling ratio values: log-spaced varying from the whole genome to few SNPs.



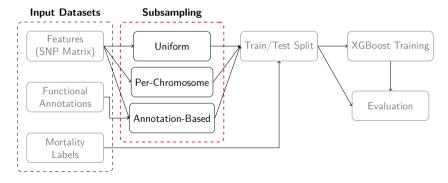
- Trained model for each combination of model and subsample rate.
- Multiple runs for each pair of parameters.
- Comparison with baseline results from a "dumb" classifier, always guessing the most common class.

Control of Randomness



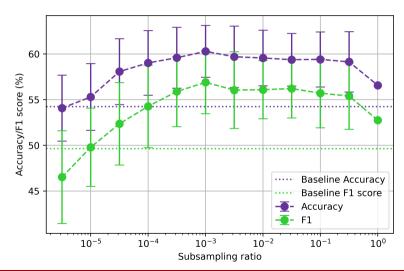
In order to limit the variance of results we impose:

- XGBoost random seed and train-test split are fixed.
- Subsampling is the only random component varying between experiments.



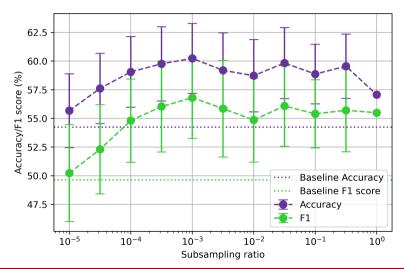
Results: uniform subsampling





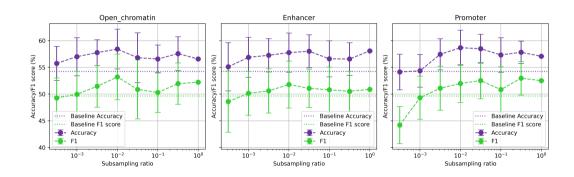
Results: subsampling uniformly on chromosomes





Results: annotated subsampling (function)

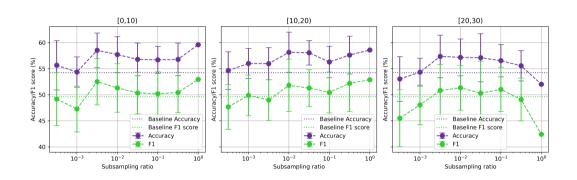




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Results: annotated subsampling (tissue number)





Conclusions



- Random SNP subsampling retains model effectiveness.
- No strong trend between rate and accuracy (outside extremes): this may be good.
- There doesn't seem to be specific regions of the genome containing the information determining the disease effects.



The results don't show any meaningful trend

We showed that subsampling allows the use of more complex models

Thank You!