Class 10: Structural Bioinformatics Pt1

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What is in the PDB database

The main repository of biomolecular structure info is the PDB < www.rcsb.org >.

Let's see what this database contains:

```
stats <- read.csv("pdb_stats.csv", row.names=1)
stats</pre>
```

	X.ray	EM	NMR	Multiple.methods	Neutron	Other
Protein (only)	161,663	12,592	12,337	200	74	32
Protein/Oligosaccharide	9,348	2,167	34	8	2	0
Protein/NA	8,404	3,924	286	7	0	0
Nucleic acid (only)	2,758	125	1,477	14	3	1
Other	164	9	33	0	0	0
Oligosaccharide (only)	11	0	6	1	0	4
	Total					
Protein (only)	186,898					
Protein/Oligosaccharide	11,559					
Protein/NA	12,621					
Nucleic acid (only)	4,378					
Other	206					
Oligosaccharide (only)	22					

```
as.numeric(stats$X.ray)
```

Warning: NAs introduced by coercion

[1] NA NA NA NA 164 11

We got to get rid of the commas. Can you find a function to get rid of the commas?

```
x <- stats$X.ray
sum(as.numeric(gsub(",", "", x)))
```

[1] 182348

I am going to turn this into a function and then use apply() to work on the entire table of data.

```
sumcomma <- function(x) {</pre>
    sum(as.numeric(gsub(",", "", x)))
  sumcomma(stats$X.ray)
[1] 182348
```

```
n.total <- sumcomma(stats$Total)</pre>
n.total
```

[1] 215684

```
apply(stats, 2, sumcomma) / sumcomma(stats$Total)
```

Multiple.methods	NMR	EM	X.ray
0.0010663749	0.0657118748	0.0872433746	0.8454405519
	Total	Other	Neutron
	1.0000000000	0.0001715473	0.0003662766

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
sum(sumcomma(stats$X.ray), sumcomma(stats$EM)) / sumcomma(stats$Total)
```

[1] 0.9326839

Q2: What proportion of structures in the PDB are protein?

```
sumcomma(stats$Total[1]) / sumcomma(stats$Total)
```

[1] 0.8665362

In Uniprot there are 258,805,733 entries which compare to PDB protein entries (186,898) means there are only $\sim 7\%$ of known sequences with a known structure.

```
186898/248805733 * 100
```

[1] 0.07511804

Visualizing the HIV-1 protease structure

Mol* (mol-star) viewer is now everywhere. The homepage is here: https://molstar.org/viewer/. I want to to insert my image from Mol* here

Working with the bio3d package

```
library(bio3d)

pdb <- read.pdb("1hsg")

Note: Accessing on-line PDB file

pdb

Call: read.pdb(file = "1hsg")

Total Models#: 1
   Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
   Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)</pre>
```

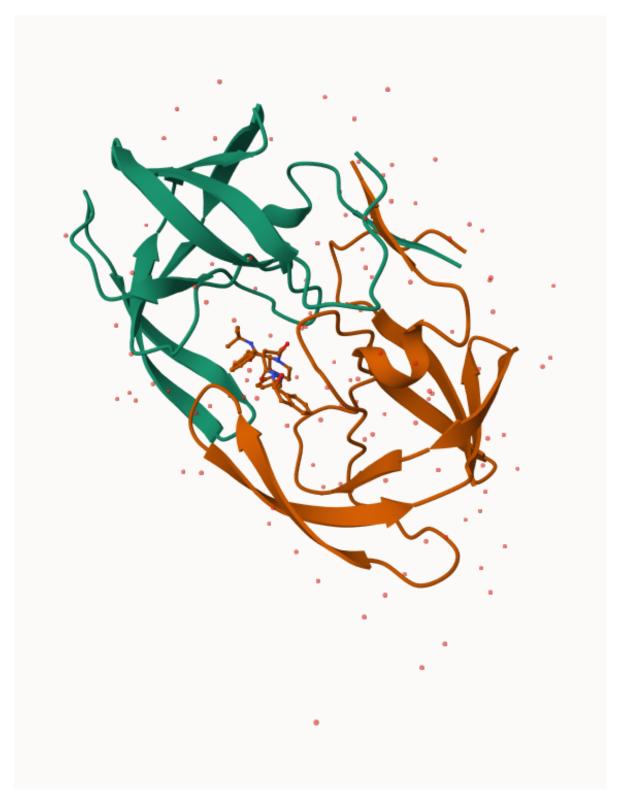


Figure 1: My first molecular image $\,$



Figure 2: 1HSG with D25 and water as spacefill

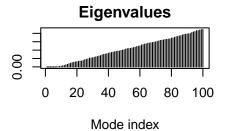
```
Non-protein/nucleic Atoms#: 172 (residues: 128)
    Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
  Protein sequence:
     PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
     QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
     ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
     VNIIGRNLLTQIGCTLNF
+ attr: atom, xyz, seqres, helix, sheet,
       calpha, remark, call
  head(pdb$atom)
 type eleno elety alt resid chain resno insert
                                                     X
                                                            У
1 ATOM
          1
                N < NA >
                         PRO
                                 Α
                                       1
                                           <NA> 29.361 39.686 5.862 1 38.10
2 ATOM
          2
               CA <NA>
                         PRO
                                 Α
                                       1 <NA> 30.307 38.663 5.319 1 40.62
3 ATOM
          3
                C <NA>
                         PRO
                                 Α
                                       1 <NA> 29.760 38.071 4.022 1 42.64
4 ATOM
          4
                         PRO
                                       1 <NA> 28.600 38.302 3.676 1 43.40
                O < NA >
                                 Α
5 ATOM
               CB <NA>
                         PRO
                                 Α
                                       1 <NA> 30.508 37.541 6.342 1 37.87
                                       1 <NA> 29.296 37.591 7.162 1 38.40
6 ATOM
          6
               CG <NA>
                         PRO
                                 Α
 segid elesy charge
1 <NA>
           N
               <NA>
2 <NA>
           C
               <NA>
3 <NA>
           C <NA>
4 <NA>
           O <NA>
5 <NA>
           C <NA>
           С
6 <NA>
               <NA>
  pdbseq(pdb)[25]
25
"D"
```

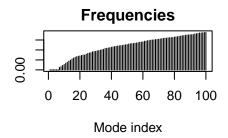
Predicting functional motions of a single structure

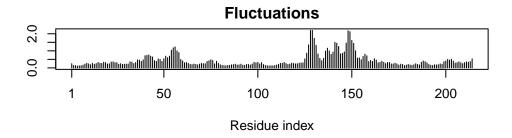
We can do bioinformatics prediction of functional motions (i.e flexibility/dynamics):

```
pdb <- read.pdb("6s36")</pre>
```

```
Note: Accessing on-line PDB file
  PDB has ALT records, taking A only, rm.alt=TRUE
  pdb
       read.pdb(file = "6s36")
  Total Models#: 1
    Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
    Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
    Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
    Non-protein/nucleic Atoms#: 244 (residues: 244)
    Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
  Protein sequence:
     MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLVT
     DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
     VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG
+ attr: atom, xyz, seqres, helix, sheet,
       calpha, remark, call
  m <- nma(pdb)
Building Hessian...
                           Done in 0.03 seconds.
Diagonalizing Hessian...
                           Done in 0.17 seconds.
  plot(m)
```







mktrj(m, file="adk_m7.pdb")