**Reserve Name** (LKS) **NERR Nutrient Metadata**

**January to December, 2023**

**Latest Update:** 6/11/2024

Note: This is a provisional metadata document; it has not been authenticated as of its download date. Contents of this document are subject to change throughout the QAQC process and it should not be considered a final record of data documentation until that process is complete. Contact the CDMO ([cdmosupport@baruch.sc.edu](mailto:cdmosupport@baruch.sc.edu)) or reserve with any additional questions.

**I. Data Set and Research Descriptors**

**1) Principal investigator(s) and contact persons –**

Hannah Ramage, Monitoring Coordinator (Laboratory Contact)

14 Marina Drive, Superior, WI 54880

705-399-4088

[Hannah.ramage@wisc.edu](mailto:Hannah.ramage@wisc.edu)

Kait Reinl, Research Coordinator

14 Marina Drive, Superior, WI 54880

715-399-4087

[kreinl@wisc.edu](mailto:kreinl@wisc.edu)

**2) Research objectives** –

The Lake Superior NERR is situated on the freshwater estuary at the confluence of the St. Louis River and Lake Superior, the largest and most pristine of the Great Lakes. The Reserve is a diverse, 16,697-acre complex that contains a variety of representative terrestrial and aquatic habitats allowing for extensive research and educational opportunities. The Reserve provides opportunities for research and monitoring, experiential learning, and training, while continuing to contribute to the protection of the ecological health of the St. Louis River Estuary and Lake Superior coastal habitats.

The Lake Superior NERR implements the NERR System-Wide Monitoring Program (SWMP) along a river-to-Lake gradient. SWMP includes a continuous meteorological station, four continuous water quality monitoring stations, and monthly nutrient/pigment sampling at those same four stations. The nutrient sampling has two programmatic parts:

1. Monthly Grab Sampling Program: Identifies nutrient difference along the river-to-Lake gradient throughout the ice-free season. Samples are collected at the four long-term SWMP water quality stations.
2. Diel Sampling Program: Lake Superior does not experience strong tides, therefore 12 diel samples are simply collected with an auto-sampler every two hours, beginning the day before or day of grab sample collections. Diel samples are collected at the same SWMP station, every month.

**3) Research methods** –

1. Monthly Grab Sampling Program

Grab samples (sequential replicates) were collected from a boat or canoe once a month at the depth of the sonde deployment (~1.5 meters beneath the surface, except at Pokegama which is shallower) using a horizontal sampler. Sample bottles are acid-washed amber one-liter poly bottles. Ambient water quality data was collected concurrent with sampling, with a YSI EXO datasonde calibrated at the LKS NERR laboratory. At each station, seasonal depth profiles (bottom, middle, and top of water column) were recorded on a field sheet. Depth profile data are only available by contacting the Reserve directly.

1. Diel Sampling Program

Diel samples were taken from the dock located at Barker’s Island SWMP station, at the same depth as the water quality datasonde, with an ISCO autosampler. The sampler was set to sample twelve times, with pre-reverse, every two hours for 24 hours beginning either the day before, or the day of monthly grab samples. Sample bottles are acid washed one-liter translucent poly bottles. Ice was added to the ISCO sample bottle container for the duration of sampling during warm summer months. Cold months inhibit use of the ISCO sampler because of ice buildup in the tubing.

Both monthly grab and diel samples were transported from the field to the laboratory in a cooler, then filtered within a few hours of arrival in the LKS NERR laboratory and at the latest 24 hours from collection. Samples were filtered in low light to prevent chlorophyll *a* degradation. Samples were frozen at ≤-20°C. Chlorophyll *a* filters were folded and enclosed in aluminum wrapped centrifuge tubes and kept in the freezer at ≤-20°C until extraction within 28 days. The LKS NERR laboratory conducted all Chlorophyll *a* and Total Suspended Solid analysis for all grab and diel samples every month in 2023.

The University of Minnesota-Duluth’s Natural Resources Research Institute (NRRI) Central Analytical Laboratory conducted nutrient analysis for all sampling in 2023. Samples were filtered at the LKS NERR then either transported via cooler to NRRI (30-minute drive) the same day and placed in a freezer at NRRI, or frozen at the LKS NERR then transported within 3 days to NRRI.

**4) Site location and character –**

|  |  |
| --- | --- |
| Site name | Oliver Bridge (OL) |
| Latitude and longitude | 46.65685, -92.20166 |
| Tidal range *(meters)* | None – experiences seiche effects of up to 0.2 m |
| Salinity range *(psu)* | 0.08 – 0.2 PPT |
| Type and amount of freshwater input | Entirely freshwater from St. Louis River |
| Water depth (*meters, MLW*) | *approximately 8m deep* |
| Sonde distance from bottom (*meters*) | *the sonde is deployed at a depth of ~1.5 m from the surface at this site* |
| Bottom habitat or type | *soft sediment* |
| Pollutants in area | Site is approximately 5.5 miles downstream of the Fond du Lac dam, there were historically paper mills above the site and there is current mining in the upper watershed |
| Description of watershed | *this site is the furthest upstream site monitored in the St. Louis River Estuary by LKS, approximately 11 miles upstream from the mouth at Lake Superior. The watershed is primarily composed of woody wetlands and mixed forest.* |

|  |  |
| --- | --- |
| Site name | Blatnik Bridge (BL) |
| Latitude and longitude | 46.748649, -92.10027 |
| Tidal range *(meters)* | None – experiences seiche effects of up to 0.3 m |
| Salinity range *(psu)* | 0.08 – 0.25 PPT |
| Type and amount of freshwater input | Entirely freshwater from St. Louis River, Lake Superior, some urban tributaries, and discharge from the Western Lake Superior Sanitary District Wastewater Treatment Plant |
| Water depth (*meters, MLW*) | *approximately 7m deep* |
| Sonde distance from bottom (*meters*) | *the sonde is deployed at a depth of ~1.5 m from the surface at this site* |
| Bottom habitat or type | *soft sediment* |
| Pollutants in area | site is located within the urban area of Superior, WI, and Duluth, MN; site is immediately downstream of the Western Lake Superior Sanitary District Wastewater Treatment Plant discharge. It is also downstream of several impaired tributaries. |
| Description of watershed | *Watershed is mostly woody wetlands and mixed forest, however this site is adjacent to areas of high intensity developed land.* |

|  |  |
| --- | --- |
| Site name | Barker’s Island (BA) |
| Latitude and longitude | 46.721772, -92.06352 |
| Tidal range *(meters)* | None – experiences seiche effects of up to 0.3 m |
| Salinity range *(psu)* | 0.08 – 0.25 PPT |
| Type and amount of freshwater input | Entirely freshwater from St. Louis River, Lake Superior, some urban tributaries, and discharge from the Superior Wastewater Treatment Facility. Just downstream of Faxon Creek. |
| Water depth (*meters, MLW*) | *approximately 2m deep* |
| Sonde distance from bottom (*meters*) | *the sonde is deployed at a depth of ~1.5 m from the surface, about 0.5m from the bottom* |
| Bottom habitat or type | *soft sediment* |
| Pollutants in area | site is downstream of Superior and Duluth’s wastewater treatment facilities, and near several storm water outfalls and Faxon Creek (an entirely urban stream) it is also adjacent to a public beach which has been closed occasionally due to E. coli standard exceedances and harmful algal blooms |
| Description of watershed | *Watershed is mostly woody wetlands and mixed forest, however this site is adjacent to areas of high intensity developed land.* |

|  |  |
| --- | --- |
| Site name | Pokegama Bay (PO) |
| Latitude and longitude | 46.672360, -92.135614 |
| Tidal range *(meters)* | None – experiences seiche effects of up to 0.2 m |
| Salinity range *(psu)* | 0.06 – 0.21 PPT |
| Type and amount of freshwater input | Entirely freshwater from the Pokagama River |
| Water depth (*meters, MLW*) | *approximately 1m deep* |
| Sonde distance from bottom (*meters*) | *the sonde is deployed 0.5m from the bottom* |
| Bottom habitat or type | *Soft bottom of glacial red clay* |
| Pollutants in area | this site is downstream of Village of Superior’s wastewater lagoons and is impaired due to Total Phosphorus exceedances |
| Description of watershed | *the Pokegama River is a tributary to the St. Louis River, entering the estuary on the Wisconsin side of Clough Island. The Pokegama River watershed measures approximately 20,144 acres, 51% of which is wetland, 37% forested, 4% developed and 6% agricultural use (the remainder is open water or bare land). This site is on a red clay tributary to the St. Louis River* |

All LKS NERR historical nutrient/pigment monitoring stations:

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Station Code** | **SWMP Status** | **Station Name** | **Location** | **Active Dates** | **Reason Decommissioned** | **Notes** |
| BA | P | LKSBAWQ | 46° 43' 18.38 N, 92° 03' 48.67 W | 05/05/2012 00:00 -current | NA | NA |
| BL | P | LKSBLWQ | 46° 44' 55.14 N, 92° 06' 0.97 W | 05/08/2012 00:00 -current | NA | NA |
| OL | P | LKSOLWQ | 46° 39' 24.66 N, 92° 12' 5.98 W | 05/08/2012 00:00 -current | NA | NA |
| PO | P | LKSPOWQ | 46° 40' 20.50 N, 92° 8' 8.21 W | 05/28/2013 00:00 -current | NA | NA |

**5) Coded variable definitions** –

lksbanut = Lake Superior NERR Barker’s Island nutrients

lksponut = Lake Superior NERR Pokegama River nutrients

lksolnut = Lake Superior NERR Oliver Bridge nutrients

lksblnut = Lake Superior NERR Blatnik Bridge nutrients

monthly grab sample program = 1

diel grab sample program = 2

**6) Data collection period** –

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **SITE** | **Barker’s Island** | **Blatnik Bridge** | **Oliver Bridge** | **Pokegama Bay** | **Barker’s Island** |
| **First Year**  **Sampled** | 2014 | 2012 | 2012 | 2013 | 2014 |
| **Monitoring Program** | Grab | Grab | Grab | Grab | Diel |
| **May** | 05/03/23 15:25 | 05/03/23 14:35 | 05/03/23 11:20 | 05/03/23 12:05 | 05/03/23 10:00 to 05/04/23 8:00 |
| **May-June** | 05/31/23 6:30 | 05/31/23 9:00 | 05/31/23 13:50 | 05/31/23 12:41 | 05/31/23 10:00 to 06/01/23 8:00 |
| **June** | 06/29/23 6:20 | 06/29/23 7:56 | 06/29/23 11:39 | 06/29/23 10:27 | 06/29/23 8:00 to 06/30/23 6:00 |
| **July** | 07/11/23 12:58 | 07/11/23 12:19 | 07/11/23 9:31 | 07/11/23 10:34 | 07/11/23 10:00 to 07/12/23 8:00 |
| **August** | 08/18/23 12:16 | 08/18/23 11:11 | 08/18/23 7:59 | 08/18/23 8:42 | 08/18/23 11:00 to 8/19/23 9:00 |
| **September** | 09/14/23 15:12 | 09/14/23 8:20 | 09/14/23 11:20 | 09/14/23 12:22 | 09/13/23 12:00 to 09/14/23 10:00 |
| **October** | 10/11/23 12:48 | 10/11/23 12:18 | 10/11/23 9:43 | 10/11/23 10:20 | 10/16/23 12:00 to 10/17/23 10:00 |
| **November** | 11/08/23 14:40 | 11/08/23 13:21 | Not collected | 11/08/23 10:25 | Not collected (ice) |

**7) Associated researchers and projects–**

As part of the SWMP long-term monitoring program, LKS NERR also monitors 15-minute meteorological and water quality data which may be correlated with this nutrient/pigment dataset. These data are available at [www.nerrsdata.org](http://www.nerrsdata.org).

In 2023 the LKS NERR collected phytoplankton samples and associated environmental parameters (nutrients, pigment, light etc.) as a part of a NERRS Science Collaborative funded project. These data will become available once the project is complete in Spring 2025. For more information visit - <https://nerrssciencecollaborative.org/project/Ramage22>

The St. Louis River Estuary is listed as an Area of Concern under the Great Lakes Water Quality Agreement. One of the impairments for which it was listed is “Excessive Loading of sediment and nutrients”. Other agencies working in the St. Louis River Estuary to remove impairments include the Wisconsin and Minnesota Department of Natural Resources, the United States Environmental Protection Agency Mid-Continent Ecology Lab, United States Fish and Wildlife Service and the United State Geological Survey. The LKS NERR participates with partnerships in the area with these agencies as well as with the City of Superior, Douglas County, and several non-profits. More information can be found at - [https://www.pca.state.mn.us/watershed-information/st-louis-river /](https://www.pca.state.mn.us/watershed-information/st-louis-river%20/)

**8) Distribution** –

NOAA retains the right to analyze, synthesize and publish summaries of the NERRS System-wide Monitoring Program data.  The NERRS retains the right to be fully credited for having collected and processed the data.  Following academic courtesy standards, the NERR site where the data were collected should be contacted and fully acknowledged in any subsequent publications in which any part of the data are used.  The data set enclosed within this package/transmission is only as good as the quality assurance and quality control procedures outlined by the enclosed metadata reporting statement.  The user bears all responsibility for its subsequent use/misuse in any further analyses or comparisons.  The Federal government does not assume liability to the Recipient or third persons, nor will the Federal government reimburse or indemnify the Recipient for its liability due to any losses resulting in any way from the use of this data.

Requested citation format:

NOAA National Estuarine Research Reserve System (NERRS). System-wide Monitoring Program. Data accessed from the NOAA NERRS Centralized Data Management Office website: www.nerrsdata.org; *accessed* 12 October 2022.

NERR nutrient data and metadata can be obtained from the Research Coordinator at the individual NERR site (please see Principal investigators and contact persons), from the Data Manager at the Centralized Data Management Office (please see personnel directory under the general information link on the CDMO home page) and online at the CDMO home page [www.nerrsdata.org](http://cfcdmo.baruch.sc.edu/). Data are available in comma separated version format.

**II. Physical Structure Descriptors**

**9) Entry verification** –

Raw results for chlorophyll-a (ug/L) and total suspended solids (mg/L) are hand recorded in laboratory notebooks or bench sheets. These results are later entered digitally into an excel spreadsheet. All data transfers from hand recorded datasheets, bench sheets, or notebooks were independently checked by a second person for copy errors.

Nutrient data from the NRRI Central Analytical laboratory are provided in an excel spreadsheet over email. These data undergo a unit conversion then are copied over to the NutrientQAQC Excel macro (see below).

Nutrient data are entered into a Microsoft Excel worksheet and processed using the NutrientQAQC Excel macro. The NutrientQAQC macro sets up the data worksheet, metadata worksheets, and MDL worksheet; adds chosen parameters and facilitates data entry; allows the user to set the number of significant figures to be reported for each parameter and rounds using banker’s rounding rules; allows the user to input MDL values and then automatically flags/codes measured values below MDL and inserts the MDL; calculates parameters chosen by the user and automatically flags/codes for component values below MDL, negative calculated values, and missing data; allows the user to apply QAQC flags and codes to the data; produces summary statistics; graphs selected parameters for review; and exports the resulting data file to the CDMO for tertiary QAQC and assimilation into the CDMO’s authoritative online database.

The NRRI Central Analytical Laboratory calculates and reports results in ppb (parts per billion). For purposes of consistency in the NERR System, Lake Superior NERR calculates the concentrations as mg/l. Therefore, Lake Superior NERR staff divides the concentrations reported by the NRRI Central Analytical Laboratory by 1000 to yield mg/l.

Hannah Ramage was responsible for these data management tasks.

**10) Parameter titles and variable names by category –**

Required NOAA NERRS System-wide Monitoring Program nutrient parameters are denoted by an asterisk “\*”.

Data Category Parameter Variable Name Units of Measure

Phosphorus and Nitrogen:

\*Orthophosphate PO4F mg/L as P

\*Ammonium, Filtered NH4F mg/L as N

\*Nitrite, Filtered NO2F mg/L as N

\*Nitrate, Filtered NO3F mg/L as N

\*Nitrite + Nitrate, Filtered NO23F mg/L as N

Dissolved Inorganic Nitrogen DIN mg/L as N

Total Nitrogen TN mg/L as N

Total Phosphorus TP mg/L as P

Plant Pigments:

\*Chlorophyll a CHLA\_N µg/L

Other Lab Parameters:

Total Suspended Solids TSS mg/L

Notes:

1. Time is coded based on a 2400 clock and is referenced to Standard Time.

2. Reserves have the option of measuring either NO2 and NO3 or they may substitute NO23 for individual analyses if they can show that NO2 is a minor component relative to NO3.

**11) Measured or calculated laboratory parameters** –

1. **Parameters measured directly**

Nitrogen species: NH4F, NO2F, NO23F, TN

Phosphorus species: PO4F, TP

Other: CHLA\_N, TSS

1. **Calculated parameters**

NO3F NO23F-NO2F

DIN NO23F+NH4F

**12) Limits of detection** –

The Method Detection Limit (MDL) is defined as the minimum measured concentration of a substance that can be reported with 99% confidence that the measured concentration is distinguishable from method blank results. The LKS NERR Laboratory revisits MDLs annually using EPA 821-R-16-006 procedures for CHLA\_N. MDLs for TSS are calculated only using blanks processed throughout the entire year. NRRI Central Analytical Laboratory, a Minnesota state certified laboratory, revisits MDL values for every parameter annually, and reports a single value for each parameter for the year.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Parameter | Start Date | End Date | MDL | Revisited |
| NH4F | 01/01/2023 | 12/31/2023 | 0.008 | Annually |
| NO23F | 01/01/2023 | 12/31/2023 | 0.005 | Annually |
| NO2F | 01/01/2023 | 12/31/2023 | 0.007 | Annually |
| PO4F | 01/01/2023 | 12/31/2023 | 0.003 | Annually |
| TN | 01/01/2023 | 12/31/2023 | 0.020 | Annually |
| TP | 01/01/2023 | 12/31/2023 | 0.004 | Annually |
| CHLA\_N | 1/01/2023 | 12/31/2023 | 0.06 | 5/24/2023 |
| TSS | 01/01/2023 | 12/31/2023 | 2.3 | 11/08/2023 |
|  |  |  |  |  |

**13) Laboratory methods** –

* 1. **Parameter: NH4F**

NRRI Central Analytical Laboratory

Method Reference: *U.S. Environmental Protection Agency, Methods for Chemical Analysis of Water and Wastes, Method 351.2. EPA-600/4-79-020, Revised March 1983.*

Method Descriptor: *When ammonium is heated with salicylate and hypochlorite in an alkaline phosphate buffer. An emerald green color is produced which is proportional to the ammonium concentration. The presence of EDTA in the buffer prevents precipitation of calcium and magnesium. The color is intensified by adding sodium nitroprusside.*

Preservation Method: *Samples filtered (0.45 µm membrane filter) and stored at 4°C for up to 24 hours or filtered and stored at -20°C for up to 28 days.*

* 1. **Parameter: NO2F and NO23F**

NRRI Central Analytical Laboratory

Method Reference: *S. Environmental Protection Agency, Methods for Chemical Analysis of Water and Wastes.*

*Method 353.2. EPA 600 4-79-020, Revised 1983.*

Method Descriptor: *Nitrate is quantitatively reduced to nitrite by passage of the sample through a copperized*

*cadmium column. The nitrite (reduced nitrate plus original nitrite) is then determined by*

*diazotizing with sulfanilamide followed by coupling with N-(1-naphthyl) ethylenediamine*

*dihydrochloride. The resulting water soluble dye has a magenta color which is read at 520 nm.*

*Nitrite alone also can be determined by removing the cadmium column.*

Preservation Method: *Samples filtered (0.45 µm membrane filter) and stored at 4°C for up to 24 hours or filtered and stored at -20°C for up to 28 days.*

* 1. **Parameter: PO4F**

NRRI Central Analytical Laboratory

Method Reference: *Standard Methods for the Examination of Water and Wastewater, Method 4500-P E. 18th Ed. 1992. American Public Health Association, Washington, D.C.*

Method Descriptor: *Ammonium molybdate and potassium antimonyl tartrate react in acid medium with orthophosphate to form a heteropoly acid (phosphomolybdic acid) that is reduced to intensely colored molybdenum blue by ascorbic acid.*

Preservation Method: *Samples filtered (0.45 µm membrane filter) and stored at 4°C for up to 24 hours or filtered and stored at -20°C for up to 28 days.*

* 1. **Parameter: CHLA\_N**

LKS NERR Laboratory Method: *SOP Chlorophyll a Non-acidification Method Rev 1 (based on EPA Method 445.0)*

Method Reference: *US.EPA 1997. Method 445.0, In Vitro Determination of Chlorophyll a and Pheophytin a in Marine and Freshwater Algae by Fluorescence, Revision 1.2, September, 1997. Arar, E.J. and Collins, G.B., National Exposure Research Laboratory, Office of Research and Development, United States Environmental Protection Agency, Cincinnati, Ohio, 45268.*

Method Descriptor: *Chlorophyll a containing phytoplankton in surface water are concentrated by filtering through a glass fiber filter (Whatman GF/F, 0.7 µm). Pigments are extracted in 90% acetone for 24 hours. The filter slurry is centrifuged for clarification and fluorescence is measured. The Turner Design Trilogy fluorimeter provides a set of very narrow bandpass excitation and emission filters that nearly eliminate the spectral interference caused by the presence of pheophytin a and chlorophyll b.*

Preservation Method: *Samples filtered (0.7 Glass Fiber) and stored at -20°C for up to 30 days, filters are placed in a foil wrapped centrifuge tube to prevent light interference.*

* 1. **Parameter: TSS**

LKS NERR Method: *SOP Solids: Total Dissolved Solids and Total Suspended Solids Rev 1 (based on Standard Methods 2540)*

Method Reference: *Standard Methods for the Examination of Water and Wastewater, 22nd Edition, Method 2540, APHA, AWWA, WEF, Port City Press, Baltimore, Maryland, 2012.*

Method Descriptor: *A well-mixed sample is filtered through a weighed standard glass fiber filter (1.5 µm). The filter and residue retained is dried to a constant weight at 103 to 105oC. The increase in weight of the filter represents the total suspended solids.*

Preservation Method: *Refrigerate sample at 4°C for no more than 7 days. Analyzed as soon as possible due to the impracticality of preservation.*

* 1. **Parameter: TP and TN**

NRRI Central Analytical Laboratory

Method References:

*Ameel, J. R. Axler, and C. Owen. 1993. Persulfate digestion for determination of total nitrogen and phosphorus in low-nutrient waters. American Environmental Laboratory. Oct. 1993.*

*Standard Methods for the Examination of Water and Wastewater, Method 4500-Norg D. (proposed). 19th Ed. 1995. American Public Health Association, Washington, D.C.*

Method Descriptor: *An alkaline persulfate digestion simultaneously oxidizes ammonia and organic N to nitrate, and liberates phosphorus compounds as ortho-phosphate. Methods for the determination of NO3 and PO4 are then performed.*

Preservation Method: *Digestion best performed within 12 hours of collection, otherwise, samples are preserved by freezing up to 6 months.*

**14) Field and Laboratory QAQC programs** –

* 1. **Precision**
     1. **Field variability** – True field replicates (successive grab samples taken within 4 minutes of one another) were collected at every SWMP station for the Monthly Grab Sampling Program. Field replicates were not collected for the Diel Sampling Program. In total,   
        29 replicates, so 20% of samples were collected in replicate for both programs combined. Variability between replicates is analyzed using Relative Percent Difference (RPD) and is summarized in the table below.

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  |  | TP | PO4F | TN | NH4 | NO23F | NO2F | CHLA\_N | TSS |
| all sites combined | min | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
|  | max | 139 | 46 | 13 | 64 | 46 | 13 | 30 | 39 |
|  | average | 16 | 11 | 3 | 8 | 6 | 1 | 7 | 11 |
|  |  |  |  |  |  |  |  |  |  |
| lksbanut | min | 2 | 0 | 0 | 0 | 1 | 0 | 0 | 0 |
|  | max | 112 | 18 | 5 | 9 | 7 | 0 | 19 | 23 |
|  | average | 24 | 8 | 3 | 5 | 3 | 0 | 5 | 10 |
|  |  |  |  |  |  |  |  |  |  |
| lksblnut | min | 0 | 0 | 0 | 1 | 0 | 0 | 1 | 0 |
|  | max | 10 | 35 | 13 | 31 | 4 | 13 | 12 | 39 |
|  | average | 5 | 10 | 3 | 6 | 2 | 3 | 6 | 14 |
|  |  |  |  |  |  |  |  |  |  |
| lksolnut | min | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 |
|  | max | 59 | 46 | 7 | 64 | 46 | 0 | 30 | 16 |
|  | average | 12 | 12 | 3 | 14 | 19 | 0 | 8 | 5 |
|  |  |  |  |  |  |  |  |  |  |
| lksponut | min | 2 | 5 | 0 | 0 | 0 | 0 | 1 | 0 |
|  | max | 139 | 29 | 3 | 27 | 6 | 0 | 20 | 37 |
|  | average | 28 | 13 | 1 | 5 | 1 | 0 | 9 | 14 |

* + 1. **Laboratory variability** – Laboratory replicates are evaluated for CHLA\_N, TSS and all dissolved nutrient parameters. In 2023, the LKS NERR laboratory analyzed 28 CHLA-N samples in duplicate (9% of samples) and 30 TSS samples in duplicate (10% of samples). These values are for all samples analyzed in the laboratory from all research projects, including SWMP. Laboratory variability (RPD) was calculated for each and is one QC parameter that determines whether data is flagged as suspect or rejected. For all dissolved nutrient parameters, the NRRI Central Analytical Laboratory performed at least 4 laboratory replicates per sample batch. SWMP samples are processed alongside other samples in an analytical run so exact calculations of the percent of LKS NERR samples that were run in duplicate cannot be directly assessed.
    2. **Inter-organizational splits** – None in 2023
  1. **Accuracy**
     1. **Sample spikes** – The NRRI, Central Analytical Laboratory reports a quality control standard, up to ten check standard values, and at least three sample spike recoveries per sample run. These are evaluated to assess whether data should be flagged. In 2023, all quality control standards were within 90 -110% recovery. All check standards were with 90 – 110 % recovery. All sample spike recoveries were within 80 – 120 %.
     2. **Standard reference material analysis –** None in 2023
     3. **Cross calibration exercises** – *None in 2023*
     4. **Blanks -** Each month at least 1 filter blank and matrix blank is assessed for contamination during the filtration or analysis process. Annually, a method blank, which is a sample of pure water treated like a sample through the whole process from collection to analysis is assessed for each parameter. In 2023, no blanks exceeded method detection limits for any parameter analyzed in the LKS NERR laboratory or sent for analysis to the NRRI Central Analytical Laboratory.

**15) QAQC flag definitions –**

QAQC flags provide documentation of the data and are applied to individual data points by insertion into the parameter’s associated flag column (header preceded by an F\_). QAQC flags are applied to the nutrient data during secondary QAQC to indicate data that are out of sensor range low (-4), rejected due to QAQC checks (-3), missing (-2), optional and were not collected (-1), suspect (1), and that have been corrected (5). All remaining data are flagged as having passed initial QAQC checks (0) when the data are uploaded and assimilated into the CDMO ODIS as provisional plus data. The historical data flag (4) is used to indicate data that were submitted to the CDMO prior to the initiation of secondary QAQC flags and codes (and the use of the automated primary QAQC system for WQ and MET data). This flag is only present in historical data that are exported from the CDMO ODIS.

-4 Outside Low Sensor Range

-3 Data Rejected due to QAQC

-2 Missing Data

-1 Optional SWMP Supported Parameter

0 Data Passed Initial QAQC Checks

1 Suspect Data

4 Historical Data: Pre-Auto QAQC

5 Corrected Data

**16) QAQC code definitions** –

QAQC codes are used in conjunction with QAQC flags to provide further documentation of the data and are also applied by insertion into the associated flag column. There are three (3) different code categories, general, sensor, and comment. General errors document general problems with the sample or sample collection, sensor errors document common sensor or parameter specific problems, and comment codes are used to further document conditions or a problem with the data. Only one general or sensor error and one comment code can be applied to a particular data point. However, a record flag column (F\_Record) in the nutrient data allows multiple comment codes to be applied to the entire data record.

General errors

GCM Calculated value could not be determined due to missing data

GCR Calculated value could not be determined due to rejected data

GDM Data missing or sample never collected

GQD Data rejected due to QA/QC checks

GQS Data suspect due to QA/QC checks

GSM See metadata

Sensor errors

SBL Value below minimum limit of method detection

SCB Calculated value could not be determined due to a below MDL component

SCC Calculation with this component resulted in a negative value

SNV Calculated value is negative

SRD Replicate values differ substantially

SUL Value above upper limit of method detection

Parameter Comments

CAB Algal bloom

CDR Sample diluted and rerun

CHB Sample held beyond specified holding time

CIP Ice present in sample vicinity

CIF Flotsam present in sample vicinity

CLE Sample collected later/earlier than scheduled

CRE Significant rain event

CSM See metadata

CUS Lab analysis from unpreserved sample

Record comments

CAB Algal bloom

CHB Sample held beyond specified holding time

CIP Ice present in sample vicinity

CIF Flotsam present in sample vicinity

CLE Sample collected later/earlier than scheduled

CRE Significant rain event

CSM See metadata

CUS Lab analysis from unpreserved sample

*Cloud cover*

CCL clear (0-10%)

CSP scattered to partly cloudy (10-50%)

CPB partly to broken (50-90%)

COC overcast (>90%)

CFY foggy

CHY hazy

CCC cloud (no percentage)

*Precipitation*

PNP none

PDR drizzle

PLR light rain

PHR heavy rain

PSQ squally

PFQ frozen precipitation (sleet/snow/freezing rain)

PSR mixed rain and snow

*Tide stage*

TSE ebb tide

TSF flood tide

TSH high tide

TSL low tide

*Wave height*

WH0 0 to <0.1 meters

WH1 0.1 to 0.3 meters

WH2 0.3 to 0.6 meters

WH3 0.6 to > 1.0 meters

WH4 1.0 to 1.3 meters

WH5 1.3 or greater meters

*Wind direction*

N from the north

NNE from the north northeast

NE from the northeast

ENE from the east northeast

E from the east

ESE from the east southeast

SE from the southeast

SSE from the south southeast

S from the south

SSW from the south southwest

SW from the southwest

WSW from the west southwest

W from the west

WNW from the west northwest

NW from the northwest

NNW from the north northwest

*Wind speed*

WS0 0 to 1 knot

WS1 > 1 to 10 knots

WS2 > 10 to 20 knots

WS3 > 20 to 30 knots

WS4 > 30 to 40 knots

WS5 > 40 knots

**17) Other remarks/notes –**

Data may be missing due to problems with sample collection or processing. Laboratories in the NERR System submit data that are censored at a lower detection rate limit, called the Method Detection Limit or MDL. MDLs for specific parameters are listed in the Laboratory Methods and Detection Limits Section (Section II, Part 12) of this document. Concentrations that are less than this limit are censored with the use of a QAQC flag and code, and the reported value is the method detection limit itself rather than a measured value. For example, if the measured concentration of NO23F was 0.0005 mg/l as N (MDL=0.0008), the reported value would be 0.0008 and would be flagged as out of sensor range low (-4) and coded SBL. In addition, if any of the components used to calculate a variable are below the MDL, the calculated variable is removed and flagged/coded -4 SCB. If a calculated value is negative, it is rejected and all measured components are marked suspect. If additional information on MDL’s or missing, suspect, or rejected data is needed, contact the Research Coordinator at the reserve submitting the data.

Note: The way below MDL values are handled in the NERRS SWMP dataset was changed in November of 2011.  Previously, below MDL data from 2007-2010 were also flagged/coded, but either reported as the measured value or a blank cell.  Any 2007-2011 nutrient/pigment data downloaded from the CDMO prior to November of 2011 will reflect this difference.

**Sample hold times for 2023:** Standard protocol at the LKS NERR is to either refrigerate samples at 4°C and analyze for dissolved nutrients within 24 hours, or freeze and store at ≤-20°C for nutrient analysis within 28 days. NERRS SOP allows dissolved nutrient samples to be held for up to 28 days (CHLA\_N for 30) at -20°C, plus allows for up to 5 days for collecting, processing, and shipping samples. TN and TP may be held at any temp for up to 6 months. Samples held beyond that time period are flagged suspect and coded CHB and are marked with an asterisk in the table below.

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | **Date Analyzed** | | | | | | | |
| **Sample Descriptor** | **TP** | **PO4F** | **TN** | **NH4F** | **NO23F** | **NO2F** | **CHLa\_N** | **TSS** |
| 5/3 – 5/4/23 all grab and diel | 5/24 | 5/16 | 5/23 | 6/2 | 5/22 | 5/30 | 5/24 | 5/24 |
| 5/31 -6/1/23 all grab and diel | 6/16 | 6/9 | 8/7/23 | 6/28 | 6/21 | 6/26 | 6/6 | 6/1 |
| 6/29 – 6/30 all grab and diel | 7/17 | 7/27 | 8/7 | 7/20 | 7/17 | 7/25 | 7/5 | 7/18 |
| 7/11 – 7/12 all grab and diel | 8/3 | 8/9 | 8/4 | 7/20 | 8/8 | 7/25 | 7/25 | 7/25 |
| 8/18 – 8/19 all grab and diel | 9/6 | 9/12 | 9/8 | 9/14 | 8/30 | 9/12 | 8/22 | 8/21 |
| 9/13 – 9/14 all grab and diel | 10/23 | 10/10 | 10/11 | 10/6 | 9/18 | 10/12 | \*10/17 | 9/15 |
| 10/11 – grab | 10/30 | 10/26 | 10/26 | 10/20 | 10/24 | 11/7 | 10/18 | 10/12 |
| 10/16 – 10/17 diel | 10/30 | 10/26 | 10/26 | 10/20 | 10/24 | 11/7 | 11/16 | 10/23 |
| 11/8 grab | 12/15 | 11/16 | 11/22 | 11/15 | 11/16 | 11/21 | 11/16 | 11/16 |

\*sample held longer than allowed by NERRS protocols

These samples were not analyzed for all parameters due broken sample containers or technical issues during filtration or analysis:

* lksbanut 5/3/2023 15:27 all nutrient parameters
* lksponut 5/3/2023 12:07 all nutrient parameters
* lksbanut 5/31/2023 6:30 NO2F
* lksbanut 5/31/2023 6:32 NO2F
* lksponut 5/31/2023 12:40 NO2F
* lksponut 5/31/2023 12:41 NO2F
* lksbanut 6/29/2023 6:20 NO2F
* lksponut 6/29/2023 10:27 NO2F
* lksbanut 8/18/2023 12:16 NO2F
* lksponut 8/18/2023 8:45 NO2F
* lksbanut 9/14/2023 15:12 NO2F
* lksponut 9/14/2023 12:22 NO2F
* lksbanut 10/11/2023 12:48 NO2F
* lksbanut 10/11/2023 12:50 NO2F
* lksponut 10/11/2023 10:20 NO2F
* lksponut 10/11/2023 10:24 NO2F
* lksolnut 11/8/2023 13:21 all parameters
* lksolnut 11/8/2023 13:23 all parameters

There was one event when diel samples were not collected in the same time frame as the grab samples (October) because of a technical issue with the autosampler. There were no rain events or drastic shifts in weather or river flow between 10/11 when grab samples were collected and 10/19-10/17 when diel samples were collected.