**Narragansett Bay (NAR) NERR Nutrient Metadata**

**January 2013 – December 2013**

**Latest Update: September 5, 2024**

# I. Data Set and Research Descriptors

1. **Principal investigator(s) and contact persons**
   1. **Reserve Contacts**

55 South Reserve Drive

Prudence Island, RI 02872

Fax: 401-682-7366

Kenneth Raposa, PhD

Research Coordinator

(401) 683-7849

[kenneth.raposa@dem.ri.gov](mailto:kenneth.raposa@dem.ri.gov)

Daisy Durant, PhD

Marine Research Specialist II

SWMP Coordinator & Data Management

Responsible for data management

(401) 683-7368

[daisy.durant@dem.ri.gov](mailto:daisy.durant@dem.ri.gov)

* 1. **Laboratory Contacts**

Nutrient analysis

Carol Pollard, Lab Manager

Virginia Institute of Marine Science

Analytical Service Center for Nutrients

PO Box 1346

Gloucester Point, VA 23062

Phone: (804) 684-7213

Email: [pollard@vims.edu](mailto:pollard@vims.edu)

Chlorophyll analysis

Laura Reed, Lab Technician

Marine Ecosystem Research Laboratory (MERL)

University of Rhode Island

Graduate School of Oceanography

Narragansett, Rhode Island 02882

Phone: (401) 874-6619

E-mail: [reedlhht@aol.com](mailto:reedlhht@aol.com)

**2) Research objectives**

Nutrient and chlorophyll samples are being collected off Prudence Island in Narragansett Bay as part of the National Estuarine Research Reserve's (NERR) System-Wide Monitoring Program (SWMP). The goal is to develop long-term datasets for representative estuarine systems in order to track changes in water quality over time. Because Prudence Island is located in the geographic center of Narragansett Bay, it is an ideal location for monitoring the status and trends in water quality in the Bay over time. One NERR monitoring station has been established at Potter Cove, on the island's northeastern shore. This area is impacted by boat traffic and storm runoff from mainland urban and residential areas. Two NERR monitoring stations are located at T-Wharf, on the southeastern shore of the island facing the open waters of Rhode Island Sound. The two T-Wharf stations (T-Wharf Surface and T-Wharf Bottom) measure conditions in the upper (~0.5 m from the surface) and bottom (~1 m from the bottom) layers of the water column, respectively. These stations are approximately 10 km south of the Potter Cove site. Boat traffic is sparse at T-Wharf and storm runoff is less likely to have a significant impact on water quality. A fourth monitoring site is located in Nag Creek, a salt marsh tidal creek which flows into the West Passage of Narragansett Bay. The addition of this site completes our representation of dominant habitat types occurring in Narragansett Bay (i.e. marsh, cove, and open water).

* 1. **Monthly grab sampling program**

Monthly grab samples are collected once per month from each of the four stations to quantify seasonal patterns of selected nutrient species and chlorophyll in different estuarine habitats (marsh creek, cove, open water surface, open water bottom).

* 1. **Diel sampling program**

Once per month, samples are collected at approximately 2 hour 15 minute intervals over a 24 hour cycle to document changes in nutrients and chlorophyll in response to tidal forcing. Previously (from 2002 to 2010) the station was located at T-Wharf. However, after analyzing the historic data from the site, no significant trend or patterns were found over time. Thus, the station was moved and has been at Potter Cove since January of 2011, in order to characterize nutrients and chlorophyll from this site.

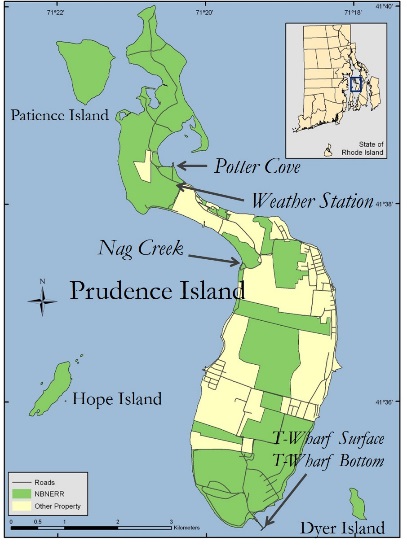
1. **Research methods**
   1. **Monthly grab sampling program**

Monthly grab samples were taken at the four permanent stations (Potter Cove, Nag Creek, T-Wharf Surface, and T-Wharf Bottom) each month throughout 2013. All grab samples were taken on the same day between 2 to 3 hrs. before slack low-water and slack low-water. No distinction was made between neap and spring tide conditions. Replicate (N=2) samples were collected by hand. All samples were collected in amber plastic wide-mouth samples bottles. All sampling bottles were previously acid washed and rinsed (3x) with distilled-deionized water, dried and followed by rinsing (3x) of ambient water prior to collection of the sample. Samples were immediately placed on ice, in the dark and returned to the laboratory for filtering. Filtered samples were frozen at -20°C and transported as quickly as possible to the Marine Ecosystems Research Laboratory (MERL) at the University of Rhode Island’s Graduate School of Oceanography (URI/GSO) for chlorophyll analyses, and mailed overnight to the Virginia Institute of Marine Science (VIMS) for nutrient analyses.

* 1. **Diel sampling program**

The diel sampling program was conducted once per month throughout 2013. For this program, an ISCO 6712 portable water sampler was deployed at the Potter Cove station. This device automatically samples 500 ml of water every 2 hrs. 15 min. All samples are pumped into polyethylene sample bottles that were previously acid washed and rinsed (3x) with distilled-deionized water and dried. At the end of the 24 hr. period, the 12 samples were kept in the dark, returned to the laboratory on Prudence Island for filtering, and then frozen at -20°C. Samples were then transported as quickly as possible to the Marine Ecosystems Research Laboratory (MERL) at the University of Rhode Island’s Graduate School of Oceanography (URI/GSO) for chlorophyll analyses, and mailed overnight to the Virginia Institute of Marine Science (VIMS) for nutrient analyses.

1. **Site location and character**

The NAR NERR consists of 4453 acres of diverse estuarine and terrestrial habitats ranging from deep water to salt marshes to forested uplands. The land holdings include approximately 65% of Prudence Island, most of nearby Patience Island, and all of Hope and Dyer Islands. The Reserve is located close to the geographic center of Narragansett Bay in Rhode Island. The Bay has a drainage basin of 1800 square miles.

Specific characteristics of the Narragansett Bay National Estuarine Research Reserve

Location: 41°38’30” N, 71°20’30” W

Tidal range: -0.2 to 1.7 meters MLW

Salinity: 15 to 32 ppt

Temperature: -1.0 to 26 C

Province: North temperate, Virginian bioregion

Specific characteristics of the Potter Cove site are:

Location: 41°38'25.984" N, 71°20'27.165" W

Depth: 0.9 to 3.9 meters

Bottom habitat: Sand, silt, some organic mud

Pollutants: Boaters’ wastes, storm runoff from mainland urban areas

Watershed: Narragansett Bay, North Prudence (4801 square km)

Specific characteristics of the Nag Creek site are:

Location: 41°37'29.458" N, 71°19'27.421" W

Depth: 0.1 to 1.4 meters

Bottom habitat: Organic mud

Pollutants: Negligible

Watershed: Narragansett Bay, West Passage

Specific characteristics of the T-Wharf Surface site are:

Location: 41°34'42.099" N, 71°19'16.049" W

Depth: 0.2 to 0.9 meters

Bottom habitat: Sand, silt, some organic mud

Pollutants: Negligible

Watershed: Narragansett Bay, South Prudence

Specific characteristics of the T-Wharf Bottom site are:

Location: 41°34'42.099" N, 71°19'16.049" W

Depth: 4.6 to 6.9 meters

Bottom habitat: Sand, silt, some organic mud

Pollutants: Negligible

Watershed: Narragansett Bay, South Prudence

1. **Coded variable definitions**

Station Code Names

narncnut = Nag Creek

narpcnut = Potter Cove

nartbnut = T-Wharf Bottom

nartsnut = T-Wharf Surface

Monthly grab sample program = 1

Diel grab sample program = 2

1. **Data collection period**

The Narragansett Bay National Estuarine Research Reserve started grab sampling at each of the four sites in the spring of 2002. Diel sampling started in 2002 at T-Wharf Bottom until 2010; it has been at Potter Cove since 2011. For both Grab and Diel sampling, time is coded based on a 2400 hour clock and is referenced to Standard Time (ST) format.

Monthly Grab Sampling

|  |  |  |  |
| --- | --- | --- | --- |
| Site | Start Date | Start Time | End Time |
| 1 narncnut | 01/09/13 |  |  |
| 1 narncnut | 02/06/13 |  |  |
| 1 narncnut | 03/11/13 |  |  |
| narncnut | 04/10/13 | 11:00 | 11:02 |
| narncnut | 05/06/13 | 10:11 | 10:13 |
| narncnut | 06/05/13 | 11:26 | 11:28 |
| narncnut | 07/05/13 | 11:03 | 11:05 |
| narncnut | 08/05/13 | 12:23 | 12:25 |
| narncnut | 09/05/13 | 12:53 | 12:55 |
| narncnut | 10/04/13 | 12:56 | 12:58 |
| narncnut | 11/04/13 | 12:00 | 12:02 |
| narncnut | 12/03/13 | 12:18 | 12:20 |
| narpcnut | 01/09/13 | 10:38 | 10:40 |
| narpcnut | 02/06/13 | 09:42 | 09:44 |
| narpcnut | 03/11/13 | 10:50 | 10:52 |
| narpcnut | 04/10/13 | 11:25 | 11:27 |
| narpcnut | 05/06/13 | 09:45 | 09:47 |
| narpcnut | 06/05/13 | 11:01 | 11:03 |
| narpcnut | 07/05/13 | 10:24 | 10:26 |
| narpcnut | 08/05/13 | 11:36 | 11:38 |
| narpcnut | 09/05/13 | 12:17 | 12:19 |
| narpcnut | 10/04/13 | 11:56 | 11:58 |
| narpcnut | 11/04/13 | 12:37 | 12:39 |
| narpcnut | 12/03/13 | 11:44 | 11:46 |

Notes

1 Sampling was not done because the creek was frozen.

Monthly Grab Sampling (cont.)

|  |  |  |  |
| --- | --- | --- | --- |
| Site | Start Date | Start Time | End Time |
| nartbnut | 01/09/13 | 10:03 | 10:05 |
| nartbnut | 02/06/13 | 09:07 | 09:09 |
| nartbnut | 03/11/13 | 11:30 | 11:32 |
| nartbnut | 04/10/13 | 10:25 | 10:27 |
| nartbnut | 05/06/13 | 11:04 | 11:06 |
| nartbnut | 06/05/13 | 12:19 | 12:21 |
| nartbnut | 07/05/13 | 09:21 | 09:24 |
| nartbnut | 08/05/13 | 10:50 | 10:52 |
| nartbnut | 09/05/13 | 11:58 | 12:00 |
| nartbnut | 10/04/13 | 11:00 | 11:03 |
| nartbnut | 11/04/13 | 13:19 | 13:21 |
| nartbnut | 12/03/13 | 13:03 | 13:05 |
| nartsnut | 01/09/13 | 09:58 | 10:00 |
| nartsnut | 02/06/13 | 09:03 | 09:05 |
| nartsnut | 03/11/13 | 11:25 | 11:27 |
| nartsnut | 04/10/13 | 10:17 | 10:19 |
| nartsnut | 05/06/13 | 10:55 | 10:57 |
| nartsnut | 06/05/13 | 12:15 | 12:17 |
| nartsnut | 07/05/13 | 09:18 | 09:20 |
| nartsnut | 08/05/13 | 10:38 | 10:40 |
| nartsnut | 09/05/13 | 11:48 | 11:50 |
| nartsnut | 10/04/13 | 10:48 | 10:50 |
| nartsnut | 11/04/13 | 13:15 | 13:17 |
| nartsnut | 12/03/13 | 12:54 | 12:56 |

Diel Sampling

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Site | Start Date | Start Time | End Date | End Time |
| narpcnut | 01/29/13 | 00:00 | 01/30/13 | 00:45 |
| narpcnut | 02/20/13 | 00:00 | 02/21/13 | 00:45 |
| narpcnut | 03/25/13 | 10:38 | 03/26/13 | 11:23 |
| narpcnut | 04/23/13 | 08:53 | 04/24/13 | 09:38 |
| narpcnut | 05/21/13 | 07:24 | 05/22/13 | 08:09 |
| narpcnut | 06/20/13 | 07:37 | 06/21/13 | 08:22 |
| narpcnut | 07/17/13 | 05:03 | 07/18/13 | 05:48 |
| narpcnut | 08/20/13 | 10:02 | 08/21/13 | 10:47 |
| narpcnut | 09/18/13 | 09:56 | 09/19/13 | 10:41 |
| narpcnut | 10/23/13 | 01:09 | 10/24/13 | 01:54 |
| narpcnut | 11/20/13 | 10:18 | 11/21/13 | 11:03 |
| narpcnut | 12/17/13 | 11:02 | 12/18/13 | 11:47 |

1. **Associated researchers and projects**

Since 2004, the SWMP nutrient monitoring program is part of a larger, multi-institution effort to monitor water quality and other associated parameters in Narragansett Bay, RI, called the Narragansett Bay Window. This includes monitoring efforts conducted by the University of Rhode Island’s Graduate School of Oceanography, the Narragansett Bay Estuary Program, and NMFS and EPA in Narragansett, RI, among others. Although the NAR NERR is no longer involved with managing data from the Bay window program, NAR NERR SWMP data are still shared with and used for analysis in the program.

sondeThe NAR NERR System-Wide Monitoring Program (SWMP) has four water quality monitoring stations around Prudence Island. The principle objective of the SWMP program is to record short-term variability and long-term changes in water quality data in order to observe trends or patterns in water quality over time. Water quality parameters have been collected since 1995 with the establishment of the first water quality monitoring station at Potter Cove. The other three water quality stations (Nag Creek, T-Wharf Surface and T-Wharf Bottom) were brought online in 2002. These stations were selected to represent a gradient in habitat types that range from salt marsh (Nag Creek station) to shallow cove (Potter Cove) to open Bay water (T-Wharf Surface and T-Wharf Bottom). Water temperature (°C), salinity (ppt), dissolved oxygen (% saturation, and mg L-1), pH, turbidity (NTU), and chlorophyll (μg L-1) data are collected at each station every 15 minutes using data loggers (see image at right) that are calibrated and swapped out at each station approximately every two weeks.

YSI 6600 V2 datalogger used to collect water quality data.

Since 2001, meteorological data has been collected as part of the SWMP at the weather station (see image below) located on Prudence Island, approximately 389 m south of Potter Cove (see map on section 4 above). Data on air temperature, relative humidity, barometric pressure, wind speed and direction, photosynthetic active radiation, and precipitation are collected. Meteorological data is continually used to complement the water quality, biological monitoring and scientific research efforts at NAR NERR and at Narragansett Bay, and to assist educational activities around the Bay.

All this information is available to any interested party through the CDMO <http://cdmo.baruch.sc.edu/>, NAR NERR <http://nbnerr.org/>, or directly contacting the Research Coordinator or the Marine Research Specialist II.

The weather station at Prudence Island has been collecting data since 2001.

1. **Distribution**

NOAA retains the right to analyze, synthesize and publish summaries of the NERRS System-wide Monitoring Program data.  The NERRS retains the right to be fully credited for having collected and processed the data.  Following academic courtesy standards, the NERR site where the data were collected should be contacted and fully acknowledged in any subsequent publications in which any part of the data are used.  The data set enclosed within this package/transmission is only as good as the quality assurance and quality control procedures outlined by the enclosed metadata reporting statement.  The user bears all responsibility for its subsequent use/misuse in any further analyses or comparisons.  The Federal government does not assume liability to the Recipient or third persons, nor will the Federal government reimburse or indemnify the Recipient for its liability due to any losses resulting in any way from the use of this data.

Requested citation format:

National Estuarine Research Reserve System (NERRS). 2012.  System-wide Monitoring Program. Data accessed from the NOAA NERRS Centralized Data Management Office website: www.nerrsdata.org; *accessed* 12 October 2012.

NERR nutrient data and metadata can be obtained from the Research Coordinator at the individual NERR site (please see Principal investigators and contact persons), from the Data Manager at the Centralized Data Management Office (please see personnel directory under the general information link on the CDMO home page) and online at the CDMO home page [www.nerrsdata.org](http://cfcdmo.baruch.sc.edu/). Data are available in comma separated version format.

**II. Physical Structure Descriptors**

1. **Entry verification**

Nutrient data are entered into a Microsoft Excel worksheet and processed using the NutrientQAQC Excel macro. The NutrientQAQC macro sets up the data worksheet, metadata worksheets, and MDL worksheet; adds chosen parameters and facilitates data entry; allows the user to set the number of significant figures to be reported for each parameter and rounds using banker’s rounding rules; allows the user to input MDL values and then automatically flags/codes measured values below MDL and inserts the MDL; calculates parameters chosen by the user and automatically flags/codes for component values below MDL, negative calculated values, and missing data; allows the user to apply QAQC flags and codes to the data; produces summary statistics; graphs selected parameters for review; and exports the resulting data file to the CDMO for tertiary QAQC and assimilation into the CDMO’s authoritative online database.

The Virginia Institute of Marine Science (VIMS) entered the nutrient data into Microsoft Excel spreadsheets, calculated and reported results in mg L-1, and sent the files electronically to the Reserve. The Marine Ecosystem Research Laboratory (MERL) entered the chlorophyll and phaeophytin data into Microsoft Excel spreadsheets, calculated and reported the results in µg L-1 and also sent the files electronically to the Reserve. For purposes of consistency in the NERR System, concentrations are calculated as mg L-1 based on atomic weights of 14.010, 30.97, and 28.09, and 12.01 for N, P, Si, and C, respectively.

Data entry verification was completed by Dr. Daisy Durant. Final verification and this metadata documentation were checked by Dr. Kenneth Raposa, before being sent to the CDMO permanent database.

**10) Parameter titles and variable names by category**

Required NOAA/NERRS System-wide Monitoring Program nutrient parameters are denoted by an asterisks “\*”.

|  |  |  |  |
| --- | --- | --- | --- |
| Data Category | Parameter | Variable Name | Units of Measure |
| Phosphorus and | \*Orthophosphate | PO4F | mg/L as P |
| Nitrogen: | \*Ammonium, Filtered | NH4F | mg/L as N |
|  | \*Nitrite, Filtered | NO2F | mg/L as N |
|  | \*Nitrate, Filtered | NO3F | mg/L as N |
|  | \*Nitrite + Nitrate, Filtered | NO23F | mg/L as N |
|  | Dissolved Inorganic Nitrogen | DIN | mg/L as N |
| Plant Pigments: | \*Chlorophyll a | CHLA\_N | µg/L |
|  | Phaeophytin | PHEA | µg/L |
| Other Lab Parameters: | Silicate, Filtered | SiO4F | mg/L as Si |
| Field Parameters: | Water Temperature | WTEM\_N | °C |

Notes:

* Time is coded based on a 2400 clock and is referenced to Standard Time format.
* Reserves have the option of measuring either NO2 or NO3 or they may substitute NO23 for individual analyses if they can show that NO2 is a minor component relative to NO3.

**11) Measured and calculated laboratory parameters**

1. **Parameters measured directly**

* Nitrogen species: NH4, NO2, NO23
* Phosphorus species: PO4F
* Other: CHLA, PHEA, SiO4, WTEM

1. **Calculated parameters**

* NO3: NO23-NO2
* DIN: NO23+NH4

**12)** **Limits of detection**

Method Detection Limits (MDL), the lowest concentration of a parameter that an analytical procedure can reliably detect, have been established by VIMS Analytical Service Center for Nutrient and by MERL for chlorophyll and Phaeophytin. The MDL is determined as 3 times the standard deviation of a minimum of 7 replicates of a single low concentration sample. These values are reviewed and revised periodically.

Method Detection Limits

|  |  |  |  |
| --- | --- | --- | --- |
| Parameter | Start Date | End Date | MDL |
| PO4F | 01/01/2013 | 12/31/2013 | 0.0020 mg L-1 |
| NH4F | 01/01/2013 | 12/31/2013 | 0.0056 mg L-1 |
| NO2F | 01/01/2013 | 12/31/2013 | 0.0016 mg L-1 |
| NO23F | 01/01/2013 | 12/31/2013 | 0.0047 mg L-1 |
| SiO4 | 01/01/2013 | 12/31/2013 | 0.0800 mg L-1 |
| CHLA\_N | 01/01/2013 | 12/31/2013 | 0.05 µg L-1 |
| PHEA | 01/01/2013 | 12/31/2013 | 0.05 µg L-1 |

**13)** **Laboratory methods**

1. **Parameter:** **Chlorophyll *a* (CHLA\_N), Phaeophytin (Phae)**
   * Marine Ecosystems Research Laboratory Method
   * Method References: Oviatt, C. A., and K. M. Hindle, 1994. Manual of biological and geochemical techniques in coastal areas. pp. 3-7.
   * Method Descriptor: Chlorophyll *a* is extracted in 10 ml 90% acetone and fluorescence is measured and recorded (Fo). Two drops of 10% hydrochloric acid are added to convert the chl to phaeopigments (Phae). The fluorescence is again measured and recorded (Fa). The concentration (μg L-1) of Chl *a* and Phae are calculated using the Fo/Fa ratio.
   * Preservation Method: 20 ml of sample is filtered onto a Whatman® Glass Microfiber Filter: Type GF/F (25 mm), treated with 2 drops of MgCO3 solution, folded in half, wrapped in aluminum foil, and stored at –20°C until analysis.
2. **Parameter: NH4F**
   * VIMS Laboratory Method: SA156-350.1 - NH3
   * Method Reference:

* U.S. EPA. 1974. Methods for Chemical Analysis of Water and Wastes, pp. 168-174.
* Standard Methods for the Examination of Water and Wastewater, 14th edition. p 410. Method 418A and 418B (1975).
* Annual Book of ASTM Standards, Part 31. "Water", Standard 1426-74, Method A, p 237 (1976).
* EPA 600/R-97/072 Method 349.0. Determination of Ammonia in Estuarine and Coastal Waters by Gas Segmented Continuous Flow Colorimetric Analysis. IN: Methods for the Determination of Chemical Substances in Marine and Estuarine Environmental Matrices -2nd Edition. National Exposure Research Laboratory, Office of Research and Development U.S. EPA, Cincinnati, Ohio 45268.
  + Method Descriptor: Ammonia reacts with alkaline phenol and hypochlorite to form indophenol blue in an amount that is proportional to the ammonia concentration. The blue color is intensified with sodium nitroprusside. The reaction is catalyzed by heat at 37°C. The range is 0.01 – 2.0 mg L-1.
  + Preservation Method: 60 ml of sample is filtered through a Whatman® Nuclepore Track-Etched Membrane, 0.4 µm disposable disk filter (47 mm), and stored at –20°C until analyzed.

1. **Parameter: NO2F, and NO2F + NO3F**
   * VIMS Laboratory Method: SA461-353.2 - NO2+3, NO2, TDN
   * Method Reference:

* U.S. EPA. 1974. Methods for Chemical Analysis of Water and Wastes, pp. 207 -212.
* Wood, E.D., F.A.G. Armstrong, and F.A. Richards. 1967. Determination of nitrate in seawater by cadmium-copper reduction to nitrite. J. Mar. Biol. Assoc. U.K. 47: 23
* Grasshoff, K., M. Ehrhardt and K. Kremling. 1983. Methods of Seawater Analysis. Verlag Chemie, Federal Republic of Germany. 419 pp.
* EPA 600/R-97/072 Method 353.4. Determination of Nitrate and Nitrite in Estuarine and Coastal Waters by Gas Segmented Flow Colorimetric Analysis. IN: Methods for the Determination of Chemical Substances in Marine and Estuarine Environmental Matrices -2nd Edition. National Exposure Research Laboratory, Office of Research and Development U.S. EPA, Cincinnati, Ohio 45268.
* Method Descriptor: Nitrate is reduced quantitatively to nitrite by cadmium metal in the form of an open tubular reactor. The nitrite thus formed plus any originally present in the sample is colorimetrically detected at 540 nm, following its diazotization with sulfanilamide, and subsequent coupling with N-1-naphthylethylenediamine dihydrochloride. The dissolved nitrate concentrations of the samples are calculated as NO3-N.
  + Preservation Method: 60 ml of sample is filtered through a Whatman® Nuclepore Track-Etched Membrane, 0.4 µm disposable disk filter (47 mm), and stored at –20°C until analyzed.

1. **Parameter:** **SIO2F**
   * VIMS Laboratory Method: Skalar Method Silicates, Catnr. 563-052 issue 101899/MH/99208255
   * Method Reference:

* U.S. EPA. 1974. Methods for Chemical Analysis of Water and Wastes, Method 370.1
* Grasshoff, K., M. Ehrhardt and K. Kremling. 1983. Methods of Seawater Analysis. Verlag Chemie, Federal Republic of Germany. Pp 374-376.
* EPA 600/R-97/072 Method 366.0 Determination of Dissolved Silicate in Estuarine and Coastal Waters by Gas Segmented Flow Colorimetric Analysis. IN: Methods for the Determination of Chemical Substances in Marine and Estuarine Environmental Matrices -2nd Edition. National Exposure Research Laboratory, Office of Research and Development U.S. EPA, Cincinnati, Ohio 45268.
  + Method Descriptor: Based on the reduction of silicomolybdate in acidic solution to “molybdenum blue” by ascorbic acid. Oxalic acid is introduced to the sample stream before the addition of ascorbic acid to eliminate interference from phosphates. The range is 0-1.1 mg Si L-1.
  + Preservation Method: Preservation Method: 60 ml of sample is filtered through a Whatman® Nuclepore Track-Etched Membrane, 0.4 µm disposable disk filter (47 mm), and stored at –4°C until analyzed.

1. **Parameter: PO4F**

* VIMS Laboratory Method SKALAR Method: O-Phosphate / Total Phosphate, Catnr. 503-365.1, issue 042993/MH/93-Demo1
* Method Reference:
  + - Murphy, J. and J.P. Riley. 1962. A modified single solution method for the determination of phosphate in natural waters. Analytica Chim. Acta 27: 31-36
    - EPA 600/R-97/072 Method 365.5 Determination of Orthophosphate in Estuarine and Coastal Waters by Automated Colorimetric Analysis. IN: Methods for the Determination of Chemical Substances in Marine and Estuarine Environmental Matrices -2nd Edition. National Exposure Research Laboratory, Office of Research and Development. U.S. EPA, Cincinnati, Ohio 45268
  + Method Descriptor: Ammonium molybdate and antimony potassium tartrate react in a sulfuric acid environment to form an antimony-phospho-molybdo complex, which is reduced to a blue colored complex by ascorbic acid. Reaction is heat catalyzed at 40°C.
  + Range is 1 - 50 ppb.
  + Preservation Method: Preservation Method: 60 ml of sample is filtered through a Whatman® Nuclepore Track-Etched Membrane, 0.4 µm disposable disk filter (47 mm), and stored at –20°C until analyzed.

**14) Field and Laboratory QA/QC Programs**

1. **Precision**
   1. **Field variability** - For the monthly grab sampling program, NAR NERR collects two successive grab samples for the determination of water mass variability.
   2. **Laboratory variability** – replicates of all nutrient samples are run
   3. **Inter-organizational splits** – none
2. **Accuracy**
   1. **Sample spikes** – blanks
   2. **Standard reference material analysis** – none
   3. **Cross calibration exercises** – A cross calibration of the Turner Designs Fluorometer was performed with Battelle Ocean Sciences.

**15) QAQC flag definitions**

QAQC flags provide documentation of the data and are applied to individual data points by insertion into the parameter’s associated flag column (header preceded by an F\_). QAQC flags are applied to the nutrient data during secondary QAQC to indicate data that are out of sensor range low (-4), rejected due to QAQC checks (-3), missing (-2), optional and were not collected (-1), suspect (1), and that have been corrected (5). All remaining data are flagged as having passed initial QAQC checks (0) when the data are uploaded and assimilated into the CDMO ODIS as provisional plus data. The historical data flag (4) is used to indicate data that were submitted to the CDMO prior to the initiation of secondary QAQC flags and codes (and the use of the automated primary QAQC system for WQ and MET data). This flag is only present in historical data that are exported from the CDMO ODIS.

-4 Outside Low Sensor Range

-3 Data Rejected due to QAQC

-2 Missing Data

-1 Optional SWMP Supported Parameter

0 Data Passed Initial QAQC Checks

1 Suspect Data

4 Historical Data: Pre-Auto QAQC

5 Corrected Data

**16) QAQC code definitions**

QAQC flags provide documentation of the data and are also applied by insertion into the associated flag column. There are three (3) different code categories, general, sensor, and comment. General errors document general problems with the sample or sample collection, sensor errors document common sensor or parameter specific problems, and comment codes are used to further document conditions or a problem with the data. Only one general or sensor error and one comment code can be applied to a particular data point. However, a record flag column (F\_Record) in the nutrient data allows multiple comment codes to be applied to the entire data record.

General errors

GCM Calculated value could not be determined due to missing data

GCR Calculated value could not be determined due to rejected data

GDM Data missing or sample never collected

GQD Data rejected due to QA/QC checks

GQS Data suspect due to QA/QC checks

Sensor errors

SBL Value below minimum limit of method detection

SCB Calculated value could not be determined due to a below MDL component

SCC Calculation with this component resulted in a negative value

SNV Calculated value is negative

SRD Replicate values differ substantially

SUL Value above upper limit of method detection

Parameter Comments

CAB Algal bloom

CDR Sample diluted and rerun

CHB Sample held beyond specified holding time

CIP Ice present in sample vicinity

CIF Flotsam present in sample vicinity

CLE Sample collected later/earlier than scheduled

CRE Significant rain event

CSM See metadata

CUS Lab analysis from unpreserved sample

Record comments

CAB Algal bloom

CHB Sample held beyond specified holding time

CIP Ice present in sample vicinity

CIF Flotsam present in sample vicinity

CLE Sample collected later/earlier than scheduled

CRE Significant rain event

CSM See metadata

CUS Lab analysis from unpreserved sample

*Cloud cover*

CCL clear (0-10%)

CSP scattered to partly cloudy (10-50%)

CPB partly to broken (50-90%)

COC overcast (>90%)

CFY foggy

CHY hazy

CCC cloud (no percentage)

*Precipitation*

PNP none

PDR drizzle

PLR light rain

PHR heavy rain

PSQ squally

PFQ frozen precipitation (sleet/snow/freezing rain)

PSR mixed rain and snow

*Tide stage*

TSE ebb tide

TSF flood tide

TSH high tide

TSL low tide

*Wave height*

WH0 0 to <0.1 meters

WH1 0.1 to 0.3 meters

WH2 0.3 to 0.6 meters

WH3 0.6 to > 1.0 meters

WH4 1.0 to 1.3 meters

WH5 1.3 or greater meters

*Wind direction*

N from the north

NNE from the north northeast

NE from the northeast

ENE from the east northeast

E from the east

ESE from the east southeast

SE from the southeast

SSE from the south southeast

S from the south

SSW from the south southwest

SW from the southwest

WSW from the west southwest

W from the west

WNW from the west northwest

NW from the northwest

NNW from the north northwest

*Wind speed*

WS0 0 to 1 knot

WS1 > 1 to 10 knots

WS2 > 10 to 20 knots

WS3 > 20 to 30 knots

WS4 > 30 to 40 knots

WS5 > 40 knots

**17)** **Other Remarks**

Data may be missing due to problems with sample collection or processing. Laboratories in the NERRS System submit data that are censored at a lower detection rate limit, called the Method Detection Limit or MDL. MDLs for specific parameters are listed in the Laboratory Methods and Detection Limits Section (Section II, Part 12) of this document. Concentrations that are less than this limit are censored with the use of a QAQC flag and code, and the reported value is the method detection limit itself rather than a measured value. For example, if the measured concentration of NO23F was 0.0005 mg/l as N (MDL=0.0008), the reported value would be 0.0008 and would be flagged as out of sensor range low (-4) and coded SBL. In addition, if any of the components used to calculate a variable are below the MDL, the calculated variable is removed and flagged/coded -4 SCB. If a calculated value is negative, it is rejected and all measured components are marked suspect. If additional information on MDL’s or missing, suspect, or rejected data is needed, contact the Research Coordinator at the Reserve submitting the data.

Note: The way below MDL values are handled in the NERRS SWMP dataset was changed in November of 2011.  Previously, below MDL data from 2007-2010 were also flagged/coded, but either reported as the measured value or a blank cell.  Any 2007-2011 nutrient/pigment data downloaded from the CDMO prior to November of 2011 will reflect this difference.

The following are descriptions of different events that happened during the collection and filtering processes at the Reserve, and explanations of the CSM codes in the dataset of 2013.

**CSM**

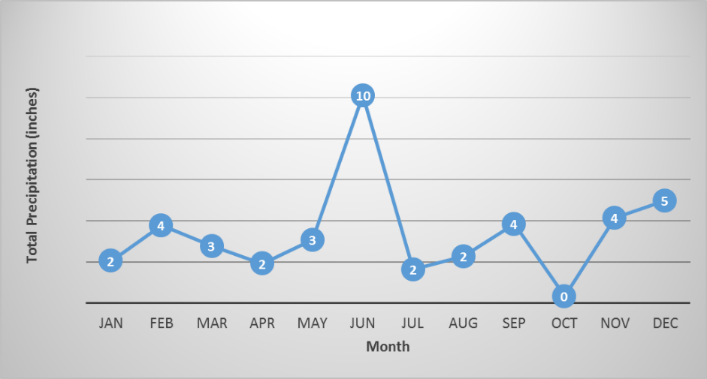
Grab samples from January 09 and February 06 and diel samples from January 29, 2013 stored at the Reserve were thawed out during a power outage due to a winter storm. These samples were not sent out for analysis as recommended by the SOP, VIMS and MERL.

**Missing samples**

Samples from Nag Creek were not collected on March 11 2013 because the creek was frozen.

Diel sample lost at the reserve from Nov 20, 2013 19:18 – only the nutrient sample was lost

Total Precipitation per month on Prudence Island for 2013. Data from NAR NERR Weather Station.



The NERRS nutrient SOP allows nutrient samples to be held for up to 28 days (CHLA for 30) at -20°C, plus allows for up to 5 days for collecting, processing, and shipping samples. Samples held beyond that time period are flagged suspect <1> and coded (CHB). If measured values were below MDL, this resulted in <-4> [SBL] (CHB) flagging/coding. Dates shown are month and day of 2013.

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| NAR NERR Sampling Dates | | VIMS: DATES OF ANALYSES | | | | | Lab total number of days to perform analyses | |
|
| DIEL | GRAB | NH4 | NO23 | NO2 | PO4 | SiO4 | DIEL | GRAB |
| 01/29 | 01/09 | \* | \* | \* | \* | \* |  |  |
|  | 02/06 | \* | \* | \* | \* | \* |  |  |
| 02/20 | 03/11 | \*\* | \*\* | \*\* | \*\* | \*\* |  |  |
| 03/25 |  | \*\* | \*\* | \*\* | \*\* | \*\* |  |  |
| 04/23 | 04/10 | 08/09 | 08/09 | 08/09 | 08/09 | 08/09 | 108 | 121 |
|  | 05/06 | 09/11 | 09/11 | 09/11 | 09/11 | 09/11 |  | 128 |
| 05/21 | 06/05 | 08/09 | 08/09 | 08/09 | 08/09 | 08/09 | 80 | 65 |
| 06/20 | 07/05 | 09/30 | 09/30 | 09/30 | 09/30 | 09/30 | 102 | 87 |
| 07/17 | 08/05 | 10/01 | 10/01 | 10/01 | 10/01 | 10/01 | 76 | 57 |
| 08/20 | 09/05 | 10/01 | 10/01 | 10/01 | 10/01 | 10/01 | 42 | 26 |
| 09/18 | 10/04 | 11/05 | 11/05 | 11/05 | 11/05 | 11/06 | 48 | 32 |
| 10/23 | 11/04 | 12/12 | 12/12 | 12/12 | 12/12 | 12/13 | 50 | 38 |
| 11/20 | 12/03 | 12/12 | 12/12 | 12/12 | 12/12 | 12/12 | 22 | 9 |
| 12/17 |  | 02/13 | 02/13 | 02/13 | 02/13 | 02/14 | 58 |  |

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| MERL: DATES OF ANALYSES (Chlorophyll and Phaeophytin) | | | | | |
| Diel Sampling | | | Grab Sampling | | |
| Sampling Date | Run Date | Lab total number of days to perform analyses | Sampling Date | Run Date | Lab total number of days to perform analyses |
| 01/29 | \* |  | 01/09 | \* |  |
| 02/20 | 03/21 | 29 | 02/06 | \* |  |
| 03/25 | 04/04 | 10 | 03/11 | 03/21 | 10 |
| 04/23 | 05/02 | 9 | 04/10 | 04/19 | 9 |
| 05/21 | 05/31 | 10 | 05/06 | 05/10 | 4 |
| 06/20 | 07/12 | 22 | 06/05 | 06/26 | 21 |
| 07/17 | 08/01 | 15 | 07/05 | 07/12 | 7 |
| 08/20 | 08/30 | 10 | 08/05 | 08/09 | 4 |
| 09/18 | 09/27 | 9 | 09/05 | 09/12 | 7 |
| 10/22 | 11/07 | 16 | 10/04 | 10/10 | 6 |
| 11/20 | 11/26 | 6 | 11/04 | 11/07 | 3 |
| 12/17 | 12/27 | 10 | 12/03 | 12/13 | 10 |

Notes

\*Samples thawed out during winter storm power outage. Analyses were not performed.

\*\*Dates were not provided by the lab.