

```
In [14]: % some housekeeping stuff
register_graphics_toolkit ("gnuplot");
available_graphics_toolkits ();
graphics_toolkit ("gnuplot")
clear
% load packages
pkg load statistics
% clear and warning off
clear
warning off
% end of housekeeping
```

Species sensitivity distribution (SSD)



Species sensitivity distributions are used to help set guidelines for metals in aquatic environments.

<https://entoxsimplified.com/2018/08/03/species-sensitivity-distribution/#comments>

- different jurisdictions have different tolerances, and safety factors
- SSD can be modified to take into account receiving water chemistry using the BLM

Biotic Ligand Model (BLM)



BLM is used for risk assessment of metals in the environment. It can even be used as part of "permit writing" (license to pollute).

- three C's
- based on chemical equilibrium modelling
- all the types of reactions we talk about PLUS reaction at the biotic ligand (such as fish gill)
- toxicity proportional to accumulation at the biotic ligand
- replaces the free ion activity model (FIAM)
- "invented" at Laurier ! (Professor Rick Playle)
- need 7 water chemistry parameters (Na, Ca, K, Mg, Cl, Alk, DOC)

reaction of metal at the biotic ligand

This is a simple metal-ligand complexation reaction. For metal consider M^{2+} and ligand BL^- . Let's assume a binding capacity (BL_{T}) of 10 nM and a logK value of 7.5 for metal complexation at the BL. Assume 50% saturation of the binding site for 50% toxicity (LA50).

LA50 does not change, but the LC50 does change.

```
In [15]:
```

```

%plot -s 600,500 -f 'svg'
BLT=10e-9; logKf=7.5; logMT=-9:0.1:-5; MT=10.^logMT;

TABLEAU=[...
%M    BL    logK    species
1     0     0      {'M'}
0     1     0      {'BL'}
1     1    logKf    {'MBL'}
];

[N,M]=size(TABLEAU); ASOLUTION=cell2mat(TABLEAU(1:N,1:M-2)); KSOLUTION=cell2mat(
SOLUTIONNAMES=strvcat(TABLEAU(1:N,M)));

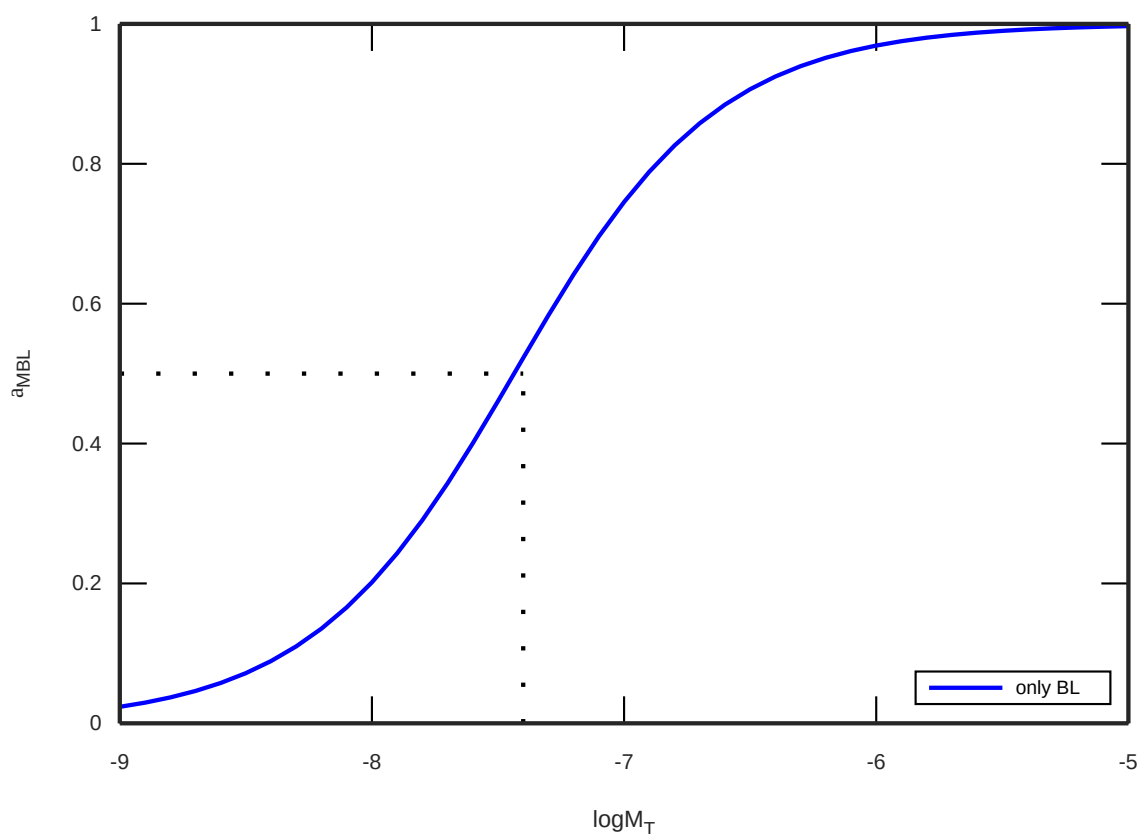
for i=1:length(MT)
    T=[MT(i); BLT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T,ASOLUTION,KSOLUTION,T);
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'b-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
legend('only BL','location','southeast');
%log10(M(index)) half way sat logM=-logKf

LC50nM=(10^halfsat)*1e9

```

LC50nM = 39.811



add proton competition

protons can also bind at the biotic ligand. If a proton binds then the metal can't bind so pH (lower) can be protective (toxicity modifying factor). Of course low pH might have its own toxic effects, but for now we will ignore that.

so consider that the biotic ligand has a pKa value of 4.5.

LA50 does not change, but the LC50 does change.

In [16]:

```
%plot -s 600,500 -f 'svg'
%BLT=10e-9; logKf=7.5; logMT=-9:0.01:-5; MT=10.^logMT;
pKa=4.5;

TABLEAU=[...
%H      M    BL    logK    species
1       0    0      0      {'H'}
0       1    0      0      {'M'}
0       0    1      0      {'BL'}
0       1    1    logKf    {'MBL'}
1       0    1    pKa     {'HBL'}
];

[N,M]=size(TABLEAU); ASOLUTION=cell2mat(TABLEAU(1:N,1:M-2)); KSOLUTION=cell2mat(
SOLUTIONNAMES=strvcat(TABLEAU(1:N,M));
```

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pH=4;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASOLUTION(:,1),Ksolution];
    T=[MT(i); BLT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T,Asolution,Ksolution,T);
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'b-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH4=(10^halfsat)*1e9

pH=5;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASOLUTION(:,1),Ksolution];
    T=[MT(i); BLT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T,Asolution,Ksolution,T);
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'r-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH6=(10^halfsat)*1e9

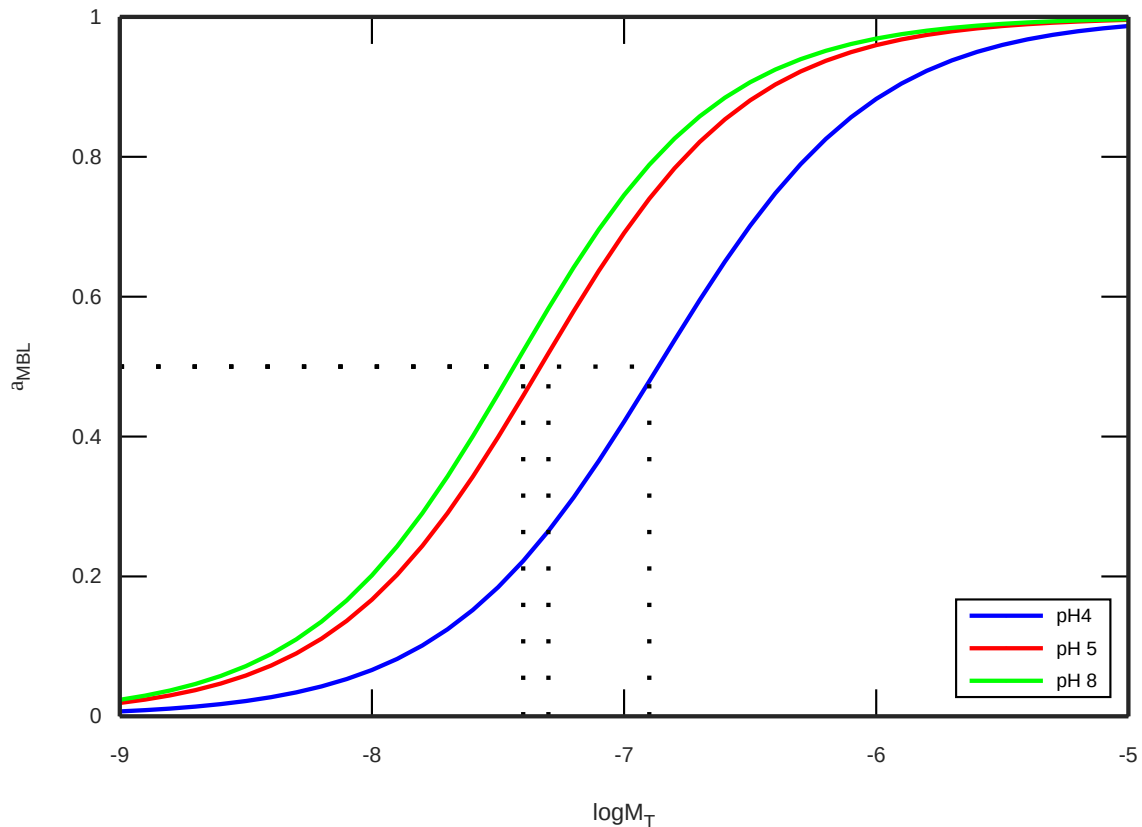
pH=8;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASOLUTION(:,1),Ksolution];
    T=[MT(i); BLT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T,Asolution,Ksolution,T);
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'g-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH8=(10^halfsat)*1e9

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legend('pH4',' ',' ','pH 5',' ',' ','pH 8','location','southeast');
```

```
LC50nMpH4 = 125.89
LC50nMpH6 = 50.119
LC50nMpH8 = 39.811
```



add strong base cation competition

other cations, Na^+ , K^+ , Mg^{2+} , Ca^{2+} , are typically non-toxic and can bind to the biotic ligand site, and prevent toxic metals from binding.

so consider that the biotic ligand has a $\log K_f$ value of 2.5 for Ca^{2+} binding. Let's simulate a high and a low calcium situation.

Again, LA_{50} does not change, but the LC_{50} does change.

In [17]:

```
%plot -s 600,500 -f 'svg'
%BLT=10e-9; logKf=7.5; logMT=-9:0.01:-5; MT=10.^logMT;%pKa=4.5;
logKfCa=2.5;
```

```
TABLEAU=[...
%H    M    BL    Ca    logK    species
1     0     0     0     0      {'H'}
0     1     0     0     0      {'M'}
0     0     1     0     0      {'BL'}
```

```

0      0      0      1      0      {'Ca'}
0      1      1      0      logKf   {'MBL'}
1      0      1      0      pKa     {'HBL'}
0      0      1      1      logKfCa  {'CaBL'}
];

[N,M]=size(TABLEAU); ASOLUTION=cell2mat(TABLEAU(1:N,1:M-2)); KSOLUTION=cell2mat(
SOLUTIONNAMES=strvcat(TABLEAU(1:N,M));
pH=7; CaT=1e-4;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASO
T=[MT(i); BLT; CaT];
[X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T./10,Asolution,Ksolution,T)
check(i)=100*(max(F./T)); %worst case error in%
% this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'g-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH7Calow=(10^halfsat)*1e9

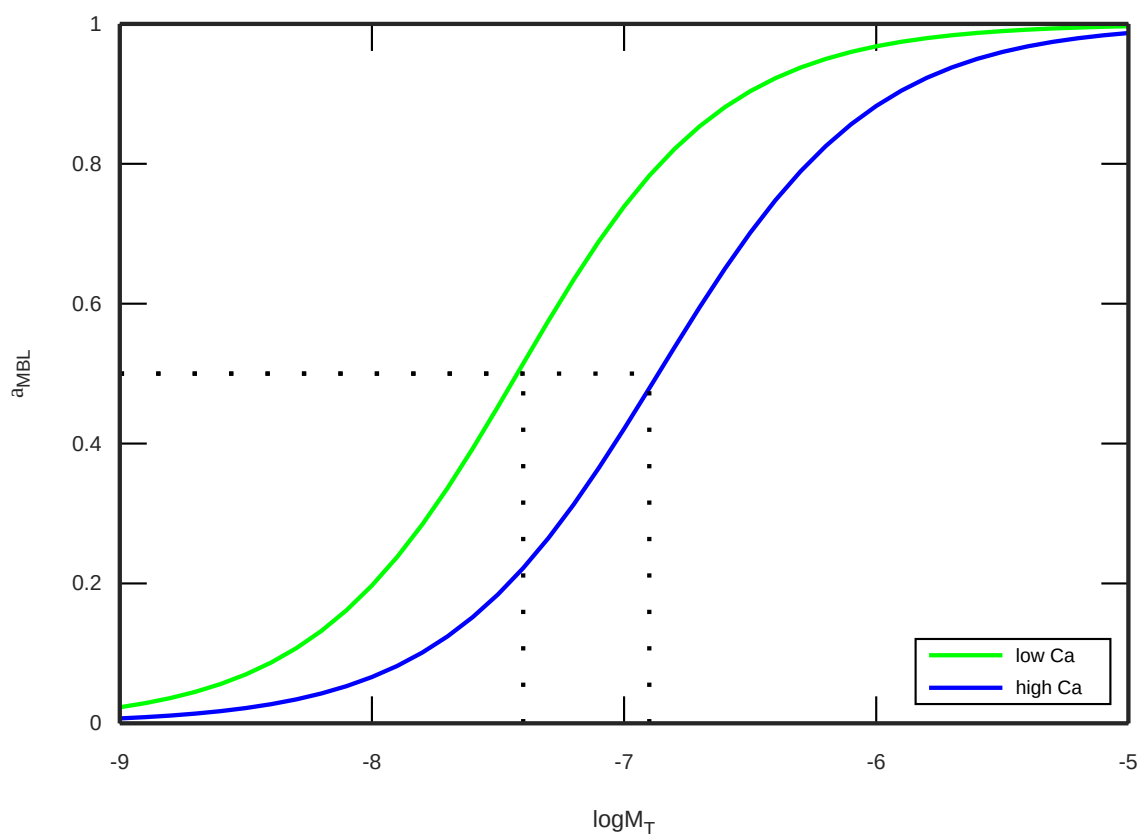
pH=7; CaT=1e-2;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASO
T=[MT(i); BLT; CaT];
[X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T./10,Asolution,Ksolution,T)
check(i)=100*(max(F./T)); %worst case error in%
% this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'b-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH7Cahigh=(10^halfsat)*1e9
legend('low Ca',' ',' ','high Ca','location','southeast');

```

LC50nMpH7Calow = 39.811

LC50nMpH7Cahigh = 125.89



add NOM complexation

other anions can bind the toxic metal and reduce accumulation at the biotic ligand. Hydroxo, chloro, carbonato complexation for example. And also natural organic matter complexation. Organic matter also will bind other cations, as well as protons.

So let's add in chloro complexation and NOM. Also, add in metal hydrolysis (MOH formation).

```
In [18]: %plot -s 600,500 -f 'svg'
%BLT=10e-9; logKf=7.5; logMT=-9:0.01:-5; MT=10.^logMT;%pKa=4.5;%logKfCa=2.5;

TABLEAU=[...
%H  M  BL  Ca  Cl  NOM  logK  species
1    0    0    0    0    0    0      {'H'}
0    1    0    0    0    0    0      {'M'}
0    0    1    0    0    0    0      {'BL'}
0    0    0    1    0    0    0      {'Ca'}
0    0    0    0    1    0    0      {'Cl'}
0    0    0    0    0    1    0      {'NOM'}
0    1    1    0    0    0    logKf    {'MBL'}
1    0    1    0    0    0    pKa      {'HBL'}
0    0    1    1    0    0    logKfCa  {'CaBL'}
0    1    0    0    1    0    3        {'MCl'}
-1   1    0    0    0    0    -8       {'MOH'}
1    0    0    0    0    1    10       {'HNOM'}
2    0    0    0    0    1    14       {'H2NOM'}
```

```

0      1      0      0      0      1      8      {'MNOM'}
0      0      0      1      0      1      3.5    {'CaNOM'}
];

[N,M]=size(TABLEAU); ASOLUTION=cell2mat(TABLEAU(1:N,1:M-2)); KSOLUTION=cell2mat(
SOLUTIONNAMES=strvcat(TABLEAU(1:N,M));
pH=7; CaT=1e-4; ClT=1e-4; NOMT=10e-6;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASO
    T=[MT(i); BLT; CaT; ClT; NOMT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T./10,Asolution,Ksolution,T)
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

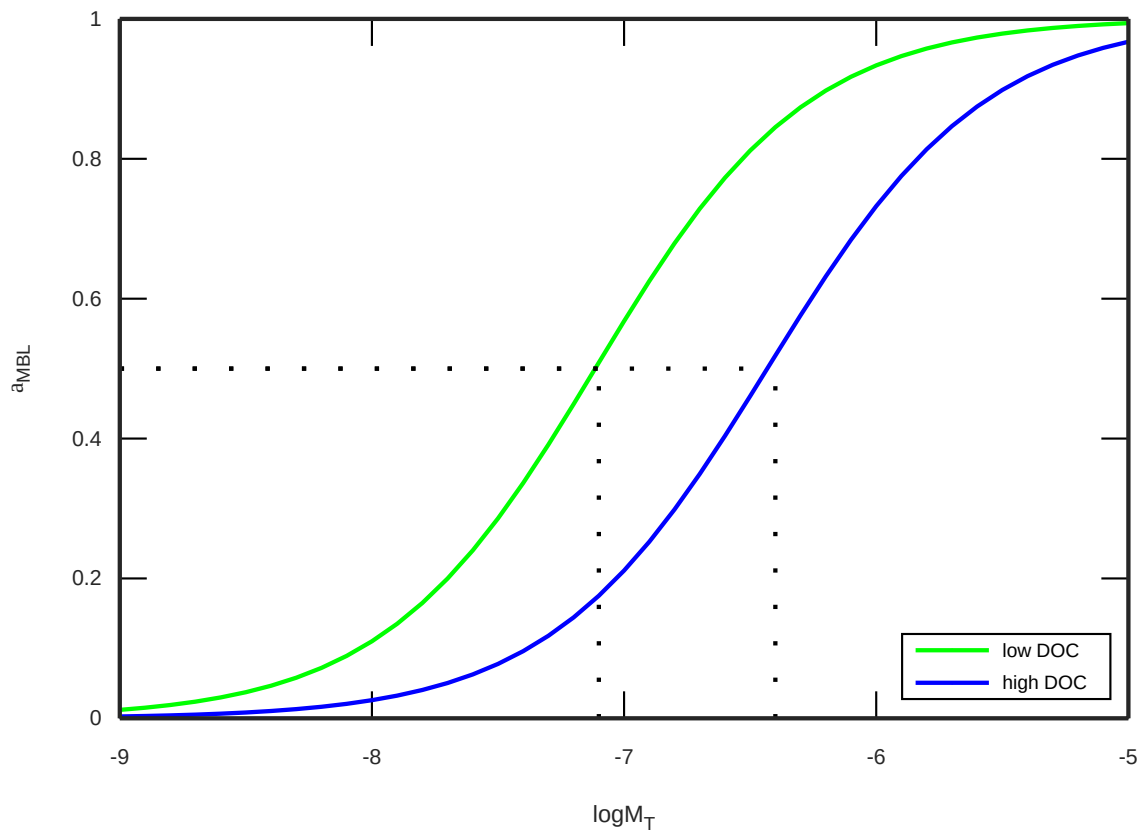
alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'g-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH7CalowClTlowNOM=(10^halfsat)*1e9

pH=7; CaT=1e-4; ClT=1e-4; NOMT=100e-6;
for i=1:length(MT)
    [N,M]=size(ASOLUTION); Ksolution=KSOLUTION-ASOLUTION(:,1)*pH; Asolution=[ASO
    T=[MT(i); BLT; CaT; ClT; NOMT];
    [X,F,J,SPECIESCONCS]=nl_massbalancerrnosolid_NR(T./10,Asolution,Ksolution,T)
    check(i)=100*(max(F./T)); %worst case error in%
    % this will generate the outputs
    for k=1:size(SPECIESCONCS,1)
        txt=[SOLUTIONNAMES(k,:), '(i)=SPECIESCONCS(k);'];eval(txt)
    end
end

alpha=MBL./BLT; [value,index]=min(abs(0.5-alpha)); halvesat=logMT(index);
plot(logMT,MBL./BLT,'b-','linewidth',2)
set(gca,'linewidth',2); xlabel('logM_T'); ylabel('\alpha_{MBL}')
hold on
plot([halfsat halvesat],[0 0.5],'k:','linewidth',2)
plot([min(logMT) halvesat],[0.5 0.5],'k:','linewidth',2)
LC50nMpH7CalowClThighNOM=(10^halfsat)*1e9
legend('low DOC',' ',' ','high DOC','location','southeast');

```

LC50nMpH7CalowClTlowNOM = 79.433
 LC50nMpH7CalowClThighNOM = 398.11



In []: