

```
In [1]: import sys
        !{sys.executable} -m pip install --user scikit-allel
```

```
Requirement already satisfied: scikit-allel in /projappl/project_2009301/s
oftware/lib/python3.10/site-packages (1.3.8)
Requirement already satisfied: numpy in /PUHTI_TYKKY_FRQGCcR/miniconda/env
s/env1/lib/python3.10/site-packages (from scikit-allel) (1.26.4)
Requirement already satisfied: dask[array] in /PUHTI_TYKKY_FRQGCcR/minicon
da/envs/env1/lib/python3.10/site-packages (from scikit-allel) (2024.4.1)
Requirement already satisfied: click>=8.1 in /PUHTI_TYKKY_FRQGCcR/minicond
a/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-allel)
(8.1.7)
Requirement already satisfied: cloudpickle>=1.5.0 in /PUHTI_TYKKY_FRQGCcR/
miniconda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit
-allel) (3.0.0)
Requirement already satisfied: fsspec>=2021.09.0 in /PUHTI_TYKKY_FRQGCcR/m
iniconda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-
allel) (2024.3.1)
Requirement already satisfied: packaging>=20.0 in /PUHTI_TYKKY_FRQGCcR/min
iconda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-al
lel) (23.2)
Requirement already satisfied: partd>=1.2.0 in /PUHTI_TYKKY_FRQGCcR/minico
nda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-alle
l) (1.4.1)
Requirement already satisfied: pyyaml>=5.3.1 in /PUHTI_TYKKY_FRQGCcR/minic
onda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-alle
l) (6.0.1)
Requirement already satisfied: toolz>=0.10.0 in /PUHTI_TYKKY_FRQGCcR/minic
onda/envs/env1/lib/python3.10/site-packages (from dask[array]->scikit-alle
l) (0.12.1)
Requirement already satisfied: importlib-metadata>=4.13.0 in /PUHTI_TYKKY_
FRQGCcR/miniconda/envs/env1/lib/python3.10/site-packages (from dask[array]
->scikit-allel) (7.1.0)
Requirement already satisfied: zipp>=0.5 in /PUHTI_TYKKY_FRQGCcR/minicond
a/envs/env1/lib/python3.10/site-packages (from importlib-metadata>=4.13.0-
>dask[array]->scikit-allel) (3.17.0)
Requirement already satisfied: locket in /PUHTI_TYKKY_FRQGCcR/miniconda/en
vs/env1/lib/python3.10/site-packages (from partd>=1.2.0->dask[array]->scik
it-allel) (1.0.0)
```

```
In [1]: import numpy as np
        import scipy
        import pandas
        import matplotlib as mpl
        import matplotlib.pyplot as plt
        %matplotlib inline
        import seaborn as sns
        sns.set_style('white')
        sns.set_style('ticks')
        sns.set_context('notebook')
        import h5py
        import allel; print('scikit-allel', allel.__version__)
```

scikit-allel 1.3.8

VCF to HDF5

```
In [2]: allel.vcf_to_hdf5('/users/mcevoysu/scratch/output/Anebrodensis/vcf_filter
```

Get data

```
In [3]: callset_var_fn = '/users/mcevoysu/scratch/output/Anebrodensis/scikit-alle  
callset_var = h5py.File(callset_var_fn, mode='r')
```

```
In [4]: calldata_var = callset_var['calldata']  
list(calldata_var)
```

```
Out[4]: ['AD', 'DP', 'GQ', 'GT', 'MIN_DP', 'PGT', 'PID', 'PL', 'PS', 'RGQ', 'S  
B']
```

```
In [5]: list(callset_var['variants'])
```

```
Out[5]: ['AC',  
        'AF',  
        'ALT',  
        'AN',  
        'BaseQRankSum',  
        'CHROM',  
        'DP',  
        'END',  
        'ExcessHet',  
        'FILTER_LowQual',  
        'FILTER_PASS',  
        'FS',  
        'ID',  
        'InbreedingCoeff',  
        'MLEAC',  
        'MLEAF',  
        'MQ',  
        'MQRankSum',  
        'POS',  
        'QD',  
        'QUAL',  
        'RAW_MQandDP',  
        'REF',  
        'ReadPosRankSum',  
        'SOR',  
        'altlen',  
        'is_snp',  
        'numalt']
```

Make datasets

```
In [6]: variants = allel.VariantChunkedTable(callset_var['variants'])  
variants
```

```
Out [6]: <VariantChunkedTable shape=(74891,) dtype=[('AC', '<i4', (3,)), ('AF', '<f4', (3,)),
('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'),
('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER_LowQual', '?'), ('FILTER_PASS', '?'), ('FS',
'<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)),
('MQ', '<f4'), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'),
('RAW_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'),
('altlen', '<i4', (3,)), ('is_snp', '?'), ('numalt', '<i4')] nbytes=12.8M cbytes=2.6M
cratio=4.9 values=h5py._hl.group.Group>
```

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	Ex
0	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	1.11	b'aalba5_s000000080'	96	-1	
1	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	0.0	b'aalba5_s000000080'	36	-1	
2	[4 -1 -1]	[0.286 nan nan]	[b'G' b'' b'']	14	0.0	b'aalba5_s000000080'	24	-1	
...									
74888	[6 -1 -1]	[0.1 nan nan]	[b'C' b'' b'']	60	0.0	b'aalba5_s01418300'	156	-1	
74889	[1 -1 -1]	[0.017 nan nan]	[b'A' b'' b'']	60	0.0	b'aalba5_s01418300'	117	-1	
74890	[1 -1 -1]	[0.017 nan nan]	[b'C' b'' b'']	60	0.765	b'aalba5_s01418300'	80	-1	

```
In [7]: variants_np = variants[:,]
rawsnps = variants_np.query('(is_snp == True)')
rawsnps
```

```
Out [7]: <VariantTable shape=(50778,) dtype=(numpy.record, [('AC', '<i4', (3,)), ('AF', '<f4', (3,)), ('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'), ('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER_LowQual', '?'), ('FILTER_PASS', '?'), ('FS', '<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)), ('MQ', '<f4'), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'), ('RAW_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'), ('altlen', '<i4', (3,)), ('is_snp', '?'), ('numalt', '<i4')])>
```

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	Ex
0	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	1.11	b'aalba5_s000000080'	96	-1	(
1	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	0.0	b'aalba5_s000000080'	36	-1	(
2	[4 -1 -1]	[0.286 nan nan]	[b'G' b'' b'']	14	0.0	b'aalba5_s000000080'	24	-1	(
...									
50775	[6 -1 -1]	[0.1 nan nan]	[b'C' b'' b'']	60	0.0	b'aalba5_s01418300'	156	-1	
50776	[1 -1 -1]	[0.017 nan nan]	[b'A' b'' b'']	60	0.0	b'aalba5_s01418300'	117	-1	
50777	[1 -1 -1]	[0.017 nan nan]	[b'C' b'' b'']	60	0.765	b'aalba5_s01418300'	80	-1	

```
In [8]: notsnp = variants_np.query('(is_snp != True)')
notsnp
```

```
Out [8]: <VariantTable shape=(24113,) dtype=(numpy.record, [('AC', '<i4', (3,)), ('AF', '<f4', (3,)), ('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'), ('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER_LowQual', '?'), ('FILTER_PASS', '?'), ('FS', '<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)), ('MQ', '<f4'), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'), ('RAW_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'), ('altlen', '<i4', (3,)), ('is_snp', '?'), ('numalt', '<i4')])>
```

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	Exc
0	[1 -1 -1]	[0.023 nan nan]	[b'*' b'' b'']	44	nan	b'aalba5_s000000180'	173	-1	
1	[2 -1 -1]	[0.04 nan nan]	[b'*' b'' b'']	50	nan	b'aalba5_s000000198'	331	-1	0
2	[2 -1 -1]	[0.04 nan nan]	[b'*' b'' b'']	50	nan	b'aalba5_s000000198'	330	-1	0
...									
24110	[5 -1 -1]	[0.083 nan nan]	[b'*' b'' b'']	60	nan	b'aalba5_s01418300'	61	-1	
24111	[5 -1 -1]	[0.083 nan nan]	[b'*' b'' b'']	60	nan	b'aalba5_s01418300'	61	-1	
24112	[5 -1 -1]	[0.083 nan nan]	[b'*' b'' b'']	60	nan	b'aalba5_s01418300'	61	-1	

Plot function

```
In [9]: def plot_hist(f, dsubset='', bins=30, ):
    if dsubset == 'var':
        x = variants[f][:]
        l = 'Variant'
    elif dsubset == 'snp':
        x = rawsnps[f][:]
        l = 'Raw SNP'
    elif dsubset == 'notsnp':
        x = notsnp[f][:]
        l = 'Raw Not SNP'
    elif dsubset == 'biallelic':
        x = biallelic_np[f][:]
        l = 'Biallelic SNP'
    elif dsubset == 'varsel':
        x = var_selection[f][:]
        l = 'Filtered Variants'
    elif dsubset == 'snpsel':
        x = snp_selection[f][:]
        l = 'Filtered SNP'
    else:
```

```

        x = bi_selection[f][:]
        l = 'Biallelic SNP'
    fig, ax = plt.subplots(figsize=(10, 5))
    sns.despine(ax=ax, offset=10)
    ax.hist(x, bins=bins)
    ax.set_xlabel(f)
    ax.set_ylabel('No. variants')
    ax.set_title('%s %s distribution' % (l, f))

```

Find Biallelic SNPS

```

In [10]: numalt = rawsnps['numalt']
         np.max(numalt)

```

Out[10]: 3

```

In [11]: count_numalt = np.bincount(numalt)
         count_numalt

```

Out[11]: array([0, 50252, 523, 3])

```

In [12]: n_multiallelic = np.sum(count_numalt[2:])
         n_multiallelic

```

Out[12]: 526

```

In [13]: filter_expression = '(numalt == 1)'
         biallelic_np = rawsnps.query(filter_expression)[: ]
         biallelic_np

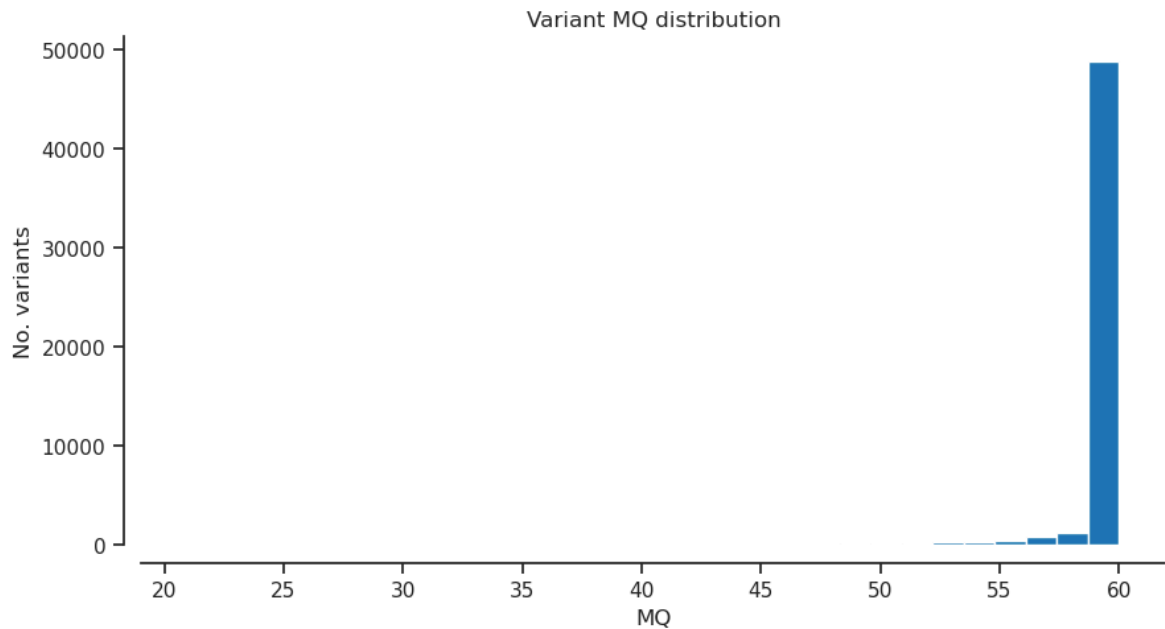
```

```
Out [13]: <VariantTable shape=(50252,) dtype=(numpy.record, [('AC', '<i4', (3,)), ('AF', '<f4', (3,)), ('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'), ('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER_LowQual', '?'), ('FILTER_PASS', '?'), ('FS', '<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)), ('MQ', '<f4'), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'), ('RAW_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'), ('altlen', '<i4', (3,)), ('is_snp', '?'), ('numalt', '<i4')])>
```

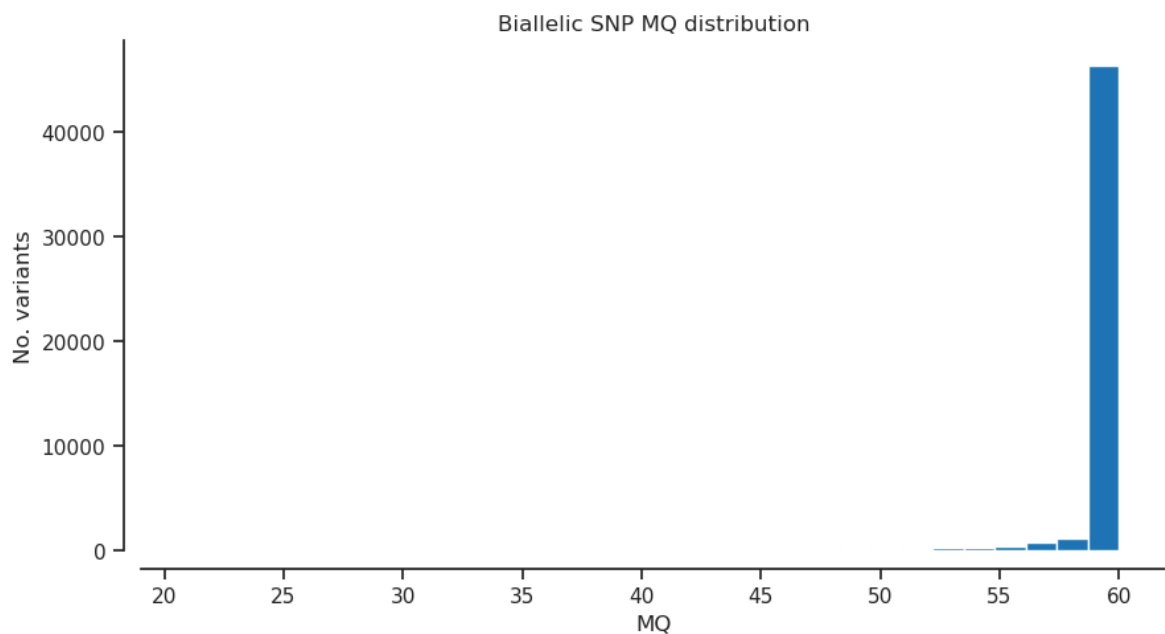
	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	Ex
0	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	1.11	b'aalba5_s000000080'	96	-1	
1	[4 -1 -1]	[0.286 nan nan]	[b'T' b'' b'']	14	0.0	b'aalba5_s000000080'	36	-1	
2	[4 -1 -1]	[0.286 nan nan]	[b'G' b'' b'']	14	0.0	b'aalba5_s000000080'	24	-1	
...									
50249	[6 -1 -1]	[0.1 nan nan]	[b'C' b'' b'']	60	0.0	b'aalba5_s01418300'	156	-1	
50250	[1 -1 -1]	[0.017 nan nan]	[b'A' b'' b'']	60	0.0	b'aalba5_s01418300'	117	-1	
50251	[1 -1 -1]	[0.017 nan nan]	[b'C' b'' b'']	60	0.765	b'aalba5_s01418300'	80	-1	

MQ - RMS mapping quality

```
In [14]: plot_hist('MQ', 'var') # RMS mapping quality
```

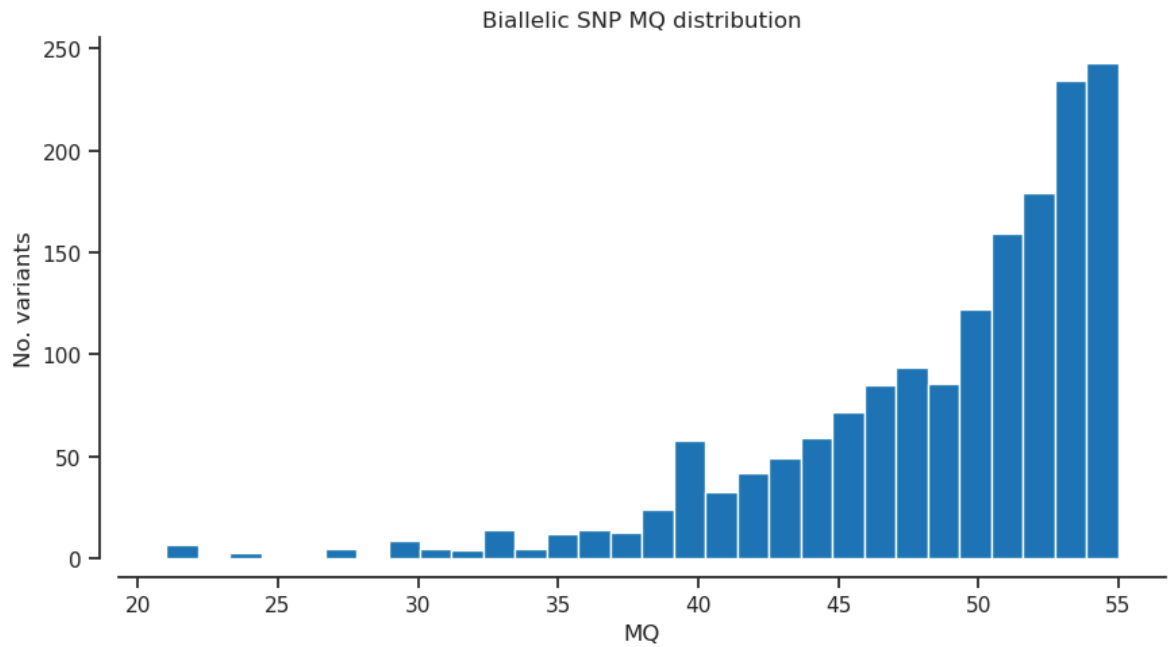


```
In [15]: plot_hist('MQ','biallelic') # RMS mapping quality
```



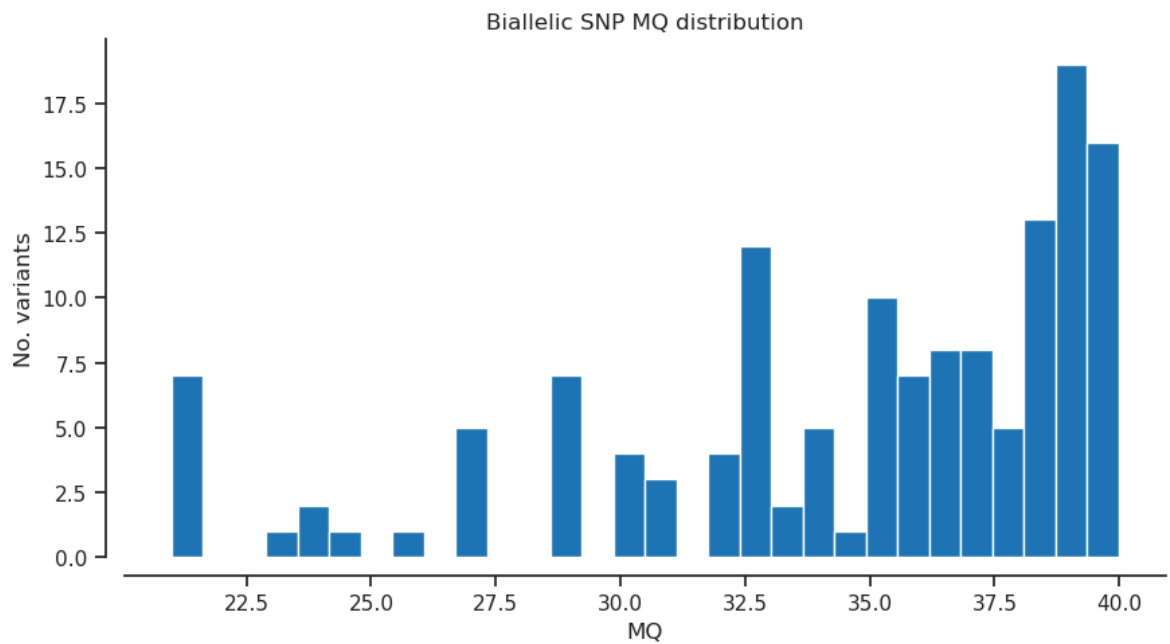
```
In [16]: filter_expression = '(MQ < 55)'
bi_selection = biallelic_np.query(filter_expression)[: ]
#np.count_nonzero(var_selection)
```

```
In [17]: plot_hist('MQ')
```

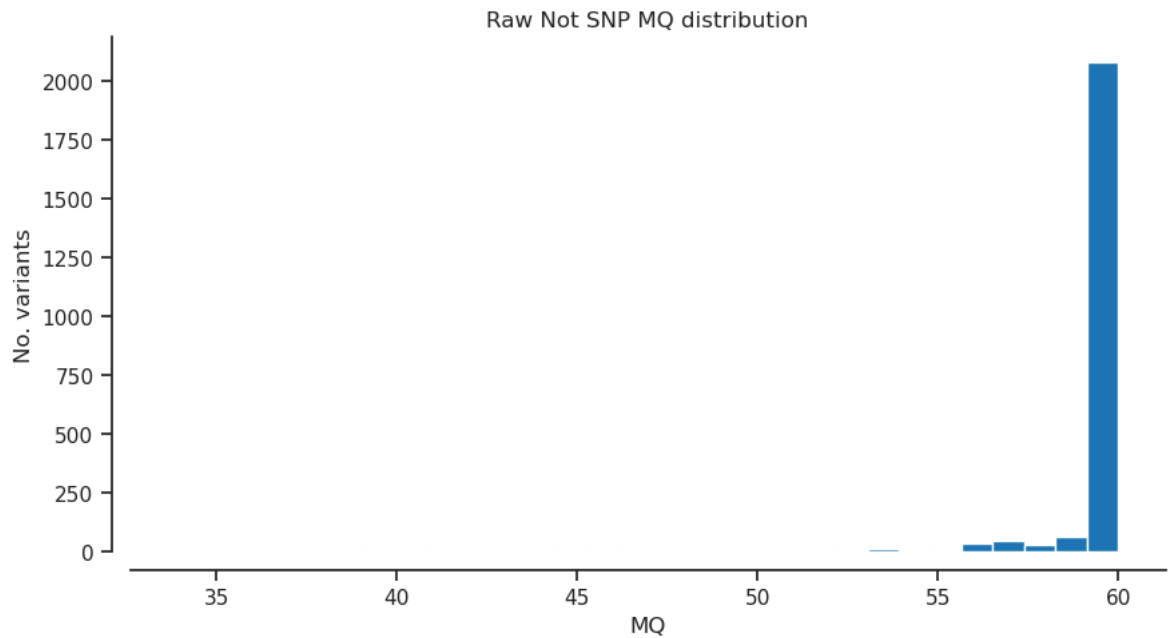



```
In [18]: filter_expression = '(MQ < 40)'  
bi_selection = biallelic_np.query(filter_expression)[:]
```

```
In [19]: plot_hist('MQ')
```

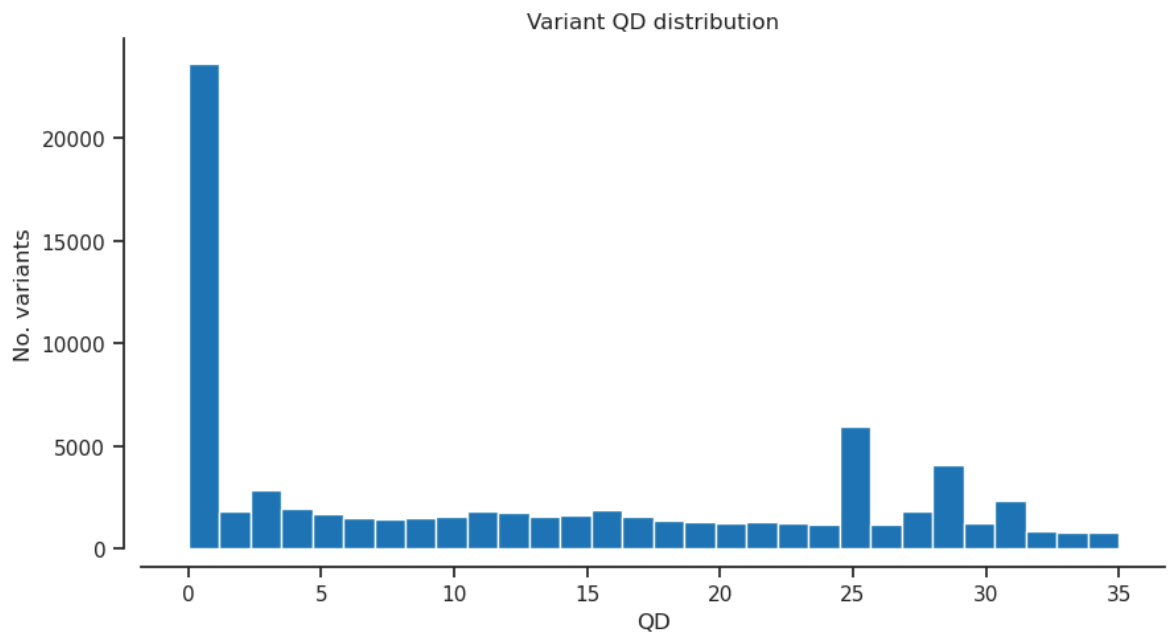


```
In [20]: plot_hist('MQ', 'notsnr')
```

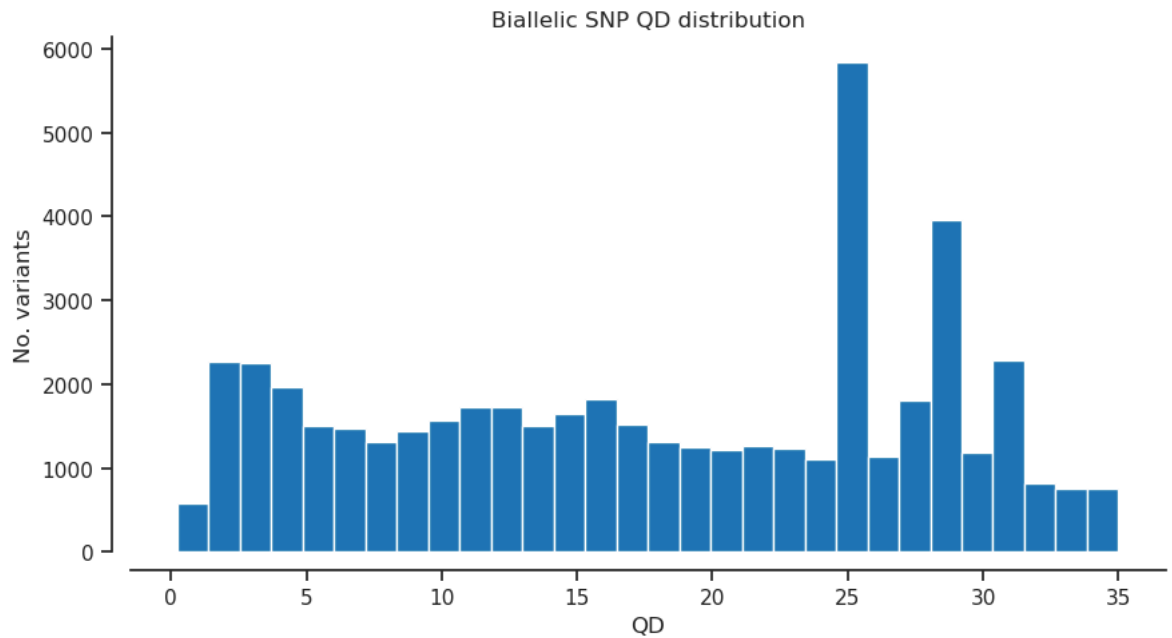


QD - Variant Confidence/Quality by Depth

```
In [21]: plot_hist('QD','var') # Variant Confidence/Quality by Depth
```

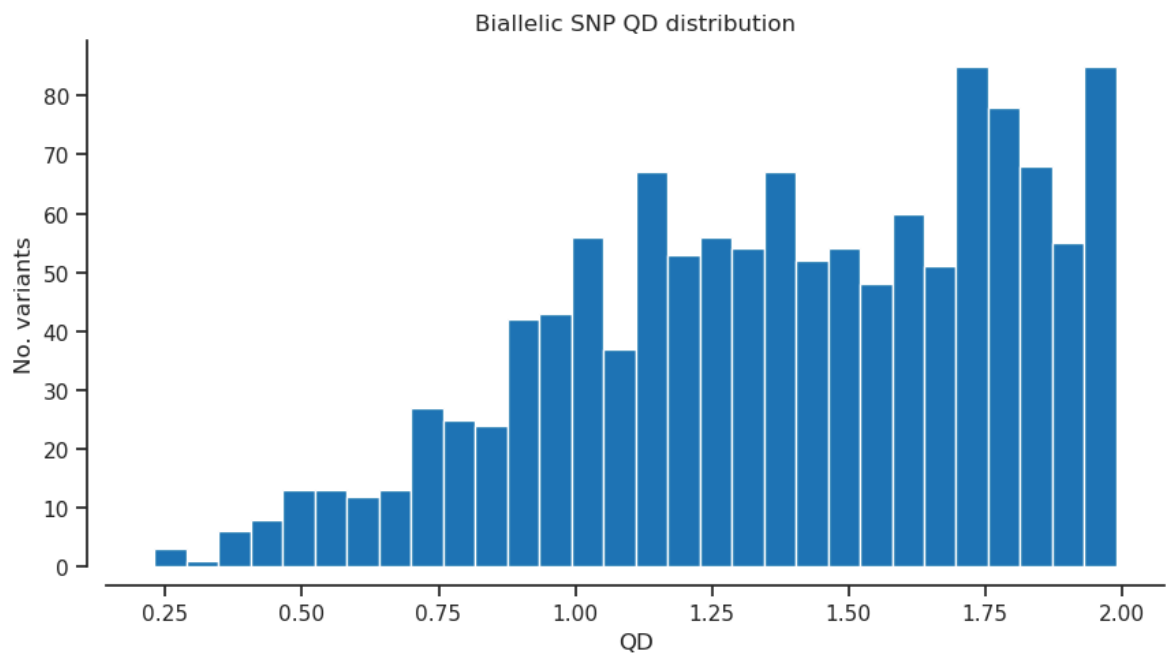


```
In [22]: plot_hist('QD','biallelic') # Variant Confidence/Quality by Depth
```

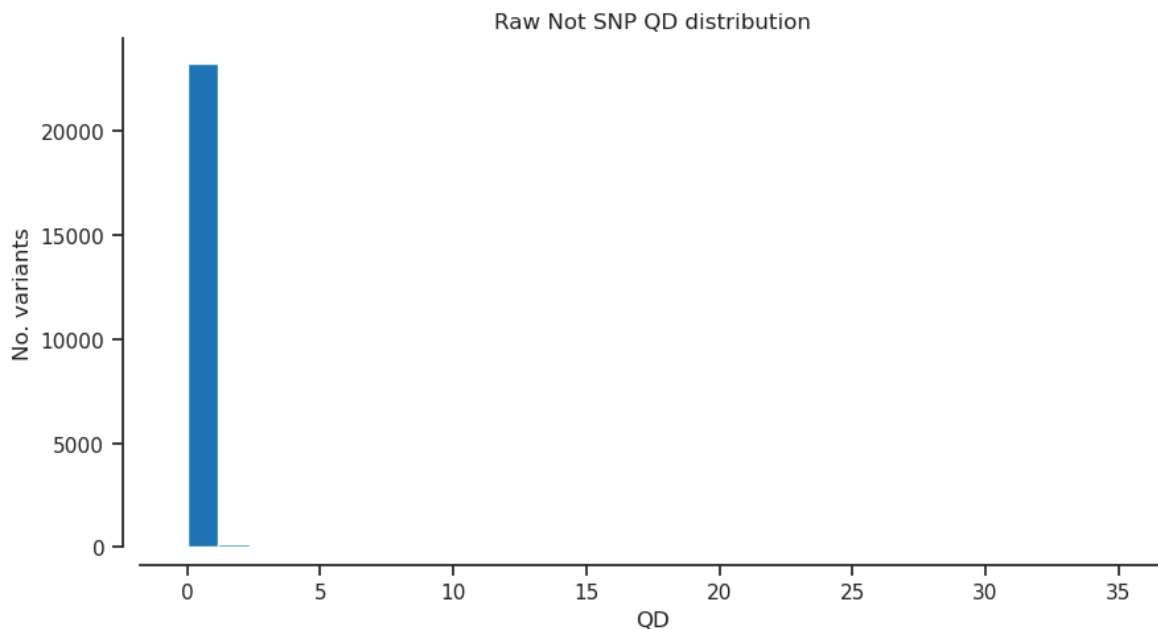


```
In [23]: filter_expression = '(QD < 2)'  
bi_selection = biallelic_np.query(filter_expression)[:]
```

```
In [24]: plot_hist('QD')
```

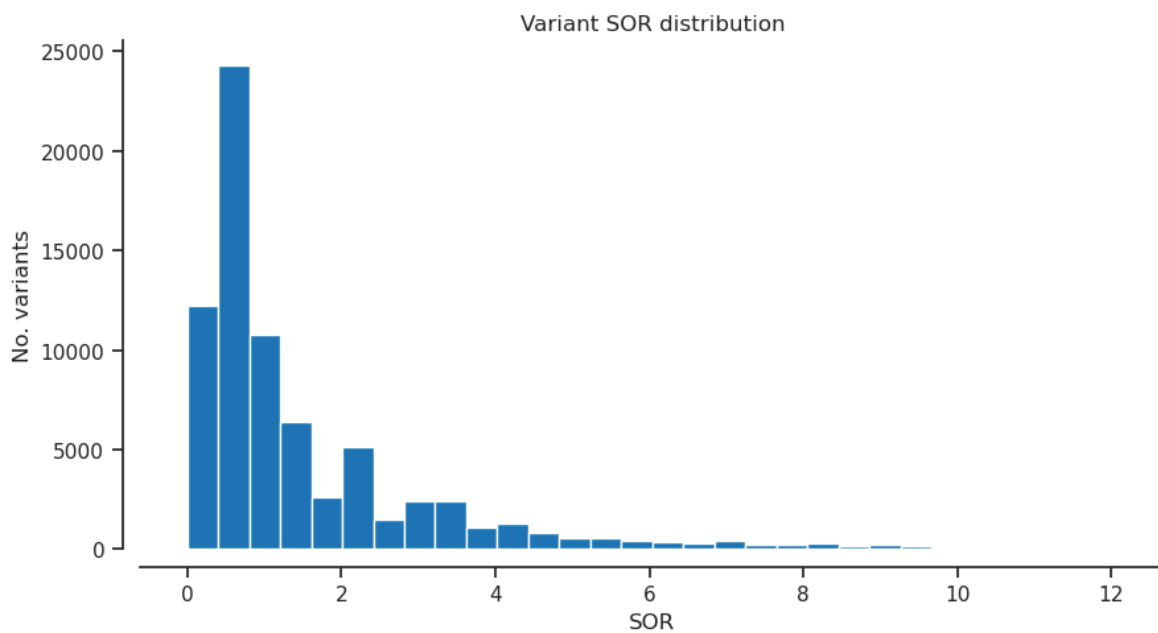


```
In [25]: plot_hist('QD', 'notsnp') # Variant Confidence/Quality by Depth
```

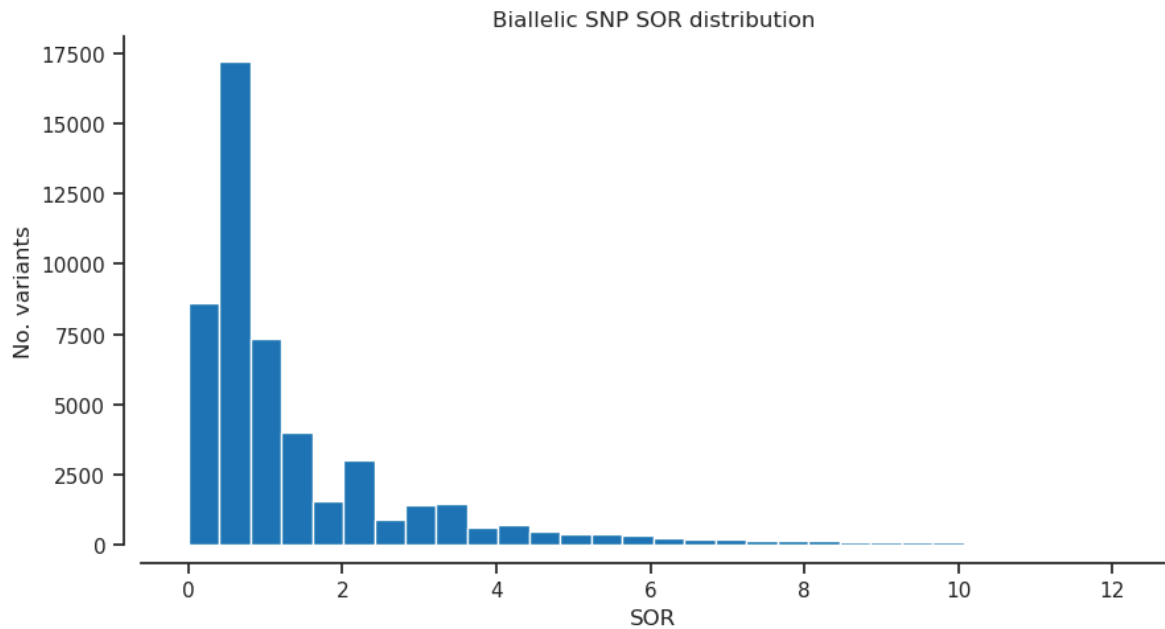


SOR - Symmetric Odds Ratio of 2x2 contingency table to detect strand bias

```
In [26]: plot_hist('SOR', 'var') # Symmetric Odds Ratio of 2x2 contingency table t
```

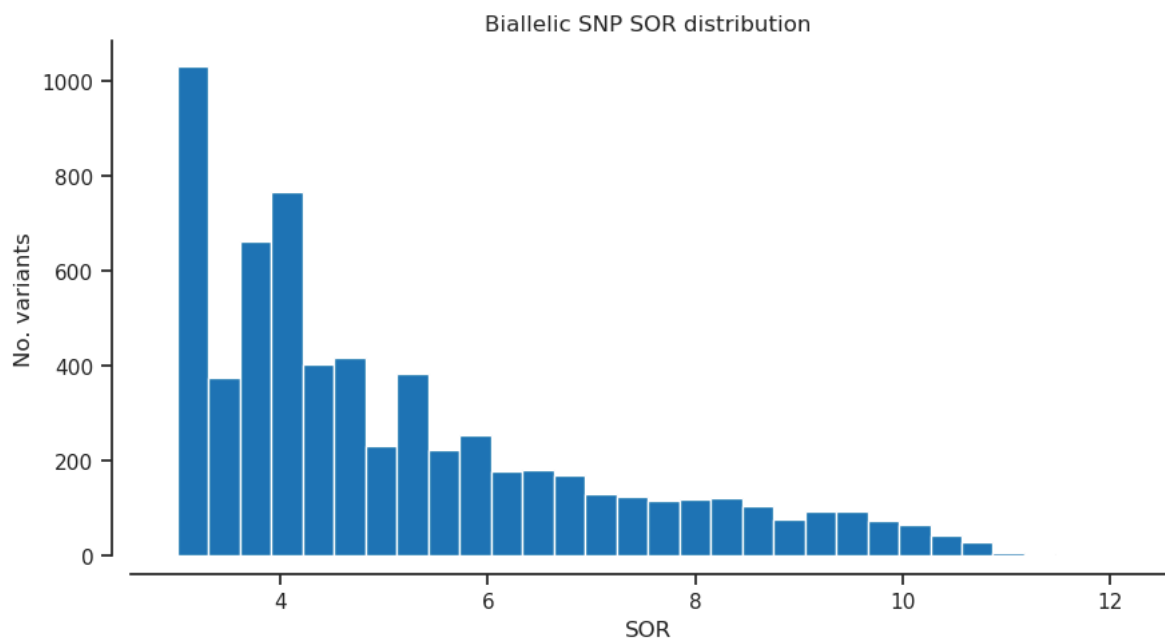


```
In [27]: plot_hist('SOR', 'biallelic') # Symmetric Odds Ratio of 2x2 contingency ta
```

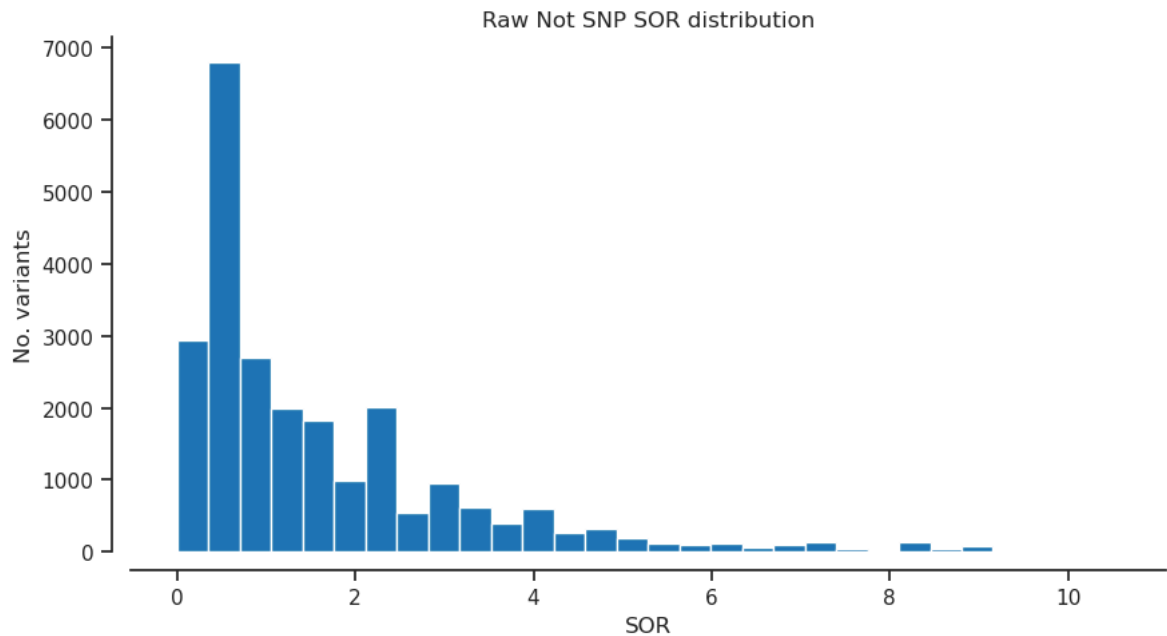


```
In [28]: filter_expression = '(SOR > 3)'
         bi_selection = biallelic_np.query(filter_expression)[:]
```

```
In [29]: plot_hist('SOR') # Symmetric Odds Ratio of 2x2 contingency table to detect
```

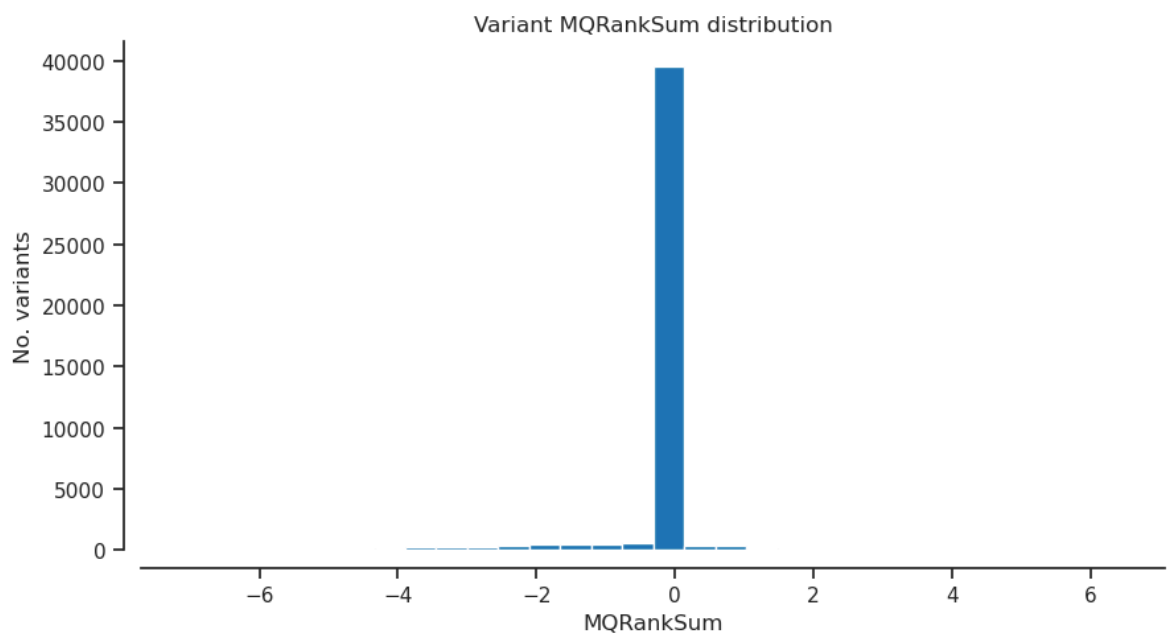


```
In [30]: plot_hist('SOR', 'notsnp') # Symmetric Odds Ratio of 2x2 contingency table
```

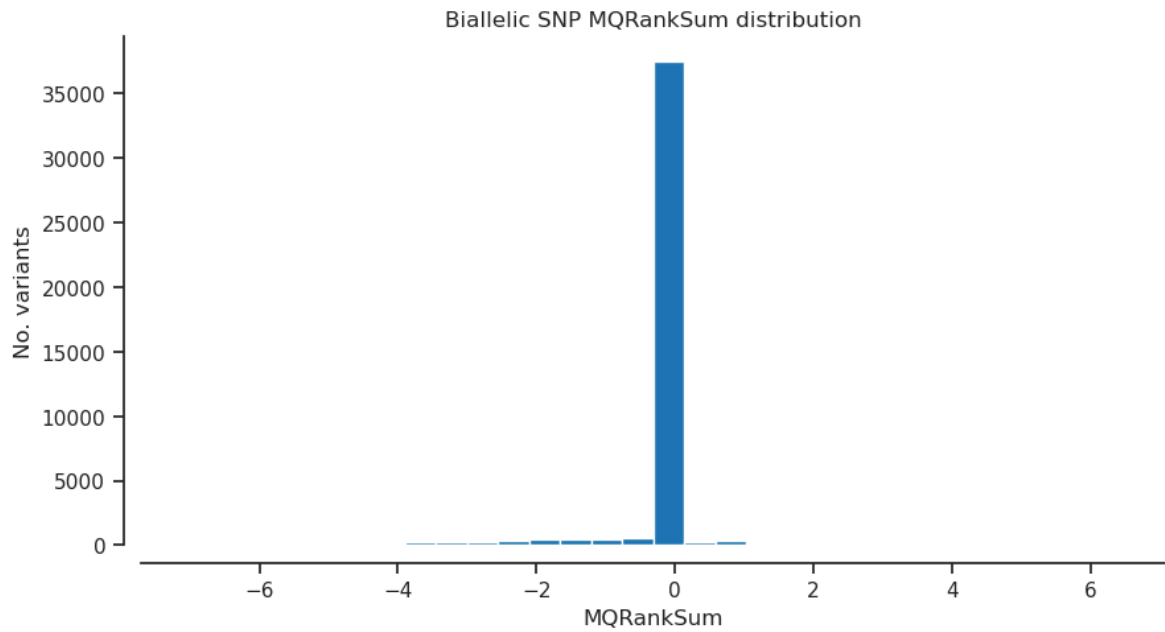


MQRankSum - Z-score From Wilcoxon rank sum test of Alt vs. Ref read mapping qualities

```
In [31]: plot_hist('MQRankSum', 'var') # Z-score From Wilcoxon rank sum test of Alt
```

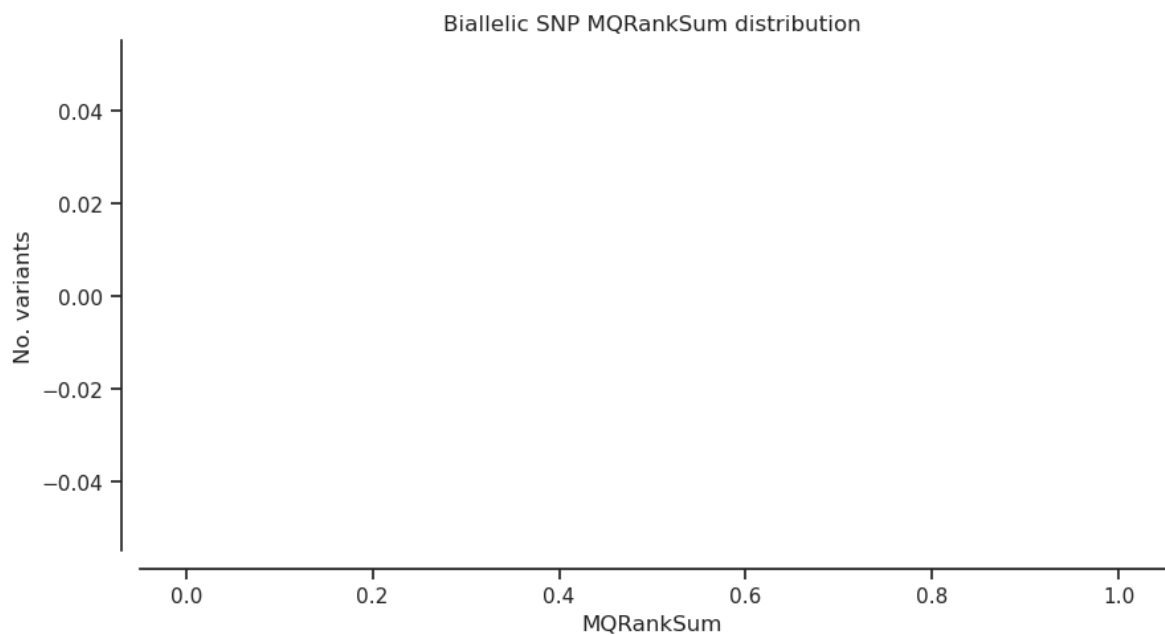


```
In [32]: plot_hist('MQRankSum', 'biallelic') # Z-score From Wilcoxon rank sum test
```

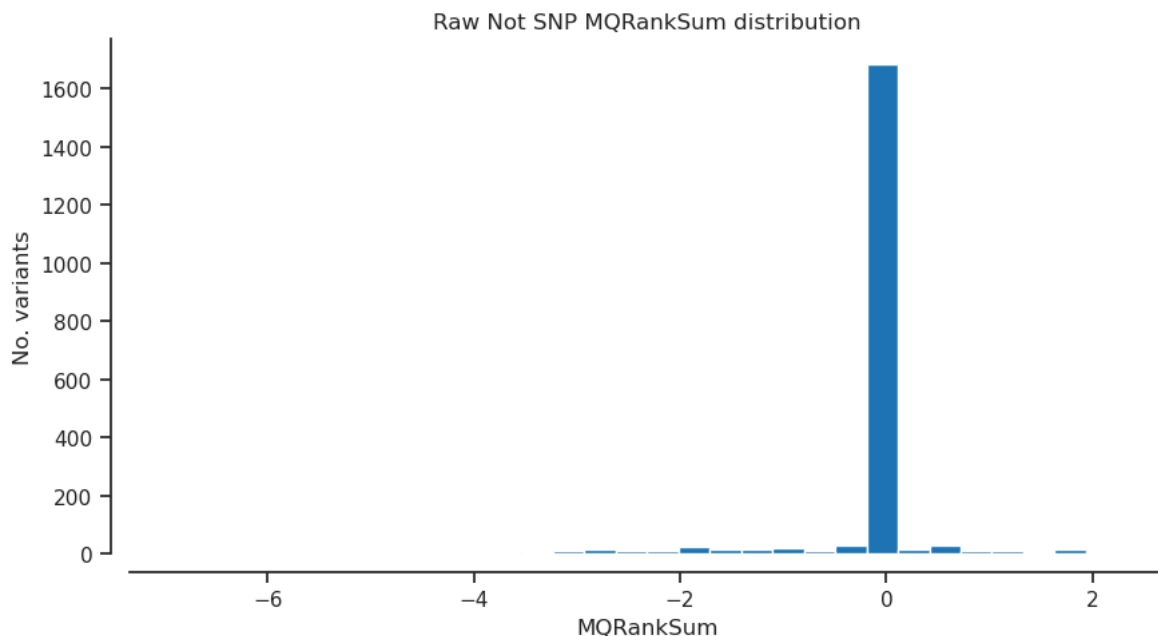


```
In [33]: filter_expression = '(MQRankSum < -12.5)'
         bi_selection = biallelic_np.query(filter_expression)[:]
```

```
In [34]: plot_hist('MQRankSum') # Z-score From Wilcoxon rank sum test of Alt vs. R
```

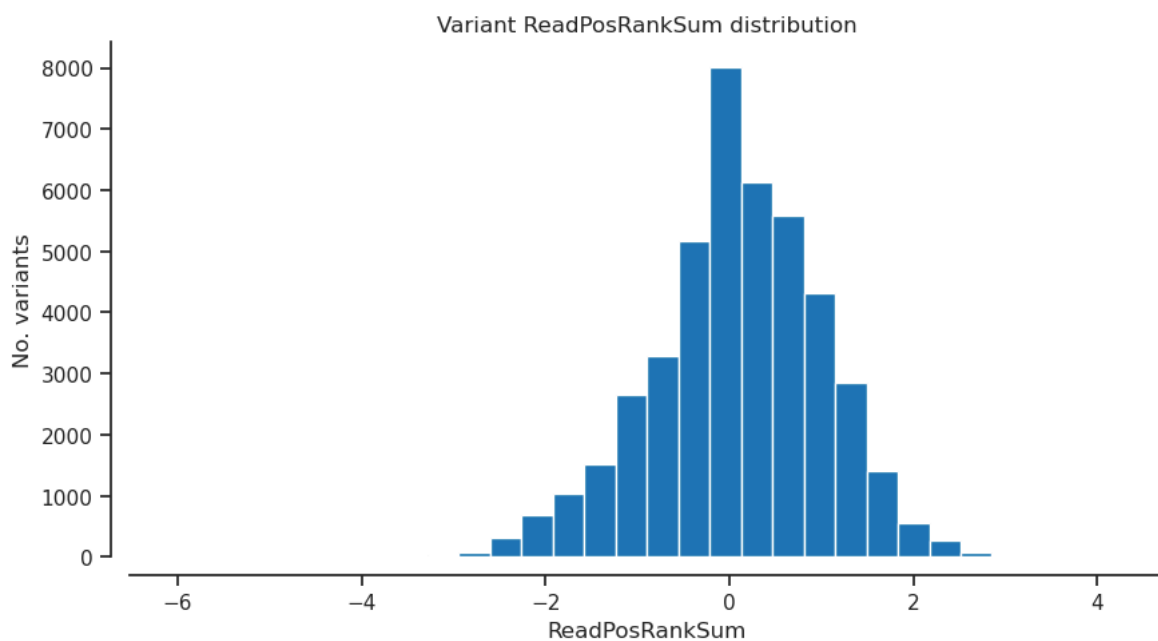


```
In [35]: plot_hist('MQRankSum', 'notsnp') # Z-score From Wilcoxon rank sum test of
```

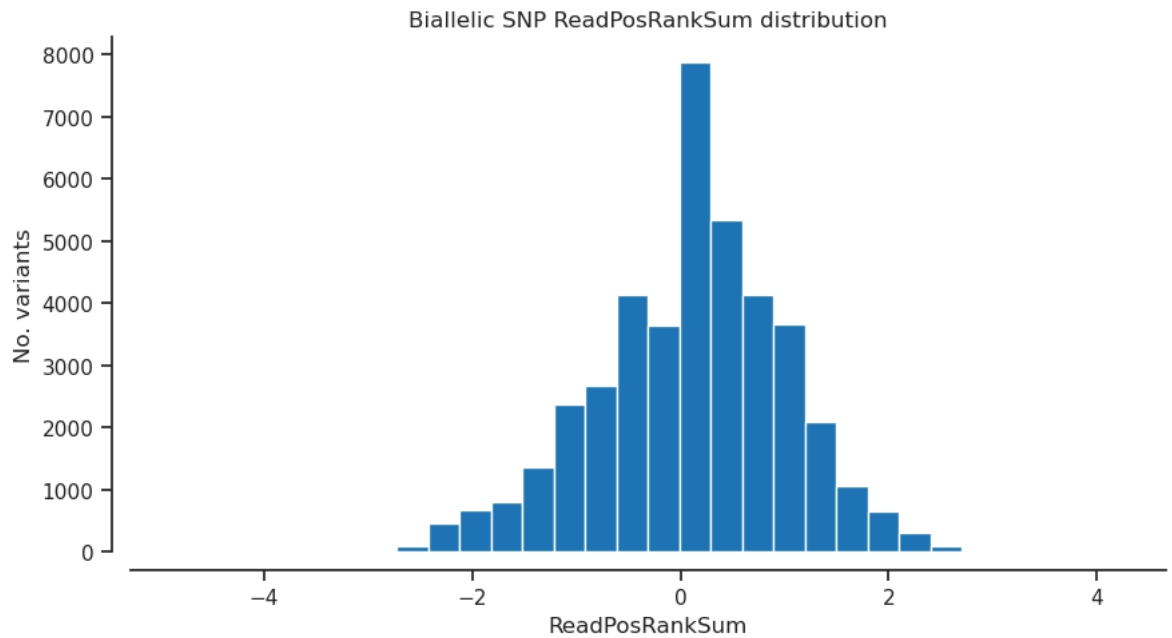


ReadPosRankSum - Z-score from Wilcoxon rank sum test of Alt vs. Ref read position bias

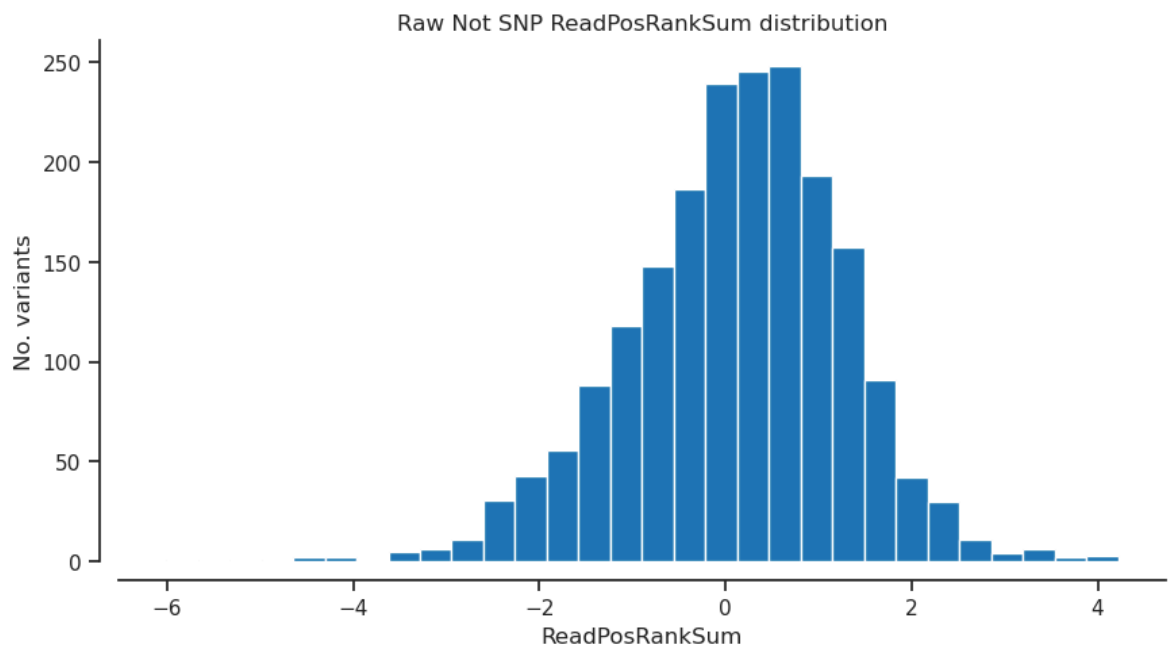
```
In [36]: plot_hist('ReadPosRankSum','var') # Z-score from Wilcoxon rank sum test o
```



```
In [37]: plot_hist('ReadPosRankSum','biallelic') # Z-score from Wilcoxon rank sum
```

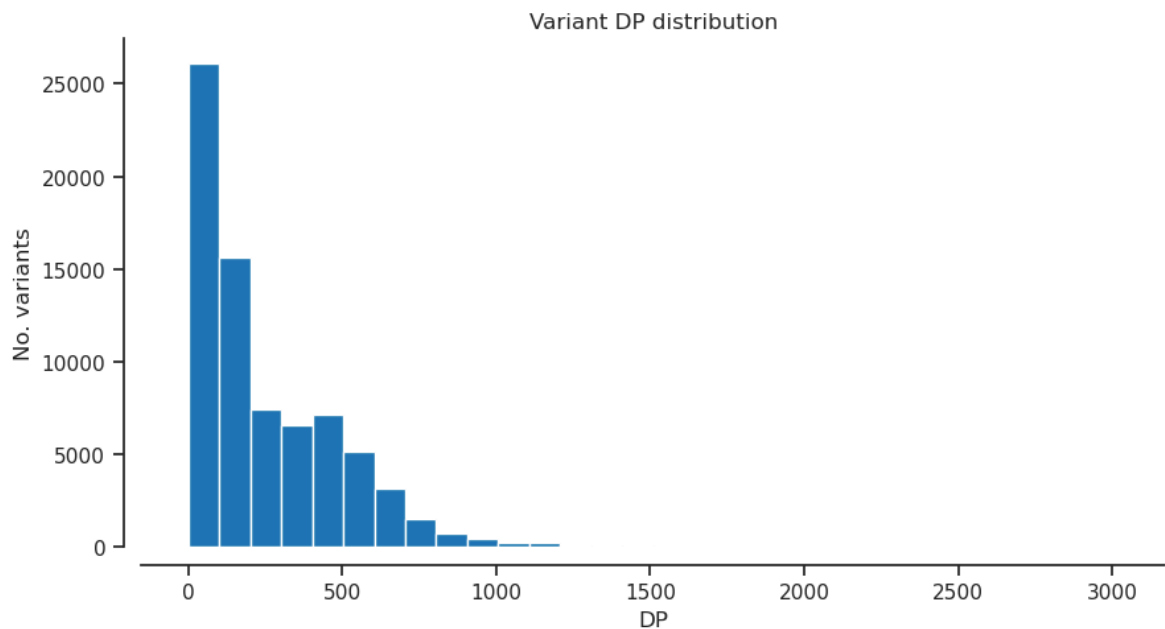



```
In [38]: plot_hist('ReadPosRankSum','notsnr') # Z-score from Wilcoxon rank sum tes
```

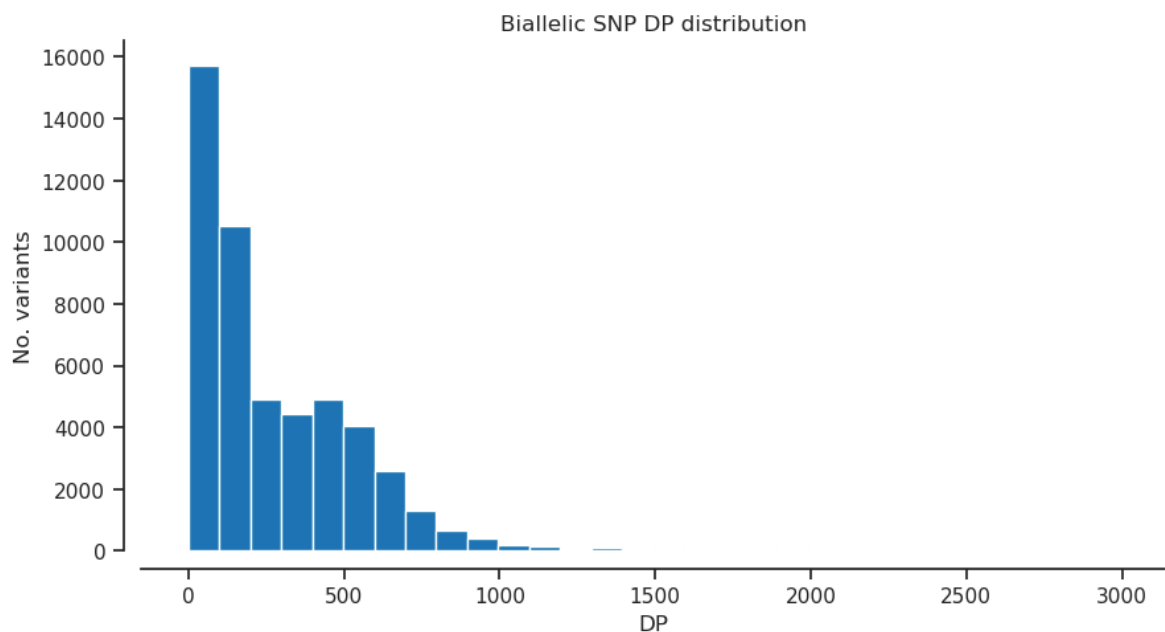


DP - Approximate read depth

```
In [39]: plot_hist('DP','var')
```

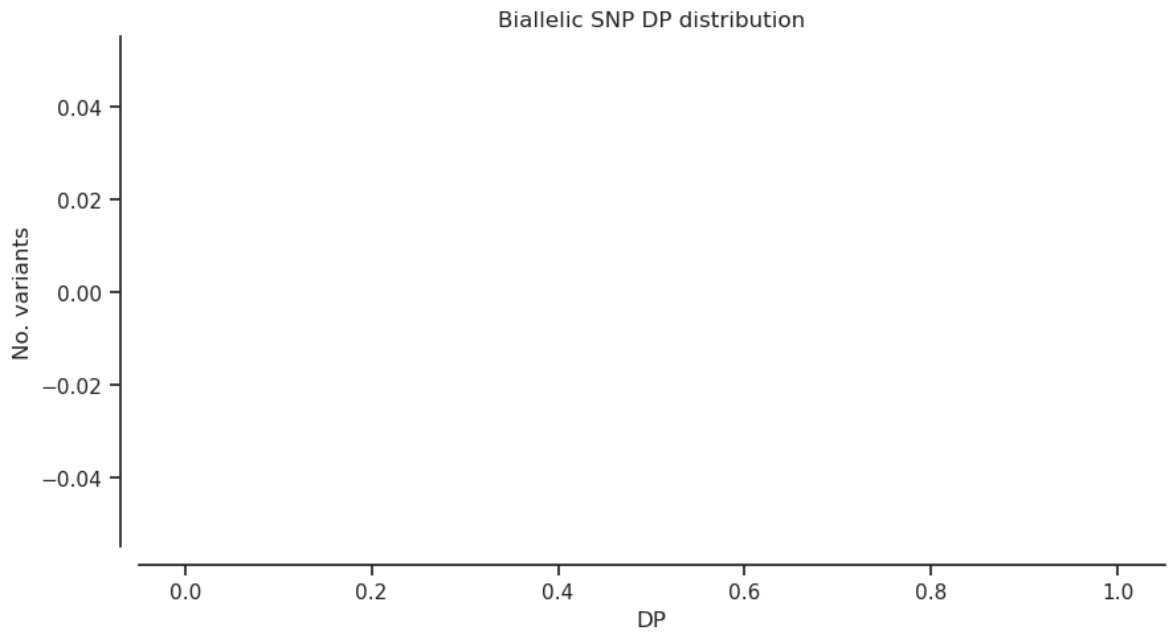


```
In [40]: plot_hist('DP', 'biallelic')
```

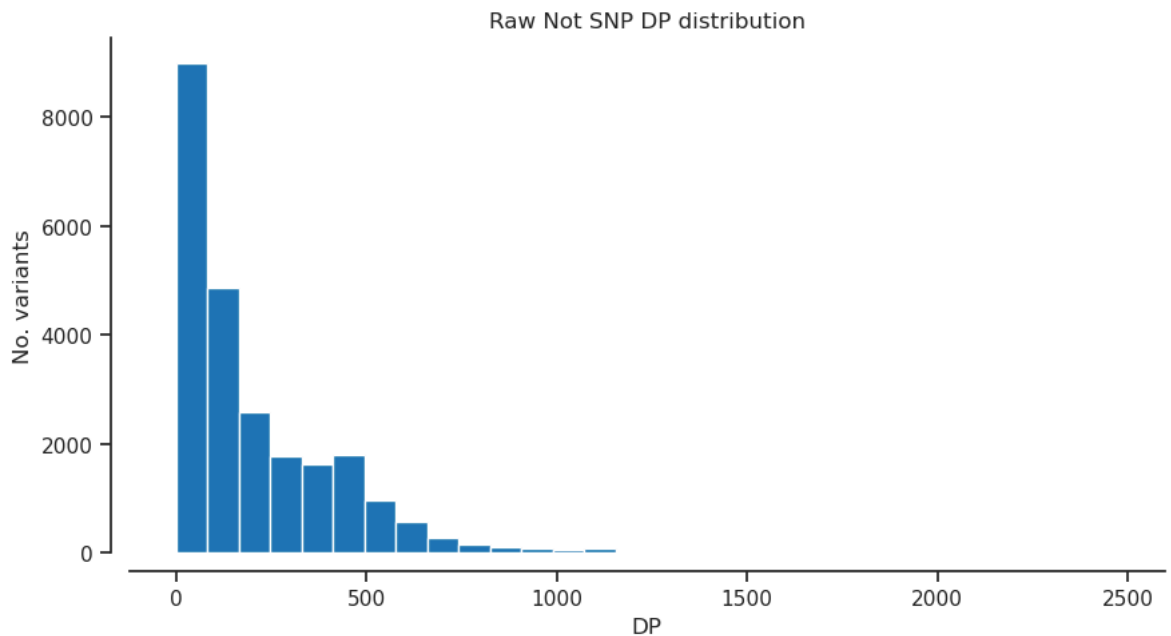


```
In [41]: filter_expression = '(DP > 20000) & (DP < 40000)'  
bi_selection = biallelic_np.query(filter_expression)[:]
```

```
In [42]: plot_hist('DP')
```

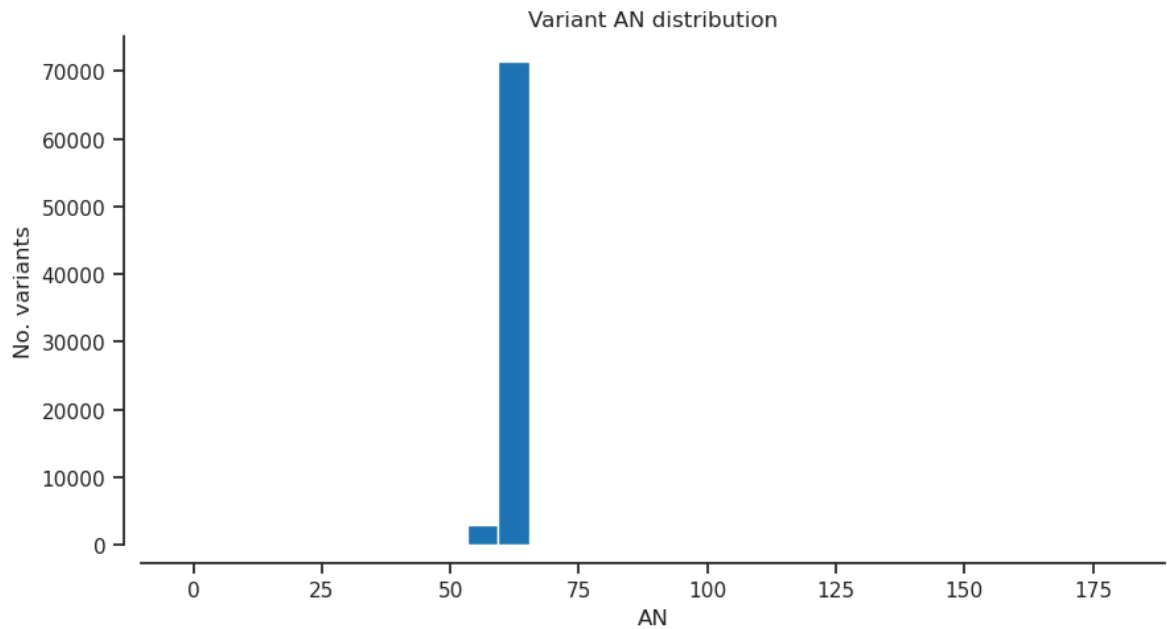


```
In [43]: plot_hist('DP','notsnp')
```

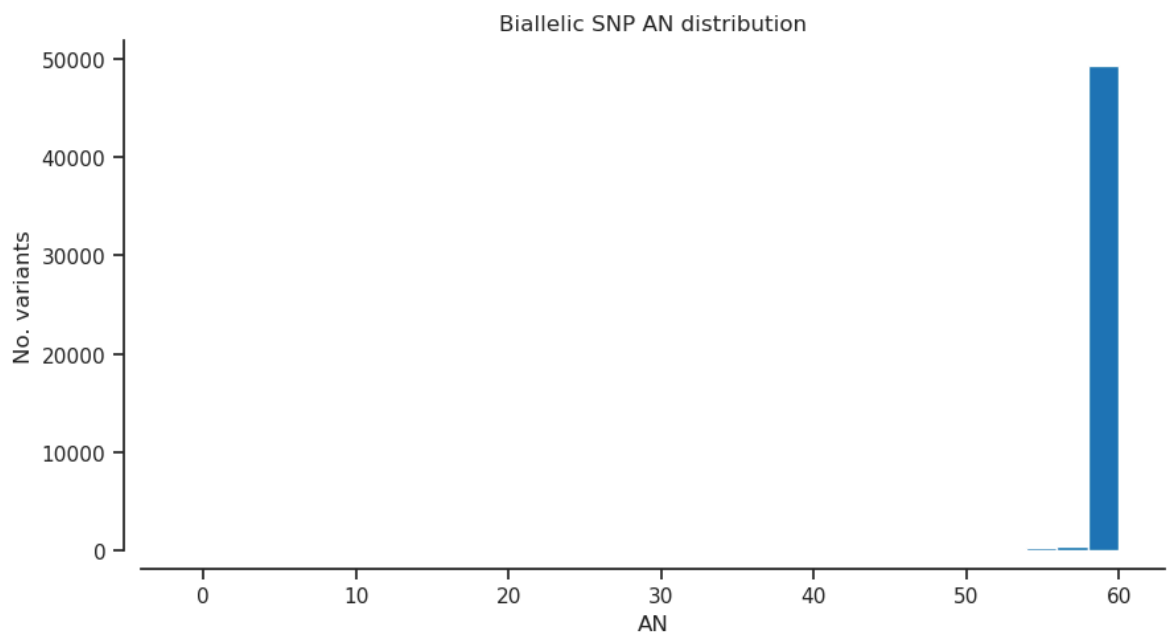


AN - Total number of alleles in called genotypes

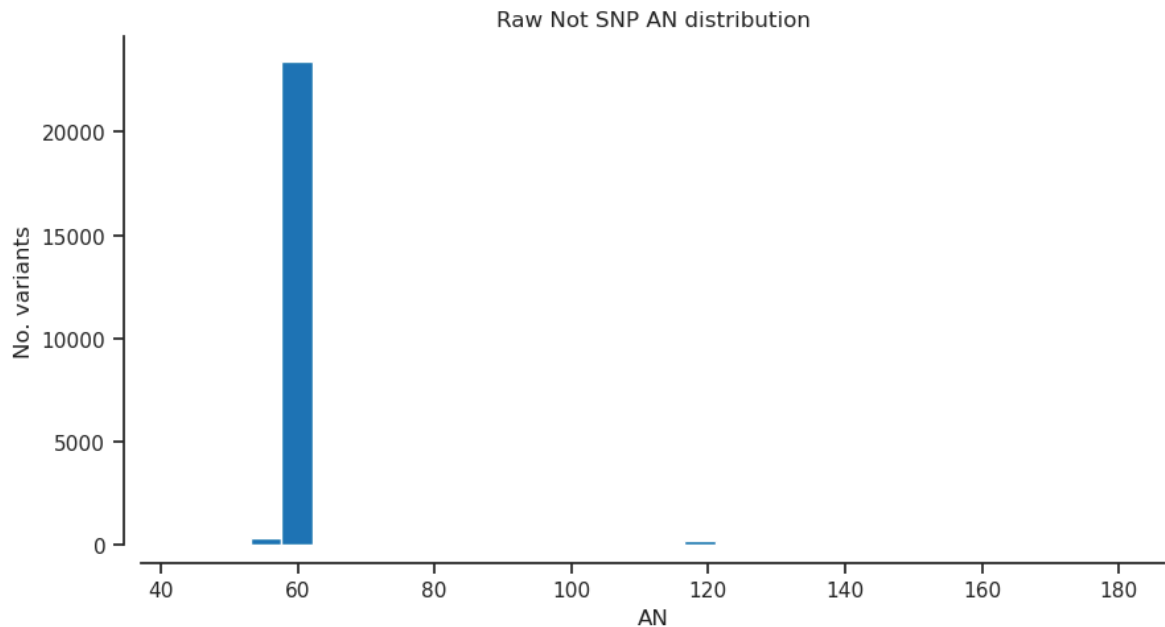
```
In [44]: plot_hist('AN','var') # Total number of alleles in called genotypes
```



```
In [45]: plot_hist('AN','biallelic') # Total number of alleles in called genotypes
```



```
In [46]: plot_hist('AN','notsnp') # Total number of alleles in called genotypes
```



Selected filter

```
In [47]: # QD: Variant Confidence/Quality by Depth
# AN: Total number of alleles in called genotypes
filter_expression = '(QD >= 2) & (MQ >= 40) & (MQRankSum >= -12.5) & (is_
variant_selection = variants_np.eval(filter_expression)[:])
np.count_nonzero(variant_selection)
```

Out[47]: 40767

Genotype

```
In [48]: calldata_var = callset_var['calldata']
list(calldata_var)
```

Out[48]: ['AD', 'DP', 'GQ', 'GT', 'MIN_DP', 'PGT', 'PID', 'PL', 'PS', 'RGQ', 'SB']

```
In [49]: genotypes_var = allele.GenotypeChunkedArray(calldata_var['GT'])
genotypes_var
```

Out [49]: <GenotypeChunkedArray shape=(74891, 30, 2) dtype=int8 chunks=(65536, 30, 2)
 nbytes=4.3M cbytes=456.4K cratio=9.6 compression=gzip compression_opts=1
 values=h5py._hl.dataset.Dataset>

	0	1	2	3	4	...	25	26	27	28	29
0	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
1	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
2	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
...	...										
74888	0/0	0/0	0/0	0/0	0/1	...	0/1	0/1	0/0	0/0	0/0
74889	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0
74890	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0

In [50]: *# using the selected filters set above*
 gt_filtered_snps = genotypes_var.subset(variant_selection)
 gt_filtered_snps

Out [50]: <GenotypeChunkedArray shape=(40767, 30, 2) dtype=int8 chunks=(5096, 30, 2)
 nbytes=2.3M cbytes=581.4K cratio=4.1 compression=blosc compression_opts=
 {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

	0	1	2	3	4	...	25	26	27	28	29
0	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
1	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
2	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
...	...										
40764	0/0	0/1	0/0	0/0	0/0	...	0/0	0/0	0/0	0/1	0/1
40765	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0
40766	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0

In [51]: *# grab the allele counts for the populations*
 ac = gt_filtered_snps.count_alleles()
 ac

Out [51]: <AlleleCountsChunkedArray shape=(40767, 4) dtype=int32 chunks=(20384, 4)
 nbytes=637.0K cbytes=66.8K cratio=9.5 compression=blosc compression_opts=
 {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

	0	1	2	3
0	10	4	0	0
1	10	4	0	0
2	10	4	0	0
...	...			
40764	54	6	0	0
40765	59	1	0	0
40766	59	1	0	0

In [52]: `ac[:]`

Out [52]: <AlleleCountsArray shape=(40767, 4) dtype=int32>

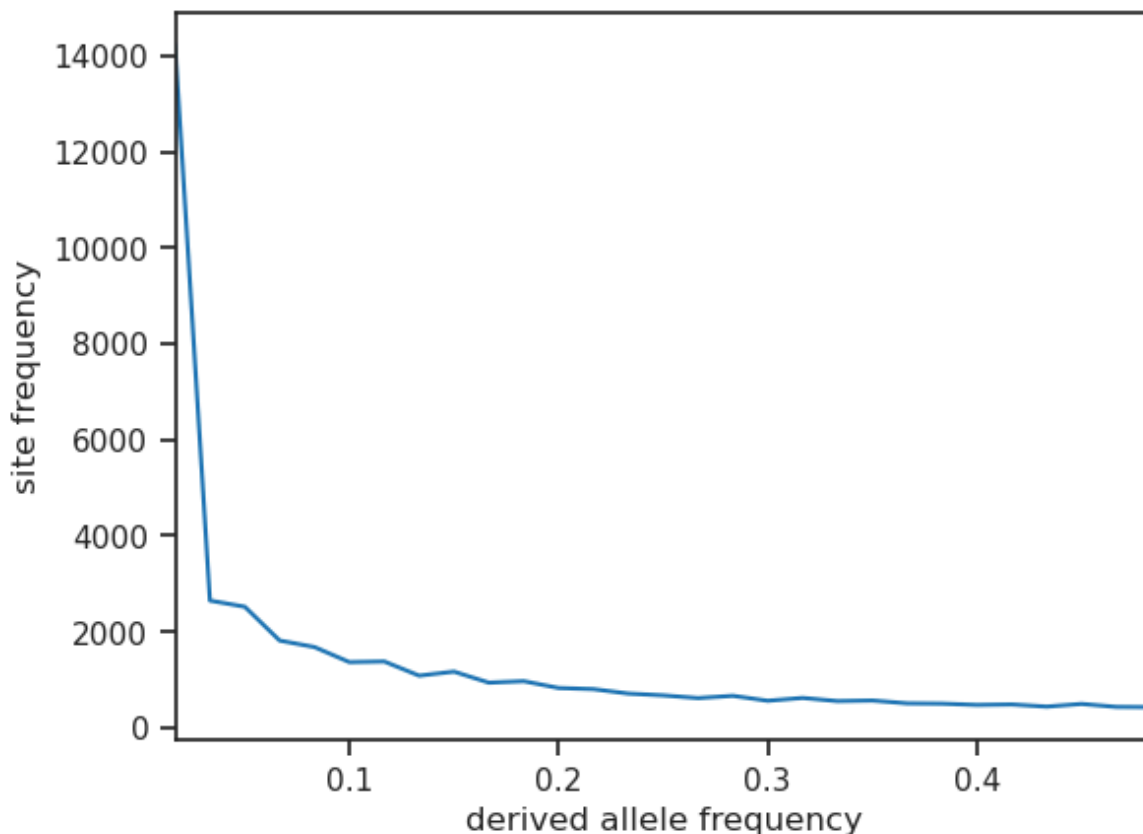
	0	1	2	3
0	10	4	0	0
1	10	4	0	0
2	10	4	0	0
...	...			
40764	54	6	0	0
40765	59	1	0	0
40766	59	1	0	0

In [53]: *# Which ones are biallelic?*
`is_biallelic_01 = ac.is_biallelic_01()[:]`
`ac1 = ac.compress(is_biallelic_01, axis=0)[: , :2]`
`ac1`
##this part of the code is only for graphing the SFS, is not useful for f

Out [53]: `array([[10, 4],
 [10, 4],
 [10, 4],
 ...,
 [54, 6],
 [59, 1],
 [59, 1]], dtype=int32)`

In [54]: *# plot the sfs of the derived allele*
`s = allel.sfs_folded(ac1)`
`allel.plot_sfs(s, yscale="linear", n=ac1.sum(axis=1).max())`

Out [54]: <Axes: xlabel='derived allele frequency', ylabel='site frequency'>



```
In [55]: biallelic = (ac.max_allele() == 1)
###This is the filter expression for biallelic sites
biallelic
```

```
Out[55]: <ChunkedArrayWrapper shape=(40767,) dtype=bool chunks=(40767,)
nbytes=39.8K cbytes=2.6K cratio=15.4
compression=blosc compression_opts={'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0}
values=zarr.core.Array>
```

```
In [56]: # select only the biallelic variants
gt_biallelic = gt_filtered_snps.compress(biallelic)
gt_biallelic
```

```
Out[56]: <GenotypeChunkedArray shape=(40326, 30, 2) dtype=int8 chunks=(5041, 30, 2)
nbytes=2.3M cbytes=570.0K cratio=4.1 compression=blosc compression_opts=
{'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>
```

	0	1	2	3	4	...	25	26	27	28	29
0	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
1	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
2	0/0	./.	./.	./.	./.	...	1/1	./.	./.	./.	./.
...						...					
40323	0/0	0/1	0/0	0/0	0/0	...	0/0	0/0	0/0	0/1	0/1
40324	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0
40325	0/0	0/0	0/0	0/0	0/0	...	0/0	0/0	0/0	0/0	0/0


```
In [57]: n_variants = len(gt_biallelic)
n_variants
```

```
Out[57]: 40326
```

```
In [58]: pc_missing = gt_biallelic.count_missing(axis=0)[:]* 100 / n_variants
pc_het = gt_biallelic.count_het(axis=0)[:]* 100 / n_variants
```

Samples

```
In [60]: samples_var = callset_var['samples']
samples_var = list(samples_var)
samples_var
```

```
Out[60]: [b'ITA00252-001',
b'ITA00252-002',
b'ITA00252-003',
b'ITA00252-004',
b'ITA00252-005',
b'ITA00252-006',
b'ITA00252-007',
b'ITA00252-008',
b'ITA00252-009',
b'ITA00252-010',
b'ITA00252-011',
b'ITA00252-012',
b'ITA00252-013',
b'ITA00252-014',
b'ITA00252-015',
b'ITA00252-016',
b'ITA00252-017',
b'ITA00252-018',
b'ITA00252-019',
b'ITA00252-020',
b'ITA00252-021',
b'ITA00252-022',
b'ITA00252-023',
b'ITA00252-024',
b'ITA00252-025',
b'ITA00252-026',
b'ITA00252-027',
b'ITA00252-028',
b'ITA00252-029',
b'ITA00252-030']
```

```
In [63]: samples_fn = '~/scratch/data/Anebrodensis/Abies_nebrodensis_sample_list_s
samples = pandas.read_csv(samples_fn, sep='\t')
samples
```

Out [63]:

	ID	Population
0	ITA00252-001	ITA00252
1	ITA00252-002	ITA00252
2	ITA00252-003	ITA00252
3	ITA00252-004	ITA00252
4	ITA00252-005	ITA00252
5	ITA00252-006	ITA00252
6	ITA00252-007	ITA00252
7	ITA00252-008	ITA00252
8	ITA00252-009	ITA00252
9	ITA00252-010	ITA00252
10	ITA00252-011	ITA00252
11	ITA00252-012	ITA00252
12	ITA00252-013	ITA00252
13	ITA00252-014	ITA00252
14	ITA00252-015	ITA00252
15	ITA00252-016	ITA00252
16	ITA00252-017	ITA00252
17	ITA00252-018	ITA00252
18	ITA00252-019	ITA00252
19	ITA00252-020	ITA00252
20	ITA00252-021	ITA00252
21	ITA00252-022	ITA00252
22	ITA00252-023	ITA00252
23	ITA00252-024	ITA00252
24	ITA00252-025	ITA00252
25	ITA00252-026	ITA00252
26	ITA00252-027	ITA00252
27	ITA00252-028	ITA00252
28	ITA00252-029	ITA00252
29	ITA00252-030	ITA00252

```
In [64]: samples.Population.value_counts()
```

Out [64]: Population
ITA00252 30
Name: count, dtype: int64

```
In [65]: populations = samples.Population.unique()
populations
###This identifiers come from the metadata file
```

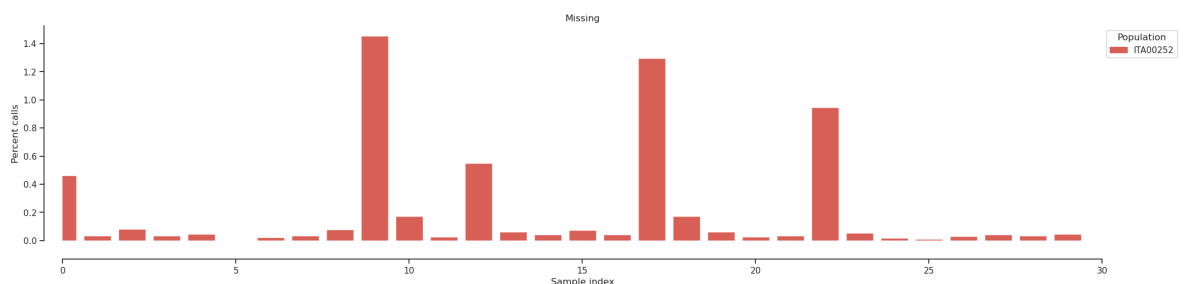
```
Out[65]: array(['ITA00252'], dtype=object)
```

Gt frequency function

```
In [66]: def plot_genotype_frequency(pc, title):
fig, ax = plt.subplots(figsize=(24, 5))
sns.despine(ax=ax, offset=24)
left = np.arange(len(pc))
palette = sns.color_palette("hls", 1)
pop2color = {'ITA00252': palette[0]}
colors = [pop2color[p] for p in samples.Population]
ax.bar(left, pc, color=colors)
ax.set_xlim(0, len(pc))
ax.set_xlabel('Sample index')
ax.set_ylabel('Percent calls')
ax.set_title(title)
handles = [mpl.patches.Patch(color=palette[0])]
ax.legend(handles=handles, labels=['ITA00252'], title='Population',
        bbox_to_anchor=(1, 1), loc='upper left')
```

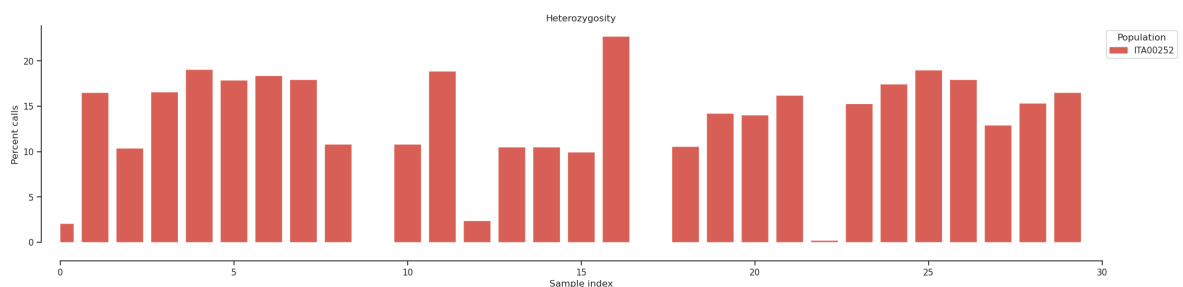
Plot missing

```
In [67]: plot_genotype_frequency(pc_missing, 'Missing')
```



Plot heterozygosity

```
In [68]: plot_genotype_frequency(pc_het, 'Heterozygosity')
```



PCA

```
In [69]: palette = sns.color_palette("hls",1)
pop_colours = {
    'ITA00252': palette[0]
}
```

```
In [70]: def plot_pca_coords(coords, model, pc1, pc2, ax, sample_population):
    sns.despine(ax=ax, offset=5)
    x = coords[:, pc1]
    y = coords[:, pc2]
    for pop in populations:
        flt = (sample_population == pop)
        ax.plot(x[flt], y[flt], marker='o', linestyle=' ', color=pop_color,
                label=pop, markersize=6, mec='k', mew=.5)
    ax.set_xlabel('PC%s (%.1f%%)' % (pc1+1, model.explained_variance_ratio[pc1]))
    ax.set_ylabel('PC%s (%.1f%%)' % (pc2+1, model.explained_variance_ratio[pc2]))

def fig_pca(coords, model, title, sample_population=None):
    if sample_population is None:
        sample_population = samples.Population
    # plot coords for PCs 1 vs 2, 3 vs 4
    fig = plt.figure(figsize=(10, 5))
    ax = fig.add_subplot(1, 2, 1)
    plot_pca_coords(coords, model, 0, 1, ax, sample_population)
    ax = fig.add_subplot(1, 2, 2)
    plot_pca_coords(coords, model, 2, 3, ax, sample_population)
    ax.legend(bbox_to_anchor=(1, 1), loc='upper left')
    fig.suptitle(title, y=1.02)
    fig.tight_layout()
```

```
In [71]: ac2 = gt_biallelic.count_alleles()
ac2
```

```
Out [71]: <AlleleCountsChunkedArray shape=(40326, 2) dtype=int32 chunks=(20163, 2)
nbytes=315.0K cbytes=58.5K cratio=5.4 compression=blosc compression_opts=
{'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>
```

	0	1
0	10	4
1	10	4
2	10	4
...
40323	54	6
40324	59	1
40325	59	1

```
In [72]: flt = (ac2[:, :2].min(axis=1) > 1)
gf = gt_biallelic.compress(flt, axis=0)
gn = gf.to_n_alt()
gn
```

```
Out[72]: <ChunkedArrayWrapper shape=(26099, 30) dtype=int8 chunks=(6525, 30)
         nbytes=764.6K cbytes=326.0K cratio=2.3
         compression=blosc compression_opts={'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0}
         values=zarr.core.Array>
```

```
In [73]: coords1, model1 = allel.pca(gn, n_components=10, scaler='patterson')
```

```
In [74]: fig_pca(coords1, model1, 'Figure 1. Conventional PCA.')
```

Figure 1. Conventional PCA.

