```
In [ ]:
        import sys
        !{sys.executable} -m pip install --user scikit-allel
In [1]:
        import numpy as np
        import scipy
        import pandas
        import matplotlib as mpl
        import matplotlib.pyplot as plt
        %matplotlib inline
        import seaborn as sns
        sns.set_style('white')
        sns.set_style('ticks')
        sns.set_context('notebook')
        import h5py
        import allel; print('scikit-allel', allel.__version__)
```

scikit-allel 1.3.8

#### VCF to HDF5

In [2]: allel.vcf\_to\_hdf5('/users/mcevoysu/scratch/output/Storminalis/vcf\_filteri

#### Get data

```
In [3]: callset_var_fn = '/users/mcevoysu/scratch/output/Storminalis/scikit-allel
    callset_var = h5py.File(callset_var_fn, mode='r')

In [4]: calldata_var = callset_var['calldata']
    list(calldata_var)

Out[4]: ['AD', 'DP', 'GQ', 'GT', 'MIN_DP', 'PGT', 'PID', 'PL', 'PS', 'RGQ', 'S
    B']

In [5]: list(callset_var['variants'])
```

```
Out[5]: ['AC',
          'AF',
          'ALT',
          'AN',
          'BaseQRankSum',
          'CHROM',
          'DP',
          'END',
          'ExcessHet',
           'FILTER_FILT',
           'FILTER_LowQual',
          'FILTER_PASS',
          'FS',
          'ID',
          'InbreedingCoeff',
          'MLEAC',
          'MLEAF',
          'MQ',
          'MQRankSum',
          'POS',
          'QD',
          'QUAL',
          'RAW_MQandDP',
          'REF',
          'ReadPosRankSum',
          'SOR',
          'altlen',
          'is_snp',
           'numalt']
```

## Make datasets

```
In [6]: variants = allel.VariantChunkedTable(callset_var['variants'])
variants
```

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	ExcessHet
0	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2059	-1	0.0971
1	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2065	-1	0.0963
2	[312 -1 -1]	[0.788 nan nan]	[b'G' b'' b'']	378	0.0	b'Chr01'	3473	-1	0.0
•••									
51347	[ 6 -1 -1]	[0.012 nan nan]	[b'C' b'' b'']	490	0.434	b'Chr17'	24021	-1	0.0008
51348	[261 -1 -1]	[0.542 nan nan]	[b'C' b'' b'']	462	0.0	b'Chr17'	6762	-1	0.7781
51349	[ 4 -1 -1]	[0.01 nan nan]	[b'T' b'' b'']	442	0.0	b'Chr17'	4997	-1	0.0891

```
In [7]: variants_np = variants[:]
    rawsnps = variants_np.query('(is_snp == True)')
    rawsnps
```

Out [7]: <VariantTable shape=(51348,) dtype=(numpy.record, [('AC', '<i4', (3,)), ('AF', '<f4', (3,)), ('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'), ('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER\_FILT', '?'), ('FILTER\_LowQual', '?'), ('FILTER\_PASS', '?'), ('FS', '<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'), ('RAW\_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'), ('altlen', '<i4', (3,)), ('is\_snp', '?'), ('numalt', '<i4')])>

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	ExcessHet
0	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2059	-1	0.0971
1	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2065	-1	0.0963
2	[312 -1 -1]	[0.788 nan nan]	[b'G' b'' b'']	378	0.0	b'Chr01'	3473	-1	0.0
•••									
51345	[ 6 -1 -1]	[0.012 nan nan]	[b'C' b'' b'']	490	0.434	b'Chr17'	24021	-1	0.0008
51346	[261 -1 -1]	[0.542 nan nan]	[b'C' b'' b'']	462	0.0	b'Chr17'	6762	-1	0.7781
51347	[ 4 -1 -1]	[0.01 nan nan]	[b'T' b'' b'']	442	0.0	b'Chr17'	4997	-1	0.0891

```
In [8]: notsnp = variants_np.query('(is_snp != True)')
notsnp
```

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	ExcessHet	FII
0	[ 2 -1 -1]	[0.006048 nan nan]	[b'*' b'' b'']	338	-2.846	b'Chr07'	4412	-1	0.0272	
1	[ 2 -1 -1]	[0.008065 nan nan]	[b'*' b'' b'']	356	nan	b'Chr07'	4626	-1	0.054	

#### Plot function

```
In [9]: def plot_hist(f, dsubset='', bins=30, ):
            if dsubset == 'var':
                x = variants[f][:]
                l = 'Variant'
            elif dsubset == 'snp':
                x = rawsnps[f][:]
                l = 'Raw SNP'
            elif dsubset == 'notsnp':
                x = notsnp[f][:]
                l = 'Raw Not SNP'
            elif dsubset == 'biallelic':
                x = biallelic_np[f][:]
                l = 'Biallelic SNP'
            elif dsubset == 'varsel':
                x = var selection[f][:]
                l = 'Filtered Variants'
            elif dsubset == 'snpsel':
                x = snp selection[f][:]
                l = 'Filtered SNP'
            else:
                x = bi selection[f][:]
                l = 'Biallelic SNP'
            fig, ax = plt.subplots(figsize=(10, 5))
            sns.despine(ax=ax, offset=10)
            ax.hist(x, bins=bins)
            ax.set_xlabel(f)
            ax.set ylabel('No. variants')
            ax.set_title('%s %s distribution' % (l, f))
```

#### Find Biallelic SNPS

```
In [10]: numalt = rawsnps['numalt']
np.max(numalt)

Out[10]: 1

In [11]: count_numalt = np.bincount(numalt)
count_numalt

Out[11]: array([ 0, 51348])

In [12]: n_multiallelic = np.sum(count_numalt[2:])
n_multiallelic

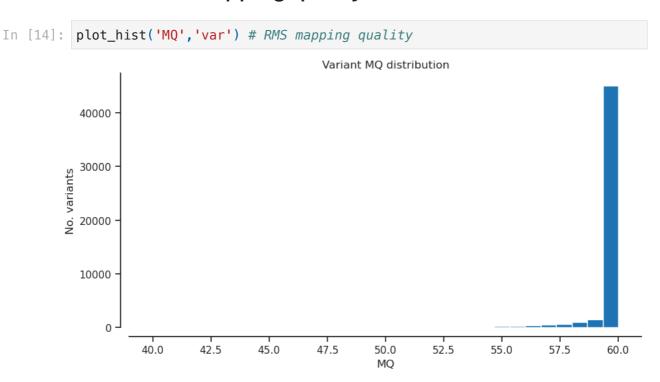
Out[12]: 0

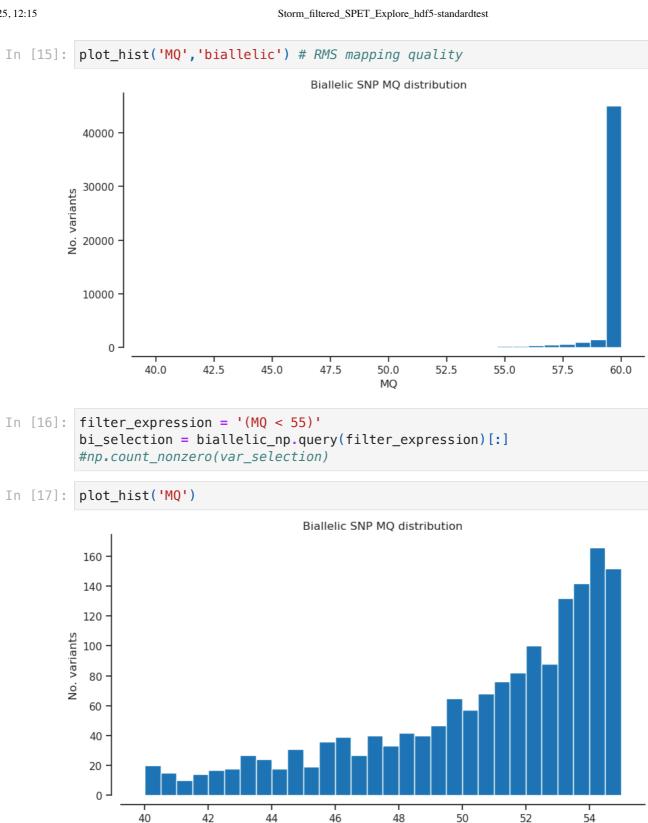
In [13]: filter_expression = '(numalt == 1)'
biallelic_np = rawsnps.query(filter_expression)[:]
biallelic_np
```

Out[13]: <VariantTable shape=(51348,) dtype=(numpy.record, [('AC', '<i4', (3,)), ('AF', '<f4', (3,)), ('ALT', 'O', (3,)), ('AN', '<i4'), ('BaseQRankSum', '<f4'), ('CHROM', 'O'), ('DP', '<i4'), ('END', '<i4'), ('ExcessHet', '<f4'), ('FILTER\_FILT', '?'), ('FILTER\_LowQual', '?'), ('FILTER\_PASS', '?'), ('FS', '<f4'), ('ID', 'O'), ('InbreedingCoeff', '<f4'), ('MLEAC', '<i4', (3,)), ('MLEAF', '<f4', (3,)), ('MQRankSum', '<f4'), ('POS', '<i4'), ('QD', '<f4'), ('QUAL', '<f4'), ('RAW\_MQandDP', '<i4', (2,)), ('REF', 'O'), ('ReadPosRankSum', '<f4'), ('SOR', '<f4'), ('altlen', '<i4', (3,)), ('is\_snp', '?'), ('numalt', '<i4')])>

	AC	AF	ALT	AN	BaseQRankSum	CHROM	DP	END	ExcessHet
0	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2059	-1	0.0971
1	[ 6 -1 -1]	[0.032 nan nan]	[b'T' b'' b'']	212	0.0	b'Chr01'	2065	-1	0.0963
2	[312 -1 -1]	[0.788 nan nan]	[b'G' b'' b'']	378	0.0	b'Chr01'	3473	-1	0.0
•••									
51345	[ 6 -1 -1]	[0.012 nan nan]	[b'C' b'' b'']	490	0.434	b'Chr17'	24021	-1	0.0008
51346	[261 -1 -1]	[0.542 nan nan]	[b'C' b'' b'']	462	0.0	b'Chr17'	6762	-1	0.7781
51347	[ 4 -1 -1]	[0.01 nan nan]	[b'T' b'' b'']	442	0.0	b'Chr17'	4997	-1	0.0891

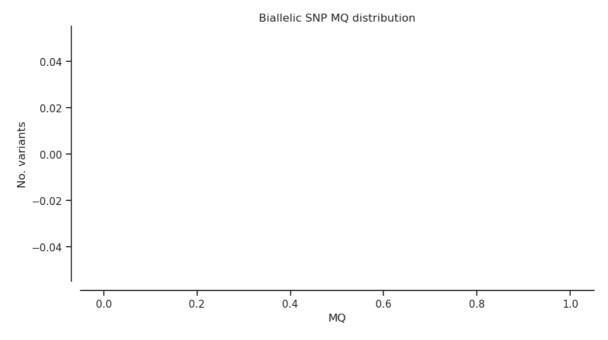
## MQ - RMS mapping quality

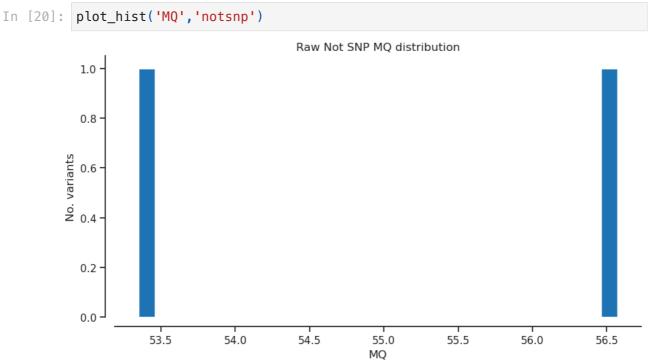




```
In [18]: filter_expression = '(MQ < 40)'
bi_selection = biallelic_np.query(filter_expression)[:]
In [19]: plot_hist('MQ')</pre>
```

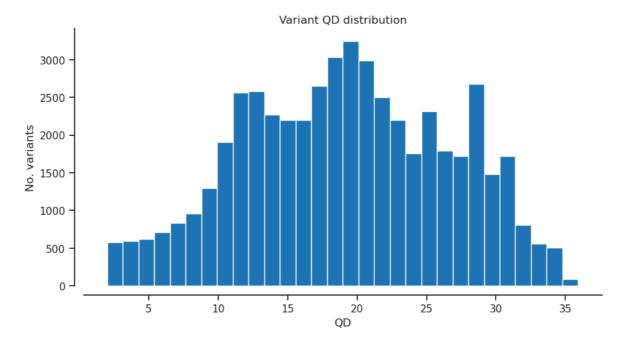
MQ



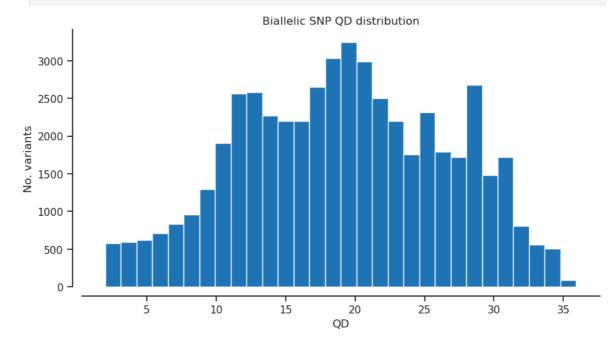


## QD - Variant Confidence/Quality by Depth

In [21]: plot\_hist('QD','var') # Variant Confidence/Quality by Depth

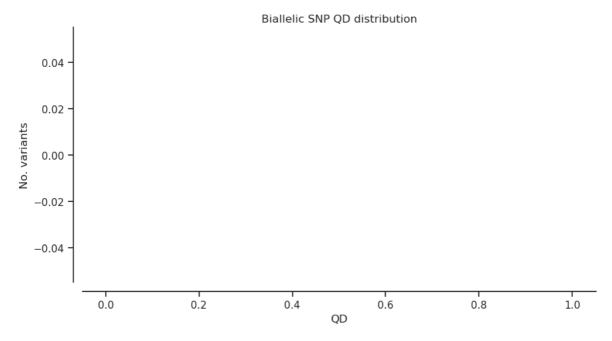


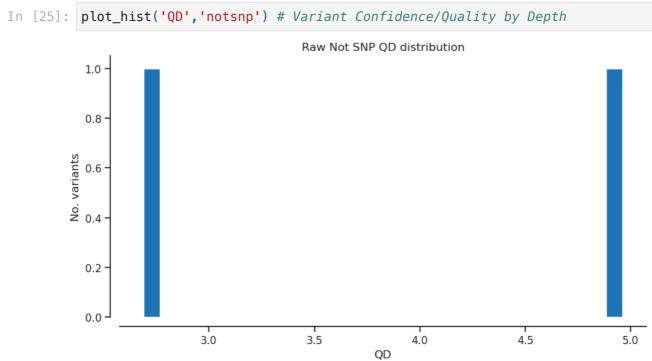
In [22]: plot\_hist('QD','biallelic') # Variant Confidence/Quality by Depth



```
In [23]: filter_expression = '(QD < 2)'
bi_selection = biallelic_np.query(filter_expression)[:]</pre>
```

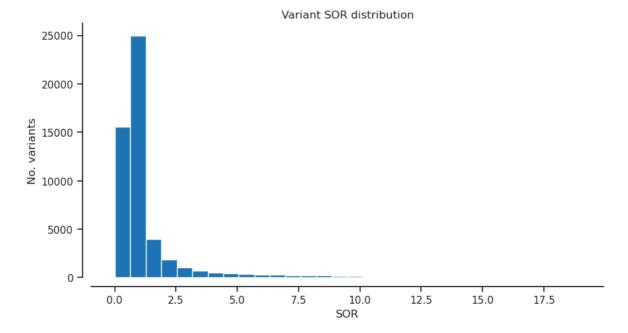
In [24]: plot\_hist('QD')



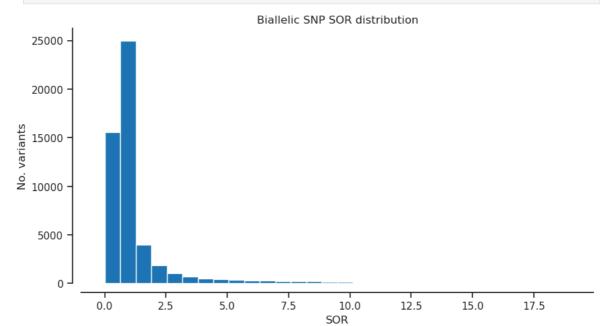


SOR - Symmetric Odds Ratio of 2x2 contingency table to detect strand bias

In [26]: plot\_hist('SOR', 'var') # Symmetric Odds Ratio of 2x2 contingency table t

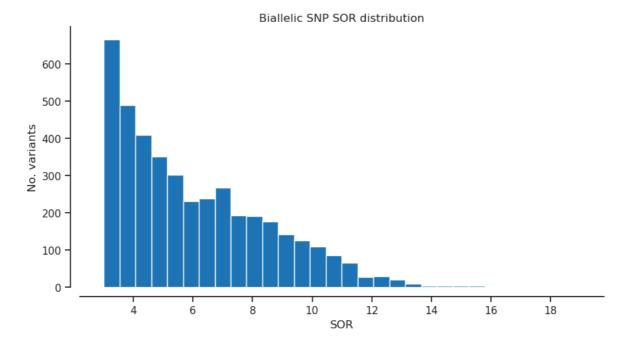




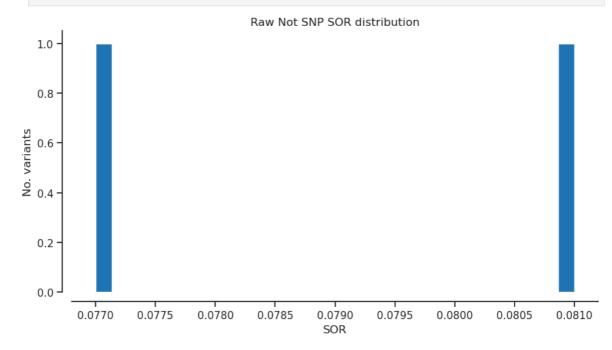


```
In [28]: filter_expression = '(SOR > 3)'
bi_selection = biallelic_np.query(filter_expression)[:]
```

In [29]: plot\_hist('SOR') # Symmetric Odds Ratio of 2x2 contingency table to detec

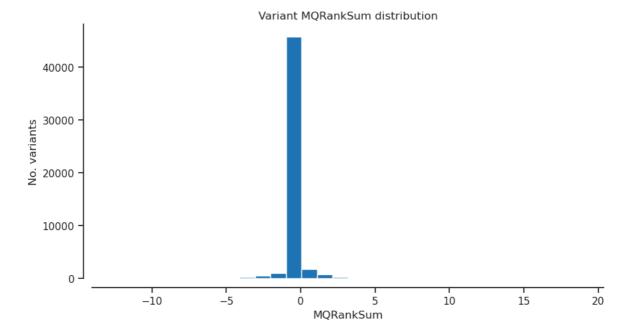


In [30]: plot\_hist('SOR', 'notsnp') # Symmetric Odds Ratio of 2x2 contingency table

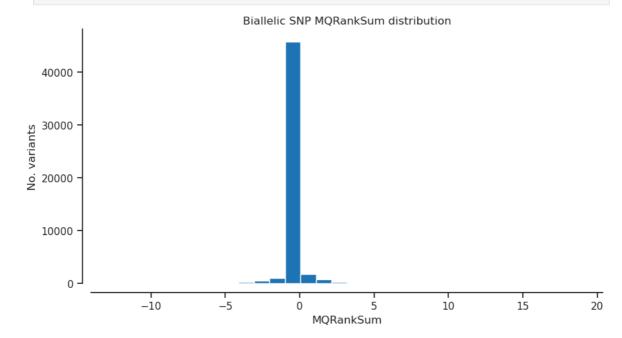


## MQRankSum - Z-score From Wilcoxon rank sum test of Alt vs. Ref read mapping qualities

In [31]: plot\_hist('MQRankSum','var') # Z-score From Wilcoxon rank sum test of Alt

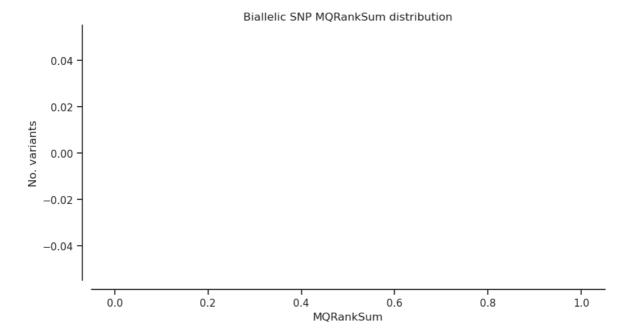


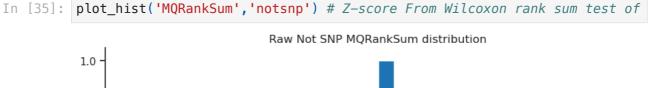
In [32]: plot\_hist('MQRankSum', 'biallelic') # Z-score From Wilcoxon rank sum test

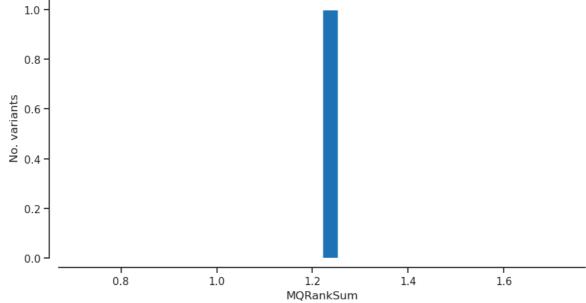


```
In [33]: filter_expression = '(MQRankSum < -12.5)'
bi_selection = biallelic_np.query(filter_expression)[:]</pre>
```

In [34]: plot\_hist('MQRankSum') # Z-score From Wilcoxon rank sum test of Alt vs. R

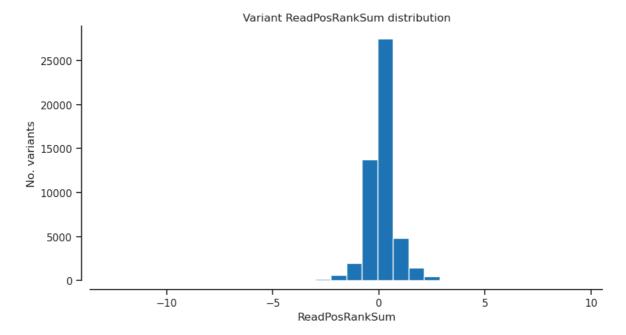




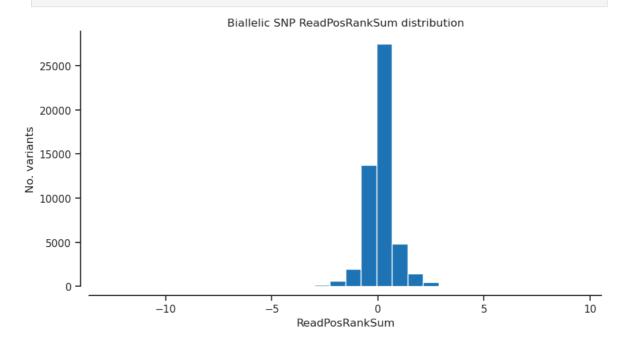


# ReadPosRankSum - Z-score from Wilcoxon rank sum test of Alt vs. Ref read position bias

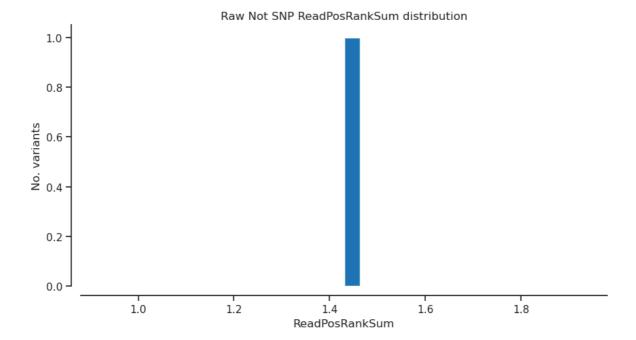
In [36]: plot\_hist('ReadPosRankSum','var') # Z-score from Wilcoxon rank sum test o



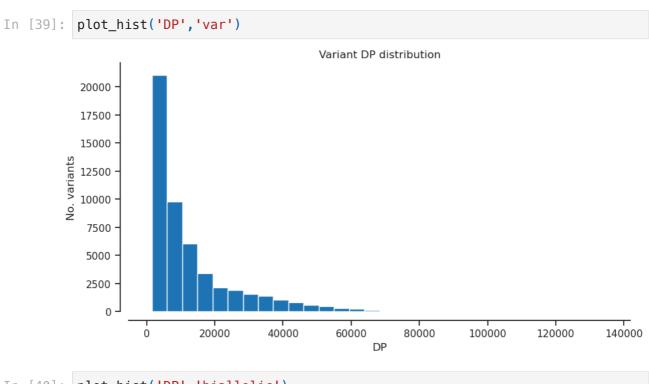
In [37]: plot\_hist('ReadPosRankSum', 'biallelic') # Z-score from Wilcoxon rank sum



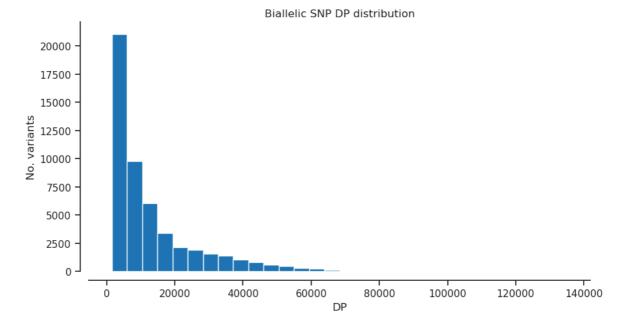
In [38]: plot\_hist('ReadPosRankSum', 'notsnp') # Z-score from Wilcoxon rank sum tes



## DP - Approximate read depth

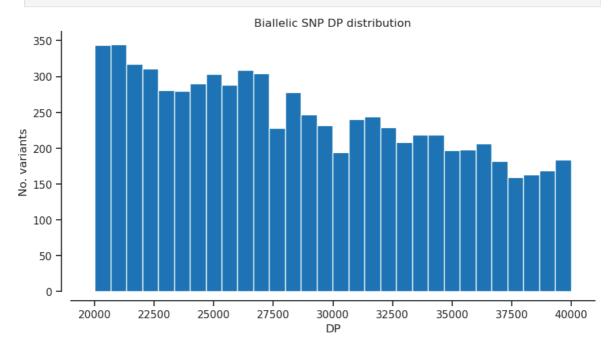


In [40]: plot\_hist('DP','biallelic')

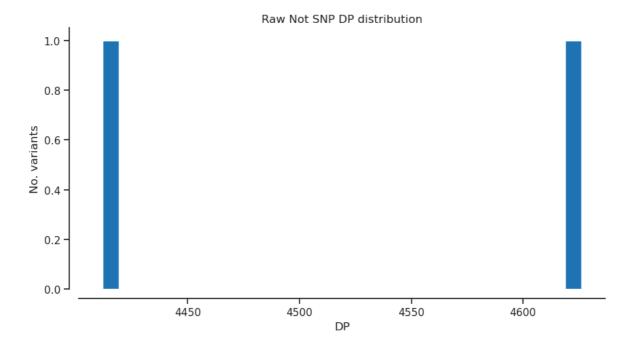


In [41]: filter\_expression = '(DP > 20000) & (DP < 40000)'
bi\_selection = biallelic\_np.query(filter\_expression)[:]</pre>

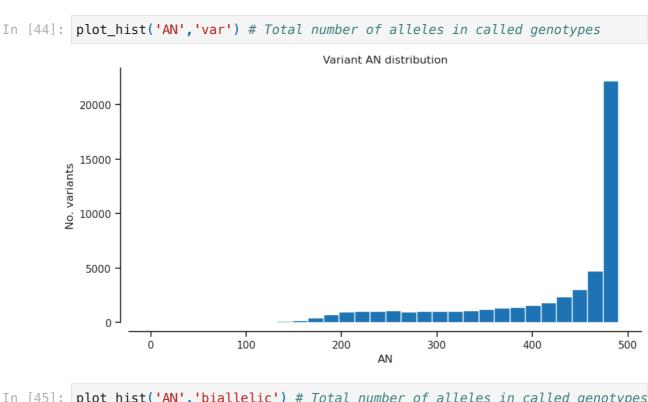




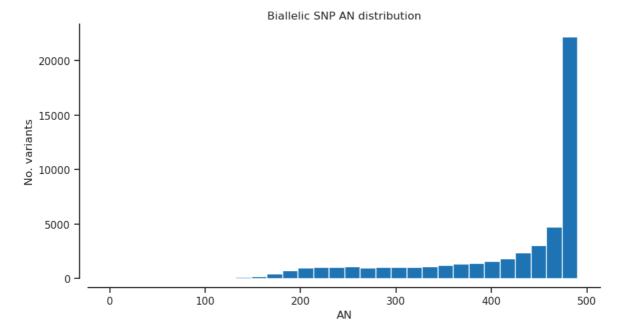
In [43]: plot\_hist('DP','notsnp')



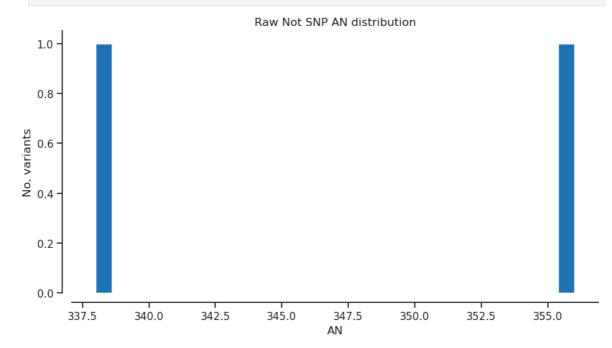
## AN - Total number of alleles in called genotypes



plot\_hist('AN','biallelic') # Total number of alleles in called genotypes







## Selected filter

```
In [47]: # QD: Variant Confidence/Quality by Depth
# AN: Total number of alleles in called genotypes
filter_expression = '(QD >= 2) & (MQ >= 40) & (MQRankSum >= -12.5) & (is_
variant_selection = variants_np.eval(filter_expression)[:]
np.count_nonzero(variant_selection)
```

Out[47]: 51180

## Genotype

```
In [48]: calldata_var = callset_var['calldata']
list(calldata_var)
```

```
Out[48]: ['AD', 'DP', 'GQ', 'GT', 'MIN_DP', 'PGT', 'PID', 'PL', 'PS', 'RGQ', 'S B']
```

Out [49]: <GenotypeChunkedArray shape=(51350, 245, 2) dtype=int8 chunks=(65536, 64, 2) nbytes=24.0M cbytes=3.5M cratio=6.9 compression=gzip compression\_opts=1 values=h5py.\_hl.dataset.Dataset>

	0	1	2	3	4	•••	240	241	242	243	244
0	./.	./.	./.	0/0	./.	•••	0/0	0/0	0/0	0/0	0/0
1	./.	./.	./.	0/0	./.	•••	0/0	0/0	0/0	0/0 0/0 0/0	0/0
2	1/1	1/1	1/1	1/1	./.	•••	0/0	1/1	1/1	0/0	0/1
•••						•					
51347	0/0	0/0	0/0	0/0	0/0	•••	0/0	0/0	0/0	0/0	0/0
51348											
51349	0/0	0/0	0/0	0/0	0/1	•••	0/0	0/0	0/0	0/0	0/0

```
In [50]: # using the selected filters set above
    gt_filtered_snps = genotypes_var.subset(variant_selection)
    gt_filtered_snps
```

Out [50]: <GenotypeChunkedArray shape=(51180, 245, 2) dtype=int8 chunks=(1600, 245, 2)
 nbytes=23.9M cbytes=6.0M cratio=4.0 compression=blosc compression\_opts=
 {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

	0	1	2	3	4	•••	240	241	242	243	244
0	./.	./.	./.	0/0	./.	•••	0/0	0/0	0/0	0/0	0/0
1	./.	./.	./.	0/0	./.	•••	0/0	0/0	0/0	0/0	0/0
2	1/1	1/1	1/1	1/1	./.	•••	0/0	1/1	1/1	0/0	0/1
•••						•	••				
51177	0/0	0/0	0/0	0/0	0/0	•••	0/0	0/0	0/0	0/0	0/0
51178	0/0	1/1	1/1	0/1	0/0	•••	0/1	0/1	0/1	0/1	0/1
51179	0/0	0/0	0/0	0/0	0/1	•••	0/0	0/0	0/0	0/0	0/0

```
In [51]: # grab the allele counts for the populations
ac = gt_filtered_snps.count_alleles()
ac
```

out [51]: <AlleleCountsChunkedArray shape=(51180, 2) dtype=int32 chunks=(25590, 2)
 nbytes=399.8K cbytes=145.5K cratio=2.7 compression=blosc compression\_opts=
 {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

	0	1				
0	206	6				
1	206	6				
2	66	312				
•••						
51177	484	6				
51178	201	261				
51179	438	4				

```
In [52]: ac[:]
```

Out [52]: <AlleleCountsArray shape=(51180, 2) dtype=int32>

```
0
              1
  0
        206
              6
  1
        206
  2
        66
             312
51177
       484
              6
51178
        201
             261
        438
              4
51179
```

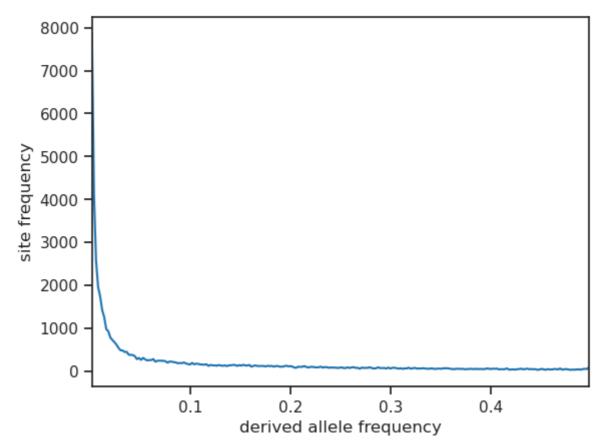
```
In [53]: # Which ones are biallelic?
  is_biallelic_01 = ac.is_biallelic_01()[:]
  ac1 = ac.compress(is_biallelic_01, axis=0)[:, :2]
  ac1
  ##this part of the code is only for graphing the SFS, is not useful for f
```

Out [53]: <AlleleCountsArray shape=(51180, 2) dtype=int32>

	0	1				
0	206	6				
1	206	6				
2	66	312				
•••	••	•				
 51177	 484	. 6				
 51177 51178	 484 201	6 261				

```
In [54]: # plot the sfs of the derived allele
s = allel.sfs_folded(ac1)
allel.plot_sfs(s, yscale="linear", n=ac1.sum(axis=1).max())
```

Out[54]: <Axes: xlabel='derived allele frequency', ylabel='site frequency'>



```
In [55]: biallelic = (ac.max_allele() == 1)
###This is the filter expression for biallelic sites
biallelic
```

```
In [56]: # select only the biallelic variants
   gt_biallelic = gt_filtered_snps.compress(biallelic)
   gt_biallelic
```

Out [56]: <GenotypeChunkedArray shape=(51180, 245, 2) dtype=int8 chunks=(1600, 245, 2) nbytes=23.9M cbytes=6.0M cratio=4.0 compression=blosc compression\_opts= {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

							240				
0	./.	./.	./.	0/0	./.		0/0	0/0	0/0	0/0	0/0
1	./.	./.	./.	0/0	./.		0/0	0/0	0/0	0/0	0/0
2	1/1	1/1	1/1	1/1	./.	•••	0/0	1/1	1/1	0/0	0/1
•••						•					
51177	0/0	0/0	0/0	0/0	0/0	•••	0/0	0/0	0/0	0/0	0/0
51178	0/0	1/1	1/1	0/1	0/0		0/1	0/1	0/1	0/1	0/1
51179	0/0	0/0	0/0	0/0	0/1	•••	0/0	0/0	0/0	0/0	0/0

```
In [57]: n_variants = len(gt_biallelic)
         n_variants
```

Out[57]: 51180

```
pc_missing = gt_biallelic.count_missing(axis=0)[:] * 100 / n_variants
In [58]:
         pc_het = gt_biallelic.count_het(axis=0)[:] * 100 / n_variants
```

## Samples

```
In [59]: samples_var = callset_var['samples']
         samples_var = list(samples_var)
         samples_var
```

```
[b'AUT00154-001',
Out [59]:
           b'AUT00154-002',
           b'AUT00154-003'
           b'AUT00154-004'
           b'AUT00154-005',
           b'AUT00154-006',
           b'AUT00154-007'
           b'AUT00154-008',
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```

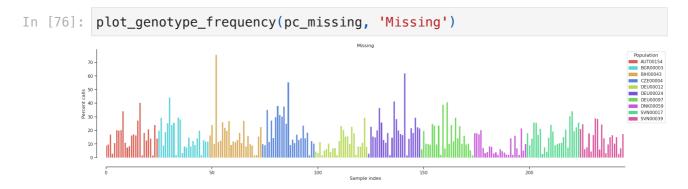
b'SVN00039-017'.

```
b'SVN00039-018',
          b'SVN00039-019',
          b'SVN00039-020',
          b'SVN00039-021']
In [72]: samples_fn = '~/scratch/data/Storminalis/Storminalis_sample_list_filtered
         samples = pandas.read_csv(samples_fn, sep='\t')
         samples
Out[72]:
                         ID Population
               AUT00154-001
                             AUT00154
              AUT00154-002
                             AUT00154
              AUT00154-003
                             AUT00154
              AUT00154-004
                             AUT00154
               AUT00154-005
                             AUT00154
         240
               SVN00039-017
                             SVN00039
          241
              SVN00039-018
                             SVN00039
         242 SVN00039-019
                             SVN00039
         243 SVN00039-020
                             SVN00039
         244 SVN00039-021
                             SVN00039
         245 rows × 2 columns
In [73]:
         samples.Population.value_counts()
Out[73]:
         Population
         AUT00154
                      25
                      25
         BIH00043
         DEU00097
                      25
          CZE00004
                      25
         DEU00012
                      25
         DEU00024
                      25
                      25
          SVN00017
         DNK00059
                      25
          BGR00003
                      24
          SVN00039
                      21
         Name: count, dtype: int64
         populations = samples.Population.unique()
In [74]:
         populations
         ###This identifiers come from the metadata file
         array(['AUT00154', 'BGR00003', 'BIH00043', 'CZE00004', 'DEU00012',
Out[74]:
                 'DEU00024', 'DEU00097', 'DNK00059', 'SVN00017', 'SVN00039'],
                dtype=object)
```

## Gt frequency function

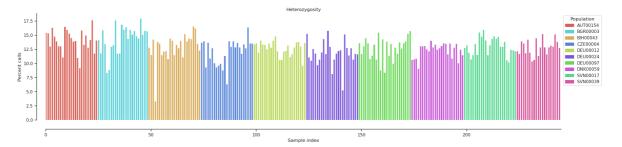
```
In [75]: def plot genotype frequency(pc, title):
             fig, ax = plt.subplots(figsize=(24, 5))
             sns.despine(ax=ax, offset=24)
             left = np.arange(len(pc))
             palette = sns.color_palette("hls", 10)
             pop2color = {'AUT00154': palette[0],
                           'BGR00003': palette[5],
                           'BIH00043': palette[1],
                           'CZE00004': palette[6],
                           'DEU00012': palette[2],
                           'DEU00024': palette[7],
                           'DEU00097': palette[3],
                           'DNK00059': palette[8],
                           'SVN00017': palette[4],
                           'SVN00039': palette[9]}
             colors = [pop2color[p] for p in samples.Population]
             ax.bar(left, pc, color=colors)
             ax.set_xlim(0, len(pc))
             ax.set_xlabel('Sample index')
             ax.set_ylabel('Percent calls')
             ax.set title(title)
             handles = [mpl.patches.Patch(color=palette[0]),
                        mpl.patches.Patch(color=palette[5]),
                        mpl.patches.Patch(color=palette[1]),
                        mpl.patches.Patch(color=palette[6]),
                        mpl.patches.Patch(color=palette[2]),
                        mpl.patches.Patch(color=palette[7]),
                        mpl.patches.Patch(color=palette[3]),
                        mpl.patches.Patch(color=palette[8]),
                        mpl.patches.Patch(color=palette[4]),
                        mpl.patches.Patch(color=palette[9])]
             ax.legend(handles=handles, labels=['AUT00154', 'BGR00003', 'BIH00043'
                 'DEU00024', 'DEU00097', 'DNK00059', 'SVN00017', 'SVN00039'], title
                       bbox to anchor=(1, 1), loc='upper left')
```

## Plot missing



## Plot heterozygosity

```
In [77]: plot_genotype_frequency(pc_het, 'Heterozygosity')
```



#### **PCA**

```
palette = sns.color_palette("hls",10)
In [78]:
         pop_colours = {
                          'AUT00154': palette[0],
                           'BGR00003': palette[5],
                           'BIH00043': palette[1],
                           'CZE00004': palette[6],
                           'DEU00012': palette[2],
                           'DEU00024': palette[7],
                           'DEU00097': palette[3],
                           'DNK00059': palette[8],
                           'SVN00017': palette[4],
                           'SVN00039': palette[9]
In [79]: def plot_pca_coords(coords, model, pc1, pc2, ax, sample_population):
             sns.despine(ax=ax, offset=5)
             x = coords[:, pc1]
             y = coords[:, pc2]
             for pop in populations:
                 flt = (sample_population == pop)
                 ax.plot(x[flt], y[flt], marker='o', linestyle=' ', color=pop_colo
                          label=pop, markersize=6, mec='k', mew=.5)
             ax.set_xlabel('PC%s (%.1f%%)' % (pc1+1, model.explained_variance_rati
             ax.set_ylabel('PC%s (%.1f%%)' % (pc2+1, model.explained_variance_rati
         def fig_pca(coords, model, title, sample_population=None):
             if sample_population is None:
                 sample_population = samples.Population
             # plot coords for PCs 1 vs 2, 3 vs 4
             fig = plt.figure(figsize=(10, 5))
             ax = fig.add_subplot(1, 2, 1)
             plot_pca_coords(coords, model, 0, 1, ax, sample_population)
             ax = fig.add_subplot(1, 2, 2)
             plot_pca_coords(coords, model, 2, 3, ax, sample_population)
             ax.legend(bbox_to_anchor=(1, 1), loc='upper left')
             fig.suptitle(title, y=1.02)
             fig.tight_layout()
         ac2 = gt_biallelic.count_alleles()
In [80]:
         ac2
```

Out [80]: <AlleleCountsChunkedArray shape=(51180, 2) dtype=int32 chunks=(25590, 2) nbytes=399.8K cbytes=145.5K cratio=2.7 compression=blosc compression\_opts= {'cname': 'lz4', 'clevel': 5, 'shuffle': 1, 'blocksize': 0} values=zarr.core.Array>

	0	1			
0	206	6			
1	206	6			
2	66	312			
•••					
51177	484	6			
51178	201	261			
51179	438	4			

```
In [81]: flt = (ac2[:, :2].min(axis=1) > 1)
   gf = gt_biallelic.compress(flt, axis=0)
   gn = gf.to_n_alt()
   gn
```

```
In [82]: coords1, model1 = allel.pca(gn, n_components=10, scaler='patterson')
```

```
In [83]: fig_pca(coords1, model1, 'Figure 1. Conventional PCA.')
```

Figure 1. Conventional PCA.

