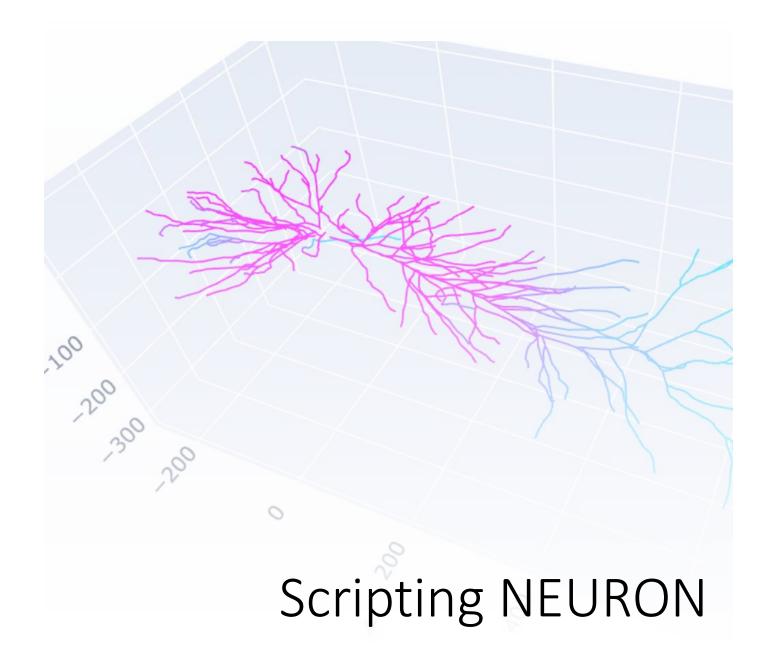
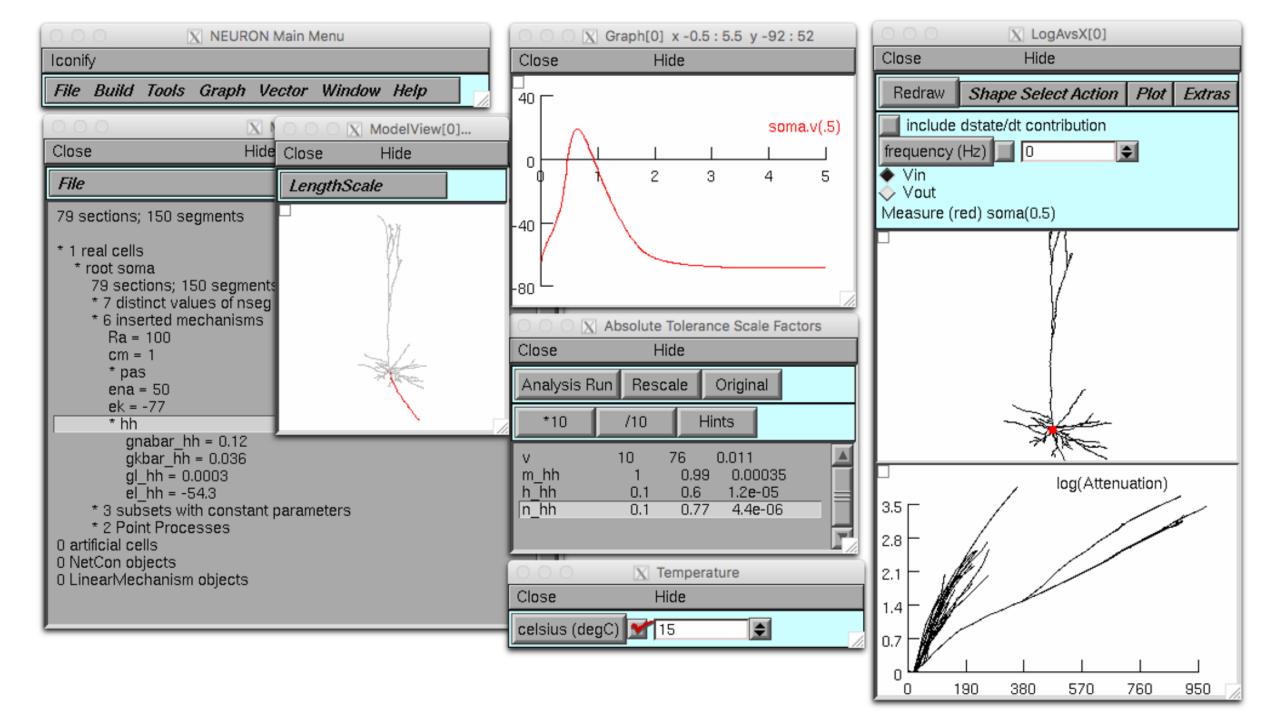
```
from neuron import h
from neuron.units import mV, ms
from matplotlib import cm
import plotly
h.load_file('stdrun.hoc')
h.load_file('c91662.ses')
h.hh.insert(h.allsec())
ic = h.IClamp(h.soma(0.5))
ic.delay = 1 * ms
ic.dur = 1 * ms
ic.amp = 10
h.finitialize(-65 * mV)
h.continuerun(2 * ms)
ps = h.PlotShape(False)
ps.variable('v')
ps.plot(plotly, cmap=cm.cool).show()
```



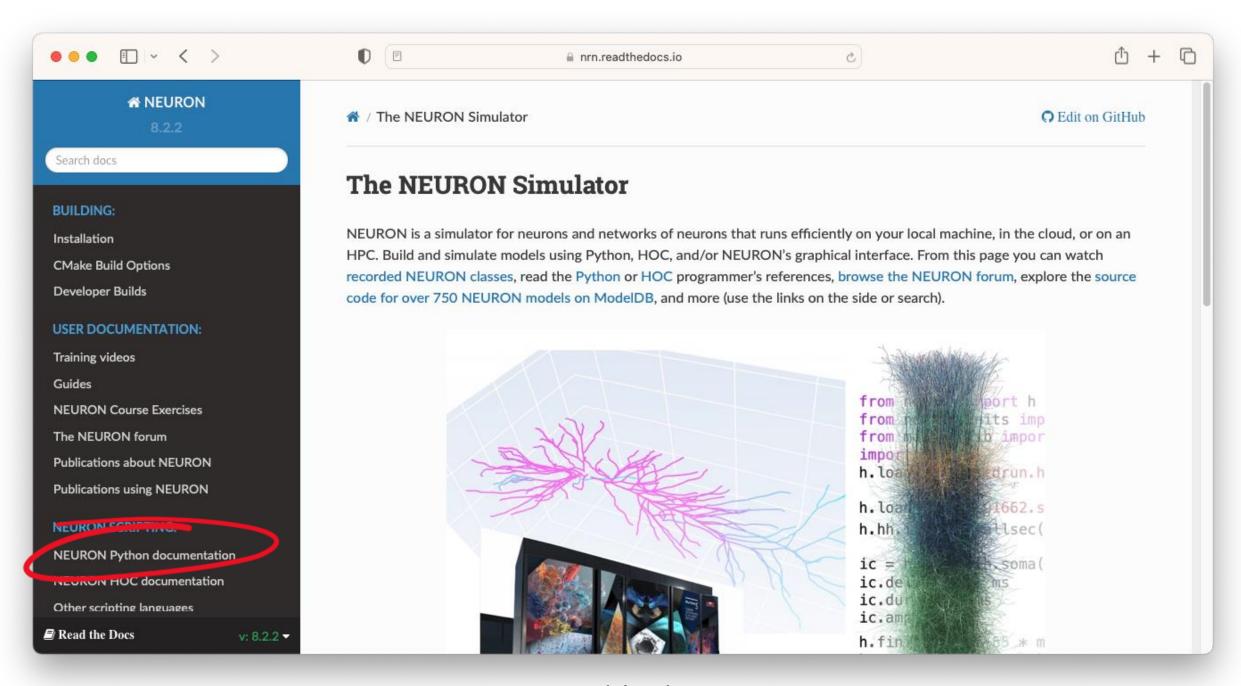


What is a script?

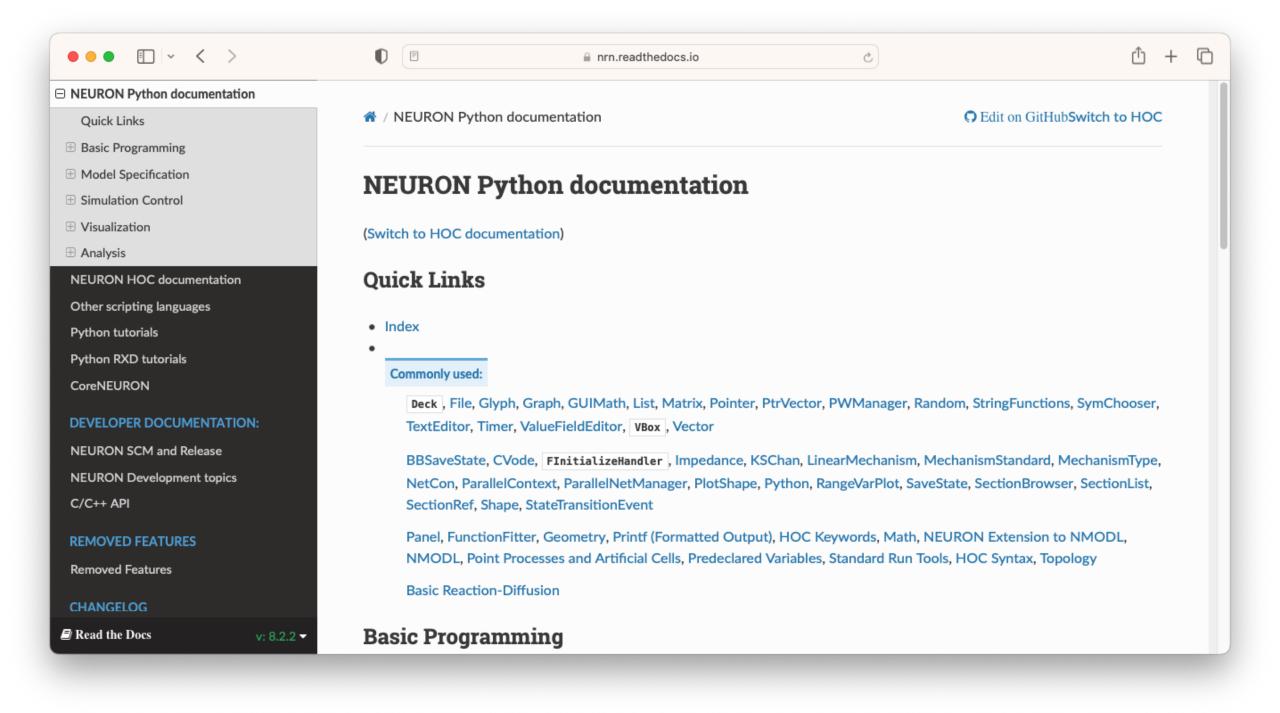
- A **script** is a file with computer-readable instructions for performing a task.
- In NEURON, scripts can:
 - set-up a module
 - define and perform an experimental protocol
 - record data
 - save and load data
 - and more ...

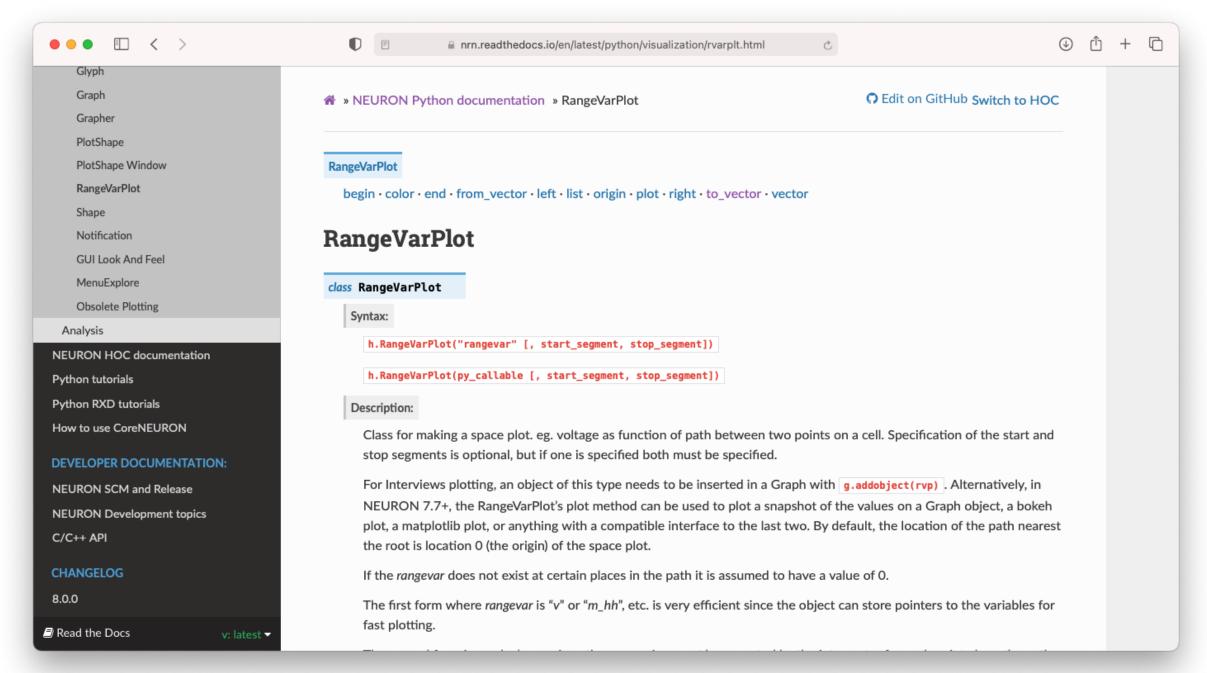
Why write scripts for NEURON?

- Automation ensures **consistency** and reduces manual effort.
- Facilitates comparing the suitability of different models.
- Facilitates **repeated experiments** on the same model with different parameters (e.g. drug dosages).
- Facilitates **re-collecting data** after change in experimental protocol.
- Provides a complete, **reproducible** version of the experimental protocol.

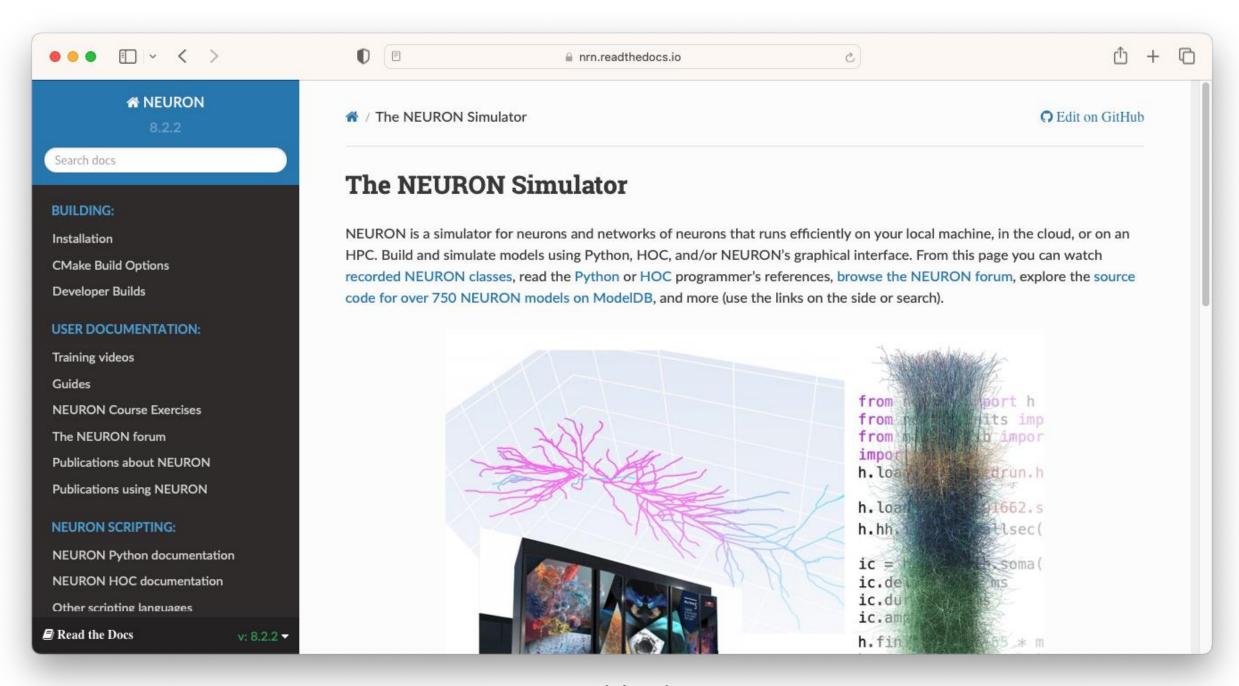


nrn.readthedocs.io

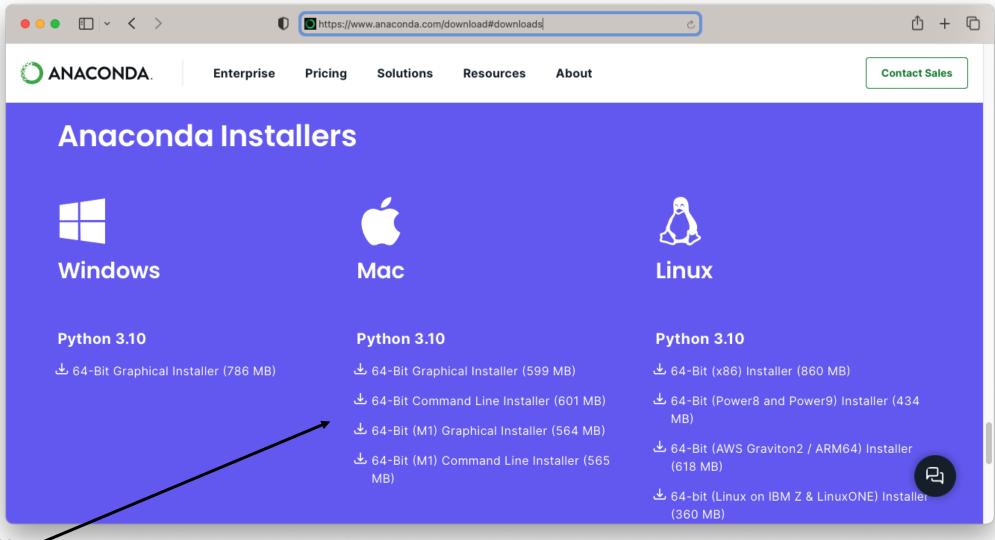




Use the "Switch to HOC" link in the upper-right corner of every page if you need documentation for HOC, NEURON's original programming language. HOC may be used in combination with Python: use h.load file to load a HOC library; the functions and classes are then available with an h. prefix.

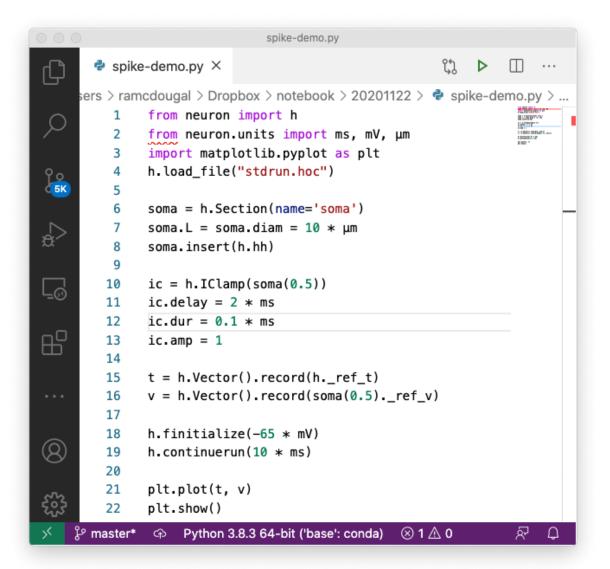


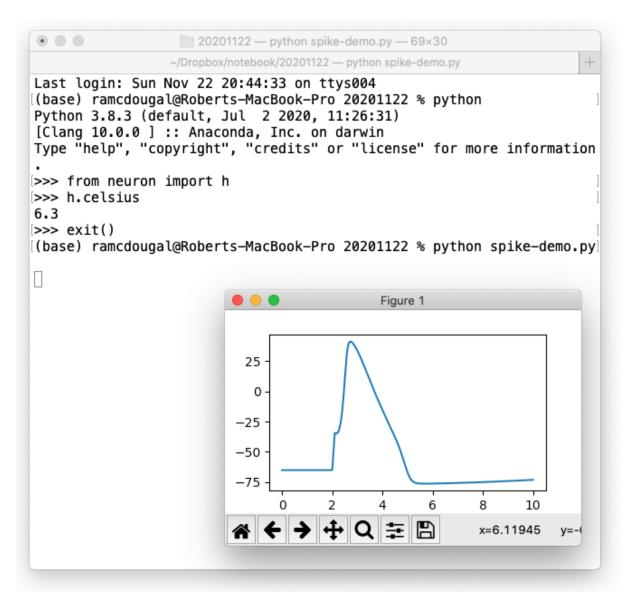
nrn.readthedocs.io

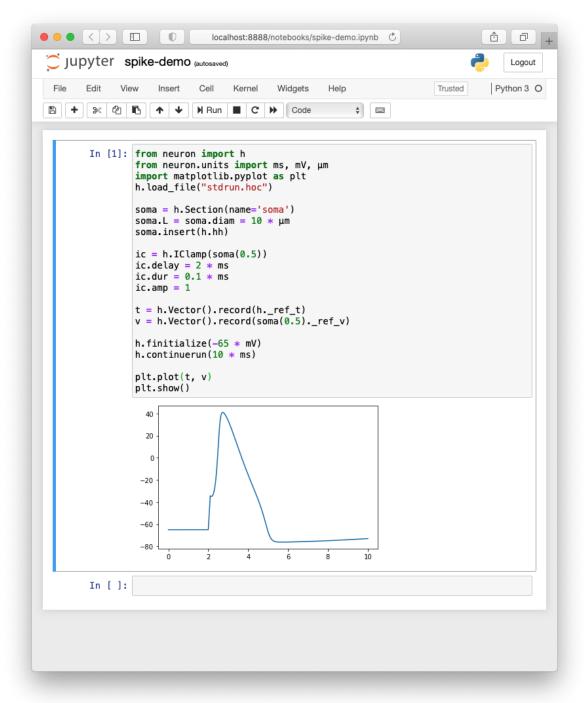


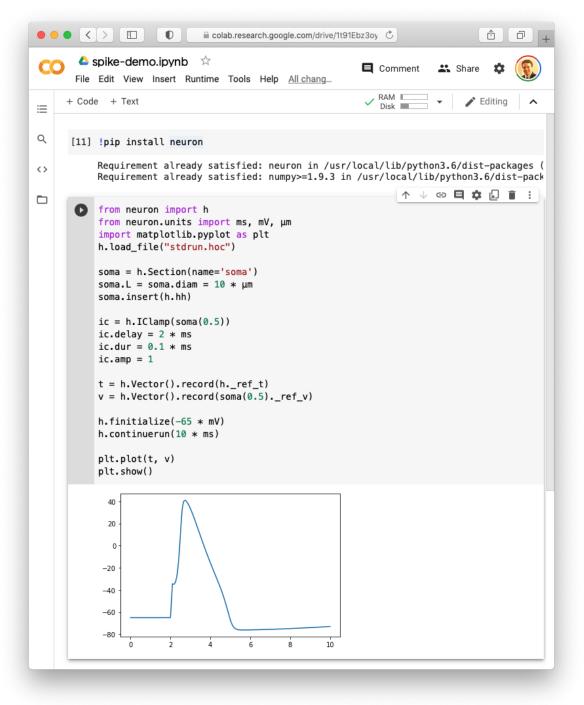
Note: on a Mac, be sure to get a Python version that matches your hardware (Intel or M1/M2)

There are many Python distributions. Any should work, but many people prefer Anaconda as it comes with a large set of useful libraries.









Introduction to Python

Displaying results: the print function

```
print("NEURON is a great tool for simulation.")
NEURON is a great tool for simulation.
print(5 * (3 + 2))
25
print(soma.diam)
10.0
```

Variables

• Give things a name to access them later:

```
diameter = 4
print("The diameter is", diameter)
print("The square of the diameter is", diameter ** 2)
The diameter is 4
The square of the diameter is 16
```

Lists and for loops

 To do the same thing to several items, put the items in a list and use a for loop:

```
cell_parts = ["soma", "apical", "basal", "axon"]
for part in cell_parts:
    print(part)
```

• Items in a list can be accessed directly using the [] notation. Note: lists start at position 0.

```
print(cell_parts[2])
basal
```

• To check if an item is in a list, use in:

```
print("brain" in cell_parts)
False
```

Dictionaries

• If there is no natural order, specify your own key-value pairs:

```
diameters = {"soma": 10, "axon": 2, "apical": 5}
print(diameters["apical"])
5
```

Loop over keys and values using .items():

```
for name, diam in diameters.items():
    print("The diameter of", name, "is", diam, "microns")

The diameter of soma is 10 microns
The diameter of axon is 2 microns
The diameter of apical is 5 microns
```

Functions

- If a calculation is used more than once, give it a name via def and refer to it by the name.
- If there is a complicated self-contained calculation, give it a name.
- Return the result of the calculation with the return keyword.

```
def volume_of_cylinder(diameter, length):
    return (3.14 / 4) * diameter ** 2 * length

vol1 = volume_of_cylinder(5, 20)
apical_vol = volume_of_cylinder(apical.diam, apical.L)
```

Libraries (aka "modules")

- Python modules provide functions, classes, and values that your scripts can use.
- To load a module, use import:

```
import math
```

Use dot notation to access a function or value from the module:

```
print(math.cos(math.pi / 3))
```

0.50000000000000001

 One can also load specific items from a module or give a short-hand name for the module:

```
from neuron import h, gui
import pandas as pd
```

Other useful Python modules

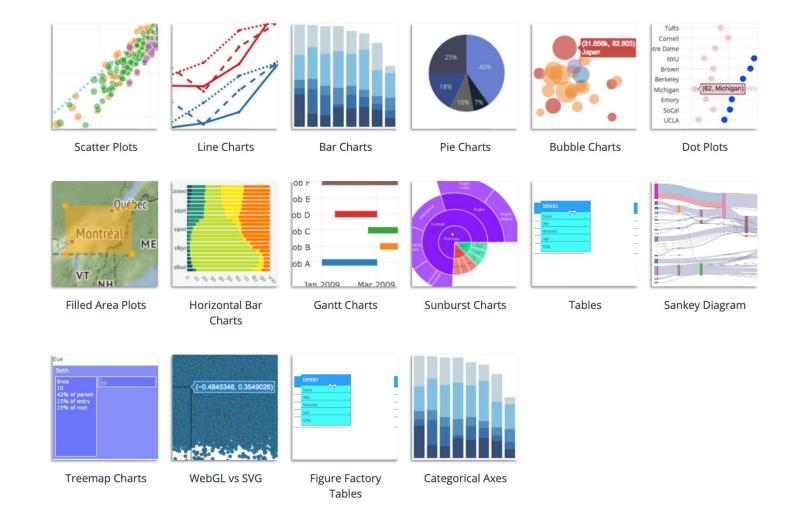
- math
 - Basic math functions
- numpy
 - Advanced math functions
- pandas
 - Basic data science and database access
- sklearn, tensorflow, pytorch, transformers
 - Machine learning
- plotly, plotnine, matplotlib, mayavi
 - Plotting

plotly

Free (MIT licensed), full-featured graphics library

Graphs are interactive and can be saved.

Supports both Python and JavaScript.



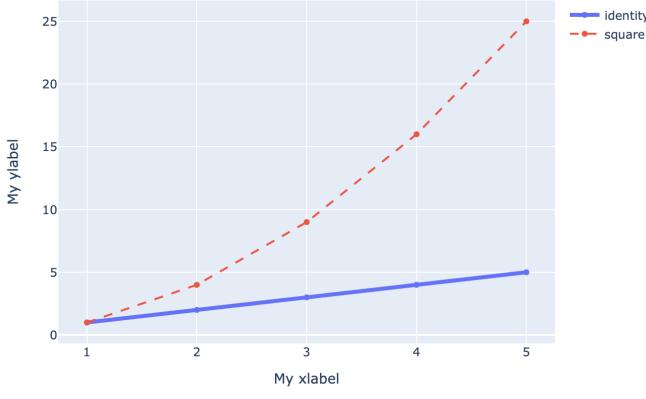
```
import plotly.graph_objects as go
```

```
fig = go.Figure()
fig.add_trace(
    go.Scatter(x=[1,2,3,4,5], y=[1,2,3,4,5], name="identity", line={"width": 4})
)
fig.add_trace(
    go.Scatter(x=[1,2,3,4,5], y=[1,4,9,16,25], name="square", line={"dash": "dash"})
)
fig.update_layout({
    "xaxis_title": "My xlabel",
    "yaxis_title": "My ylabel"
})
```

For NEURON built-in graphs, we'll just use:

import plotly

fig.show()



String formatting

- We'll often want to insert variables into text
 - labeling time points in graphs, storing parameters in data filenames, ...
- In Python, this is done using an f-string:

```
tstop = 10
my_string = f"We should stop at t = {tstop} ms"
```

 Formatting can be specified e.g. to round to a certain number of digits.

```
f"pi is approximately {pi:.5}"

f"pi is approximately {pi:.7}"

pi is approximately 3.1416

pi is approximately 3.141593
```

Getting help

• To get a list of functions, etc. in a module (or class) use dir:

```
from neuron import h
print(dir(h))

['APCount', 'AlphaSynapse', 'AtolTool', 'AtolToolItem', 'BBSaveState',
'CVode', 'DEG', 'Deck', 'E', 'ExecCommand', 'Exp2Syn', 'ExpSyn', 'FARAD
AY', 'FInitializeHandler', 'Family', 'File', 'GAMMA', 'GUIMath', 'Glyph
', 'Graph', 'HBox', 'IClamp', 'Impedance', 'Inserter', 'IntFire1', 'Int
Fire2', 'IntFire4', 'KSChan', 'KSGate', 'KSState', 'KSTrans', 'L', 'Lin
earMechanism', 'List', 'Matrix', 'MechanismStandard', 'MechanismType',
```

Getting help

• To see help information for a specific function, use help:

Getting help

Python is widely used, and there are many online resources available, including:

- docs.python.org the official documentation
- Stack Overflow a general-purpose programming forum
- The NEURON programmer's reference NEURON documentation
- The NEURON forum for NEURON-related programming questions



Basic NEURON Scripting

Loading NEURON

Core NEURON functionality

```
from neuron import h
```

Unit definitions

```
from neuron.units import mV, ms, um
```

Chemical dynamics

```
from neuron import rxd
```

You will almost always need these.

NEURON run control library

```
h.load file("stdrun.hoc")
```

stdrun.hoc loads NEURON's "standard run" system, which provides the h.continuerun function for running a simulation until a specific time.

Creating and naming sections

- A Section in NEURON is an unbranched stretch of e.g. dendrite.
- To create a Section, use h. Section and assign to a variable:

```
apical = h.Section("apical")
```

A single Section can have multiple references to it.

```
a = apical
print(a == apical)
True
```

 Printing a Section displays its name. Use str (section) to get the name as a string:

```
s = str(apical)
print(apical)
apical
```

Basic unit: h.Section

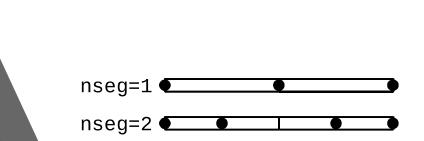
soma = h.Section('soma')

Length: soma.L

Diameter: soma.diam

Discretization: soma.nseg

TIP: Always use an odd value of nseg (puts a point at the center), and test convergence by multiplying by an odd number (all old centers will still exist).



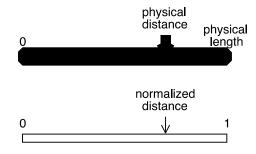
nseg=3

Inside a cell class, specify the cell argument as well:

soma = h.Section('soma', self)

Can also explicitly label name='soma', cell='self', then can be in either order

TIP: Provide a
__repr__ method
for the cell class to
allow Sections to
work with
NEURON's built-in
GUI tools



The connect method joins Section objects to define arbitrary morphologies.

Looping over a Section gives the Segments

```
dend = h.Section("dend")
dend.L = 3
dend.diam = 2
dend.nseg = 3

for seg in dend:
    print(seg, seg.area())
```

```
dend(0.166667) 6.283185307179586
dend(0.5) 6.283185307179586
dend(0.833333) 6.283185307179586
```

Getting x and Section

```
seg = dend(0.5)
print(seg.x, seg.sec == dend)
```

```
0.5 True
```

Select specific Segments; they can have different properties

```
dend(0.5).diam = 4

for seg in dend:
    print(seg, seg.area())

dend(0.166667) 6.283185307179586
dend(0.5) 12.566370614359172
dend(0.833333) 6.283185307179586
```

Not limited to cylinders

```
dend.nseg = 1
dend.pt3dclear()
dend.pt3dadd(0, 0, 0, 1)
dend.pt3dadd(10, 0, 0, 5)
dend(0.5).volume()
```

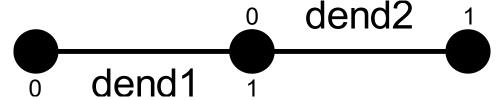
81.15781021773631

Connecting sections

To construct a neuron's full branching structure, individual sections must be connected using .connect:

```
dend2.connect(dend1(1))
```

Each section is oriented and has a 0- and a 1-end. In NEURON, traditionally the 0-end of a section is attached to the 1-end of a section closer to the soma. In the example above, dend2's 0-end is attached to dend1's 1-end.



To print the topology of cells in the model, use h.topology().

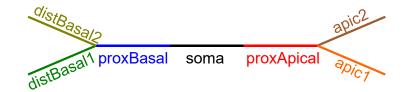
Example

```
from neuron import h
# define sections
soma = h.Section("soma")
papic = h.Section("proxApical")
apic1 = h.Section("apic1")
apic2 = h.Section("apic2")
pb = h.Section("proxBasal")
db1 = h.Section("distBasal1")
db2 = h.Section("distBasal2")
# connect them
papic.connect(soma)
pb.connect(soma(0))
apic1.connect(papic)
apic2.connect(papic)
db1.connect(pb)
db2.connect(pb)
# list topology
h.topology()
```

Output:

```
|-| soma(0-1)
    `| proxApical(0-1)
    `| apic1(0-1)
    `| apic2(0-1)
    `| proxBasal(0-1)
    `| distBasal1(0-1)
    `| distBasal2(0-1)
```

Morphology:



Length, diameter, and position

Set a Section's length with .L and diameter with .diam:

```
sec.L = 20 * um

sec.diam = 2 * um 

diameter may also be specified per segment
```

If no units are specified, NEURON assumes µm. ← um and µm are both recognized

To specify the (x, y, z; d) points a section sec passes through, use e.g. sec.pt3dadd(x, y, z, d). The section sec has sec.n3d() 3D points; their i^{th} x-coordinate is sec.x3d(i). The methods .y3d, .z3d, and .diam3d work similarly.

Caution: Squid

NEURON's defaults are based on the squid giant axon.

sec.diam: 500 μm

 $\text{sec.Ra: } 35.4~\Omega~\text{cm}$

h.celsius: 6.3 C



Tip: define classes of cells not individual cells

Consider the code

```
class Pyramidal:

def __init__(self):
    self.soma = h.Section("soma", self)
    self.soma.L = self.soma.diam = 10 * \mu m
```

• The __init__ method is run whenever a new Pyramidal cell is created; e.g. via

```
pyr1 = Pyramidal()
```

• The soma can be accessed using dot notation:

```
print(pyr1.soma.diam)
10.0
```

Tip: define classes of cells not individual cells

 By defining a cell in a class, once we are happy with it, we can a copy of the cell in a single line of code:

```
pyr2 = Pyramidal()
```

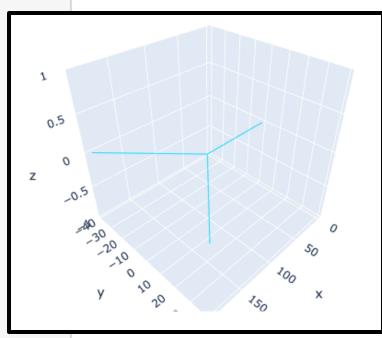
• Or even many copies:

```
pyrs = [Pyramidal() for i in range(1000)]
```

• For network models, helpful to associate a number (a gid) with each cell.

Viewing the morphology with h. PlotShape

```
from neuron import h
from neuron.units import um
import plotly
class Cell:
    def __init__(self):
        main = h.Section(name="main", cell=self)
        dend1 = h.Section(name="dend1", cell=self)
        dend2 = h.Section(name="dend2", cell=self)
        dend1.connect(main)
        dend2.connect(main)
        main.diam = 10 * um
        dend1.diam = 2 * um
        dend2.diam = 2 * um
        # important: store the sections
        self.main = main; self.dend1 = dend1; self.dend2 = dend2
        self.all = main.wholetree()
my_cell = Cell()
ps = h.PlotShape(False)
ps.plot(plotly).show()
```



Passing True instead of False will plot in an InterViews window instead.

The InterViews windows can be saved as postscript using e.g.

```
ps.printfile("filename.eps")
```

Viewing voltage, sodium, etc...

• Suppose we make the voltage ∨ nonuniform which we can do via:

```
my_cell.main.v = 50
my_cell.dend1.v = 0
my_cell.dend2.v = -65
```

• We can create a PlotShape that color-codes the sections by voltage:

```
ps = h.PlotShape(False)
ps.variable("v")
ps.scale(-80, 80)
ps.plot(plotly).show()
```

Viewing voltage, sodium, etc...

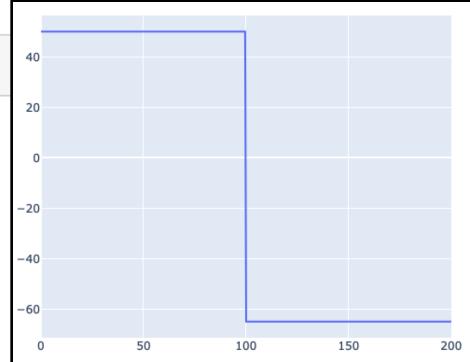
After increasing the spatial resolution:

```
for sec in my_cell.all: sec.nseg = 101
```

• We can plot the voltage as a function of distance from

main(0) **to** dend2(1):

```
rvp = h.RangeVarPlot('v', my_cell.main(0), my_cell.dend2(1))
rvp.plot(plotly).show()
```



Viewing voltage, sodium, etc...

Variable	Value	Pointer (e.g. for recording)	With PlotShape or RangeVarPlot
Voltage	seg.v	segref_v	"V"
Na ⁺ (inside membrane)	seg.nai	segref_nai	"nai"
Na ⁺ (outside membrane)	seg.nao	segref_nao	"nao"
Na ⁺ (current)	seg.ina	segref_ina	"ina"
Na ⁺ (reversal potential)	seg.ena	segref_ena	"ena"
d(sodium current)/dv	seg.dina_dv_	segref_dina_dv_	"dina_dv_"

Potassium is the same as for sodium, except with "k" replacing "na"; Chloride is the same except with "cl"; Calcium is the same except with "ca", etc... ions may only be accessed when a mechanism using them is present or when they are explicitly inserted via sec.insert or rxd.

Distributed mechanisms

• Insert a distributed mechanism (e.g. from a mod file) into a section or list of sections with .insert:

```
h.hh.insert(apical)
h.hh.insert([apical, soma, basal])
h.hh.insert(h.allsec())
```

 Mechanisms may also be inserted one-at-a-time into a single section via e.g.

```
apical.insert(h.hh)
```

Ion Channels

- Specify using insert method.
- Built-in: Hodgkin-Huxley (h.hh), passive (h.pas)
- Hundreds more on ModelDB (.mod files)
- Compile mod files via: nrnivmodl

Model

Hodgkin-Huxley cable equations

$$\frac{D}{4R_a}\frac{\partial^2 V}{\partial x^2} = C_m \frac{\partial V}{\partial t}$$

$$\begin{split} &+ \overline{g} \, m^3 \, h \cdot \big(V - E_{na} \big) + \overline{g}_k \, n^4 \cdot \big(V - E_k \big) + g_l \cdot \big(V - E_l \big) \\ &\frac{dm}{dt} = -\alpha_m m + \beta_m (1 - m) \quad \alpha_m = \frac{0.1 (V + 40)}{1 - \mathrm{e}^{-0.1 (V + 40)}} \quad \beta_m = 4 \mathrm{e}^{-(V + 65)/18} \\ &\frac{dh}{dt} = -\alpha_h h + \beta_h (1 - h) \quad \alpha_h = 0.07 \mathrm{e}^{-0.05 (V + 65)} \quad \beta_h = \frac{1}{1 + \mathrm{e}^{-0.1 (V + 35)}} \\ &\frac{dn}{dt} = -\alpha_n n + \beta_n (1 - n) \quad \alpha_n = \frac{0.01 (V + 55)}{1 - \mathrm{e}^{-0.1 (V + 55)}} \quad \beta_n = 0.125 \, \mathrm{e}^{-(V + 65)/80} \end{split}$$

Simulation

Representation

Defining ion channels, synapses, etc

tinyurl.com/hhmodfile tinyurl.com/expsyn

```
squid sodium, potassium, and leak channels
TITLE hh.mod
COMMENT
 This is the original Hodgkin-Huxley treatment for the set of sodium,
 potassium, and leakage channels found in the squid giant axon membrane.
 ("A quantitative description of membrane current and its application
  conduction and excitation in nerve" J.Physiol. (Lond.) 117:500-544 (1952).)
 Membrane voltage is in absolute mV and has been reversed in polarity
 from the original HH convention and shifted to reflect a resting potential
 of -65 \text{ mV}.
 Remember to set celsius=6.3 (or whatever) in your HOC file.
 See squid.hoc for an example of a simulation using this model.
 SW Jaslove 6 March, 1992
ENDCOMMENT
UNITS {
        (mA) = (milliamp)
        (mV) = (millivolt)
        (S) = (siemens)
? interface
NEURON {
        SUFFIX hh
        REPRESENTS NCIT: C17145
                                 : sodium channel
        REPRESENTS NCIT: C17008
                                 : potassium channel
        USEION na READ ena WRITE ina REPRESENTS CHEBI:29101
```

Compile mod files on your local machine using: nrnivmodl

Hundreds of mod files from published work are available at modeldb.science

Point processes

• To insert a point process, specify the segment when creating it, and save the return value. e.g.

```
pp = h.IClamp(soma(0.5))
```

• To find the segment containing a point process pp, use

```
seg = pp.get_segment()
```

• The section is then seg.sec and the normalized position is seg.x.

• The point process is removed when no variables refer to it.

Setting and reading parameters

- In NEURON, each section has normalized coordinates from 0 to 1.
- To read the value of a parameter defined by a range variable at a given normalized position, use: sec(x) . MECHANISM. VARNAME e.g.

```
gkbar = apical(0.2).hh.gkbar
```

Setting variables works the same way:

```
apical(0.2).hh.gkbar = 0.037
```

Setting and reading parameters

• To specify how many evenly-sized pieces (segments) a section should be broken into (each potentially with their own value for range variables), use section.nseg:

```
apical.nseg = 11
```

• To specify the temperature, use h.celsius:

```
h.celsius = 37
```

Setting and reading parameters

• Often you will want to read or write values on all segments in a section. To do this, use a for loop over the Section:

```
for seg in apical:
    seg.hh.gkbar = 0.037
```

• The above is equivalent to apical.gkbar_hh = 0.037, however the first version allows setting values nonuniformly, e.g.

```
for sec in h.allsec():
    for seg in sec:
        seg.hh.gkbar = some_function(h.distance(seg, soma(0.5)))
is an iterable of all sections
```

 A list comprehension can be used to create a Python list of all the values of a given property in a segment:

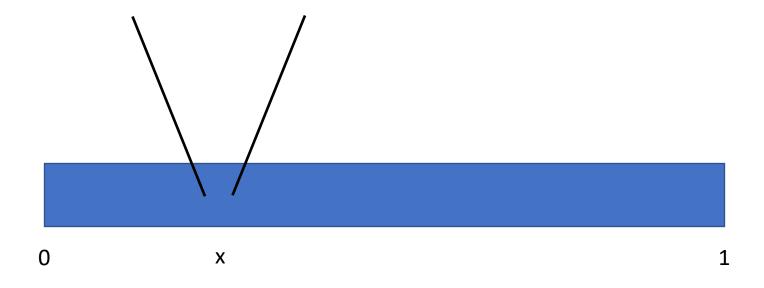
```
apical_gkbars = [segment.hh.gkbar for segment in apical]
```

Note: looping over a Section only returns true Segments. If you want to include the voltage-only nodes at 0 and 1, iterate over, e.g. apical.allseg() instead. HOC's for (x,0) and for (x) are equivalent to looping over a section and looping over allseg, respectively.

Recording Results

We can read the instantaneous membrane potential at a location via, e.g.

To record this value over time, we use an h. Vector and pass in the pointer (prefixed with _ref_) to the record method.



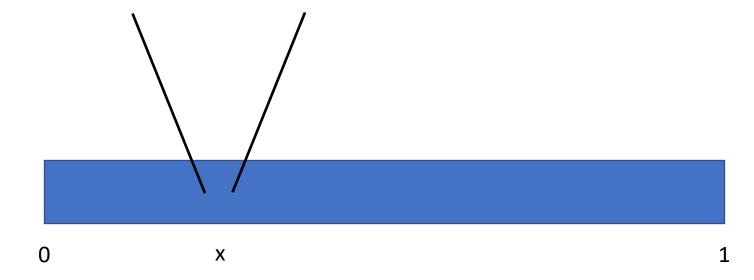
```
v = h.Vector().record(axon(x)._ref_v)
t = h.Vector().record(h._ref_t)
```

Note: Vector objects also have a play method that can be used to input stored values into the model.

Recording Results II

NetCon objects can be used as shown to detect the times when a variable crosses a threshold from below.

As the name suggests, a
NetCon can be used to
connect cells together in a
network. To do this, pass in a
synapse as the second
argument or use
ParallelContext.



```
spike_times = h.Vector()
nc = h.NetCon(axon(0.1)._ref_v, None, sec=axon)
nc.threshold = 0 * mV
nc.record(spike_times)
```

Stimulating a model

Set potential

- soma(0.5).v = 10 * mV
- Voltage clamp
 - cl = h.SEClamp(soma(0.5))
 - cl.amp1 = -65 * mV
 - cl.dur1 = 10 * ms
 - Similarly for .amp2, .amp3, .dur2, .dur3
 - Could also: vec.play(cl._ref_amp2, tvec)
 - SEClamp single electrode
 - VClamp two electrode

Current Clamp

- ic = h.IClamp(soma(0.5))
- ic.delay = 5 * ms
- ic.dur = 0.1 * ms
- ic.amp = 1 # nA

Synaptic input

- ns = h.NetStim()
 - ns.number = 1
 - ns.start = 5 * ms
 - ns.noise = False
 - ns.interval = 20 * ms
 - Only matters for number > 1
- sy = h.ExpSyn(soma(0.5))
 - sy.tau = 5 * ms
 - sy.e = 0 * mV
- nc = h.NetCon(ns, sy)
 - nc.weight[0] = 1

Running simulations: the basics

For convenience, we use a high-level simulation control functions defined in the stdrun.hoc library. Load this via:



Initialize to -65 mV:

h.finitialize(-65 * mV)



Run until time 10 ms:

h.continuerun(10 * ms)

Running simulations: the basics

For convenience, we use a high-level simulation control functions defined in the stdrun.hoc library. Load this via:



Initialize to -65 mV:

h.finitialize(-65 * mV)



Advance one timestep:

h.fadvance()

Running simulations: improving accuracy

Increase time resolution (by reducing time steps) via, e.g.

$$h.dt = 0.01 * ms$$

Enable variable step (allows error control):

Set the absolute tolerance to e.g. 10^{-5} :

$$h.CVode().atol(1e-5)$$

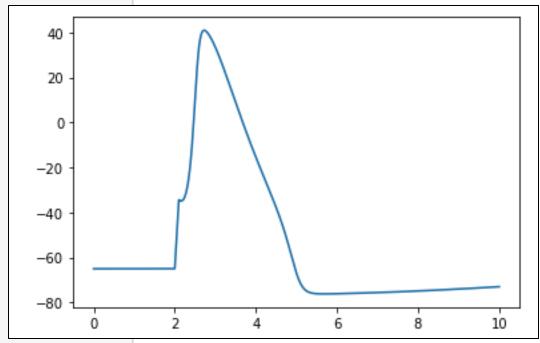
Increase spatial resolution by e.g. a factor of 3 everywhere:

```
for sec in h.allsec(): sec.nseg *= 3
```

Example: Hodgkin-Huxley

Note: Here we trigger the action potential by injecting a current. We could alternatively include a model of a synapse and trigger the synapse using an h.NetStim. See the documentation for more information.

```
from neuron import h
from neuron.units import ms, mV, µm
import matplotlib.pyplot as plt
h.load_file("stdrun.hoc")
soma = h.Section(name='soma')
                                                                  40
soma.L = soma.diam = 10 * \mu m
h.hh.insert(soma)
                                                                  20
ic = h.IClamp(soma(0.5))
ic.delay = 2 * ms
                                                                   0
ic.dur = 0.1 * ms
ic.amp = 1
                                                                 -20
t = h.Vector().record(h._ref_t)
                                                                 -40
v = h.Vector().record(soma(0.5)._ref_v)
                                                                 -60
h.finitialize(-65 * mV)
h.continuerun(10 * ms)
                                                                 -80
plt.plot(t, v)
                                                                               2
plt.show()
```



Example: spike detection

```
from neuron import h
         from neuron.units import ms, mV, µm
         import matplotlib.pyplot as plt
         h.load_file("stdrun.hoc")
         axon = h.Section(name='axon')
         h.hh.insert(axon)
                                                                                  40
         iclamps = []
         for input_time in [2 * ms, 13 * ms, 27 * ms, 40 * ms]:
                                                                                  20
              ic = h.IClamp(axon(0.5))
 Many
              ic.delay = input_time
 inputs
              ic.dur = 0.5 * ms
              ic.amp = 50
              iclamps.append(ic)
                                                                                 -20
         t = h.Vector().record(h._ref_t)
         v = h.Vector().record(axon(0.5)._ref_v)
                                                                                 -40
         nc = h.NetCon(axon(0.5)._ref_v, None, sec=axon)
Recording
         spike times = h.Vector()
                                                                                 -60
 spikes
         nc.record(spike_times)
         h.finitialize(-65 * mV)
                                                                                 -80
         h.continuerun(49.5 * ms)
                                                                                                10
                                                                                                          20
                                                                                                                    30
                                                                                                                             40
         print("spike times:", list(spike_times))
         plt.plot(t, v)
         plt.show()
          spike times: [3.225000000100012, 28.2000000009893, 41.7000000010092]
```

Networks of neurons

• Suppose we have the simple model:

```
from neuron import h
from neuron.units import ms, mV

class Cell:
    def __init__(self):
        self.soma = h.Section(name="soma", cell=self)
        self.all = self.soma.wholetree()
        h.hh.insert(self.all)
```

• and two cells:

```
neuron1 = Cell()
neuron2 = Cell()
```

Networks of neurons

- If the first cell has a sufficient current clamp injection, we know that it will fire, but how can we get that to send a signal to another cell?
- We do this with a synapse.
- On the post-synaptic side:

```
postsyn = h.ExpSyn(neuron2.soma(0.5))
postsyn.e = 0 # reversal potential
```

• On the pre-synaptic side, specify a source pointer, the corresponding post-synaptic side, the transmission delay, and synaptic weight:

```
syn = h.NetCon(neuron1.soma(0.5)._ref_v, postsyn, sec=neuron1.soma)
syn.delay = 1
syn.weight[0] = 5
```

Networks of neurons

Record, run, and plot as normal:

```
due to the synapse
t = h.Vector().record(h._ref_t)
v1 = h.Vector().record(neuron1.soma(0.5)._ref_v)
v2 = h.Vector().record(neuron2.soma(0.5)._ref_v)
                                                        40
h.finitialize(-65 * mV)
h.continuerun(10 * ms)
                                                        20
plt.plot(t, v1, t, v2)
plt.xlim((0, 10))
plt.show()
                                                       -20
                                                       -40
                             due to the iclamp
                                                       -60
                              (code not shown)
                                                       -80
```

Tip:

For parameter sweeps with changes only after an initial period, use h. SaveState

```
s = h.SaveState()
s.save()
# NOTE: calling s.save() stores the state
# to the s object; it does not store the
# state to a file; use s.fwrite(file_obj)
# for that and s.fread(file obj) to read
# state from a file before restoring.
h.continuerun(1000 * ms)
# go back to the way things were when we
# called s.save()
s.restore()
```

Storing and loading data with pandas

Saving as CSV with pandas:

```
import pandas as pd
pd.DataFrame({"t": t, "v": v}).to_csv("data.csv", index=False)
```

Loading from CSV with pandas:

```
import pandas as pd
data = pd.read_csv("data.csv")
t = h.Vector(data["t"])
v = h.Vector(data["v"])
```

t and v are h. Vector instances

Saving to a database table

```
import pandas as pd
import sqlite3
import time
# do all the following inside a loop
start = time.perf_counter()
# do the simulation here
data = pd.DataFrame(
  "dx": [dx],
  "L": L,
  "diam": diam,
  "alpha": alpha,
  "temperature": h.celsius,
  "spike half width": spike half width,
  "calculated_value": calculated_value,
  "runtime": time.perf counter() - start,
with sqlite3.connect(DB_FILENAME) as conn:
 data.to_sql("data", conn,
        if_exists="append", index=False)
```

DB FILENAME

is a string

```
import pandas as pd
import sqlite3
import time

with sqlite3.connect(DB_FILENAME) as conn:
  data = pd.read_sql("SELECT * FROM data", conn)
```

Read from a database table

```
with sqlite3.connect(DB_FILENAME) as conn:
data = pd.read_sql(
    """

SELECT dx, L, diam, temperature, alpha FROM data
    WHERE calculated_value > 7
    ORDER BY temperature
    """, conn)
```



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NEURON Resources

Unified documentation

• <u>nrn.readthedocs.io</u>

Forum

• <u>tinyurl.com/neuron-forum</u>

NEURON models on ModelDB

• tinyurl.com/nrn-models

Video tutorials

<u>tinyurl.com/nrn-videos</u>